

**Enzymatic characterization of AMP phosphorylase and ribose-1,5-bisphosphate
isomerase functioning in an archaeal AMP metabolic pathway**

*Running title: Archaeal enzymes involved in nucleotide metabolism

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ABSTRACT

AMP phosphorylase (AMPpase), ribose-1,5-bisphosphate (R15P) isomerase, and type III ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco) have been proposed to constitute a novel pathway involved in AMP metabolism in the Archaea. Here we performed a biochemical examination of AMPpase and R15P isomerase from *Thermococcus kodakarensis*. R15P isomerase was specific for the α -anomer of R15P and did not recognize other sugar compounds. We observed that activity was extremely low with the substrate R15P alone, but is dramatically activated in the presence of AMP. Using AMP-activated R15P isomerase, we re-evaluated the substrate specificity of AMPpase. AMPpase exhibited phosphorylase activity toward CMP and UMP in addition to AMP. The [S]- v plot (plot of velocity versus substrate) of the enzyme toward AMP was sigmoidal, with an increase in activity observed at concentrations higher than approximately 3 mM. The behavior of the two enzymes toward AMP indicates that the pathway is intrinsically designed to prevent excess degradation of intracellular AMP. We further examined formation of 3-phosphoglycerate from AMP, CMP, UMP in *T. kodakarensis* cell extracts. 3-Phosphoglycerate generation was observed from AMP alone, and from CMP or UMP in the presence of dAMP, which also activates R15P isomerase. 3-Phosphoglycerate was not formed when 2-carboxy-arabinitol 1,5-bisphosphate, a Rubisco inhibitor, was added. The results strongly suggest that these enzymes are actually involved in the conversion of nucleoside monophosphates to 3-phosphoglycerate in *T. kodakarensis*.

1 INTRODUCTION

2 Archaea comprise the third domain of life and exhibit unique metabolic features
3 that are not found in bacteria and eukaryotes. The metabolic enzymes and pathways utilized
4 for glycolysis and pentose biosynthesis in many archaea differ from the classical Embden-
5 Meyerhoff (EM)/Entner-Doudoroff (ED) pathways (24, 27, 33, 35) and pentose phosphate
6 pathway (11, 19, 28), respectively. A previous study of the hyperthermophilic archaeon
7 *Thermococcus kodakarensis* KOD1 (3, 9) suggested the presence of a novel pathway
8 involved in nucleic acid metabolism (25). The pathway consists of a type III ribulose-1,5-
9 biphosphate carboxylase/oxygenase (Rubisco) and two novel enzymes, AMP phosphorylase
10 (AMPPase) and ribose-1,5-biphosphate (R15P) isomerase. In the first reaction of this
11 pathway, catalyzed by AMPPase, the adenine base of AMP is released and replaced by a
12 phosphate group to generate R15P. In the following R15P isomerase reaction, R15P is
13 converted to ribulose 1,5-biphosphate (RuBP). In the final reaction, Rubisco catalyzes the
14 conversion of RuBP, CO₂, and H₂O to 2 molecules of 3-phosphoglycerate (3-PGA), which is
15 an intermediate of central sugar metabolism. Genome sequences indicate that this pathway is
16 distributed broadly among the Archaea, including all members of Thermococcales,
17 Archaeoglobales, Methanomicrobiales, and Methanosarcinales. The pathway is also found in
18 several members of Halobacteriales, Methanococcales, Desulfurococcales, and
19 Thermoproteales. The pathway at present seems to be confined to the Archaea, since a
20 complete set of genes cannot be found in any of the genomes of the Bacteria and Eucarya.

21 AMPPase is a unique enzyme in terms of its substrate specificity. It is the first
22 enzyme identified to catalyze a phosphorolysis reaction on a nucleotide. This is distinct from
23 the structurally related phosphorylases that have been characterized previously, which act
24 specifically towards nucleosides (6, 12, 20, 30). R15P isomerase catalyzes the isomerization
25 of R15P, an aldopentose with a phosphorylated 1-hydroxy group. The reaction is of interest

1 because opening of the furanose ring cannot precede isomerization due to the phosphate
2 group on the C-1 carbon. The structure of R15P isomerase revealed that the enzyme
3 displayed a hexameric assembly (18). Crystal structures bound to substrate/product and
4 mutational studies indicated that Cys133 and Asp202 act as active site residues in the enzyme
5 and that the reaction proceeds via a *cis*-phosphoenolate intermediate. R15P isomerase is
6 structurally related to 5-methylthioribose-1-phosphate (MTR1P) isomerase. MTR1P
7 isomerase functions in the methionine salvage pathway and catalyzes a similar reaction, the
8 isomerization of MTR1P, an aldopentose with a phosphorylated 1-hydroxy group, to 5-
9 methylthioribulose 1-phosphate (2, 26). Enzymatic (22) and structural (5, 29) examinations
10 have been carried out on the enzymes from *Bacillus subtilis* and *Saccharomyces cerevisiae*.
11 One of the reaction mechanisms that have been proposed for this enzyme also proceeds via a
12 *cis*-phosphoenolate intermediate (29).

13 The enzymatic and structural features of type III Rubiscos have been reported
14 previously (1, 7, 8, 15-17, 34). As mentioned above, we have recently determined the 3-
15 dimensional structure of *T. kodakarensis* R15P isomerase (18). However, no detailed
16 biochemical characterization of the two novel enzymes, AMPpase and R15P isomerase, has
17 been reported. Since unregulated breakdown of AMP would be detrimental to the cells, one
18 can suppose that the pathway should be regulated so that activity is rapidly shut down once
19 intracellular levels of AMP become low. Here we performed the first detailed biochemical
20 analysis of AMPpase and R15P isomerase. The enzymes from *T. kodakarensis* were examined
21 by focusing mainly on their substrate specificity and kinetic behavior as well as how their
22 activities are regulated at the protein level in response to the presence of various metabolites.
23 We have also confirmed the conversion of nucleoside monophosphates (NMPs) to 3-PGA in
24 cell-free extracts, providing strong evidence that the pathway functions in *T. kodakarensis*.

MATERIAL AND METHODS

Strains and growth conditions

Escherichia coli strains DH5 α and BL21-CodonPlus (DE3)-RIL were used for plasmid construction and heterologous gene expression, respectively. These strains were cultivated at 37°C in Luria-Bertani medium containing 100 μ g/ml ampicillin (23). *T. kodakarensis* KOD1 was cultivated under strictly anaerobic conditions at 85°C in a nutrient-rich growth medium based on artificial sea water (ASW) (21). Briefly, ASW-YT medium was composed of 0.8 x ASW, 0.5% yeast extract, and 0.5% tryptone. Either 0.2% elemental sulfur (ASW-YT-S⁰), 0.5% pyruvate (ASW-YT-Pyr), or 0.5% pyruvate and 2.5 mM of a mixture of adenosine, guanosine, cytidine, and uridine at 2.5 mM (ASW-YT-Pyr-Nuc) were added.

Preparation of the recombinant proteins

Recombinant His₆-tagged R15P isomerase and Rubisco were produced in *E. coli* and were purified as described previously (18). The recombinant AMPpase was expressed as described elsewhere (25). In order to purify recombinant AMPpase, cells were harvested, resuspended in 50 mM Tris-HCl (pH 7.5), and sonicated. Soluble proteins were incubated for 10 min at 80°C and centrifuged (at 20,000 x *g*, 30 min) to remove thermolabile proteins deriving from the host cells. The supernatant was applied to an anion exchange column (Resource Q; GE Healthcare, Little Chalfont, Buckinghamshire, United Kingdom), and proteins were eluted with a linear gradient of NaCl (0 to 1.0 M) in 50 mM Tris-HCl (pH 7.5). After concentrating the fractions containing AMPpase were concentrated with an Amicon Ultra centrifugal filter unit (molecular weight cutoff [MWCO], 100,000; Millipore, Billerica, MA), the sample was applied to a gel filtration column (Superdex 200; GE Healthcare) with a mobile phase of 150 mM NaCl in 50 mM Tris-HCl (pH 7.5) at room temperature.

Enzymatic synthesis of ribose 1,5-bisphosphate with AMP phosphorylase

Enzymatic preparation of the substrate R15P, used for activity measurements of R15P isomerase, was performed as follows. The AMPpase reaction was performed in a 500 μ l mixture containing 560 nM of purified AMPpase, 100 mM Tris-HCl (pH 7.5), 30 mM sodium phosphate (pH 7.5), and 30 mM AMP. After pre-incubation at 85°C for 3 min, the reaction was initiated by the addition of AMP. After a further 10 min incubation, the reaction was terminated by rapid cooling on ice for 5 min, and AMPpase was removed by ultrafiltration with an Amicon Ultra centrifugal filter unit (MWCO 30,000; Millipore). The resulting R15P mixture was concentrated by vacuum drying and centrifugation and was used as the substrate for the R15P isomerase reaction. By use of high-performance liquid chromatography (HPLC) with a refractive index detector, the concentration of R15P was calculated to be approximately 5 mM. This enzymatically prepared substrate mixture is designated as E-R15P.

Measurement of ribose-1,5-bisphosphate isomerase activity using coupling enzymes

R15P isomerase activity was measured either with coupling enzymes or by HPLC (see below). The former procedure was carried out as follows. First, the R15P isomerase reaction was performed coupled with the carboxylase reaction of Rubisco. The reaction mixture (100 μ l) was composed of 81 nM of purified R15P isomerase, 1.0 μ M of purified Rubisco (90 mU), 100 mM NaHCO₃, 30 μ l of E-R15P (1.5 mM) or 5 mM chemically synthesized R15P (C-R15P; Tokyo Chemical Industry Co., Tokyo, Japan), in 10 mM MgCl₂ and 100 mM Bicine-NaOH (pH 8.3). NAD⁺, which had been included in the reaction mixture previously (25), was found to have no effect on activity (see below) and was excluded from the reaction mixture. When C-R15P was used as the substrate, AMP was added to the reaction mixture in order to activate the enzyme. After preincubation at 85°C for 3 min, the reaction was initiated by the addition of NaHCO₃ and R15P. The reaction was carried out for 5 min at 85°C and was

terminated by rapid cooling on ice for 5 min, and the enzymes were removed with an Amicon Ultra centrifugal filter unit (MWCO, 30,000). After appropriate dilution, the amount of 3-PGA synthesized by the reaction was determined by a second coupling reaction, described elsewhere (7). The second reaction mixture (100 μ l) was composed of 5 mM ATP, 0.2 mM NADH, 8 mM $MgCl_2$, 80 mM Bicine-NaOH (pH 8.3), 20 μ l of coupling enzymes solution, and an aliquot of the R15P isomerase reaction mixture. The coupling enzymes solution contained 563 U ml^{-1} 3-phosphoglycerate phosphokinase, 125 U ml^{-1} glyceraldehyde-3-phosphate dehydrogenase, 260 U ml^{-1} triose-phosphate isomerase, 22.5 U ml^{-1} glycerophosphate dehydrogenase, 5 mM reduced glutathione, 0.1 mM EDTA, and 20 % glycerol in 50 mM Bicine-NaOH (pH 8.0). After pre-incubation at 25°C for 3 min, the reaction was initiated with the addition of the enzymes. The decrease in absorbance at 340 nm due to the consumption of NADH was measured.

The effects of various compounds on the R15P isomerase reaction were examined by adding one of the following compounds to the reaction mixture: 18 mM sodium phosphate, 0.4 mM adenine, 0.4 mM NAD^+ , or 3 mM of AMP, CMP, GMP, UMP, TMP, ADP, ATP, adenosine, dAMP, 5'-methylthioadenosine (MTA), *S*-adenosylmethionine (SAM), *S*-adenosylhomocysteine (SAH), phosphoribosylpyrophosphate (PRPP), ribose 5-phosphate (R5P), fructose 1,6-bisphosphate (FBP), fructose 6-phosphate (F6P), 3-PGA, glucose, or pyruvate. We could not use the enzyme coupling method to measure the effects of ADP and 3-PGA, because ADP inhibits 3-phosphoglycerate phosphokinase, and 3-PGA is the product of the R15P isomerase/Rubisco reactions. In these cases, HPLC was applied for activity measurements (described below).

Measurement of ribose-1,5-bisphosphate isomerase activity using HPLC

When activity was measured with HPLC, the reaction mixture (100 μ l) was composed of 140 nM of purified R15P isomerase, 3 mM AMP, and 5 mM C-R15P in 10 mM MgCl_2 and 100 mM Bicine-NaOH (pH 8.3). After pre-incubation at 85°C for 3 min, the reaction was initiated by the addition of C-R15P, followed by incubation for 3, 5, and 7 min, and was terminated by rapid cooling on ice for 5 min. R15P isomerase was removed with an Amicon Ultra centrifugal filter unit (MWCO 30,000). After the addition of an equal volume of 600 mM sodium phosphate (pH 4.4) to the filtrate in order to adjust the phosphate concentration to that of the HPLC mobile phase, the sample was analyzed on an amino column (Asahipak NH2P-50 4E column; Shodex, Tokyo, Japan), with 300 mM sodium phosphate buffer (pH 4.4) as the mobile phase. When the effects of ADP and 3-PGA on R15P isomerase activity were investigated, activity was calculated by measuring the decrease of R15P levels. Isomerase activity towards 40 mM PRPP, R5P, ribose, FBP, F6P, glucose 1,6-bisphosphate (G16P), glucose 6-phosphate (G6P), or glucose 1-phosphate (G1P) was examined by monitoring substrate consumption and/or product generation in the presence or absence of R15P isomerase. When activity towards PRPP was examined, a C_{18} column (COSMOSIL 5C₁₈-PAQ; Nacalai Tesque) was utilized with 50 mM NaH_2PO_4 (pH 4.3) as the mobile phase. Column temperatures were set at 40°C, and compounds were detected with a refractive index detector in all cases.

Measurement of AMP phosphorylase activity

The phosphorylase reaction of nucleoside monophosphates (NMPs) was performed in a mixture (100 μ l) containing 100 mM Tris-HCl (pH 7.5), 20 mM sodium phosphate (pH 7.5), 190 nM of purified AMPpase, and 20 mM NMP. After preincubation at 85°C for 3 min, the reaction was initiated by the addition of an NMP. The reaction was carried out at 85°C for 5 min and was terminated by rapid cooling on ice for 5 min, and AMPpase was removed with

an Amicon Ultra centrifugal filter unit (MWCO, 30,000). The R15P generated in the AMPpase reaction was then converted to 3-PGA by the isomerase activity of R15P isomerase and the carboxylase activity of Rubisco. The reaction mixture (100 μ l) was composed of 1.3 μ M of purified R15P isomerase, 1.0 μ M of purified Rubisco (90 mU), 10 mM AMP, 100 mM NaHCO₃, and 10 μ l of the AMPpase reaction mixture. After preincubation at 85°C for 3 min, the reaction was initiated by the addition of NaHCO₃ and the AMPpase reaction mixture. The reaction was carried out for 10 min at 85°C, terminated by rapid cooling on ice for 5 min, and the enzymes were removed with an Amicon Ultra centrifugal filter unit (MWCO, 30,000). The 3-PGA generated in this reaction was quantified by the second coupling reaction described above.

In order to examine the substrate specificity of AMPpase, phosphorylase activity towards the following substrates were measured by HPLC; 20 mM of dNMP, adenosine, cytidine, uridine, ADP, ATP, SAM, SAH, PRPP, or R5P, or 2 mM MTA. The reactions were carried out for 3, 5, or 7 min. Compounds were detected with a refractive index detector and/or a UV detector (A₂₅₄). The phosphorylase activity was quantified by substrate consumption and/or nucleobase release.

Examination of 3-phosphoglycerate synthesis in cell-free extracts

Cell-free extracts (CFE) of *T. kodakarensis* KOD1 were prepared as follows. Cells cultivated in ASW-YT-S⁰ medium for 17 h were harvested by centrifugation (5,000 x g, 15 min, 4°C) and were lysed in 50 mM Tris-HCl (pH 7.5) containing 0.1% of Triton X-100 at a volume of 1/500 of the culture. After mixing with a vortex for 30 min, the supernatant after centrifugation (20,000 x g, 30 min, 4°C) was used as CFE. Examination of 3-PGA synthesis with the CFE was performed in a mixture (100 μ l) containing 100 mM Bicine-NaOH (pH 8.3), 10 mM MgCl₂, the CFE (corresponding to 100 μ g protein), 20 mM sodium phosphate

(pH 7.5), 100 mM NaHCO₃, and 20 mM NMP. When necessary, 20 mM dAMP (to activate R15P isomerase) and/or 20 mM 2-carboxyarabinitol 1,5-bisphosphate (CABP) (to inhibit Rubisco) was also added to the reaction mixture. After preincubation at 85°C for 3 min, the reaction was initiated by the addition of NaHCO₃ and an NMP. The reaction was carried out at 85°C for 30 min, terminated by rapid cooling on ice for 5 min, and proteins were removed with an Amicon Ultra centrifugal filter unit (MWCO, 3,000). The 3-PGA generated in this reaction was quantified by a second coupling reaction, described above.

Western blot analysis.

Cell-free extracts from *T. kodakarensis*, grown in ASW-YT-Pyr or ASW-YT-Pyr-Nuc, were prepared as described above, and proteins were separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) (12.5% acrylamide) and were electroblotted onto a Hybond-P membrane (GE Healthcare). After blocking with blocking reagents (GE Healthcare), membranes were hybridized with a rabbit antiserum containing polyclonal anti-AMPpase, anti-R15P isomerase, or anti-Rubisco antibodies, washed, hybridized with horseradish peroxidase (HRP)-conjugated recombinant protein G (dilution, 1:100,000; Zymed Laboratories, San Francisco, CA), and washed again. For signal detection, the ECL Advance Western Blotting Detection System (GE Healthcare) and a Lumi Vision PRO 400EX image analyzer (Aisin, Kariya, Japan) were used.

RESULTS

Identification of compounds that activate ribose-1,5-bisphosphate isomerase

In previous studies, the substrate (R15P) used in measuring R15P isomerase activity was prepared enzymatically from AMP and sodium phosphate using AMPpase (18, 25). R15P

1 isomerase displayed a specific activity of about $35 \mu\text{mol min}^{-1} \text{mg}^{-1}$, a level similar to those
2 observed in the previous studies, when the AMPpase reaction mixture, which includes R15P
3 as well as AMP, adenine, and phosphate (referred to here as E-R15P), was used as the
4 substrate (Fig. 1A). In order to remove the effects of the other components in the
5 enzymatically prepared substrate, we used chemically synthesized R15P (C-R15P) as the
6 substrate here. Intriguingly, we could observe only very low levels of R15P isomerase
7 activity by using C-R15P. This suggested that R15P isomerase was activated by a
8 component(s) present in the AMPpase reaction mixture. R15P isomerase activity with C-
9 R15P was thus measured in the presence of individual components of the AMPpase reaction
10 mixture: AMP, sodium phosphate, or adenine. In the presence of AMP, R15P isomerase
11 displayed activity with C-R15P at a level comparable to that observed with E-R15P. When
12 R15P was excluded from the reaction mixture in the presence of AMP, RuBP synthesis was
13 not observed. Furthermore, HPLC analyses of the R15P isomerase reaction with AMP
14 revealed a time-dependent decrease in R15P level, whereas consumption of AMP was not
15 detected (see Fig. S1 in the supplemental material). These results indicated that AMP itself
16 was not a substrate of R15P isomerase, but an activator. On the other hand, the addition of
17 sodium phosphate and adenine had no effect on R15P isomerase activity. We further
18 examined the possibility that AMP activated Rubisco, which was utilized as a coupling
19 enzyme in the R15P isomerase activity measurements, consequently enhancing the
20 conversion of R15P to 3-PGA. We found that the addition of AMP had no effect on Rubisco
21 activity (data not shown). These results clearly indicate that AMP dramatically activates R15P
22 isomerase, with an increase of >40-fold in activity levels. All further analyses using R15P
23 were performed with chemically synthesized R15P.

24 Considering the effects of AMP, we next examined the effects of other nucleotides on
25 R15P isomerase activity. Among CMP, GMP, UMP, TMP, and NAD^+ , we observed a slight

1 increase in R15P isomerase activity in the presence of GMP. In order to compare the extents
2 of activation brought about by AMP and GMP, R15P isomerase activity was measured in the
3 presence of various concentrations of AMP or GMP. As shown in Fig. 1B, higher degrees of
4 activation were observed for AMP than for GMP at lower concentrations, suggesting that
5 AMP is the major activator of R15P isomerase *in vivo*.

6 We further explored the possibilities of other compounds acting as activators of R15P
7 isomerase. Various compounds were added at a concentration of 3 mM, and in order to
8 expand the range of compounds that we could examine, we also performed activity
9 measurements with HPLC when necessary. This allows us to analyze the effects of
10 compounds such as ADP and 3-PGA, which would be difficult with the enzyme coupling
11 assays. The enzyme coupling assay and the HPLC analysis revealed that the addition of 3
12 mM ADP resulted in increases in R15P isomerase activity that were comparable to those
13 observed with AMP. Other compounds with an adenosyl moiety also exhibited activating
14 effects, but at higher concentrations. On the other hand, R15P isomerase was not activated by
15 R5P, PRPP, FBP, F6P, glucose, pyruvate, or 3-PGA. These results suggested a tendency for
16 compounds with an adenosyl moiety to activate R15P isomerase. To compare the extent of
17 activation, R15P isomerase activity was measured in the presence of varying concentrations
18 of these compounds. As shown in Fig. 1C, the effects with SAM, SAH, and ATP were
19 relatively small, while significant activation was detected in the presence of dAMP, adenosine,
20 and MTA. However, in terms of the degree of activation and the concentration range within
21 which activation was observed, AMP had the most dramatic effect. At low concentrations of
22 250 μ M, which may better represent physiological concentrations, activity with AMP was
23 more than 3-fold higher than that observed with ADP. The dissociation constant values of the
24 activators calculated from the HPLC measurements for AMP and ADP (Fig. 1D) were 217

1 μM (K_{AMP}) and 667 μM (K_{ADP}). The effects of other compounds at this concentration were
2 negligible.

4 **Kinetic analysis and substrate specificity of ribose-1,5-bisphosphate isomerase**

5 Kinetic analysis of R15P isomerase was carried out in the presence of 10 mM AMP by
6 using the enzyme coupling method (Fig. 2). The isomerase reaction from R15P to RuBP
7 followed Michaelis-Menten kinetics with a K_m of 0.6 ± 0.1 mM for R15P and a k_{cat} of $29.2 \pm$
8 0.7 s^{-1} at 85°C (Table 1).

9 We next examined the substrate specificity of R15P isomerase. Among the two
10 anomers of R15P, we observed previously that the enzyme utilizes only the α -anomer
11 compound (18). Here we investigated whether R15P isomerase can recognize other
12 phosphorylated sugars. Activities towards the following substrates were investigated by
13 HPLC: R5P, ribose, G16P, G6P, G1P, FBP, F6P, and PRPP. We did not observe any decrease
14 in the levels of substrates or synthesis of products dependent on R15P isomerase, with any of
15 the compounds except PRPP. Although a decrease in PRPP levels and an increase in RuBP
16 levels were observed, the decrease in PRPP levels was not dependent on the R15P isomerase
17 enzyme. We presume that PRPP displayed thermal degradation during the incubation at 85°C ,
18 resulting in the generation of R15P, the substrate of R15P isomerase. These results indicated
19 that among the compounds examined, R15P isomerase can utilize only α -R15P, implying that
20 the substrate specificity of this enzyme is strict.

22 **Substrate specificity of AMP phosphorylase**

23 AMPpase was previously reported to catalyze an AMP-specific phosphorylase
24 reaction generating R15P and adenine. Relevant levels of phosphorylase activity could not be
25 observed with other nucleoside monophosphates, such as UMP, GMP, CMP, and TMP (25).

1 However, because the activity measurements were performed with coupling enzymes that
2 included R15P isomerase, we realized that phosphorylase activities toward nucleoside
3 monophosphates other than AMP might have been overlooked or underestimated due to the
4 low activity levels of R15P isomerase in the absence of AMP. Therefore, we re-evaluated the
5 substrate specificity of AMPpase by using the coupling enzymes, but under the condition that
6 R15P isomerase was activated with saturating concentrations of AMP. As a result, we found
7 that AMPpase exhibited phosphorylase activity not only toward AMP but also CMP and UMP,
8 while only low phosphorylase activity was observed with GMP and IMP (Table 2).

9 In order to investigate whether AMPpase could catalyze the phosphorolysis of other
10 compounds related to nucleotides/nucleosides, activity measurements with HPLC were
11 performed on the following compounds: dAMP, dCMP, dGMP, dUMP, TMP, adenosine,
12 cytidine, uridine, ADP, ATP, MTA, SAM, SAH, PRPP, and R5P. We observed significant
13 levels of cytosine released from dCMP during the reaction. On the other hand, the use of
14 dAMP, dGMP, or dUMP resulted in the generation of only trace amounts of their
15 corresponding nucleobases, adenine, guanine, or uracil, respectively (Table 2). No nucleobase
16 synthesis and no substrate consumption were observed when dCMP, dAMP, dGMP, and
17 dUMP were incubated in the absence of the enzyme or phosphate, indicating that the
18 nucleobases were not the product of thermal degradation but dependent on the enzyme
19 activity of AMPpase. We did not detect enzyme activities toward TMP, adenosine, cytidine,
20 uridine ADP, ATP, MTA, SAM, SAH, PRPP, or R5P.

22 **Kinetic analysis of AMP phosphorylase**

23 Kinetic analyses of the activities of AMPpase toward AMP, CMP, GMP, and UMP
24 were carried out (Fig. 3A). Toward CMP, GMP, and UMP, the plots of velocity versus
25 substrate concentration ($[S]$ - v plots) followed Michaelis-Menten kinetics with the kinetic

parameters shown in Table 1. Although activity toward GMP was relatively low, activity with CMP and UMP were as high as or higher than that observed with AMP. These results confirmed that AMPpase could convert not only AMP but also CMP and UMP. It should be noted that the [S]- v plot with AMP as the substrate displayed a sigmoidal curve, indicating regulation of AMPpase by AMP (Fig. 3B). A number of kinetic models including cooperativity were considered, but no equation that fit our data well could be identified. Kinetic analysis of AMPpase activity toward dCMP was also carried out by HPLC (Fig. 3C). The [S]- v plot was very similar to that obtained with AMP, suggesting a similar mode of regulation. In order to confirm that the activity levels obtained by measurements with HPLC and coupling enzymes could be accurately compared with one another, we performed a kinetic analysis on CMP again using HPLC. No large differences were observed between the two procedures.

Kinetic analysis of AMPpase toward its other substrate, Pi (Fig. 3D), revealed that this phosphorylase reaction followed Michaelis-Menten kinetics with a K_m of 2.8 ± 0.1 mM and a k_{cat} of 15.0 ± 0.2 s⁻¹ at 85°C (Table 1).

Conversion of nucleoside monophosphates to 3-phosphoglycerate in cell-free extracts

We indicated previously that the recombinant AMPpase, R15P isomerase, and Rubisco could catalyze sequential reactions converting AMP to 3-PGA *in vitro* (25). In order to gain insight on whether this was also the case *in vivo*, we investigated whether NMPs were converted to 3-PGA in cell-free extracts (CFE). When AMP was used as the substrate, we clearly observed the generation of 3-PGA in the CFE (Table 3). We could not observe 3-PGA synthesis when CMP or UMP was added alone. We presumed that this was due to the fact that R15P isomerase was not activated. We then added dAMP to the reaction mixture, since this compound activates R15P isomerase but does not serve as a substrate to generate 3-PGA

(Table 3). With the addition of dAMP, we detected significant levels of 3-PGA synthesis from CMP and UMP. Since the amounts of 3-PGA generated with CMP and UMP were lower than that observed for AMP, we investigated whether activation of R15P isomerase by dAMP was not as effective as that by AMP. We added R15P directly to the CFE and measured 3-PGA synthesis in the presence or absence of dAMP. The addition of dAMP led to a dramatic increase in 3-PGA formation, from 12.1 to 202 μ M. The fact that the addition of dAMP enhanced 3-PGA generation strongly suggests that the conversion of AMP, CMP and UMP to 3-PGA is brought about by the three enzymes AMPpase, R15P isomerase, and Rubisco. In order to gain further support, CABP, an inhibitor of Rubisco, was added to the reaction mixture. In the presence of CABP, the significant levels of 3-PGA observed from AMP, CMP, and UMP in the experiments described above could not be detected (Table 3), providing further indications that AMP, CMP, and UMP are converted to 3-PGA via the archaeal AMP metabolic pathway in *T. kodakarensis*.

Protein levels of AMPpase, R15P isomerase and Rubisco

To investigate whether the three enzymes AMPpase, R15P isomerase, and Rubisco are subject to regulation at the transcriptional/translational level in addition to their responses to AMP at the activity level, we raised polyclonal antibodies against each protein and performed Western blot analyses. Since we had observed previously that nucleosides are assimilated well in *T. kodakarensis* (19), we grew the cells in ASW-YT-Pyr medium with or without a mixture of nucleosides (Fig. 4). We observed increases in the protein levels of R15P isomerase and Rubisco in the cells cultivated with nucleosides. In contrast, protein levels of AMPpase were more or less equivalent in cells grown in the presence or absence of nucleosides. The result suggests that the archaeal AMP metabolic pathway, in addition to its

response at the activity level to AMP, also responds to nucleosides or related metabolites at the transcriptional and/or translational level.

DISCUSSION

In this study, we performed the first detailed biochemical characterization of two novel enzymes functioning in an archaeal AMP metabolic pathway, AMPpase and R15P isomerase. Examination of the substrate specificity of AMPpase indicated that this enzyme utilized not only AMP but also CMP, UMP and dCMP. However, since intracellular concentrations of dNMPs are in general much lower than those of NMPs (32), it is likely that the major substrates of AMPpase in *T. kodakarensis* cells are the ribonucleotides AMP, CMP and UMP. We found that 3-PGA was synthesized from AMP, CMP, and UMP in *T. kodakarensis* cell-free extracts, suggesting that the three enzymes AMPpase, R15P isomerase, and Rubisco most likely function as a metabolic pathway *in vivo*. Kinetic analysis of AMPpase toward CMP, UMP, and Pi revealed that the [S]-v plots for these substrates followed Michaelis-Menten kinetics. In contrast, the [S]-v plot for AMP displayed a sigmoidal curve, indicating that this enzyme is regulated by AMP. This behavior toward AMP may act as a regulation mechanism to prevent excess degradation of AMP in *T. kodakarensis* cells. There may also be an unidentified mechanism to regulate the degradation of CMP and UMP, a hypothesis supported by the observation that the levels of 3-PGA synthesized from CMP and UMP in CFE were lower than that from AMP. We cannot rule out the possibility that CMP and UMP are rapidly consumed in other pathways present in CFE, but we think this unlikely, since we added the substrates at high concentrations. On the other hand, the equilibrium constant of the AMPpase reaction ($[R15P] [adenine] / [AMP] [Pi]$) has been determined previously as $6.02 \times 10^{-3} \pm 0.46 \times 10^{-3}$ (25), indicating that the reaction, from a thermodynamic point of view, favors AMP synthesis. This is also the case for other

nucleoside phosphorylases (4, 13, 14, 20). Since the reaction toward other NMPs can also be expected to favor NMP synthesis, AMPase may be involved in nucleotide interconversion between NMPs via R15P and nucleobases, regulating the intracellular NMP ratio when the *in vivo* concentration of AMP is low and R15P is not actively consumed by R15P isomerase. Although nucleoside interconversion has been found in Bacteria and Eucarya (10, 31), nucleotide interconversion has not been reported until now, and this may represent a novel regulation mechanism in nucleotide metabolism. The fact that the protein levels of AMPase were constitutive and did not respond to addition of nucleosides in the medium may be related to this mechanism.

Through enzymatic characterization of R15P isomerase, we clarified that, among various compounds we used, R15P isomerase was activated more than 40-fold in the presence of 1 mM AMP. This property of R15P isomerase suggests that the flux of the AMP metabolic pathway responds to the intracellular concentrations of AMP. The regulation of R15P isomerase activity responding to AMP basically answers our question as to how excess degradation of intracellular AMP by the AMP degradation pathway is prevented. The recently reported crystal structure of R15P isomerase did not include AMP (18). However, this enzyme form without AMP displays activity, although it is low, as shown in this study and by the fact that co-crystallization of the wild-type enzyme with the substrate (R15P) led to a protein structure bound to the product (RuBP). Further studies will be necessary to determine how AMP-binding affects enzyme structure and activity. MTR1P isomerase, a protein homologous to R15P isomerase, displays a three-dimensional structure resembling the dimer unit of R15P isomerase (5, 18, 29). This enzyme has not been examined for activation by metabolites (22), and the question of whether this enzyme is also regulated similarly or not is of interest.

Based on the novel insight obtained in this study, our present understanding of the archaeal AMP metabolic pathway and its regulation is illustrated in Fig. 5. AMP phosphorylase and R15P isomerase catalyze the reversible reactions between AMP (or CMP, UMP) and RuBP. The equilibrium of the AMP phosphorylase reaction is greatly skewed toward nucleoside monophosphate formation, but the irreversible reaction catalyzed by Rubisco results in the unidirectional formation of 3-PGA by this pathway. Activation of R15P isomerase by AMP stimulates the conversion between AMP and RuBP, and this activation, together with Rubisco, promotes 3-PGA formation. A decrease in AMP concentration reduces the activity levels of R15P isomerase, thus preventing excess degradation of AMP. Low levels of AMP would also prevent the breakdown of CMP or UMP via the regulation of R15P isomerase. We have shown previously that *T. kodakarensis* can assimilate exogenous nucleosides (19). In this study, we have shown that R15P isomerase and Rubisco are up-regulated by the addition of nucleosides. Therefore, the pathway may be involved in this nucleoside assimilation/degradation by directing the ribose moieties of nucleosides to central carbon metabolism (3-PGA) via AMP, CMP, or UMP. This is possible either through conventional routes that involve nucleoside phosphorylase, phosphopentomutase, PRPP synthetase, and nucleobase phosphoribosyltransferase, or through a direct conversion via nucleoside kinases. The pathway may also be involved in other metabolic conversions other than exogenous nucleoside/nucleotide breakdown, and the presence of unidentified metabolic pathways that are linked to these three enzymes via the nucleoside monophosphates should be examined.

Acknowledgments

This work was also partially supported by the Japan Science and Technology Agency (to H.A., T.I., K.M.), the Ministry of Education, Culture, Sports, Science, and Technology

(Targeted Proteins Research Program to K.M., T.I., H.A.), and Japan Society for the Promotion of Science [KAKENHI 22750150 to T.S.].

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Table 1. Kinetic parameters of R15P isomerase and AMPpase^a.

Enzyme	Substrate	K_m (mM)	k_{cat} (s ⁻¹)	k_{cat} / K_m (s ⁻¹ mM ⁻¹)
R15P isomerase	R15P	0.6 ± 0.1	29.2 ± 0.7	48.7
AMPpase	CMP	6.2 ± 0.5	39.1 ± 1.2	6.3
	UMP	4.4 ± 0.5	10.5 ± 0.4	2.4
	GMP	18.5 ± 1.3	2.7 ± 0.1	0.1
	Pi	2.8 ± 0.1	15.0 ± 0.2	5.4

^aThe kinetic parameters of the R15P isomerase reaction were examined in the presence of 10 mM AMP. The kinetic parameters of the AMPpase reaction toward NMPs and Pi were investigated in the presence of 20 mM Pi and 20 mM AMP, respectively. The kinetic parameters of the AMPpase reaction toward AMP and dCMP were not determined, because kinetic equations that fit well to our data could not be identified.

1 **Table 2. Substrate specificity of AMPpase towards NMPs and dNMPs.**

Substrate	Phosphorylase activity ($\mu\text{mol min}^{-1} \text{mg}^{-1}$) ^a determined by the following method:	
	Coupling enzymes	HPLC
AMP	15.9 ± 0.7	—
CMP	37.5 ± 1.4	35.2 ± 0.7
UMP	10.8 ± 0.5	—
GMP	1.8 ± 0.2	—
IMP	0.3 ± 0.1	—
dAMP	—	1.5 ± 0.02
dCMP	—	15.7 ± 0.5
dUMP	—	0.4 ± 0.02
dGMP	—	0.4 ± 0.03

2 ^aActivities were investigated in the presence of 20 mM NMP and 20 mM Pi.

3 —; not performed.

4

Table 3. 3-Phosphoglycerate formation from NMPs in *T. kodakarensis* cell-free extracts.

Substrate ^a	Additional component ^a		3-PGA concn (μM) ^b
	dAMP	CABP	
—	—	—	< 10
	+	—	< 10
	—	+	< 10
RuBP	—	—	1240
	—	+	15.1
AMP	—	—	94.3
	—	+	< 10
CMP	—	—	< 10
	+	—	42.4
	+	+	< 10
UMP	—	—	< 10
	+	—	14.4
	+	+	< 10
R15P	—	—	12.1
	—	+	< 10
	+	—	202

^aThe substrate, dAMP, and CABP were each used at a concentration of 20 mM. —, absence; +, presence.

^bDetected after a 30-min reaction with the substrates and the additional components along with 100 mM NaHCO₃ and 20 mM Pi in the cell-free extract (100 μg protein).

FIGURES LEGENDS

FIGURE 1. Effect of various compounds on the activity of R15P isomerase. (A) R15P isomerase activity was measured by enzyme coupling method with enzymatically prepared or chemically synthesized R15P (E-R15P or C-R15P, respectively). With C-R15P, the effects of the following compounds were examined: 3 mM AMP, 18 mM sodium phosphate (Pi), 0.4 mM adenine, 3 mM CMP, 3 mM GMP, 3 mM UMP, 3 mM TMP, or 0.4 mM NAD⁺. (B) Activation of R15P isomerase in the presence of varying concentrations of AMP or GMP. The initial velocities of R15P isomerase measured with coupling enzymes in the presence of AMP (*circles*) or GMP (*triangles*) are shown. (C) Activation of R15P isomerase in the presence of various compounds with an adenosyl moiety. The initial velocities of R15P isomerase were measured with coupling enzymes at varying concentrations of the following compounds: AMP (*filled diamonds*), deoxy AMP (*filled triangles*), methylthioadenosine (*filled circles*), adenosine (*plus sign*), *S*-adenosylhomocysteine (*filled squares*), *S*-adenosylmethionine (*asterisks*), or ATP (*open circles*). AMP (*open diamonds*) and ADP (*open squares*) are the results of measurements by HPLC. (D) Results of AMP and ADP measurements by HPLC in panel C up to 2.5 mM. In all experiments, C-R15P was used at a concentration of 5 mM.

FIGURE 2. Kinetic analysis of R15P isomerase. Initial velocities of R15P isomerase were measured with coupling enzymes in the presence of varying concentrations of C-R15P. AMP (10 mM) was present for all measurements.

FIGURE 3. Kinetic analyses of AMPpase. (A) Kinetic analysis of AMPpase toward NMPs. Initial velocities of AMPpase were measured with coupling enzymes in the presence of varying concentrations of AMP (*circles*), CMP (*squares*), GMP (*triangles*), or UMP (*diamonds*). The concentration of Pi was constant at 20 mM. (B) Enlarged view of the results

1 shown in panel A. (C) Kinetic analysis of AMPpase toward dCMP. Initial velocities of
2 AMPpase were measured in the presence of varying concentrations of dCMP (*triangles*),
3 AMP (*circles*), or CMP (*squares*). The results for CMP were obtained by both the enzyme
4 coupling method (*open symbols*) and HPLC (*filled symbols*). The concentration of Pi was
5 constant at 20 mM. (D) Dependence of AMPpase activity on Pi concentrations. Initial
6 velocities were measured in the presence of varying concentrations of Pi and 20 mM AMP.

7
8 **FIGURE 4.** Protein levels of AMPpase, R15P isomerase and Rubisco in *T. kodakarensis* cells.
9 Western blot analyses using an antiserum containing polyclonal anti-AMPpase, anti-R15P
10 isomerase, or anti-Rubisco antibodies were performed against cell-free extracts of *T.*
11 *kodakarensis* cells grown in ASW-YT-Pyr and ASW-YT-Pyr-Nuc medium.

12
13 **FIGURE 5.** Substrate specificities and regulatory properties of AMPpase, R15P isomerase
14 and Rubisco in *T. kodakarensis*.

Fig. 1. Aono *et al.*

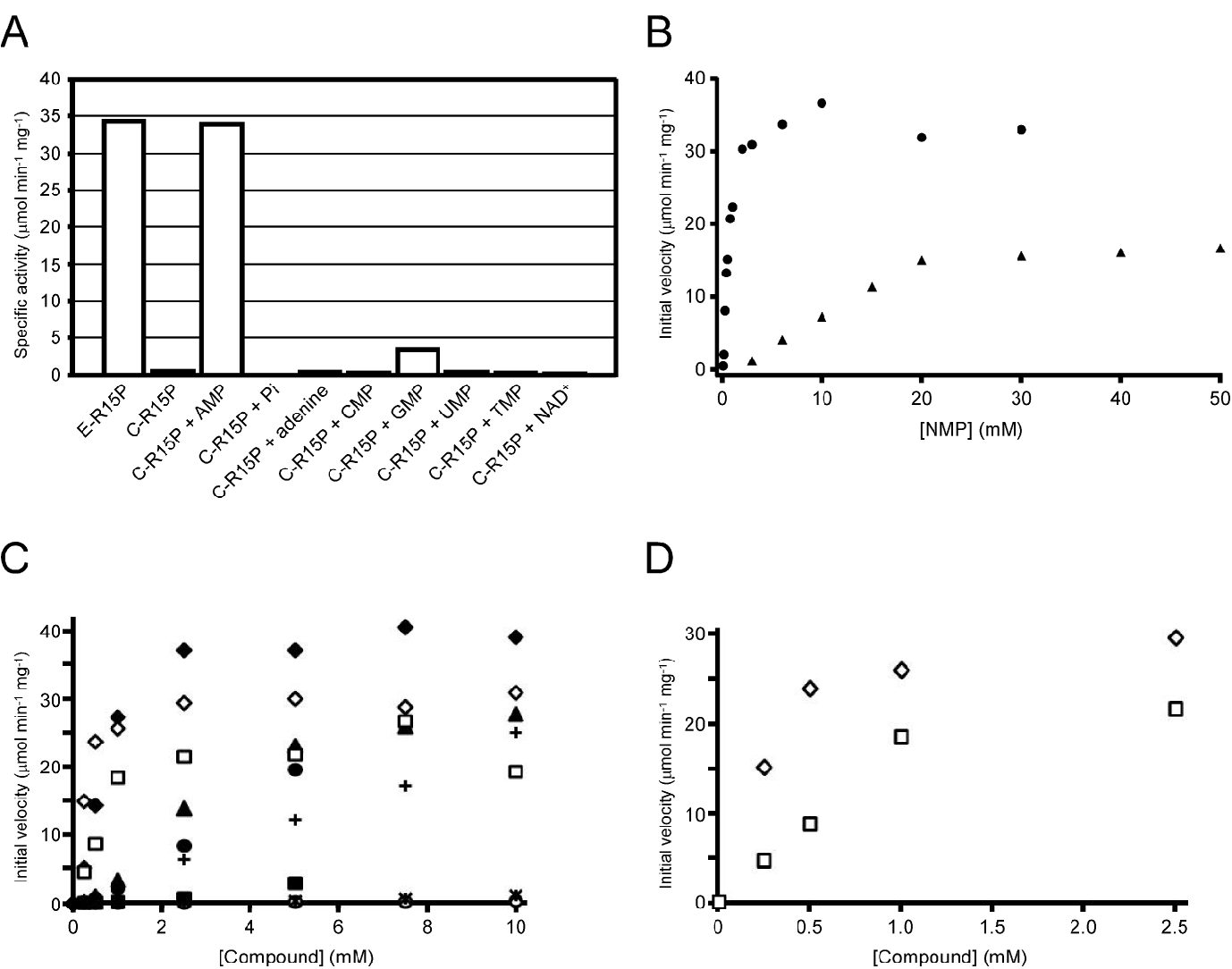


Fig. 2. Aono *et al.*

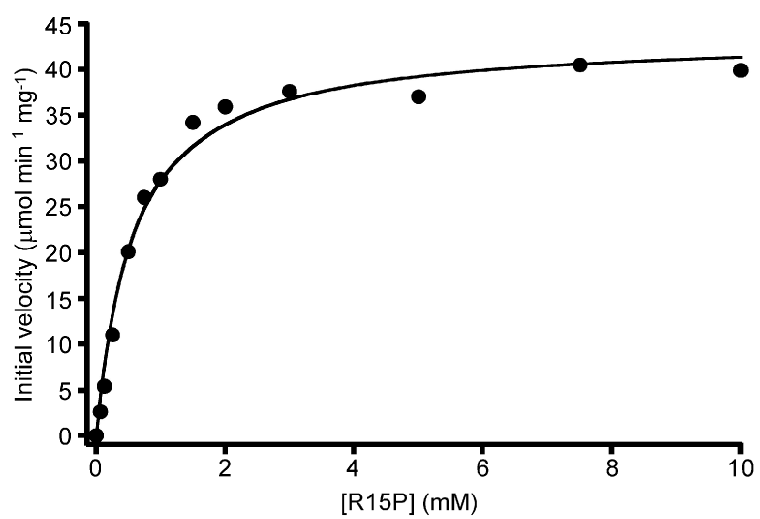


Fig. 3. Aono *et al.*

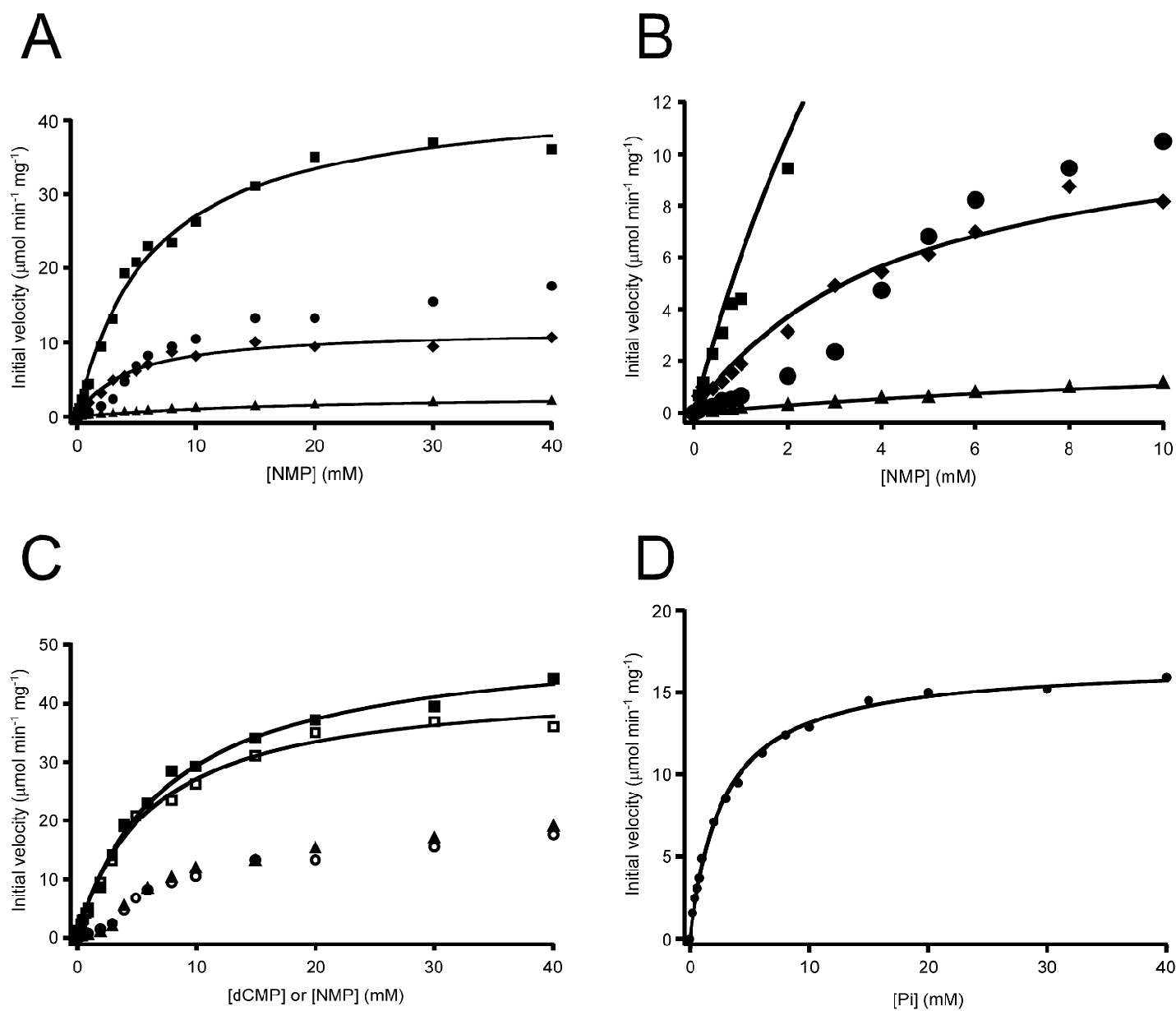


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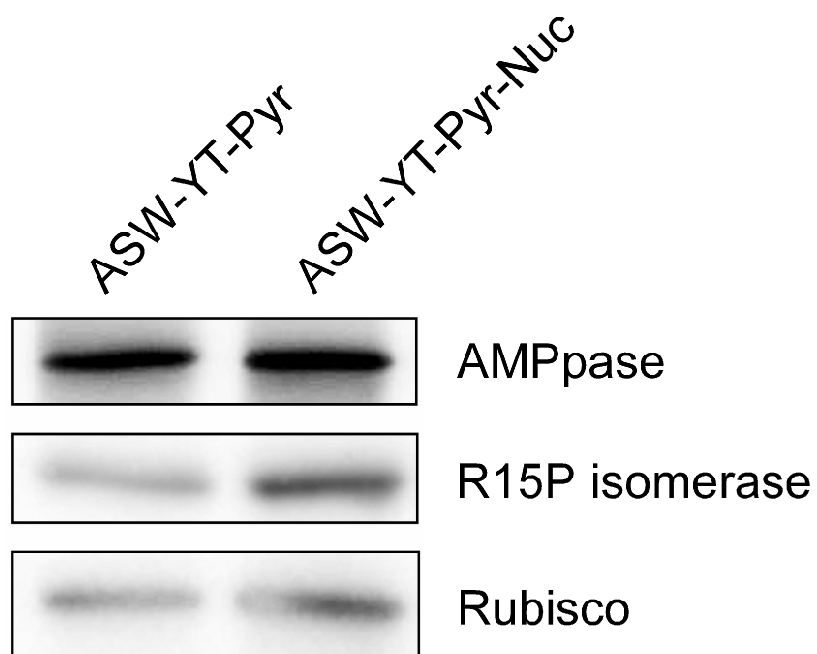
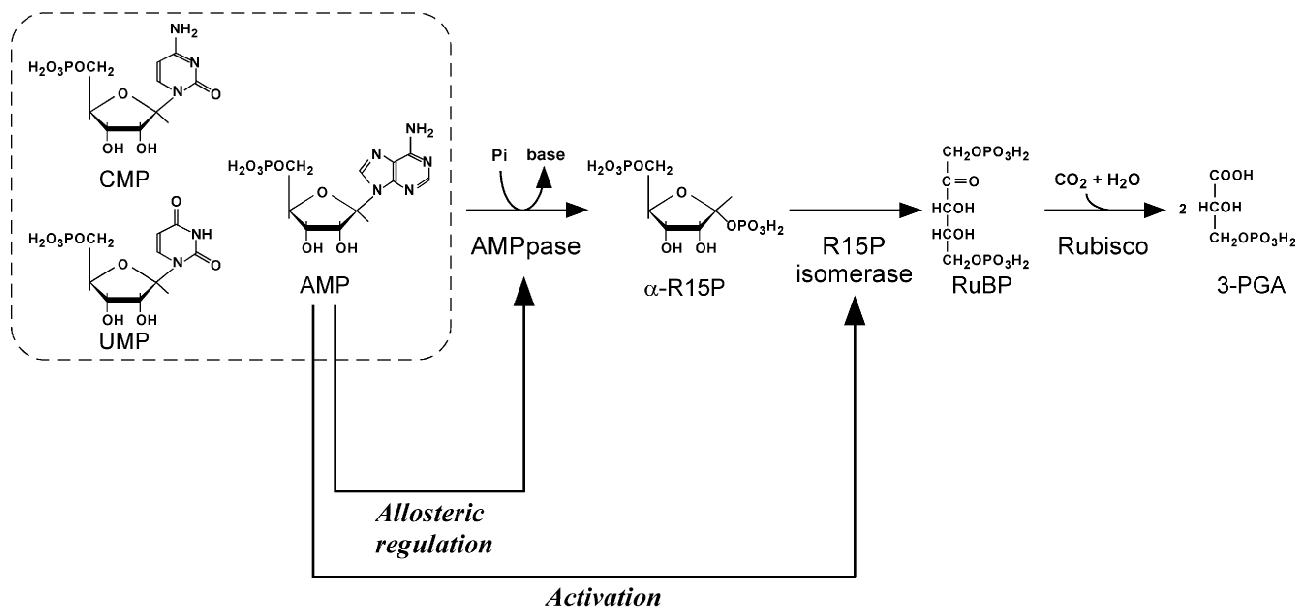


Fig. 5. Aono *et al.*



SUPPLEMENTAL MATERIAL

Enzymatic characterization of AMP phosphorylase and ribose-1,5-bisphosphate isomerase functioning in an archaeal AMP metabolic pathway

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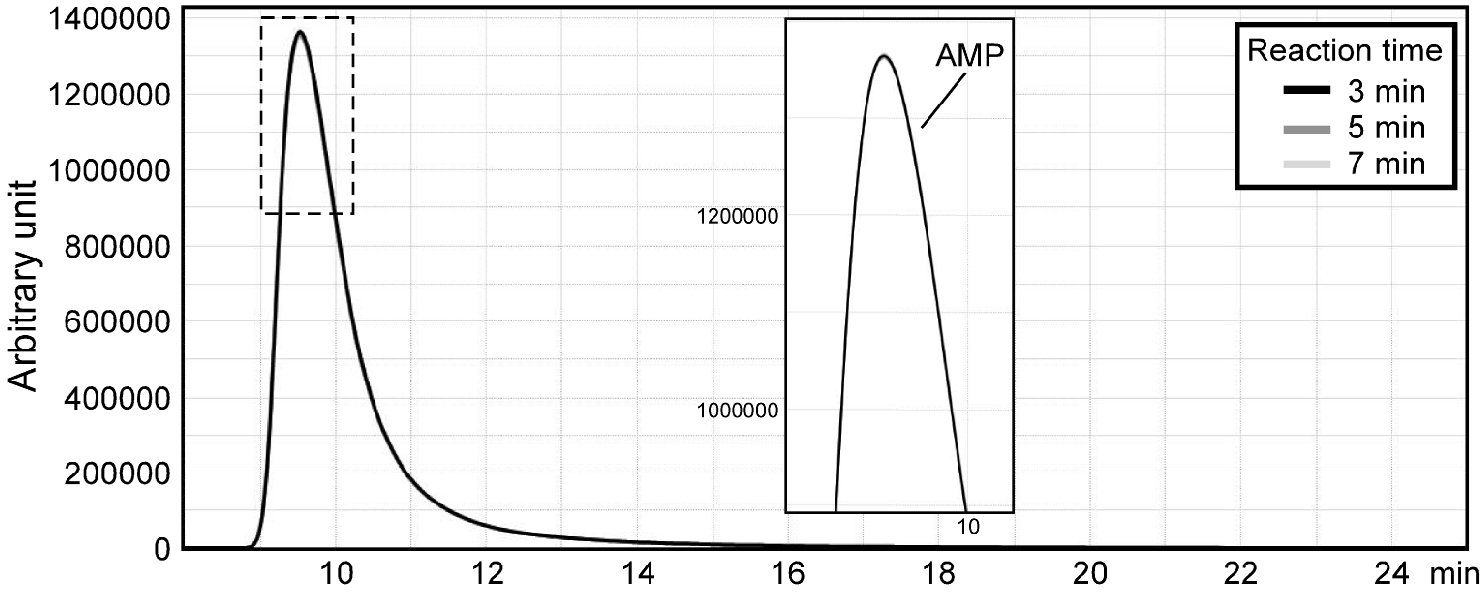
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LEGEND TO FIGURE

Fig. S1. HPLC analyses of the R15P isomerase reaction in the presence of AMP. (A) Quantification of AMP after 3, 5, and 7 min of the R15P isomerase reaction. AMP was measured with a UV detector (A_{254}). (B) Quantification of R15P after 3, 5, and 7 min of the R15P isomerase reaction. R15P was measured with a refractive index detector. Insets are enlarged views of the regions boxed with dotted lines.

Fig. S1. Aono *et al.*

A



B

