

1 **“*Mamonoviridae*”, a proposed new family of the phylum *Nucleocytoviricota***

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13 Abstract

14 *Acanthamoeba castellanii* medusavirus J1 is a giant virus isolated from a hot spring in Japan
15 in 2019. Recently, a close relative of this virus was also isolated in Japan and named
16 medusavirus stheno T3. Here, we describe their morphological, genomic and gene content
17 similarities and also propose to create a new family “*Mamonoviridae*”, a new genus,
18 “*Medusavirus*”, and two species, “*Medusavirus medusae*” and “*Medusavirus sthenus*” to
19 classify these two viruses within the phylum *Nucleocytoviricota*.

20 Introduction

21 Various amoeba-infecting giant viruses have been isolated during the last 20 years [1–6].
22 They are characterized by the large size of their genomes and particles. This group of viruses
23 have been classified in the phylum *Nucleocytoviricota* [7]. Nucleo-Cytoplasmic Virus
24 Orthologous Groups (NCVOGs) and Giant Virus Orthologous Groups (GVOGs) have been
25 widely used for core gene identification and to conduct comprehensive classification of
26 these viruses [8, 9]. As a result, the structure of the phylum *Nucleocytoviricota* has been
27 recently challenged and an expansion from seven currently recognized families
28 (*Mimiviridae*, *Phycodnaviridae*, *Ascoviridae*, *Iridoviridae*, *Marseilleviridae*, *Asfarviridae*,
29 and *Poxviridae*) to 32 families has been proposed [8, 10, 11].

30 Recently, two viruses were discovered by co-culture with *Acanthamoeba castellanii*. The
31 first one was isolated from a hot spring in Japan and was named *Acanthamoeba*
32 *castellanii* medusavirus J1 (ACMV-J1), because the host amoebae tend to form cysts upon
33 infection with this virus and this phenomenon is reminiscent of Medusa in Greek mythology
34 [4]. The second isolate was a close relative of the first virus and named medusavirus stheno
35 T3 (MVS-T3), because Stheno is a sister of Medusa [5]. Both viruses show substantial
36 morphological and genomic similarities with the members of the phylum *Nucleocytoviricota*.
37 However, these viruses were not phylogenetically close to any member of established
38 families within *Nucleocytoviricota*. Thus, in order to officially classify these two viruses
39 within the ICTV framework Virus Taxonomy, we propose to create two species,
40 “*Medusavirus medusae*” (typified by ACMV-J1) and “*Medusavirus sthenus*” (with an
41 exemplar isolate MVS-T3), to classify them in the new genus and family, “*Medusavirus*”

42 and “*Mamonoviridae*” respectively, which belongs to the class *Megaviricetes* of the phylum
43 *Nucleocytoviricota*.

44 **Etymology of taxa nomenclature**

45 Species and genus nomenclature was inspired by the two Gorgon sisters from Greek
46 mythology (Medusa and Stheno), while the family name originates from Japanese word
47 “mamono” (魔物), meaning “monster”.

48 **Infection cycle**

49 ACMV-J1 was shown to enter the host cell by endocytosis and then, enter the host nucleus
50 approximately one hour post infection (hpi). The virus gradually transformed the host
51 nucleus into a viral factory without disrupting the nuclear membrane. At around 10 hpi, the
52 cytoplasm was filled with empty viral capsids and eventually, viral particles were released
53 outside the cell in a non-lytic way at around 14 hpi [4, 12] (Fig. 1a).

54 **Genomic and proteomic features**

55 The ACMV-J1 virion shows an icosahedral shape with a diameter of approximately 260 nm,
56 including surface spikes, as revealed by cryo-electron microscopy (cryo-EM) [12] (Fig
57 1b&1c). It encapsidates a linear, double-stranded DNA (dsDNA) genome of 381,277 bp
58 with a high G+C content (61.7%) [4]. A total of 461 open reading frames (ORFs) have been
59 predicted in the genome. ACMV-J1 genome encodes five of the seven core genes of
60 *Nucleocytoviricota* that are frequently used in phylogeny [8]. These are major capsid protein
61 (MCP), superfamily II helicase (SFII), DNA polymerase family B (PolB), A32-like
62 packaging ATPase (A32), and virus late transcription factor (VLTf3). However, the virus
63 is unique among other amoeba-infecting giant viruses in encoding a full set of histone
64 proteins (i.e., linker histone H1, and core histones H2A, H2B, H3, and H4) and lacking two
65 of the core genes, namely, RNA polymerase and DNA topoisomerase II (TopoII).

66 MVS-T3 was isolated in 2021 [5]. This virus shows icosahedral particles similar to those of
67 ACMV-J1, and has a G+C-rich (62.64%), 362,811 bp-long dsDNA genome. The average

68 nucleotide identity (ANI) between ACMV-J1 and MVS-T3 is 79.5%. MVS-T3 has the same
69 set of core genes as ACMV-J1 and also encodes a full set of histones, but the genes for H3
70 and H4 are fused into a single gene.

71 **Phylogenomics**

72 To clarify the relationship between medusaviruses and other members of the
73 *Nucleocytoviricota*, we used the seven core genes from GVOGs, which have been argued to
74 have the optimum performance for phylogenetic analysis of *Nucleocytoviricota* (i.e., PolB,
75 SFII, A32, VLTF3, TopoII, TFIIB, RNAPL) [8]. The two medusaviruses formed a clade in
76 the phylum *Nucleocytoviricota* with a high branch support (SH-aLRT = 100%, Ultrafast
77 bootstrap = 100%) (Fig. 2), consistent with a previous study that demonstrated that ACMV-
78 J1 does not belong to any virus group identified so far [4]. In the tree, medusaviruses are
79 close to Feldmannia species virus, Ectocarpus siliculosus virus 1, coccolithoviruses,
80 pandoraviruses and molliviruses, which were previously suggested to form a putative and
81 yet non-recognized order, “*Pandoravirales*” [8]. However, the branch support for this clade
82 was weak (SH-aLRT = 97.9% and Ultrafast bootstrap = 58%). Thus, here we focus on
83 position of the two medusaviruses and propose to create two new species in a new genus
84 and a new family.

85 **Relationship between medusaviruses and clandestinovirus**

86 Recently, another giant virus named clandestinovirus was isolated by co-culture with
87 another host, *Vermamoeba vermiformis*, in France [6]. The clandestinovirus shows a larger
88 genome, more genes and a lower G+C content (581 kbp, 617 genes, 43.5%) than
89 medusaviruses. In terms of core genes, clandestinovirus encodes all core genes
90 that medusaviruses have and additionally encodes RNA polymerase and TopoII. Alike
91 medusaviruses, clandestinovirus also induces a nucleo-cytoplasmic infection, and enters and
92 turns the host nucleus into the viral factory. A previous study has shown that the closest
93 relative of clandestinovirus is ACMV-J1 in terms of the core genes [6].

94 Here, we used a quantitative way to draw a family-level boundary to figure out the
95 relationship between clandestinovirus and medusaviruses. We compared these three viruses

96 in terms of the nucleotide level similarity, including ANI and tetra-nucleotide similarity
97 (TETRA), phylogenomic distance by calculating the distance between tips on the
98 phylogenomic tree, and number of shared OGs. We then compared these metrics between
99 them to the inter- and intra-family metrics for other virus families. As a result, the
100 relationship between medusaviruses and clandestinovirus lies in the middle of inter-family
101 and intra-family levels.

102 In terms of phylogenomic tree, the clandestinovirus branched together with medusaviruses
103 with a high branch support (Ultrafast bootstrap = 100%, SH-aLRT = 98.8%) (Fig. 2).
104 However, the tip distances (3.92 to ACMV-J1 and 3.95 MVS-T3) lay between mean values
105 for intra-family (2.46) and inter-family distances (7.30) (Fig. 3a).

106 In terms of genome-level nucleotide similarity, ANI and TETRA were calculated by python
107 package pyani [13]. The ANI between clandestinovirus and the two medusaviruses were
108 both 0, whereas the average of intra- and inter-family were 0.36 and 0.01, respectively. In
109 addition, only kaumoebavirus had non-zero ANI (0.68) against clandestinovirus. The
110 TETRA between medusaviruses and clandestinovirus were both 0.32, which was lower than
111 the average of inter-family TETRA (0.38). In addition, TETRA between clandestinovirus
112 and medusaviruses only ranked 134th and 139th among 220 comparisons between
113 clandestinovirus and other viruses. (Fig. 3b, 3c).

114 We then used Orthofinder v.2.5.2 to identify OGs and calculated the gene-sharing level S_{ij}
115 based on the number of shared OGs between viral genomes [14]. The number of shared OGs
116 was normalized by the total number of OGs of each virus under comparison using the
117 following formula:

$$118 \quad S_{ij} = \frac{OG_{ij}}{\sqrt{OG_i \times OG_j}}$$

119 Here, OG_{ij} is the number of shared OGs between virus i and j , and OG_i is the total number
120 of OGs in virus i . The gene-sharing level between clandestinovirus and medusaviruses (0.16
121 to ACMV-J1, ranked 30th among all comparisons between clandestinovirus and other

122 viruses; 0.17 to MVS-T3, 25th) lay between the mean values for intra- and inter-family
123 levels (0.47 and 0.07, respectively) (Fig. 3d).

124 Among known viruses, clandestinovirus is the closest relative of medusaviruses. However,
125 they show large divergence that places their phylogenetic relationships in the middle of
126 intra- and inter-family levels. Thus, at this moment we do not include the clandestinovirus
127 into the proposed new family “*Mamonoviridae*”.

128 Finally, we propose the following simple and ready-made criteria for species, genus and
129 family demarcations under the family “*Mamonoviridae*”. If a virus shares >95% ANI,
130 similar morphology, and comparable genome size to the members of two proposed species
131 (e.g., “*Medusavirus medusae*” and “*Medusavirus sthenus*”) in the genus “*Medusavirus*”, it
132 should be classified in one of these two taxa. The average of intra-genus ANI is 70% within
133 five families of the phylum *Nucleocytoviricota* (i.e., *Mimiviridae*, *Ascoviridae*,
134 *Phycodnaviridae*, *Poxviridae* and *Iridoviridae*). By taking this statistic in consideration, we
135 propose that if a virus shares >70% ANI, similar morphology, and comparable compositions
136 of core genes to the members of the proposed genus “*Medusavirus*”, it should be classified
137 in this genus. For a virus distantly related to the members of this proposed family
138 “*Mamonoviridae*”, its inclusion in or exclusion from the family should be considered based
139 on phylogenomic analyses like those we presented in this study. We acknowledge that these
140 criteria are subject to updated according to the progress of analytical methods and
141 discoveries of new traits in viruses.

142 **Conclusion**

143 Medusaviruses are amoeba-infecting giant viruses that carry out a nucleo-cytoplasmic
144 infection cycle and are unique among known viruses by encoding a full set of histone genes.
145 Currently, there are two well-characterized but not yet officially classified medusaviruses
146 (ACMV-J1 and MVS-T3). Our phylogenomic analysis revealed that this group of viruses
147 does not branch within any groups of viruses. Thus, based on overall characteristics of the
148 two currently known medusaviruses, in particular genome features and phylogenomics, here
149 we propose creation of two species, “*Medusavirus medusae*” and “*Medusavirus sthenus*” in

150 a new genus, “*Medusavirus*” and a new family “*Mamonoviridae*” to classify ACMV-J1 and
151 MVS-T3, respectively. We propose that the new family is included in the class
152 *Megaviricetes* of the phylum *Nucleocytoviricota*. This article is related to a taxonomic
153 proposal, recently officially submitted to the ICTV for consideration, but not yet
154 approved/ratified at the time of publication. Therefore, taxa proposed in this paper are not
155 part of the official ICTV taxonomy.

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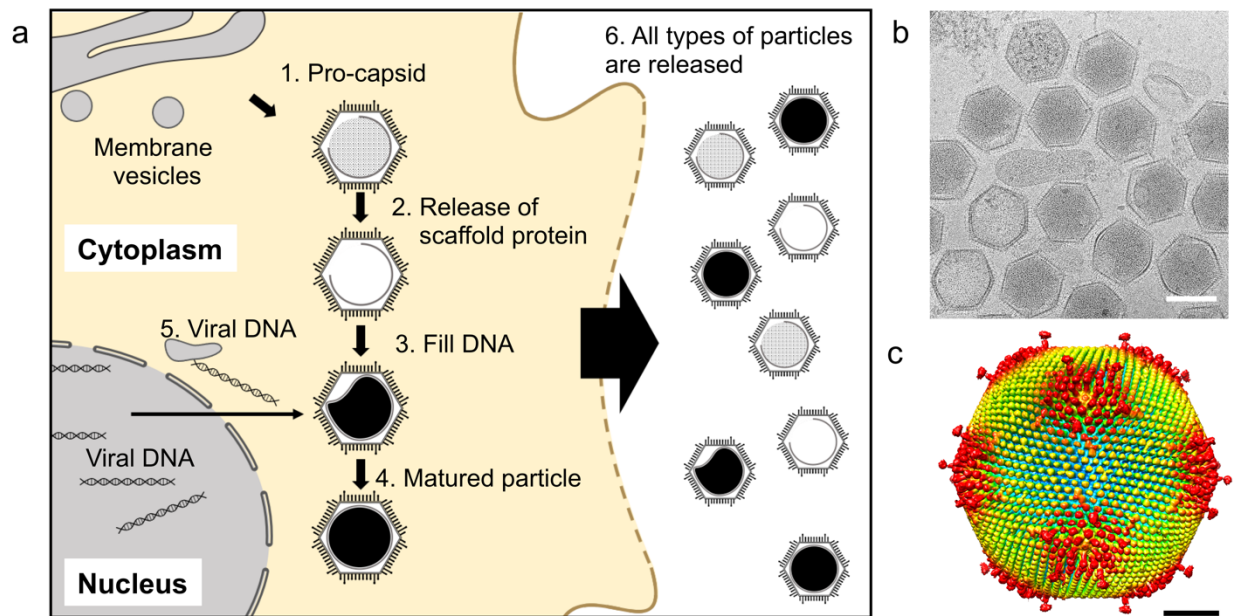
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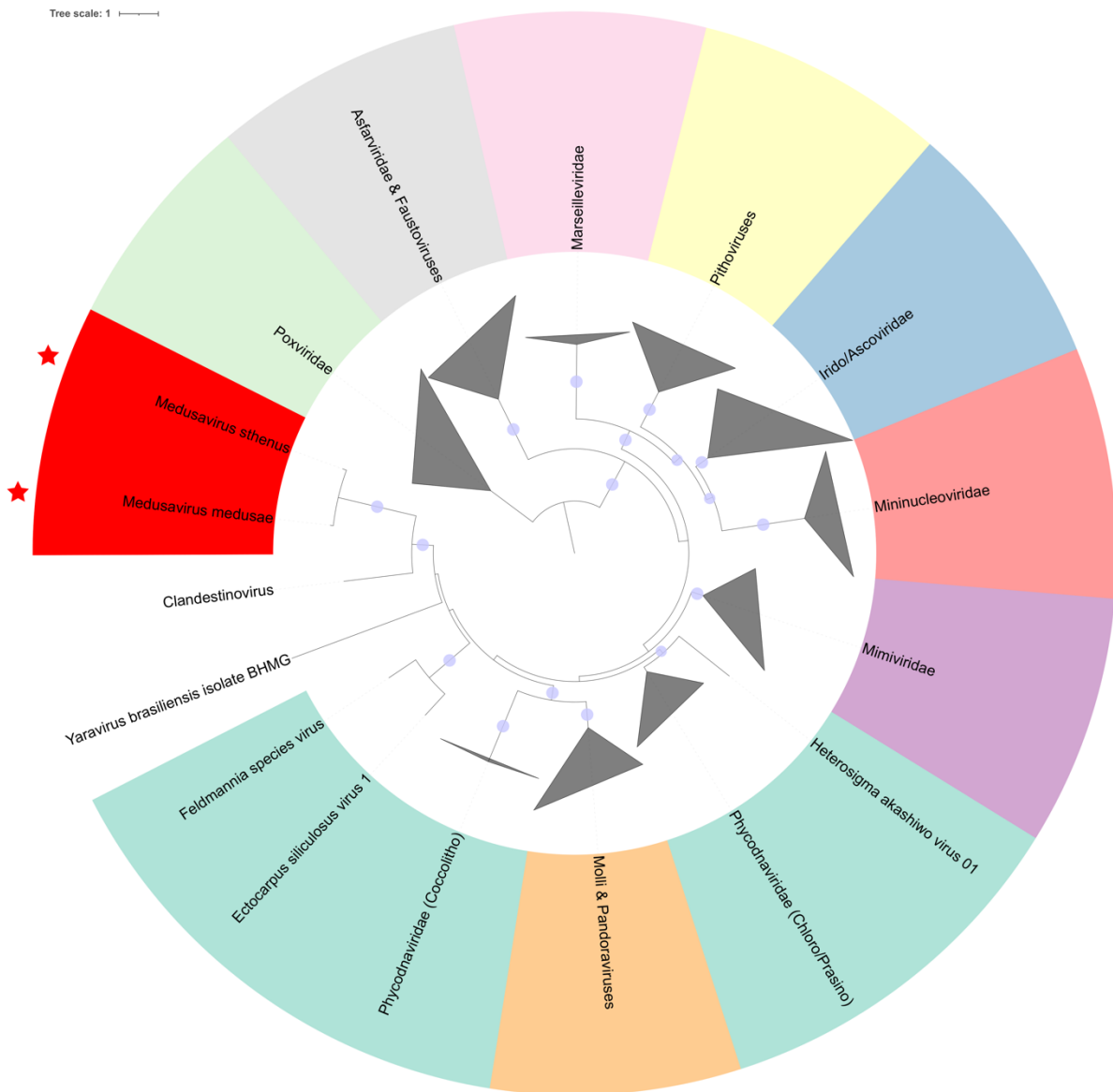
224 **Figures**

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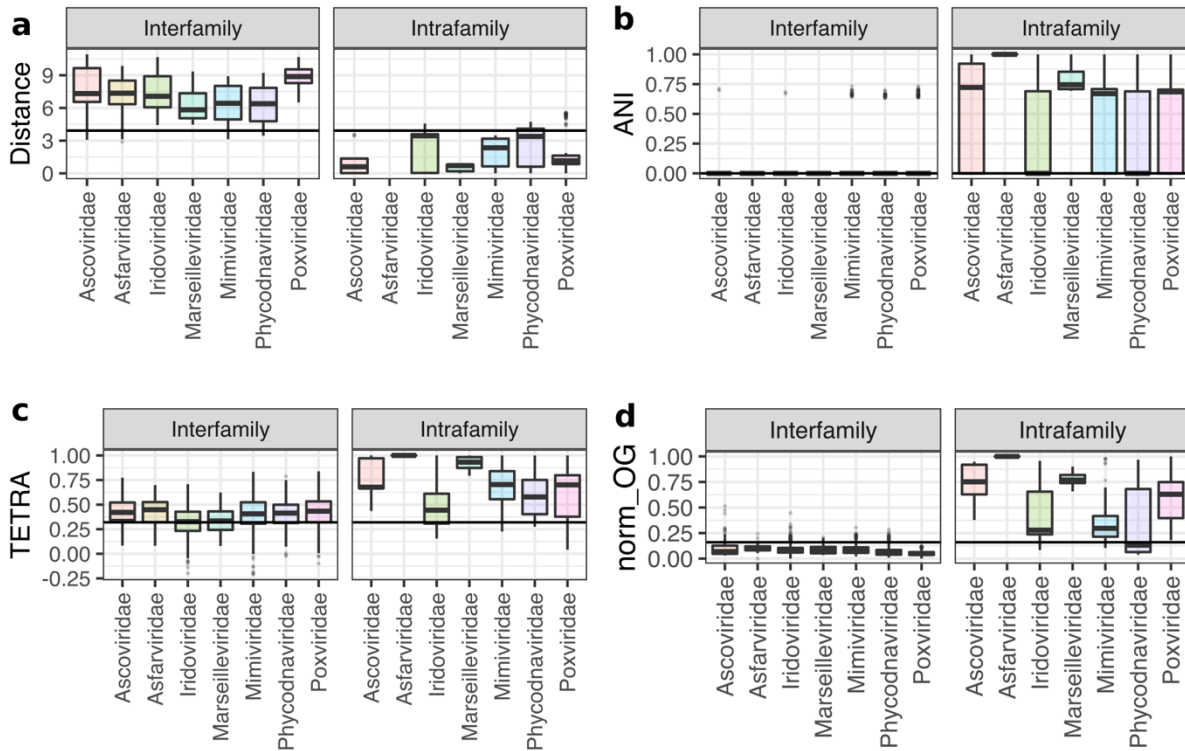
227 Fig. 1 *Acanthamoeba castellanii* medusavirus J1 (ACMV-J1) replication and its particle feature
 228 [12]. (a) ACMV-J1 replication in amoeba cell after infection. (b) A cryo-EM image of ACMV-J1.
 229 Scale 200 nm. (c) A 3D reconstruction of ACMV-J1 virion. Scale 50 nm.



230

231 Fig 2 Maximum-likelihood phylogenetic tree of *Nucleocytoviricota*. The tree was based on a
 232 concatenated amino acid sequence alignment of seven marker genes constructed using MAFFT
 233 (v.7.471) and trimAl (v.1.4.1) and was built using IQ-TREE 2 (v.2.1.3) [15–17]. The model was
 234 LG+F+R8 selected by the built-in Modelfinder of IQ-TREE 2 [18]. The branch supports were
 235 computed by 1000 ultrafast bootstrap and SH-aLRT [19]. The tree was visualized by iTOL,
 236 the round labels on branches represent high confidence supports with Ultrafast bootstrap \geq

237 95%, SH-aLRT \geq 80%. Position of proposed family “*Mamonoviridae*” is reported in red
 238 background and marked with stars.



239

240 Fig. 3 Boxplots for (a) tip distance, (b) ANI, (c) TETRA, and (d) normalized OGs sharing level.
 241 The horizontal black line represents the value between clandestinovirus and Acanthamoeba
 242 castellanii medusavirus J1 (ACMV-J1), a member of proposed species “*Medusavirus*
 243 *medusae*”.