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A Monolithic Dual-Color Total-Internal-Reflection-Based Chip for Highly Sensitive and High-Resolution Dual-Fluorescence Imaging

Nam Cao Hoai Le, Dzung Viet Dao, Ryuji Yokokawa, Member, IEEE, John C. Wells, and Susumu Sugiyama, Member, IEEE

Abstract—We report a dual-color total-internal-reflection (TIR)-based chip that can generate two overlapping evanescent fields with different wavelengths for simultaneous imaging of two types of fluorophores. We derived a general relationship among the dimensions of the components of the chip to guarantee the overlap of two evanescent fields. Optical simulation results also confirm the generation and overlap of two evanescent fields. Using Si bulk micromachining and poly(dimethylsiloxane) (PDMS) casting, our fabrication method integrates all miniaturized optical components into one monolithic PDMS chip. Thus, assembly is unnecessary, and misalignment is avoided. Our PDMS chip can be employed with various sample delivery platforms, such as glass slide, flow cell, microchannel, etc. We first demonstrated the capability of the chip by imaging TIR fluorescent spots of a mixture of two fluorophores, namely, fluorescein and tetramethylrhodamine. We then employed the chip to observe the Brownian motion of a mixture of nile-red and dragon-green 500-nm microbeads. Our chip could potentially be integrated into a micro-total analysis system for highly sensitive and high-resolution dual-fluorescence imaging applications.

Index Terms—Dual-color total-internal-reflection fluorescent microscopy (TIRFM), dual-fluorescence imaging, evanescent fields, micro-total analysis systems (μ-TAS), poly(dimethylsiloxane) (PDMS).

I. INTRODUCTION

AMONG MANY fluorescence-based investigation techniques, such as epifluorescence microscopy, confocal fluorescence microscopy, fluorescence correlation spectroscopy, etc., total-internal-reflection fluorescence microscopy (TIRFM) has received increasing recognition in the past decades due to its unique capabilities. The working principle of TIRFM relies on the evanescent-field illumination generated by TIR at a liquid/dielectric interface [1]. This evanescent field, whose depth of penetration is on the order of 100–300 nm, is capable of selectively exciting fluorophores in the liquid region very close to the dielectric material. The extremely thin excitation depth greatly reduces the background noise, which is often the biggest problem in fluorescence detection and imaging, since the fluorophores in the bulk liquid are not excited. Fluorescent images obtained by this evanescent excitation typically possess very high signal-to-noise ratio and virtually no out-of-focus signal from the bulk liquid.

Within the last two decades, TIRFM has been increasingly applied by many researchers for studying the dynamics of biological systems from cell membranes to single biomolecules [2]–[8] and for characterizing micro-/nanofluidic flows [9]–[13]. Very recently, some groups have proposed several configurations of dual-color TIRFM that can generate two overlapping evanescent fields with different excitation wavelengths for simultaneous detection of two different fluorophores either inside a cell or on a glass surface. In cell imaging, Tengholm et al. [14] reported a prism-type dual-color TIRFM system for the simultaneous detection of recruitment and dissociation of two differently labeled fluorescent proteins to and from the plasma membrane. Schmoranzer and Simon [15] used an objective-type dual-color TIRFM system to observe the transport, docking, and fusion of vesicles along microtubule cytoskeletons in a cell membrane. Similarly, Ikuko et al. [16] utilized an objective-type dual-color TIRFM system to study the colocalization of two single molecules in living cells. For application on a glass surface, using a prism-type dual-color TIRFM, Kang et al. [17] and Lee et al. [18] reported the detection of the hybridization and interaction of two single biomolecules on
the surface of a glass chip. Leutenegger et al. [19] combined an objective-type dual-color TIRFM system with a cross-correlation spectroscopy to study the binding of two dual-labeled double-stranded DNA molecules on glass surfaces.

Although those conventional objective- and prism-type dual-color TIRFM systems have been powerful tools in the aforementioned fields, they are highly costly, bulky, and elaborate and time consuming in assembly and alignment. Miniaturized and integrated dual-color TIR-based devices might offer lower cost and easier setup than conventional dual-color TIRFM systems. Furthermore, such devices might allow integration with micro-total analysis systems (μ-TAS) for highly sensitive dual-fluorescence detection and imaging applications. Recently, some groups, including ours, have proposed various configurations of integrated, miniaturized, and low-cost TIR-based devices [20]–[25]. However, these devices can only provide one excitation wavelength for the detection of one type of fluorophores.

We propose here, to our knowledge, the first integrated and miniaturized dual-color TIR-based device fabricated using Si micromachining and poly(dimethylsiloxane) (PDMS) casting technology. This paper presents the design, analysis, simulation, fabrication, and evaluation of the proposed chip. We derived a set of equations governing the overlap condition of two evanescent fields in a given chip configuration. A simple yet effective fabrication method has enabled us to integrate all miniaturized optical components including a prism, cylindrical microlenses, and fiber alignment channels into one monolithic PDMS chip. Demonstrations of the chip for the simultaneous imaging of mixtures of two fluorescent dyes and two different fluorescent microbeads will be presented.

II. DESIGN, ANALYSIS, AND SIMULATION

A. Design Concept

The cross section of the chip with the optical path is schematically shown in Fig. 1, while the nomenclature of the chip dimensions is listed in Table I. The monolithic PDMS chip consists of a prism in the center, two cylindrical microlenses, and fiber alignment channels into one monolithic PDMS chip. Demonstrations of the chip for the simultaneous imaging of mixtures of two fluorescent dyes and two different fluorescent microbeads will be presented.

![Cross-sectional view (not to scale) of the dual-color TIR-based chip with glass flow cell. The nomenclature of the chip dimensions are listed in Table I. The incident angles of green and blue excitations are 68.1° and 70.0°, respectively. They are both larger than the critical angle θC = 61.° (for the glass/water interface), satisfying the condition for TIR for both excitation lasers. The overlapping evanescent fields simultaneously excite the mixture of two different fluorophores. The emission is passed to an objective lens, chromatically separated by a specialized filter setup and imaged side by side on a CCD camera.]

![Table I: Nomenclature of the Principal Parameters of the Device](image-url)

TABLE I: NOMENCLATURE OF THE PRINCIPAL PARAMETERS OF THE DEVICE

<table>
<thead>
<tr>
<th>Symbol</th>
<th>Quantity</th>
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<tbody>
<tr>
<td>lP</td>
<td>Length of base of prism</td>
</tr>
<tr>
<td>WP</td>
<td>Width of base of prism</td>
</tr>
<tr>
<td>lB</td>
<td>Length of chip base</td>
</tr>
<tr>
<td>WB</td>
<td>Width of chip base</td>
</tr>
<tr>
<td>tB</td>
<td>Thickness of chip base</td>
</tr>
<tr>
<td>tG</td>
<td>Glass slide thickness</td>
</tr>
<tr>
<td>hP</td>
<td>Height of prism</td>
</tr>
<tr>
<td>tC</td>
<td>Height of core center of optical fiber</td>
</tr>
<tr>
<td>tL</td>
<td>Height of cylindrical lens and fiber alignment channel</td>
</tr>
<tr>
<td>r1</td>
<td>Rear radius of curvature of cylindrical microlens</td>
</tr>
<tr>
<td>r2</td>
<td>Front radius of curvature of cylindrical microlens</td>
</tr>
<tr>
<td>f</td>
<td>Rear focal length of cylindrical microlens</td>
</tr>
<tr>
<td>tM</td>
<td>Thickness of cylindrical microlens</td>
</tr>
<tr>
<td>tW</td>
<td>Effective width of cylindrical microlens</td>
</tr>
<tr>
<td>c1</td>
<td>Rear opening length of fiber alignment channel</td>
</tr>
<tr>
<td>c2</td>
<td>Contact length of fiber alignment channel</td>
</tr>
<tr>
<td>c3</td>
<td>Front opening length of fiber alignment channel</td>
</tr>
<tr>
<td>c4</td>
<td>Rear opening gap of fiber alignment channel</td>
</tr>
<tr>
<td>c5</td>
<td>Contact gap of fiber alignment channel</td>
</tr>
<tr>
<td>c6</td>
<td>Front opening gap of fiber alignment channel</td>
</tr>
<tr>
<td>c7</td>
<td>Gap of throat of fiber alignment channel</td>
</tr>
<tr>
<td>c8</td>
<td>Gap of exit of fiber alignment channel</td>
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cells. This feature is very useful since the chip can be reused, while the sample delivery platforms can be disposable. It also facilitates surface treatments or immobilizations on the sample delivery platforms before observations. Furthermore, by simply turning the device upside down, it is possible to use it with both upright and inverted fluorescent microscopes [25].

B. Optical Ray Analysis

For simplicity, optical analysis only considered the central ray of the beam. We assumed that the bottom surface of the glass capillary touches the base, so the central ray of the beam will be at a height \( t_c \) from the chip base. Since the chip is symmetric, optical ray analysis from the green excitation side was performed first. Using Snell’s law and geometric relationships from Fig. 1, the following equations could easily be derived:

\[
\begin{align*}
\theta_1 &= 90^\circ - 54.7^\circ = 35.3^\circ \\
\theta_2 &= \sin^{-1} \left( \frac{n_A}{n_P} \sin \theta_1 \right) \\
\theta_3 &= 90^\circ - \theta_1 + \theta_2 = 90^\circ - \theta_1 + \sin^{-1} \left( \frac{n_A}{n_P} \sin \theta_1 \right) \\
\theta_1 &= \sin^{-1} \left( \frac{n_P}{n_G} \sin \theta_3 \right) \\
\theta_1 &= \sin^{-1} \left[ \frac{n_P}{n_G} \sin \left( 90^\circ - \theta_1 + \sin^{-1} \left( \frac{n_A}{n_P} \sin \theta_1 \right) \right) \right]
\end{align*}
\]

where \( n_A, n_P, \) and \( n_G \) are the refractive indices of air, PDMS, and glass, respectively. It should be noted that the refractive index of PDMS depends on wavelength, mixing ratio, and curing temperature [27]. As an approximation, we employed Conrady’s formula with three index–wavelength pairs (i.e., 1.465 at 460 nm, 1.422 at 610 nm [27], and 1.43 at 580 nm [28]) of PDMS to obtain a continuous fit [29]. The refractive indices of PDMS at 543 and 473 nm were then estimated to be 1.44 and 1.46, respectively. The refractive index of the cover glass is specified by the manufacturer (Matsunami Co., Japan) to be 1.52 and is negligibly dependent on wavelength. For green excitation (543 nm), substituting \( n_P = 1.44, n_G = 1.52, n_A = 1, \) and \( \theta_1 = 35.3^\circ \) into (5), we have \( \theta_1 = 68.1^\circ \). The same equation (5) can also be used if we consider the other side of the chip, i.e., blue excitation (473 nm), where \( n_P = 1.46, n_G = 1.52, n_A = 1, \) and \( \theta_1 = 35.3^\circ \), resulting in \( \theta_1 = 70.0^\circ \). Clearly, the incident angles of both excitation wavelengths are greater than the critical angle of \( \theta_c = 61^\circ \) (for the glass/water interface), so the condition for TIR is satisfied for both excitation lasers [24].

As mentioned earlier, we must ensure the overlap of the centers of two evanescent fields. From Fig. 1, it can be observed that this condition will be met if the centers of the green and blue evanescent fields both fall on the center of the prism projected on the cover glass of the flow cell. Considering again the green-excitation side of the chip, this condition can be expressed by

\[
l_1 = \frac{l_p}{2} - \frac{t_c}{\tan 54.7^\circ}.
\]
Again from Fig. 1, (7) and (8) can be derived as

\[ l_2 = t_g \tan \theta_1 \tag{7} \]
\[ t_b + t_c = \frac{l_1 - l_2}{\tan \theta_3}. \tag{8} \]

Substituting (7) into (8), and then \( l_1 \) from (8) into (6), we have

\[ t_b = \frac{l_p}{2 \tan \theta_3} - t_g \tan \theta_1 \tan \theta_3 - t_c \left( \frac{1}{\tan \theta_3 \tan 54.7^\circ} + 1 \right). \tag{9} \]

The cover glasses have a nominal thickness \( t_g \) of 170 \( \mu \)m. The O.D. of the glass capillaries is 250 \( \mu \)m, yielding a \( t_c \) of 125 \( \mu \)m. This value of \( t_c \) is obviously smaller than both prism height \( t_p \), typically in the range of 250–270 \( \mu \)m according to the wet-etching time, and cylindrical-lens height \( t_l \) defined by the 300-\( \mu \)m-thick Si wafers used in fabrication. While \( t_c \) must be smaller than both \( t_l \) and \( t_p \), it is necessary for the central ray of the beam to hit the prism in the midside node so that the divergent marginal rays, defined by the \( NA \) of the fiber, will still be confined within the height of the prism. Finally, \( t_b \) was plotted against varying \( l_p \)’s by putting the relevant parameters into (9) for green and blue excitations (Fig. 3). These lines show a linear relationship between chip base thickness \( t_b \) and chip prism size \( l_p \) in order for the center of the evanescent fields to fall in the center of the prism projected on the flow cell.

The smallest prism base lengths would be 2235 and 2288 \( \mu \)m for green and blue excitations, respectively, for a chip base thickness \( t_b \) of zero, i.e., when the prism and lenses are directly attached to the glass slide. However, a finite chip base thickness \( t_b \) is necessary to support and integrate the components of the chip. Considering the chip size with respect to the glass flow-cell size, the number of chips on a 4-in wafer, and the availability of the sizes of O-ring for the wet anisotropic etching jig, we chose \( l_p = 5042 \mu \text{m} \), yielding \( t_b = 599 \) and \( 611 \mu \text{m} \) for green and blue excitations, respectively. Choosing \( w_p = 2966 \mu \text{m} \) and taking into account the total length and width of the supporting wings of the fiber alignment channel of 5205 and 9966 \( \mu \text{m} \), respectively, the resulting chip size would be 10 mm \( \times \) 20 mm \( \times \) 0.9 mm in \( L \times W \times T \). This chip would be sufficiently small to attach wholly to the cover glass of the flow cell (24 mm \( \times \) 36 mm in \( L \times W \)) yet large enough to be handled manually.

C. Simulation

Other than ray analysis, the ZEMAX EE 2007\(^1\) optical simulation package was also employed to confirm our design concept and to find the dimensions of the cylindrical microlens, as well as the design parameters of the chip. First, the sequential mode was used to solve a three-variable (i.e., focal length, lens thickness, and radius of curvatures) optimization problem with target values of Gaussian beam radius and beam divergence angle to obtain optimal microlens parameters. For simplicity, we used the following assumptions and parameters in the simulation: The beam from the single-mode fiber had a Gaussian intensity distribution; the NA and mode field diameter (MFD) of the fiber were 0.12 and 9.2 \( \mu \text{m} \), respectively, obtained from the manufacturer (Sumitomo Inc., Japan); the excitation wavelength was 543 nm; and the number of analysis rays was 1000. It should be noted that, however, a standard single-mode fiber would typically have a cutoff wavelength near 1300 nm [30]. Thus, at a wavelength of 543 nm (or 473 nm) used in this paper, the single-mode fibers might transmit not only a fundamental mode, which has a Gaussian distribution, but also higher modes, unless a mechanism to filter out these additional modes is introduced [31]. The 1/e beam radius \( r = 30 \mu \text{m} \) (at the imaging plane that is 140 \( \mu \text{m} \) after the microlens where the prism base is located), together with the beam 0° divergence angle, was chosen as the target value for the optimization problem.

The obtained optimal microlens is of convex–convex type with front radius of curvature \( r_1 = -457 \mu \text{m} \), rear radius of curvature \( r_2 = 1628 \mu \text{m} \), thickness \( w_1 = 557 \mu \text{m} \), and rear focal length of \( f = 488 \mu \text{m} \) [Figs. 2 and 4(a)]. Fig. 4(b) shows the spot diagram obtained in the imaging plane. Since the cylindrical microlens can only collimate the laser beam in the horizontal direction, the beam diverges in the vertical direction when propagating along the chip. The elliptical shape of the spot diagram has a root-mean-square (rms) radius of 75 \( \mu \text{m} \), which is smallest compared to other lens configurations (data not shown). The widths of the beam obtained by using ZEMAX EE in both horizontal and vertical directions [Fig. 4(b)] agree very well with the ray analysis results obtained by using Snell’s law in Fig. 4(c) and (d), respectively. The 243-\( \mu \text{m} \) width of the beam in the vertical direction requires the use of a glass capillary to increase the height of the fiber core to 125 \( \mu \text{m} \). Since the prism height \( t_p \) is larger than 250 \( \mu \text{m} \), we can expect that all rays of the beam will enter the prism. We also solved the problem with blue wavelength (473 nm) and found that the lens dimensions differ negligibly from the case of green wavelength (543 nm).

Thus, these dimensions were applied for both cylindrical lenses.

In order to confirm the existence and overlap of two evanescent fields, the nonsequential mode in ZEMAX EE was used. We used the same parameters mentioned earlier but with an additional blue laser beam with a wavelength of 473 nm. The cover-glass thickness \( t_g \) and the height of the fiber core \( t_c \) were also fixed at 170 and 125 \( \mu \text{m} \), respectively. A detector surface, whose size is equal to the base of the prism, was inserted between the interface of glass and water to collect the incoherent irradiance of green and blue excitations. In previous work, the

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incoherent irradiance spots at the glass/water interface were used to estimate the size and shape of the evanescent fields [24]. In this work, we could control the overlap of two evanescent fields by finding the \( t_b \) for which the centers of two incoherent irradiance spots fall on the center of the prism while varying the prism length \( l_p \). Fig. 5(a) shows the extreme case when the chip base thickness \( t_b \) is zero, i.e., when glass directly is attached to the prism and the lenses, while Fig. 5(b) shows the design case when prism length \( l_p \) is 8042 \( \mu m \), leading to \( t_b \) of 600 \( \mu m \). In both cases, the two incoherent irradiance spots overlap at the center of the prism projected onto the glass flow cell. The size of the evanescent field in the design case is estimated to be 2500 \( \mu m \) in length and 190 \( \mu m \) in width [Fig. 5(b)]. The simulation results of \( t_b \) versus \( l_p \) are shown in Fig. 6, agreeing very well with the analysis results in Fig. 3. The small discrepancy might be attributed to the center ray versus whole beam tracing of optical ray analysis and simulation, respectively, and the capability of ZEMAX EE to take into account the index–wavelength dependence automatically in the simulation. Since the evanescent fields are quite long, a small difference in \( t_b \) for green and blue excitations would have negligible effect on their overlap. In fabrication, \( t_b \) is controlled by the volume of PDMS during casting.

### III. Fabrication Process

The polymer chip was fabricated using the standard Si bulk micromachining and PDMS casting techniques adapted from previous work [24]. The process flow is shown in Fig. 7(a)–(g). First, a 1.3-\( \mu m \)-thick oxide layer was formed on both sides of a 300-\( \mu m \) (100) single-crystal Si wafer by pyrogenic oxidation [Fig. 7(a)]. In the second step, the prism patterns on the front side and the microlens and the fiber channel patterns on the back side were defined by photolithography and SiO\(_2\) etching by buffered HF (BHF) solution [Fig. 7(b)]. The wafer was then diced into single chips (27 mm \( \times \) 27 mm) for the next steps. The third step was TMAH anisotropic wet etching [Fig. 7(c)] on the opened window of the prism on the front side. A Teflon jig was
used to protect the back side, containing opened Si patterns for the microlens and the fiber alignment channel during TMAH etching. A TMAH etch for 6 h was sufficient to etch up to 270-μm depth for an approximate etching rate of 0.75 μm/min.

The fourth step was through-wafer deep reactive-ion etching (DRIE) on the back side to create the cavities for the microlens and the fiber alignment channels [Fig. 7(d)]. It has been reported that a through-wafer DRIE process gives a very complex roughness topography at different depths on the sidewalls [32]–[34]. We have employed a thick SiO2 mask with a constant trench width of 50 μm to ensure smooth and straight sidewalls in through-wafer Si etching by DRIE [33]. With an optimized DRIE process, we could etch through 300-μm-thick Si chips within 90 min, yielding relatively smooth and vertical sidewalls. Both roughness average (Ra) and rms roughness were below 50 nm. The chips were then cleaned in Piranha for 30 min, and the remaining SiO2 was completely removed by BHF etching. We further reduced the roughness of the Si molds by three cycles of oxidation (the SiO2 thicknesses were 370, 230, and 230 nm, respectively) and BHF etching [Fig. 7(e)] [35]. The SiO2 growth has been shown to depend greatly on the feature geometry, meaning that convex corners grow faster than concave corners [36]. Accordingly, the sharp and high roughness features will be more oxidized and will be removed faster during BHF etching. With this process, we could achieve the Ra and rms roughness of the Si molds below 25 nm. This order of roughness, when transferred to PDMS, would be sufficiently small to fulfill Marechal’s criterion for scattering for both excitation wavelengths [37]. Fig. 8(a) and (b) shows a photograph and the scanning-electron-microscope (SEM) micrographs of the Si mold, respectively.

Next, in the sixth step, the Si mold was sealed to a PDMS film (thickness of 160 μm) stretching on top of a glass substrate to form a close mold [Fig. 7(f)]. The mold was then placed onto a flat hot plate. A PDMS (Silpot184W/C, Dow Corning, Japan) mixture (1:10 curing agent: prepolymer) was poured on the mold and was allowed to self-planarize for 1 h. Curing on the hot plate was done at 90 °C for 30 min. Finally, the PDMS chip replica was carefully removed from the Si mold [Fig. 7(g)]. As mentioned in Section II-A, the chip base thickness was controlled by the volume of liquid PDMS mixture during casting. For a Si mold of 27 mm × 27 mm, the volume of liquid PDMS required was 0.3 mL to achieve a tbd of approximately 600 μm (Fig. 9). Using a single Si mold, the dual-color TIR-based chips can be replicated by repeated PDMS castings. Fig. 8(c) and (d) shows a photograph and the SEM micrographs of the PDMS chip, respectively.

IV. EXPERIMENTAL SETUP AND MATERIALS

Fig. 10(a) shows the experimental setup using the dual-color TIR-based chip. A blue laser (DPBL-9050, Photop Suwtech, China, maximum power of 50 mW, 473 nm) and a green laser (05-LGR-173, Melles Griot, U.S., maximum power of 2 mW, 543 nm) were coupled into two single-mode optical fibers (core/cladding = 9.2 μm/125 μm and NA = 0.12, Sumitomo Inc., Japan). The chip was placed so that the glass flow cell facing an objective lens (10× or 60×, Olympus Inc., Japan) of a inverted microscope (IX71, Olympus Inc., Japan) equipped with an electron-multiplier CCD (EMCCD) camera (iXon EM+ DU-897, Andor Technology plc., Northern Ireland). A Dual-View (Optical Insights Inc., U.S.) was inserted between the emission port of the microscope and the C-mount of the camera to split the emission with a wavelength that was larger than 560 nm [mostly from tetramethylrhodamine (TMR) and nile red (NR)] to the emission filter ET 585/40 against the emission with wavelength smaller than 560 nm [mostly from fluorescein (Fl) and dragon-green (DG)] to the emission filter ET 515/30 [Fig. 10(b)].

A mixture of TMR (C-1171, Ex/Em = 532 nm/556 nm, Molecular Probes Inc., U.S.) and Fl (CI 45350, Ex/Em = 494 nm/521 nm, Wako, Japan) in DMSO at concentrations of 0.3 and 3.3 mM, respectively, were used in the observation of evanescent-field fluorescent spots, i.e., TIR fluorescent spots. A mixture of suspensions of carboxyl fluorescent NR beads (CFP-0556-2, Ex/Em = 532 nm/556 nm, 400–600 nm in diameter, Spherotech, Inc., U.S.) and DG beads (CP01F/8206, Ex/Em = 480 nm/520 nm, 510 nm in diameter, Bangs Laboratories, Inc., U.S.) dispersed in Tween20/DI water solution (0.05%), both at 0.05% w/v concentration, was used for the observation of the Brownian motion. The chip setup was fixed on the stage of the microscope, as shown in Fig. 10(c). In each experiment, a fluorescent sample with a volume of 10 μL was injected into the flow cell. For all experiments, the power of the lasers from both fibers were set to 0.52 mW measured by a laser power meter (PM-221, Neoark Corp., Japan). Fluorescent images were acquired and analyzed by image processing software (Andor IQ, Andor Technology plc., Northern Ireland). All experiments were performed in a dark room.

V. RESULTS AND DISCUSSION

A. Imaging of Overlapping TIR Fluorescent Spots From Two Fluorescent Dyes

Fig. 11(a)–(c) shows the fluorescent images of TIR fluorescent spots on the glass flow cell deposited with a mixture of TMR and Fl dyes on the ET 585/40 and ET 515/30 emission channels. Although the fluorescent dye solutions were spread...
Fig. 7. Fabrication process of the dual-color TIR-based chip. (a) Oxidation. (b) Back- and front-side SiO$_2$ patterning. (c) Front-side TMAH wet anisotropic etching. (d) Back-side DRIE. (e) SiO$_2$ removal and smoothing lens by oxidation and BHF etching. (f) PDMS casting. (g) PDMS chip removal.

Fig. 8. (a) Photograph of the Si mold of the dual-color TIR-based chip. (b) SEM micrographs of one side of the Si mold. (c) Photograph of the PDMS chip (the dimensions of the chip are 20 mm $\times$ 10 mm $\times$ 0.9 mm in $L \times W \times T$). (d) SEM micrographs of one side of the PDMS chip with glass capillary inserted.

Fig. 9. Chip base thickness $t_b$ versus PDMS volume during casting for the Si mold with a size of 27 mm $\times$ 27 mm.

all over the surface of the glass flow cell, only the dyes that were inside the evanescent fields were excited and emitted to form the TIR fluorescent spots. It is clearly seen that the TIR fluorescent spots from green and blue excitations have been separately generated when only one laser was turned on [Fig. 11(a) and (b)] and overlapped when both lasers were turned on [Fig. 11(c) and (d)]. The green excitation was targeted to excite the TMR dye, while the blue excitation was targeted to excite the FI dye. However, due to its broad emission spectrum [Fig. 11(e)], fluorescence emission from FI can go through both ET 585/40 and ET 515/30 emission channels, as can be seen from Fig. 11(b). Although the lengths of the TIR fluorescent spots are beyond the field of view of a 10$\times$ objective, the widths of the TIR fluorescent spots are measured to be approximately 90 $\mu$m for both green and blue excitations [Fig. 11(a) and (b)]. There is a large discrepancy between the widths of the evanescent fields estimated by simulation (approximately 190 $\mu$m) as compared to the widths of the TIR fluorescent spots (approximately 90 $\mu$m) measured from the fluorescent images. This discrepancy could be due to the possibility that the single-mode fibers might have allowed transmission of more than one fundamental mode at 543 or 473 nm. At these wavelengths, the fundamental mode is much more confined to the core of the fiber and thus has a smaller MFD than the ideal MFD of 9.2 $\mu$m, which was used in the simulation [38]. As can be seen in Fig. 11(a)–(d), there are a high-intensity region in the center excited by the fundamental mode and two much lower
Fig. 10. (a) Experimental setup with Olympus IX71 inverted microscope with 10× or 60× objective, Dual-View, iXon EM+ DU-89 EMCCD camera, and two single-mode fibers (core/cladding = 9.2 μm/125 μm) coupling optics with Melles Griot laser diode (543 nm, 2 mW), Photop Suwtech laser diode (473 nm, 50 mW), four mirrors, and two coupling lenses. (b) Construction of Dual-View adapted from http://www.magbiosystems.com/files/PDF/appnotes/dualview_appnote.pdf. (c) Close-up view of the chip setup on the stage of the microscope.

Fig. 11. Fluorescent images of the TIR fluorescent spots of a mixture of TMR dye (0.3 mM) and FI dye (3.3 mM) on the ET 585/40 and ET 515/30 emission channels when (a) only green excitation was turned on, (b) only blue excitation was turned on, and (c) both green and blue excitations were turned on are used. (d) Overlay pseudocolor image of ET 585/40 and ET 515/30 channels in (c). The fiber laser powers and exposure times in (a), (b), and (c) were all 0.52 mW and 300 ms, respectively. (e) (Dashed lines) Absorption and (solid lines) emission spectra of TMR and FI and their responses to emission filters and excitation laser sources, adapted from http://probes.invitrogen.com.

intensity regions on two sides of the high-intensity region that might be excited by the second mode. There have been several approaches to filter out the transmission of the higher modes in the single-mode fibers [31], [39], [40]. A small misalignment on the positions of the optical fibers along the longitudinal and vertical axes of the chip could also cause the variation on the size of the TIR fluorescent spots.

In previous work, the roughness transferred from the Si mold to the PDMS chip during the casting process has caused some fringe patterns of the TIR fluorescent spots [41]. In this paper, due to the smoothing of the Si lens mold fabricated by optimized DRIE and oxidation and BHF etching, we could achieve uniform excitation by the fundamental mode, as observed in the TIR fluorescent spots. This uniformity is essential to ensure the uniform excitation of all fluorophores within the field of view. It is, however, necessary to eliminate the higher modes in the optical fibers so that the evanescent fields could be excited by only the fundamental mode.

B. Imaging of Brownian Motion of Mixture of Suspensions of Fluorescent Microbeads

Fig. 12(a)–(c) shows the fluorescent snapshots of the Brownian motion of a mixture of suspensions of NR and DG
microbeads captured using the dual-color TIR-based chip on the ET 585/40 and ET 515/30 emission channels when the green laser was turned on, the blue laser was turned on, and when both lasers were turned on, respectively. The overlay of the highlighted regions from the two channels of Fig. 12(c) is shown in Fig. 12(d) with pseudocolors. Similar to imaging of fluorescent dyes, green and blue excitations were primarily aimed to excite the NR and DG microbeads, respectively. The broad emission spectrum of DG [Fig. 12(e)] also caused its fluorescence emission to go through both ET 585/40 and ET 515/30 channels, when both lasers were turned on, respectively. Overlay pseudocolor image of two regions of the ET 585/40 and ET 515/30 channels indicated in (c). The fiber laser powers and exposure times in (a), (b), and (c) were all 0.52 mW and 150 ms, respectively. (e) (Dashed lines) Absorption and (solid lines) emission spectra of NR and DG and their responses to emission filters and excitation laser sources, adapted from http://probes.invitrogen.com.

VI. CONCLUSION

A novel dual-color TIR-based chip for highly sensitive and high-resolution dual-fluorescence imaging has been presented. Optical analysis and simulation to confirm the overlap of two evanescent fields have been conducted. One could rely on the derived governing equations and vary the parameters to design the chips with different configurations. A low-cost simple fabrication method has been developed to integrate all miniaturized optical components into a monolithic PDMS chip. The monolithic PDMS chip can easily be integrated with various sample delivery platforms commonly used in μ-TAS, namely glass slides, flow cells, microchannels, etc. The capability of the chip for simultaneous dual-fluorescence imaging of two types of fluorescent dyes and two types of fluorescent microbeads has been experimentally demonstrated. The smooth PDMS lens has contributed to the uniform illumination of the evanescent fields, which is essential for the uniform excitation of all fluorophores in the field of view. Our device could be an alternative to conventional dual-color TIRFM. It could also potentially serve as a dual-color evanescent-excitation-based platform integrated into μ-TAS for dual-fluorescence imaging.

REFERENCES


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