LECTIN IMMUNOHISTOCHEMICAL EVALUATION OF HUMAN BLADDER CARCINOMAS: A COMPARISON OF CARNOY'S AND FORMALIN FIXATION

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A lectin immunohistochemical analysis of 51 human bladder carcinomas, including 44 cases of transitional cell carcinoma (TCC) (G1, 15 cases; G2, 17 cases; G3, 12 cases) and 7 cases of squamous cell carcinoma (SCC), was performed. Tissues were obtained by cold punch biopsies, fixed in Carnoy's or 10% formalin solution, stained for binding of 10 different lectins, and evaluated under the light microscope. The lectins used were concanavalin agglutinin (Con A), soybean agglutinin (SBA), Lotus tetragonolobus agglutinin (LTA), Dolichos biflorus agglutinin (DBA), peanut agglutinin (PNA), Ricinus communis agglutinin I (RCA I), Ulex europaeus agglutinin I, II (UEA-I, II), wheat germ agglutinin (WGA), and Pisum sativum agglutinin (PEA).

TCC prepared with Carnoy's fixation tended to show moderately positive Con A, UEA-I, and WGA reactions for G1, and strongly positive reactions for G2 and G3 lesions. UEA-II was mainly negative in G1, but tended to increase to become moderate in G3. DBA tended to show a moderately positive reaction in G1 and G2, but was mainly negative in G3. With formalin fixation, only RCA I demonstrated grade specific variation, tendency to react moderately in the G1 and G2 cases, and strongly in G3. There were no further differences among the histopathological grades of TCC for other lectins. Thus, Carnoy's fixation appears superior for distinguishing between grades of lesions. SCC tended to react more strongly than TCC with all the various lectins except PEA, independent of fixation.

(Key words: Bladder cancers, Lectin immunohistochemistry, Carnoy's fixation)

INTRODUCTION

Lectins, glycoproteins, found primarily in plants and in vertebrates, can recognize specific carbohydrates on cell surfaces. Because of this feature, they have been widely used in investigations of changes in carbohydrate moieties between normal and malignant states, and for grading various tissues, such as those of the alimentary tract, bronchial mucosa, uterus or ovary, thyroid gland, prostate, and others. Most previous studies were performed using formalin fixation, although a few investigators such as Walker have pointed out the importance of the fixation...
method. Several papers have in fact described different staining patterns obtained with the same lectins using different fixation methods\textsuperscript{3,6,11).}

Human bladder carcinomas have been investigated intensively using lectin immunohistochemistry because of difficulties in grading solely on the basis of histopathological findings. This is particularly important given the possibility of recurrence\textsuperscript{11-14} and the associated high risk of invasion\textsuperscript{15,16).

In the present study, we compared lectin binding patterns in carcinomas of the human bladder using two different methods of fixation, using Carnoy's and formalin solutions, to assess their relative merits.

\textit{MATERIALS AND METHODS}

\textbf{Materials}

A total of 51 specimens of urinary bladder carcinoma, from 42 men and 9 women, age 30 to 87 years, were examined. All specimens were obtained by cold punch biopsies and fixed in either Carnoy's solution (100% ethanol: chloroform: gracial acetic acid = 6:3:1) for 2 hours (31 specimens) or 10% formalin solution for 12 hours (20 specimens), then routinely embedded in paraffin. Serial paraffin sections measuring 4 \mu m thick were cut and one stained with H&E for histopathological evaluation. The tumors consisted of 44 transitional cell carcinomas (G1, 15 cases; G2, 17 cases; and G3, 12 cases) and 7 squamous cell carcinomas as summarized in Table 1.

Histopathological classification of the bladder tumors was made according to the "General Rules for Clinical and Pathological Studies on Bladder Cancer".

\textbf{Immunohistochemical methods}

The lectins studied were ConA, SBA, LTA, DBA, PNA, RCA\textsubscript{1}, UEA-I, UEA-II, WGA, and PEA. A key to these abbreviations is given in Table 2, where their origins and specificities are also specified. Deparaffinized sections were treated as follows (using the horseradish peroxidase conjugated lectin staining method\textsuperscript{17,18}):

1. Rinsed in phosphate buffered saline (PBS) for 10 min;
2. Immersed in 0.3% \textsubscript{3}HO\textsubscript{2}/methanol solution for 30 min;
3. Rinsed in PBS for 15 min;
4. Allowed to react with horseradish peroxidase-conjugated lectins for 30 min;
5. Rinsed in PBS for 5 min;
6. Immersed in 0.03% 3,3'-diaminobenzidine tetrahydrochloride/\textsubscript{3}HO\textsubscript{2} solution for 10 min; and

\begin{table}
\centering
\caption{Histology and fixation of 51 bladder cancers}
\begin{tabular}{|c|c|c|c|}
\hline
Fixation & TCC & SCC \\
\hline
Formalin & 6 & 5 & 3 & 6 \\
Carnoy & 9 & 12 & 9 & 1 \\
Total & 15 & 17 & 12 & 7 \\
\hline
\end{tabular}
\end{table}

\begin{table}
\centering
\caption{Lectins used in this study and their sugar specificities}
\begin{tabular}{|l|l|l|l|}
\hline
Lectin & Abbreviation & Origin & Sugar specificity \\
\hline
Concanavalin A & Con A & Jack bean & \textalpha-D-Man, \textalpha-D-Glc \\
Glycine maximum & SBA & Soybean & D-GalNAc, D-Gal \\
Lotus tetragonolobus A & LTA & Asparagus pea & \textalpha-L-Fuc \\
Dolichos biflorus & DBA & Horse gram & \textalpha-D-GalNAc \\
Arachis hypogaea & PNA & Peanut & \textbeta-D-Gal(1-3)-D-GalNAc, \textbeta-D-Gal \\
Ricinus communis & RCA\textsubscript{1} & Castor bean & \textbeta-D-Gal \\
Ulex europaeus agglutinin I & UEA- I & Gorse & L-Fuc \\
Ulex europaeus agglutinin II & UEA- II & Gorse & (D-GlcNAc)\textsubscript{2} \\
Triticum vulgaris & WGA & Wheat germ & (D-GlcNAc)\textsubscript{2} Sialic acid \\
 Pisum sativum & PEA & Pea & \textalpha-D-Man, \textalpha-D-Glc \\
\hline
\end{tabular}
\end{table}
Table 3. Results for immunohistochemical staining of various lectins in human bladder cancers: Carnoy's fixation

<table>
<thead>
<tr>
<th>Lectin</th>
<th>TCC</th>
<th>SCC</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>G1</td>
<td>G2</td>
</tr>
<tr>
<td></td>
<td>-</td>
<td>±</td>
</tr>
<tr>
<td>Con</td>
<td>0 2 5 2 +</td>
<td>0 0 5 7 2+</td>
</tr>
<tr>
<td>UEA-1</td>
<td>0 3 4 2 +</td>
<td>0 3 2 7 2+</td>
</tr>
<tr>
<td>WGA</td>
<td>0 4 5 0 +</td>
<td>0 0 5 7 2+</td>
</tr>
<tr>
<td>UEA-II</td>
<td>6 2 1 0 -</td>
<td>0 7 5 0 ±</td>
</tr>
<tr>
<td>PEA</td>
<td>1 2 5 1 +</td>
<td>0 4 5 3 +</td>
</tr>
<tr>
<td>DBA</td>
<td>1 1 5 2 +</td>
<td>1 5 6 0 +</td>
</tr>
<tr>
<td>SBA</td>
<td>2 5 2 0 ±</td>
<td>1 3 6 2 +</td>
</tr>
<tr>
<td>RCA</td>
<td>1 4 3 1 ±</td>
<td>0 2 6 4 +</td>
</tr>
<tr>
<td>LTA</td>
<td>8 1 0 0 -</td>
<td>1 1 0 0 -</td>
</tr>
<tr>
<td>PNA</td>
<td>7 1 0 1 -</td>
<td>1 1 0 1 -</td>
</tr>
</tbody>
</table>

*) - , negative; ±, weakly positive; +, moderately positive; 2+, strongly positive.

Fig. 1. Grade 1 transitional cell carcinoma of a human bladder (Carnoy’s fixation). A: H.E. ×100. B: Immunohistochemical staining with Ulex europaeus agglutinin I demonstrating a moderately positive reaction (×200)

Fig. 2. Grade 3 transitional cell carcinoma of a human bladder (Carnoy’s fixation). A: H.E. ×200. B: Immunohistochemical staining with Ulex europaeus agglutinin I demonstrating a strongly positive reaction (×200)

7) Rinsed in PBS for 15 min.

Tumor reactions were classified into four different categories; (-): negative, (±): weakly positive, (+): moderately positive, (2+): strongly positive, and averaged over each grade. These grades were
Table 4. Results for immunohistochemical staining of various lectins in human bladder cancers: formalin fixation

<table>
<thead>
<tr>
<th>Lectin</th>
<th>G1</th>
<th>G2</th>
<th>G3</th>
<th>SCC</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>± 2+ Average</td>
<td>± 2+ Average</td>
<td>± 2+ Average</td>
<td>± 2+ Average</td>
</tr>
<tr>
<td>Con A</td>
<td>0 0 2 4 2+</td>
<td>0 0 2 3 2+</td>
<td>0 0 3 2+</td>
<td>0 2 4 2+</td>
</tr>
<tr>
<td>UEA-I</td>
<td>0 0 4 2 +</td>
<td>0 1 2 2 +</td>
<td>0 0 2 1 +</td>
<td>0 0 6 2+</td>
</tr>
<tr>
<td>WGA</td>
<td>0 6 0 0 ±</td>
<td>0 5 0 0 ±</td>
<td>0 2 1 0 ±</td>
<td>0 2 4 2+</td>
</tr>
<tr>
<td>UEA-II</td>
<td>0 0 2 4 2+</td>
<td>0 1 1 3 2+</td>
<td>0 1 0 2 2+</td>
<td>0 3 3 2+</td>
</tr>
<tr>
<td>PEA</td>
<td>0 0 2 4 2+</td>
<td>0 0 2 3 2+</td>
<td>0 0 0 3 2+</td>
<td>5 0 1 0</td>
</tr>
<tr>
<td>DBA</td>
<td>0 4 2 0 ±</td>
<td>0 4 1 0 ±</td>
<td>0 3 0 0 ±</td>
<td>0 3 3 2+</td>
</tr>
<tr>
<td>SBA</td>
<td>0 0 4 2 +</td>
<td>0 2 4 0 +</td>
<td>0 1 1 1 +</td>
<td>0 0 6 2+</td>
</tr>
<tr>
<td>RCA1</td>
<td>0 2 3 1 +</td>
<td>0 0 4 1 +</td>
<td>0 0 3 2+</td>
<td>0 4 2  +</td>
</tr>
<tr>
<td>LTA</td>
<td>1 4 1 0 ±</td>
<td>0 5 0 0 ±</td>
<td>0 2 1 0 ±</td>
<td>4 1 1 0</td>
</tr>
<tr>
<td>PNA</td>
<td>0 1 3 2 +</td>
<td>0 1 4 1 +</td>
<td>1 0 2 0 +</td>
<td>0 3 3 2+</td>
</tr>
</tbody>
</table>

*) –, negative; ±, weakly positive; +, moderately positive; 2+, strongly positive.

**RESULTS**

1. Carnoy's fixation (Table 3)

Transitional cell carcinoma (TCC): Both LTA and PNA gave average negative values in all grades of TCC. The other lectins demonstrated the tendency of differences in intensity between grades, with ConA, UEA-I, and WGA giving moderately positive reactions in G1 (Fig. 1), and strongly positive reactions in G2 and G3 (Fig. 2). UEA-II was mainly negative in G1, but tended to increase the reactivity to moderate in G3. PEA and DBA tended to give a moderately positive reaction in G1 and G2, but mainly a weak or negative reaction in G3. SBA and RCA1 tended to react either weakly or moderately in TCC.

Squamous cell carcinoma (SCC): PEA was negative in SCC, while LTA and PNA reacted moderately. All other lectins gave strongly positive reactions.

2. Formalin fixation (Table 4)

TCC: Almost all TCC gave positive reactions for TCC. WGA, DBA, and LTA had the tendency of weak reaction in all grades. UEA-I, SBA, PNA had the
tendency to react moderately, and ConA, UEA-II, and PEA tended to react strongly with all TCC. RCAI tended to show a moderately positive reaction in G1 and G2, and a strongly positive reaction in G3. Compared with Carnoy’s fixation, lectin staining intensities did not vary between grades.

SCC: PEA and LTA were mainly negative for SCC, and RCAI tended to show a moderately positive reaction. All other lectins tended to react strongly (Fig. 3).

**DISCUSSION**

The present findings suggest that the degree of staining of ConA, UEA-I, and WGA tended to differ between G1, G2, and G3 in TCC, at least when fixed in Carnoy’s, with more advanced lesions demonstrating a more intense reaction. With formalin fixation, however, there were no differences between the three different grades for these lectins. ConA tended to be stained strongly, UEA-I moderately, and WGA weakly in all the TCC. Similarly, in the UEA-II case, the degree of staining tended to increase gradually from negative to moderate according to the grade of TCC in Carnoy fixed tissues, whereas all TCC tended to react strongly after formalin fixation. DBA tended to react moderately with TCC G1 and G2, but was mainly negative for G3 in the case of Carnoy’s fixation. But mainly demonstrated a homogenous weak binding to all TCC fixed in formalin. Thus, the tumor staining patterns between Carnoy’s and formalin fixation differed considerably.

In their formalin fixation studies of human bladder carcinomas, Lehman et al. found that PNA binding was decreased in TCC G312). The discrepancy with the present results again suggests the importance of fixation. Similar to our cases, Sumiya et al. reported that low grade tumors were DBA-positive and high grade tumors were ConA-positive13). By contrast, Langkide et al. mentioned that a gradual loss of PNA and WGA binding correlated with higher grades of atypia. They also described the loss of WGA binding to correlate significantly with both tumor aneuploidy and invasive characters14). In another study they demonstrated a marked decrease of binding of DBA, WGA, PNA, and UEA-I in invasive carcinomas15). However, Yagi reported PNA a high binding rate in high grade tumors, while the DBA binding rate was low10). An explanation for the variation in findings is awaited.

ConA agglutinability has been shown in animal experiments to characterize rat bladder cells following exposure to carcinogens19-23). Fukushima et al. reported that PNA, SBA, and WGA bound strongly to rat TCC induced by N-butyl-N-(4-hydroxybutyl) nitrosamine (BBN), while PNA and SBA stained even SCC or preneoplastic lesions24). Using a model of rat bladder cancer development induced by intravesical instillation of N-nitroso-N-methyl-urea, Langkilde et al. reported that PNA and WGA staining patterns changed from the cytoplasm to the cell membrane during progression to carcinoma25). On the other hand, Takai et al. reported that the location of ConA, WGA, RCA, and PNA binding changed from the cell membrane to the cytoplasm during the development of rat bladder carcinomas induced by BBN23). Thus the two reports gave opposing results.

Nevertheless, both clinical and animal studies have demonstrated many differences in lectin binding patterns. Of major interest, is the correspondence to our findings: the positive ConA in bladder carcinomas, and the decrease of reactivity of DBA in high grade carcinomas. Using this specificity of ConA, Akaza et al. made a bleomycin/concanavalin-A conjugate, and found it promising as an intravesical chemotherapeutic agent against an experimental bladder tumor26).

Neal et al. demonstrated differences in lectin binding patterns between cryostat sections and formalin-fixed tissues of human bladder carcinomas11), with a significant diminution of staining being apparent after formalin fixation. Abel et al. also mentioned a reduction of lectin staining after formalin fixation in cases of benign prostatic hypertrophy6). Using human bronchial mucosa, Mazzuca et al. compared three different fixations; 4% parafor-
We did not examine the same tissues by and found Carnoy's solution to give the method used for fixation would have to confirm our findings. The method of Neal et al. most reliable results and DBA staining might be usefully applied to Carnoy's fixed tissue. However, since we did not examine the same tissues by both fixation methods, further investigations to allow direct comparisons, as with the method of Neal et al. are planned to confirm our findings. Optimization of the method used for fixation would have tangible benefits.

REFERENCES


