Development of Effective Removal Methods of PFCs (Perfluorinated Compounds) in Water by Adsorption and Coagulation

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Development of Effective Removal Methods of PFCs (Perfluorinated Compounds) in Water by Adsorption and Coagulation

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Abstract

Perfluorinated compounds (PFCs) have been used for decades to make products that resist heat, oil, stains, grease and water. In the past, PFCs were not regulated. Common uses include nonstick cookware, stain-resistant carpets and fabrics, components of fire-fighting foam, industrial applications, coatings for packaging, such as milk cartons, cosmetic additives, and other personal products. Huge amount of PFCs have been already released into the environment, which have been trickled down up to the glaziers in Antarctica. The chemical structure of PFCs makes them extremely resistant to breakdown in the environment.

Although PFCs have been used for decades, limited studies on human health effects have been carried out. In animal studies, high concentrations PFCs have shown adverse effects on the liver and other organs.

It is a reported fact by researchers that the conventional water and wastewater treatment facilities cannot eliminate PFCs. Many studies have showed a positive correlation between the PFOS concentration in raw water and tap water samples suggesting minimum removal efficiency of conventional water purification systems. The main reason for the persistancy of PFCs is C-F bonds.

The C-F bond length is shorter than any other carbon–halogen bonds, and shorter than the C-N and the C-O bonds. The high electro-negativity of fluorine gives the carbon–fluorine bond a significant polarity/dipole moment.

The overall objective of this research was to investigate on possible techniques to eliminate PFCs in water. Since PFCs are negatively charged by its atomic structure, the strategy was to eliminate them by adsorption process. In this experiment, seven granular materials and nine coagulants were studied in detail for PFCs eliminations.

Six kind of synthetic resins and GAC were tested with a series of batch experiments for PFCs adsorption (Chapter 4). The batch experiment for kinetic and isotherm characteristics of each material was carried out in a shaker with 100 hrs shaking duration. Out of seven materials five were first time tested for PFCs. Faster kinetic characteristics were observed for GAC and ion exchange resins than non ion exchange resins. Considering the adsorption isotherms, synthetic resins were identified as better filter materials (in terms of adsorption capacity) to eliminate PFCs in water at a low concentration (1 µg/L).
The magnitude of Freundlich isotherm constants \((K_f)\) decreases in the following order for most of the long chain and the medium chain PFCs tested: Ion-exchange polymers > Non ion-exchange polymers > GAC, sometimes at a further low equilibrium concentration (100 ng/L) non ion-exchange polymers showed higher adsorption capacity than other adsorbents. Amb IRA- 400 was identified as the best filter material to eliminate PFOS at equilibrium concentrations of >1 \(\mu\)g/L. Considering both adsorption isotherms and adsorption kinetics, Amb XAD 4 and Dow MarathonA were best candidate materials for eliminating PFOS at ng/L equilibrium concentrations. From the results for kinetic experiments, chemisorption was identified as the main attaching process in PFCs adsorption.

The process of coagulation was studied by a series of jar tests for PFCs elimination in this research (Chapter 5). There is limited published data on PFCs coagulation, especially by organic coagulants. Six long chain cationic coagulants were tested for anionic PFCs coagulation and the results were compared with three conventional inorganic coagulants. The results of the experiments with deionized water and wastewater spiked with PFCs showed that the PFCs coagulation by organic coagulants is double than that of inorganic coagulants. Among organic coagulants FL 2749 was identified as the best candidate material to eliminate PFCs. Jar test results with actual PFCs related industrial wastewater indicated that organic coagulants are not effective (as it showed in wastewater spiked with PFCs) to coagulate PFCs. The PFCs that appear in real wastewater seems to be incorporated with other polar molecules in wastewater. Organic coagulation followed by microfiltration was identified as an effective combination to eliminate PFCs for some wastewaters, but further studies have to be done on this topic.

The results of batch experiments (chapter 4 and 5) were further consolidated by the long run continuous experiments (60 days and 130 days). Non ion exchange polymers were further studied with a column experiment (Chapter 6) (60 days continuous run) and Amb XAD 4 was recognized as the best candidate among the tested four filter materials to eliminate PFOS. Amb XAD 4 removed 99.99% PFOS up to 23,000 bed volumes pass through with the condition of 10 \(\mu\)g/L inflow concentration and flow rate of 15 ml/min (0.75 bed volumes/min). At the end of the column test, PFOS adsorbed granular materials were used to examine the material regeneratability with an organic solvent. Within 80 min, all most 100% PFOS was recovered from synthetic resins suggesting that
synthetic polymers can be effectively regenerated by an organic solvent.

A long run continuous experiment (130 days) showed the combined treatment process of coagulation (by organic coagulants) followed by adsorption and filtration is an excellent method to treat PFCs in water. The combination with some adsorbents, more than 99% removal was obtained even after 100 days of continuous run. Economical analysis with different scenarios for the level of treatment and regenerations indicated that AmbXAD 4 was the cheapest option for PFOS adsorption.

This experiment was limited for six synthetic resins and six long chain cationic organic coagulants. The results highly recommend to repeat the same experiments with more adsorbants and coagulants for material optimization. The materials tested in this experiment were limited for lab scale models and the same materials to be tested in the field. As the first step a filter column with Amb XAD4 can be installed at the discharge point of a wastewater treatment plant in a PFCs related industry.

**Keyword**

Perfluorinated compounds (PFCs), industrial wastewater, adsorption, coagulation, synthetic resins, organic coagulants
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<td>AC</td>
<td>activated carbon</td>
<td>PFBuS</td>
<td>perfluorobutane sulfonate</td>
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<td>aqueous fire fighting foam</td>
<td>PFCA</td>
<td>perfluorocarboxylates</td>
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<td>electrospray ionization</td>
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<td>perfluoropentanoic acid</td>
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<td>Calgon filtrasorb 400</td>
<td>PFTeDA</td>
<td>perfluorotetradecanoic acid</td>
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<td>GAC</td>
<td>granular activated carbon</td>
<td>PFUnA</td>
<td>perfluoroundecanoic acid</td>
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<td>LC</td>
<td>liquid chromatograph</td>
<td>POPs</td>
<td>persisted organic pollutants</td>
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<td>MS/MS</td>
<td>tandem mass spectrometer</td>
<td>RE</td>
<td>removal efficiency</td>
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<td>MTBE</td>
<td>methyl t-butyl ester</td>
<td>RO</td>
<td>reverse osmosis</td>
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<tr>
<td>PBT</td>
<td>Persistence, bioaccumulation and toxicity</td>
<td>SF</td>
<td>sand filtration</td>
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<td>pore diffusion</td>
<td>UV</td>
<td>ultra violet</td>
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<td>perfluoroalkyl sulfonates</td>
<td>WWTP</td>
<td>wastewater treatment plant</td>
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<td>PFBA</td>
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Chapter 1  Introduction

1.1 Introduction

Industrialization and technological development processes have led to the introduction of new hazardous chemicals into the water environment. Most of these chemicals are synthesized with unique characteristics for a specific application, but ultimately those contribute to pollute water environment. Perfluorinated compounds (PFCs) are such tracer level emerging water pollutants, which are synthesized by replacing all C-H bonds of a hydrocarbon by C-F bond. The C-F bond is the strongest bond in organic chemistry and fluorine always prefers to have -1 oxidation state. The ultimate result of replacing all C-H bonds by C-F bond in the synthesis process is a PFCs molecule with polarity, (negatively charged) which is strong, difficult to oxidize and sustainable in the environment (O'Hagan., 2008).

The exact toxicity of each PFCs for human being is still unclear. Some researchers have reported adverse health effects of some PFCs. Perfluorooctane Sulfonate (PFOS) were recently categorized as a POP by the Stockholm convention (Earth 2009).

On the other hand, development in instrumental chemistry has facilitated the measurement of new pollutants more accurately at low concentrations. Both tap water and discharge industrial wastewater quality standards are expanding into wider spectrum of parameters. As emerging persistent toxic pollutants, PFCs are now discussed to include drinking water quality parameters and researchers are working on how to treat them at water and wastewater purification plants.

Researchers have already identified that conventional tap water and wastewater purification plants could not remove PFCs (Fujii et al, 2007 and Takagi et al, 2008). Previous studies have identified adsorption as an effective process to eliminate PFCs from water. Different kind of granular activated carbon (GAC), some ion-exchange resins, high silica zeolite, and anaerobic sludge have been tested to evaluate the PFCs adsorption capacity (Qiang et al. 2009: Ochoa-Herrera et al. 2008). In general, PFCs traumaent research has been limited for unrealistic PFCs concentrations at mg/L levels.

The overall objective of this research was to investigate on possible PFCs treatment techniques. The research was limited for wastewater as it was identified as the main PFCs polluter. Figure 1.1 shows the flow of PFCs through different treatment processes and this research studied some physico-chemical treatment processes for PFCs elimination. The main
objectives of this study can be summarized as:

**1.2 Research objectives**

1. To study the oxidation characteristics of PFCs by ferrate techniques.
2. To determine the PFCs adsorption characteristics of ion-exchange polymers, non ion-exchange polymers and activated carbon while establishing a methodology for the batch experiment.
3. To simulate the real application of selected polymers by a long run column experiment to adsorb PFCs.
4. To identify possible candidate coagulants to eliminate PFCs for wastewater treatment process.
5. To examine the improvement of adsorption process by prior coagulation with organic coagulants.

A main part of this study was dedicated to investigate the PFCs adsorption characteristics of non ion-exchange polymers, ion-exchange polymers and granular activated carbon at low concentrations. Seven synthetic polymer materials and one GAC material were investigated in this study. Both batch experiments (with shaking time of 100 hrs) and column experiments (operation time of 100 days) were conducted to determine sorption isotherms, sorption kinetics and the applicability in the field. Out of the eight materials investigated in this study, six materials were tested for the first time for PFCs and the experiment conditions were set such that it simulates real ground situation with low PFCs concentrations. Since the PFCs are negatively charged, long chain polymeric cationic coagulants were tested and results were compared with the conventional inorganic coagulants for PFCs elimination. The individual positive results obtained for adsorption and coagulation were combined at the latter part of the study and the effect on adsorption by coagulation was observed.

**1.3 Dissertation framework**

This dissertation consists of eight chapters. Overall content of the thesis is schematically shown in **Figure 1.2**.

Chapter 1 gives a brief introduction of this research, including research background, objectives and dissertation structures.
Chapter 2 reviews current available literature on PFCs, mainly recent developments on treatment of PFCs and management strategies to control PFCs.

Chapter 3 describes ferrate oxidation of PFCs. PFCs oxidation by conventional oxidants in water and wastewater treatment is already known. Ferrate has been identified as an emerging oxidative agent to apply in water and wastewater purification industry with highest redox potential among all conventional oxidants. This study was conducted as there is no prior data available for PFCs oxidation by ferrate.

Chapter 4 shows the characteristics of PFCs adsorption onto polymers. Batch experiments were conducted to obtain adsorption isotherms and kinetics for eight granular materials including ion-exchange polymers, non ion-exchange polymers and GAC. Published data on the six polymers tested is very limited. Freundlich equation and pseudo second-order model were successfully applied to explain isotherm and kinetic data.

Chapter 5 shows coagulation of PFCs. A comparative study of polymer type organic coagulant and conventional inorganic coagulants was carried out in a series of Jar tests. Six organic coagulants and three conventional inorganic coagulants were used in the study. Published data on PFCs coagulation by organic coagulants is rarely available.

Chapter six is dedicated for the column experiment with non ion-exchange polymers. The polymers used in this experiment were selected by the results of Chapter 5. A set of columns with four non ion-exchange granular materials were continuously run for sixty days. After the experiment, the same materials in the columns were used to determine the PFOS regeneratability by organic solvents.

Chapter 7 explains combine treatment of PFCs (Adsorption + Coagulation). A treatment process of coagulation followed by adsorption was simulated by a lab scale model and it was continuously run for 130 days. The combined performance was compared with the individual performances as obtained in Chapter 5 and Chapter 6. Later part of the chapter is reserved for economical analysis for synthetic polymers and GAC.

Chapter 8 gives conclusions of this study and recommendations for further research.
Degradation
Sonication
UV, VUV
irradiation
Sonochemical degradation

PFCs and its precursors producing industries
PFCs and its precursors consuming industries
Product consumers

Air/ water environment

Treatment for PFCs

△ Biological treatment process

Physical and Chemical process

Basic unit operation
- Screening
- Clarification
- Sedimentation
- Filtration
△ Membrane processes

Coagulation
- Role of floc formation and growth
- Materials optimization
- Applications

Adsorption
- Biological effects
△ Materials optimization
- Materials regeneration
- Applications

Oxidation
- Chlorine
△ Ozone
△ Hydrogen peroxide
△ Super critical oxidation
- Ferrate

Degradation
- Sonication
△ UV, VUV irradiation
△ Sonochemical degradation

Combined treatment process
- Combined physico-chemical processes
- Combined biological processes
- Combined biological physico-chemical processes

△ Studied by our research group
△ Studied by other research groups
- Covered by this study

Fig. 1.1 Scope of the study

Water environment
Chapter 1
Introduction

Chapter 2
Literature review
(Latest development in PFCs treatment)

Chapter 3
Ferrate oxidation of PFCs

Chapter 4
Adsorption of PFCs
Isotherm and kinetic study for eight granular materials including ion-exchange polymers, non ion-exchange polymers and GAC

Chapter 5
Coagulation of PFCs
Comparative study of polymer type organic coagulant and conventional inorganic coagulants

Chapter 6
Column test for selected polymers
1. Column test for non ion-exchange polymers (60 days continues run)
2. Material regeneration by organic solvent

Chapter 7
Combine treatment of PFCs
(Adsorption + Coagulation)
1. Column test for non ion-exchange polymers (130 days continues run)
2. Effect of TOC
3. Economical analysis

Chapter 8
Conclusion and recommendation

Fig. 1.2 Structure of the thesis
Chapter 2  Literature survey

2.1 Introduction
The entire story in this study begins with the carbon–fluorine bond which is a common component of all PFCs. This bond is the strongest bond in organic chemistry. The length of the carbon–fluorine bond is typically about 1.35 angstrom (1.39 Å in fluoromethane), which is shorter than any other carbon–halogen bonds, and shorter than carbon–nitrogen and carbon–oxygen bonds. The short length of the bond can also be attributed to the ionic or electrostatic attractions between the partial charges on carbon and fluorine. The bond also strengthens and shortens as more fluorines are added to the identical carbon on a chemical compound. The high electronegativity of fluorine (4.0 for F vs. 2.5 for C) gives the carbon–fluorine bond a significant polarity/dipole moment. The electron density is concentrated around the fluorine, leaving the carbon relatively electron poor. This introduces ionic character to the bond through partial charges (C\(\delta^+\)—F\(\delta^-\)). The partial charges on the fluorine and carbon are attractive, contributing to the unusual bond strength of the carbon–fluorine bond (O'Hagan, 2008). Carbon–fluorine bonds can have a bond dissociation energy (BDE) of up to 130 kcal/mol (Lemal, 2004). The BDE is higher than other carbon–halogen and carbon–hydrogen bonds. For example, the molecule represented by CH\(_3\)X has a BDE of 115 kcal/mol for carbon–fluorine while values of 104.9, 83.7, 72.1, and 57.6 kcal/mol represent carbon–X bonds to hydrogen, chlorine, bromine, and iodine, respectively (Blanksby and Ellison, 2003). The ultimate result of all the characteristics is a strong polar organic compound of PFCs, which is persistent in environment and toxic to the living beings.

2.2 Manmade fluorinated compounds
There are about 30 natural organofluorine molecules in the environment (Hekster et al., 2003). These molecules contain only one fluorine atom per molecule. In contrast, man-made fluorinated organic compounds often contain many fluorine atoms, thus they are called polyfluorinated (Key et al. 1997). In perfluorinated compounds (PFCs), all the carbon–hydrogen bonds are replaced with carbon–fluorine bonds (Renner, 2001) except one bonding site at the end of chain. PFCs are composed of a carbon–fluorine chain and generally have side moieties attached such as carboxylic acids or sulfonic acids (Giesy and Kannan 2002). The strong carbon–fluorine bond in PFCs gives thermal and chemical stability to many PFCs (So et al., 2004). The stability that makes fluorinated
compounds desirable for commercial use also makes them potentially significant environmental contaminants due to their resistance to natural breakdown processes, that is, their persistence (Key et al., 1997). Research over the past few years has shown that the PFCs are now widespread environmental contaminants and found in living organisms globally.

2.3 Applications of PFCs
PFCs are manufactured because of their specific physical and chemical properties such as chemical and thermal inertness and special surface-active properties (Hekster et al., 2003). They repel both water and oil and act as surfactants, that is, they reduce surface tension and do so better than other surfactants. These properties have led to the use of perfluorinated compounds in a wide variety of applications. For instance, their unique properties of repelling both water and oil has led to their use as coatings for carpet protection, textile protection, leather protection, and paper and board protection. They are also used in firefighting foams and as polymerisation aids. In addition they are used as speciality surfactants, for example, in cosmetics, electronics, etching, medical use and plastics (Hekster et al., 2003, So et al., 2004. They are also widely employed in different industrial processes such as wiring insulation for telecommunications, aerospace, electronics (semiconductors), and medical use (Martin et al., 2004b; Prevedouros et al., 2006). In particular, PFOA is used as adjuvant in the production of fluoropolymers such as PTFE, Teflon or similar products, and occurs in these applications as aqueous and gaseous process emission (Fricke and Lahl, 2005: Davis et al., 2007). Point source manufacturing facilities of fluorochemicals are one of the largest sources of emissions for PFCs (Prevedouros et al., 2006; Davis et al., 2007).

2.4 Occurrence of PFCs
PFCs, especially PFOS and PFOA, have been found in surface waters in Japan (Saito et al., 2004), USA (Hansen et al., 2002: Boulanger et al., 2004, 2005), and Europe (Skutlarek et al., 2006; Loos et al., 2007; McLachlan et al., 2007). PFOA has been found in relatively high concentration levels near fluorochemical manufacturing facilities in USA, (500 ng/L) in the Tennessee River (Hansen et al., 2002), in the Ohio River (Davis et al., 2007), and in Germany up to 56ng /L in the River Alz (near a fluorochemical factory). In addition, it has been found in drinking waters, up to 519 ng/L in Germany (Schaefer, 2006: Skutlarek et al.,
2006) and in ground water near the Ohio River, USA (Davis et al., 2007). Moreover, PFOS and PFOA have also been analyzed in ocean waters with detected concentrations up to 100 pg /L and marine mammals (Kannan et al., 2002; Yamashita et al., 2004, Haukas et al., 2007). An OECD hazard assessment from the year 2002 (OECD, 2002) identified PFOS as a PBT-chemical (persistent, bioaccumulative, toxic). In December 2006, the European Parliament and the Council decided to restrict marketing and use of PFOS with a few exceptions by amending Council Directive 76/769/EC on dangerous substances for PFOS (European Commission, 2006). It is currently being discussed if PFOA should be included in this Directive. In addition, it is investigated to introduce PFOS and PFOA as so-called priority substances regulated by the Water Framework Directive (WFD).

2.5 Properties of PFCs

2.5.1 Persistence

3M Company published its reports on hydrolysis and aqueous photolytic degradation (Hatfield, 2001) of PFOA, which showed rather long half-life times in natural environment. PFOS also showed its resistance to advanced oxidation processes including ozone, ozone/UV, ozone/H₂O₂ and Fenton reagent (Schröder and Meesters, 2005). Also it has been reported that almost all PFCs had no response to oxidation by chromium potassium oxide (Cr₂O₇²⁻) or potassium permanganate acid (MnO₄⁻), which confirmed their stabilities in critic environment.

2.5.2 Bioaccumulation

K_{ow}, known as partition coefficient between octane and water phases, is useful to estimate bioaccumulation potentials. However, it is not available for most of PFCs because PFCs are surfactants and third layer is formed during measurement (OECD, 2002; US EPA, 2002). Bioaccumulation factors (BAFs) or bioconcentration factors (BCFs) are calculated by dividing the average concentrations in organism by the concentrations in water environment. BAFs represent accumulation potentials of organics from environment to organisms.

Previous researchers have reported that their preliminary study showed dietary BAFs of PFOS were 2,796 in bluegill sunfish and 720 in carp (OECD,
respectively. BAFs of PFOA were about 2 in fathead minnow and 3~8 in carp, which are quite lower than PFOS. First survey of PFOS in Japan showed quite high BAFs of PFOS to fishes in Tokyo Bay, which were as high as 1,260~19,950 (Taniyasu, et al., 2002). Survey in Great Lakes has also shown that BAFs of PFOS is about 1,000 for benthic invertebrates (Kannan, et al., 2005). Another research in Ai river of Japan showed rather high BAF of PFOS in wild turtles as 10,964 and very low BAF of PFOA equal to 3.2. Similar situation occurred for rainbow trout under synthetic PFCs spiking waters. Dietary BAFs of PFCAs have ranged from 0.038 to 1.0, while BAFs of PFASs have ranged from 4 to 23,000, both increased with length of carbon chain (Martin, et al., 2003). The research on lake trout in Great Lakes in North America also showed similar trend of increasing BAFs by carbon chain length. BAFs of PFCA showed linear increase from 3.2 to 3.9, while BAFs of PFAS changed from 500 to 12,600 (Furdui, et al., 2007). Increase of BAFs by ascendant PFCs carbon chain length indicated that long-chained PFCs or more hydrophobic PFCs had stronger bioaccumulative potentials and bioconcentration levels. Furthermore, BAFs of PFASs were greater than PFCAs in equivalent carbon chain length, which indicated that acid function groups also played important role to determine accumulation potentials.

Biomagnification factors (BMFs) are calculated by dividing the average concentrations in predators to those in preys. BMFs can be applied to estimate accumulation potentials of organics in food chains. One study in Great Lakes reported BMF of PFOS to be 10~20 for mink or bald eagles to their prey items (Kannan, et al., 2005). Another survey of US coastal marine food chain showed that BMFs of bottlenose dolphin are 0.1-13.0 for PFCA and 0.8~14 for PFAS, which were also increased by ascendant carbon chain length (Houde, et al., 2006). It can be concluded that BMFs of PFCs were around 10 in aquatic ecosystems.

### 2.5.3 Toxicity

Since PFCs differ from normal lipophilic toxic substances, their toxicokinetics is still unknown in mechanism and need more studies to elucidate the profile. Although a lot of researches have been conducted on the toxicity of PFCs, the results were in diverse qualities. Generally, medium and long-chained PFCs may be more toxic than short-chained ones by their longer
half life in rodents. Review on PFOS toxicity showed that subchronic exposure led to significant weight loss accompanied by hepatotoxicity, and reductions of serum cholesterol and thyroid hormones (OECD, 2002). Reviews also revealed the influence of PFOS on generations of laboratory rodents (Lau, et al., 2003; Thibodeaux, et al., 2003) and on human birth in Japan (Inoue, et al., 2004a).

US EPA reviewed studies on toxicity of PFOA, which showed diverse behavior between gender and generations of laboratory animals. Average half life of PFOA in human body was estimated to be 4.37 years (US EPA, 2003). PFOA behaved in different ways between species and sexes due to renal clearance, and undergoes enterohepatic circulation. However, it was also reported that PFOA can be eliminated and detoxified in the human body (Kudo and Kawashima, 2003). Pharmacokinetic profiles of PFOA in the adults may be different from those in sexually immature rats (Lau, et al., 2004).

Studies on toxicity of PFCs other than C8 were still very few. PFBS has much shorter half-life time than PFOS, therefore seems less toxic. PFHxS was detected in human serum in similar level of PFOA and 10 times less than PFOS (verbal communication with US-EPA). Both PFNA and PFDA were peroxisome proliferators, and PFDA has longer half-life time in rats and shows quite high toxic potency (Lange, et al., 2006). Pharmacokinetic and structure-activity studies on the relationship between toxicity and carbon chain length would arouse interests of toxicologists in future.

2.5.4 Physiochemical properties of PFCs
PFCs are either in liquid form or in solid form depending on the chain length. They are heavier than water and miscible with most organic solvents (ethanol, acetonitrile, methanol). They have relatively low solubility in water, and water has a very low solubility in them. The number of carbon atoms in a PFCs molecule largely determines most physical properties. The greater the number of carbon atoms, the higher the boiling point, density, viscosity, surface tension, critical properties, vapor pressure and refractive index. Some physical properties of PFCs considered for this study is shown in Table 2.1 and 2.2.
Table 2.1 Basic information of PFCs considered in this study

<table>
<thead>
<tr>
<th>Abbr. name</th>
<th>Full name</th>
<th>Molecular Structure</th>
<th>m.w</th>
<th>CAS No.</th>
</tr>
</thead>
<tbody>
<tr>
<td>PFBA</td>
<td>Perfluorobutyric Acid</td>
<td>CF₃(CF₂)₂COOH</td>
<td>214</td>
<td>375-22-4</td>
</tr>
<tr>
<td>PFHxA</td>
<td>Perfluorohexanoic Acid</td>
<td>CF₃(CF₂)₄COOH</td>
<td>314</td>
<td>307-24-4</td>
</tr>
<tr>
<td>PFHpA</td>
<td>Perfluoroheptanoic Acid</td>
<td>CF₃(CF₂)₅COOH</td>
<td>364</td>
<td>375-85-9</td>
</tr>
<tr>
<td>PFOA</td>
<td>Perfluorooctanoic Acid</td>
<td>CF₃(CF₂)₆COOH</td>
<td>414</td>
<td>335-67-1</td>
</tr>
<tr>
<td>PFDA</td>
<td>Perfluorodecanoic Acid</td>
<td>CF₃(CF₂)₈COOH</td>
<td>514</td>
<td>335-76-2</td>
</tr>
<tr>
<td>PFOS</td>
<td>Perfluorooctane Sulfonate</td>
<td>CF₃(CF₂)₇SO₃</td>
<td>500</td>
<td>2795-39-3</td>
</tr>
</tbody>
</table>

Abbr. name – Name abbreviated  m.w – molecular weight

Table 2.2 Basic physiochemical properties of PFCs

<table>
<thead>
<tr>
<th>PFCs</th>
<th>pkₐᵇᵃ</th>
<th>Melting pointᵇ°C</th>
<th>Boiling pointᶜ°C</th>
<th>Specific Gravity</th>
<th>Texture</th>
</tr>
</thead>
<tbody>
<tr>
<td>PFBA</td>
<td></td>
<td>120.8-121.0</td>
<td>1.65</td>
<td>yellow liquid</td>
<td></td>
</tr>
<tr>
<td>PFHxA</td>
<td></td>
<td>159-160</td>
<td>1.76</td>
<td>clear liquid</td>
<td></td>
</tr>
<tr>
<td>PFHpA</td>
<td></td>
<td>175</td>
<td>1.79</td>
<td>crystalline</td>
<td></td>
</tr>
<tr>
<td>PFOA</td>
<td>2.50</td>
<td>55-56, 37-50</td>
<td>189</td>
<td>white powder</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>white powder</td>
<td></td>
</tr>
<tr>
<td>PFDA</td>
<td></td>
<td>83-85, 88</td>
<td>218</td>
<td>white powder</td>
<td></td>
</tr>
<tr>
<td>PFOS</td>
<td>3.27</td>
<td>&gt;400, 277</td>
<td>2.05</td>
<td>white powder</td>
<td></td>
</tr>
</tbody>
</table>

Note: a = PFOA (OECD, 2002), PFOS (US EPA, 2002); b = melting point, data from material safety data sheet (MSDS) of Wako Company and ExFluor Company; c = boiling point, from ExFluor MSDS.
2.5.5 Molecular structure of PFCs

PFCs molecules consist of a perfluorinated carbon chain and one special functional group at a corner of the chain. This functional group is ether carboxylic or sulfonic group (Figure 2.1a,b). Fully fluorinated carbon chain of the PFCs makes it hydrophobic and functional group at the end makes PFCs hydrophilic. Different hydrophilic functional groups show diverse behavior in aqueous environment. The perfluorinated tail is covered by strongest C-F bonds, and surrounded by fluorine atoms which have similar size with the carbon atom.

**Fig 2.1.a** molecular structure PFOS  **Fig 2.1.b** molecular structure PFOA

2.6 PFCs and Stockholm Convention

The Stockholm Convention on Persistent Organic Pollutants (POPs) is a global treaty to protect human health and the environment from chemicals that remain intact in the environment for long periods, become widely distributed geographically and accumulate in the fatty tissue of humans and wildlife. This convention was adopted in 2001 and entered into force in 2004. By signing this convention the parties agree to take measures to eliminate or reduce the release of POPs into the environment. The Convention is administered by the United Nations Environment Programme and is based in Geneva, Switzerland.

A separate committee has been established (POPRC) as a subsidiary body to the Stockholm Convention for reviewing new chemicals. One of the known PFCs of PFOS was reviewed by the 4th POPs review committee and recommended to categorized it as a POPs (http://chm.pops.int/Convention/POPsReviewCommittee/hrPOPRCMeetings/POPRC4/POPRC4ReportandDecisions/tabid/450/language/en-US/Default.aspx).
This recommendation was seriously considered at the fourth meeting of the Conference of the Parties (COP4) to the Stockholm Convention and decided to amend part I of Annex B of the Convention to list perfluorooctane sulfonic acid (PFOS), its salts and perfluorooctane sulfonyl fluoride with the acceptable purposes and specific exemptions (decision SC-4/17).

2.6.1 Acceptable purposes

Eventhough PFOS was categorized as a POPs by the Stockholm convention, provision was given to further use of PFOS for the following applications. Photo-imaging, photo-resist and anti-reflective coatings for semi-conductor, etching agent for compound semi-conductor and ceramic filter, aviation hydraulic fluids, metal plating (hard metal plating) only in closed-loop systems, certain medical devices (such as ethylene tetrafluoroethylene copolymer (ETFE) layers and radio-opaque ETFE production, in-vitro diagnostic medical devices, and CCD colour filters), fire-fighting foam, insect baits for control of leaf-cutting ants from Atta spp. and Acromyrmex spp.

2.6.2 Specific exemptions

A special exemption was given to use PFOS for the following application by Stockholm convention. Photo masks in the semiconductor and liquid crystal display (LCD) industries, metal plating (hard metal plating, decorative plating), electric and electronic parts for some color printers and color copy machines, insecticides for control of red imported fire ant, and termites, chemically driven oil production, carpets, leather and apparel, textiles and upholstery, paper and packaging, coatings and coating additives, rubber and plastics.

2.7 Treatment of PFCs

2.7.1 PFCs removal at conventional water treatment process

Few researchers have already reported a positive correlation between the PFOS concentration in raw water and tap water samples suggesting minimum removal efficiency at conventional water purification systems (Fujii et al, 2007; and Takagi et al, 2008).

Ground water and drinking water around a Teflon manufacturer in Virginia, USA was found to be contaminated with PFOA up to 10 μg/L (ENDS 2004).
Drinking water samples in Japan were contaminated at maximum concentration of 40 ng/L for PFOA in Osaka area (Saito et al. 2004) and at about the same level for PFOS in Tokyo. Both authors indicated that the sources of tap water were from surface water which was contaminated at the same levels of PFOS and PFOA concentration. A study of surface water and tap water in Ruhr (Germany) (Skutlarek et al. 2006) found drinking water contamination up to 519 ng/L for PFOA and 22 ng/L for PFOS. Also there is published data to show that the concentrations found in drinking waters decreased with the concentrations of the corresponding raw water samples along the flow direction of the Ruhr river (from east to west) and were not significantly different from surface water concentrations. This indicates that perfluorinated surfactants are at present not successfully removed by water treatment steps. By measuring concentrations in tap water and surface water in an area of Germany, Skutlarek et al. revailed that perfluorinated surfactants are at present not successfully removed by water treatment steps (Skutlarek et al. 2006).

Dr Lien, a researcher worked with our research team also reported the same observation of similar PFOS levels in tap water and surface water in a given area. She reported that her case study in Kinki Region (Japan) and Istanbul (Turkey) had demonstrated similar levels of both PFOS and PFOA in tap water to those of water supply sources and these results suggested that PFOS and PFOA, at the levels of several ng/L to several tens ng/L were not effectively removed through water treatment steps.

### 2.7.2 Membrane filtration

Tang et al. (2006) investigated the feasibility of using reverse osmosis (RO) membranes for treating semiconductor wastewater containing PFOS. They reported that the RO membranes generally rejected 99% or more of the PFOS (feed concentrations 0.5 - 1500 ppm). Also they found that the rejection was better for tighter membranes, but was not affected by membrane zeta potential. Flux decreased with increasing PFOS concentration.

E. S. Darling and M. Reinhard (2008) measured the rejections of 15 perfluorochemicals (PFCs), five perfluorinated sulfonates, nine perfluorinated carboxylates, and one perfluorooctane sulfonamide (FOSA)s by four nano filtration membranes (NF270, NF200, DK, and DL). They reported that the rejections for anionic species were >95% for MW > 300 g/mol. FOSA (MW 499 g/mol), which is uncharged at the pH of deionized water (5.6), was rejected as
little as 42% (DL membrane). Lowering the pH to less than 3 reduces rejection by up to 35%. An alginate fouling layer increases transmission by factors of 4-8. Based on rejection kinetics and the extent of sorption, they suggested that two different sorption processes are significant: charged species adsorb quickly to the membrane surface, whereas the uncharged species absorbs within the membrane matrix in a much slower process.

2.7.3 PFCs adsorption

In 2007, our research group has investigated (Yong 2007) the adsorption characteristics of PFCs onto GAC by batch experiments and the results had been interpreted by the Freundlich equation and homogenous surface diffusion model (HSDM). Isothermal and kinetics experiments have implied that PFCs adsorption onto GAC was directly related with their carbon chain lengths. By ascending the carbon chain length, adsorption capacity for specific PFC had been increased, and diffusion coefficient ($D_s$) decreased. $D_s$ of GAC adsorption had been decreased gradually in smaller GAC diameters. Coexisted natural organic matters (NOMs) have reduced adsorption capacities by mechanisms of competition and carbon fouling.

It was found that Carbon fouling reduces the adsorption capacity much more intensively than by organics. Acidic bulk solution was slightly helpful for adsorption of PFCs. However adsorption velocity or kinetics was affected by NOM and pH significantly. GAC from Wako Company has showed the best performance among four kinds of GACs, and Filtra 400 from Calgon Company has considered more suitable to removal all PFCs among the commercial GACs. The results implied that background organics broke through fixed GAC bed much earlier than trace level of PFCs. Medium-chained PFCs was found to be effectively removed by fixed bed filtration without concerning biological processes.

Valeria and Reyes (2008) have evaluated the three anionic PFC surfactants, i.e., perfluorooctane sulfonate (PFOS), perfluorooctanoic acid (PFOA) and perfluorobutane sulfonate (PFBS), for the ability to adsorb onto activated carbon. Additionally, the sorptive capacity of zeolites and sludge for PFOS was compared to that of granular activated carbon. They determined the adsorption isotherms at constant ionic strength in a pH 7.2 phosphate buffer at 30°C. They reported that the sorption of PFOS onto activated carbon was stronger than PFOA and PFBS, suggesting that the length of the fluorocarbon chain and the
nature of the functional group influenced sorption of the anionic surfactants. Among all adsorbents evaluated in their study, activated carbon (Freundlich $K_f$ values = 36.7-60.9 [($mg$ PFOS/g sorbent)($mg$ PFAS/l)$^{-n}$]) showed the highest affinity for PFOS at low aqueous equilibrium concentrations, followed by the hydrophobic, high-silica zeolite NaY (Si/Al 80, $K_f$ = 31.8[($mg$ PFOS/g sorbent)($mg$ PFAS/l)$^{-n}$]), and anaerobic sludge ($K_f$ = 0.9-1.8[($mg$ PFOS/g sorbent)($mg$ PFAS/l)$^{-n}$]). Activated carbon also displayed a superior sorptive capacity at high soluble concentrations of the surfactant (up to 80 mg/L). These findings indicate that activated carbon adsorption is a promising treatment technique for the removal of PFOS from dilute aqueous streams.

Qiang Yu et al., (2009) have investigated the feasibility of using powdered activated carbon (PAC), granular activated carbon (GAC) and anion-exchange resin (AI400) to remove PFOS and PFOA from water, with regard to their sorption kinetics and isotherms. They have reported that the adsorbent size greatly influenced the sorption velocity, and both the GAC and AI400 required over 168 hrs to achieve the equilibrium, much longer than 4 hrs for the PAC. They have adopted kinetic models to describe the experimental data, and the pseudo-second-order model well described the sorption of PFOS and PFOA on the three adsorbents. The sorption isotherms have showed that the GAC had the lowest sorption capacity both for PFOS and PFOA among the three adsorbents, while the PAC and AI400 possessed the highest sorption capacity of 1.04 mMol g/L for PFOS and 2.92 mMol g/L for PFOA according to the Langmuir equation. Based on the sorption behaviors and the characteristics of the adsorbents and adsorbates, they have concluded that ion exchange and electrostatic interaction as well as hydrophobic interaction were involved in the sorption, and some hemi-micelles and micelles possibly formed in the intraparticle pores.

But in real scale application it is reported that the usage of activated carbon to ensure the removal of perfluorinated surfactants is not effective. In some German water treatment plants with carbon filters has shown poor performances to eliminate PFCs. (Schaefer et al.,2006). Table 2.3 and Table 2.4 show the summary of studies done to evaluate different materials to eliminate PFCs, mainly PFOS and PFOA.
2.7.4 PFCs oxidation

PFCs are recalcitrant towards oxidation due to the complete substitution of fluorine (C⁻F bond) for hydrogen (C⁻H bond). Fluorine will retain its electrons at any cost. Fluorine is nearly always found in the (-1) valence state with the only exception being F₂ where its oxidation state is (0). The fluorine atom is the most powerful inorganic oxidant known with a reduction potential of 3.6 V (Eq. 1.1) (Wardman, 1989) and thus is thermodynamically unfavorable to create the fluorine atom with any other one-electron oxidant.

\[
\text{F}^- + \text{e}^- \rightarrow \text{F}^- \text{ (} E_0 = 3.6 \text{V)} \tag{2.1}
\]

Perfluorination will also reduce the oxidizability of the ionic headgroup (SO₃⁻ for PFOS and CO₂⁻ for PFOA) since it inductively reduces headgroup electron density. Thus PFCs are quite resistant to oxidation as compared with their alkyl analogs.

2.7.4.1 Advanced oxidation processes (AOPs) - Oxygen-containing radicals

For particularly persistent organics, AOPs which utilize the hydroxyl radical, ozone, or O-atom are a viable solution (Oppenladter, 2003; Pera-Titus et al., 2004; Legrini, 1993). Hydroxyl radical can be generated through hydrogen peroxide photolysis (Kochyany, 1992), ozonation (Hoigne et al., 1983), photo-Fenton’s (Zepp et al., 1992), sonolysis (Hua et al., 1997), and peroxone chemistry (Acero et al., 2001). Hydroxyl radical normally reacts with saturated organics through an H-atom abstraction to form water (Eq. (1.2)) and will react with unsaturated organics primarily via an addition reaction. The hydroxyl radical reacts with most aliphatic and aromatic organics at near diffusion-controlled rates (Buxton et al., 1988). At environmentally relevant pH levels, PFOS and PFOA contain no hydrogens to abstract, thus the hydroxyl radical must act through a direct electron transfer to form the less thermodynamically favored hydroxyl ion (Eq. (1.3)).
Table 2.3 Summary of previous studies on materials to adsorb PFOS

<table>
<thead>
<tr>
<th>Material</th>
<th>Condition mg L(^{-1}) of PFOS</th>
<th>Freundlich constants</th>
<th>Langmuir constants</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>(K_f)</td>
<td>(n)</td>
<td>(q_m)</td>
</tr>
<tr>
<td>GAC</td>
<td>20-250</td>
<td>0.43 mmol(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>0.18</td>
<td>0.37 mmol g(^{-1})</td>
</tr>
<tr>
<td>PAC</td>
<td>20-250</td>
<td>1.27 mmol(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>0.18</td>
<td>1.04 mmol g(^{-1})</td>
</tr>
<tr>
<td>AI400 (Ion exchange resin)</td>
<td>20-250</td>
<td>0.52 mmol(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>0.17</td>
<td>0.42 mmol g(^{-1})</td>
</tr>
<tr>
<td>GAC (Calgon F400)</td>
<td>15-150</td>
<td>960.90 mg(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>0.28</td>
<td>236.40 mg g(^{-1})</td>
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<tr>
<td>GAC (Calgon F300)</td>
<td>15-150</td>
<td>38.50 mg(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>0.332</td>
<td>196.20 mg g(^{-1})</td>
</tr>
<tr>
<td>GAC (Calgon URV-MODI)</td>
<td>15-150</td>
<td>36.70 mg(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>0.371</td>
<td>211.60 mg g(^{-1})</td>
</tr>
<tr>
<td>NaY Zeolite</td>
<td>15-150</td>
<td>0.01 mg(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>1.577</td>
<td>-</td>
</tr>
<tr>
<td>13X Zeolite</td>
<td>15-150</td>
<td>0.73 mg(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>0.507</td>
<td>12.00 mg g(^{-1})</td>
</tr>
<tr>
<td>NaY 80 Zeolite</td>
<td>15-150</td>
<td>31.80 mg(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>0.339</td>
<td>114.70 mg g(^{-1})</td>
</tr>
<tr>
<td>Anaerobic sludge</td>
<td>15-150</td>
<td>0.95 mg(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>1.083</td>
<td>-</td>
</tr>
</tbody>
</table>

Table 2.4 Summary of previous studies on materials to adsorb PFOA

<table>
<thead>
<tr>
<th>Material</th>
<th>Condition mg L(^{-1}) of PFOA</th>
<th>Freundlich constants</th>
<th>Langmuir constants</th>
<th>Ref</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>(K_f)</td>
<td>(n)</td>
<td>(q_m)</td>
</tr>
<tr>
<td>GAC</td>
<td>20-250</td>
<td>0.47 mmol(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>0.28</td>
<td>0.39 mmol g(^{-1})</td>
</tr>
<tr>
<td>PAC</td>
<td>20-250</td>
<td>0.83 mmol(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>0.20</td>
<td>0.67 mmol g(^{-1})</td>
</tr>
<tr>
<td>AI400 (Ion exchange resin)</td>
<td>20-250</td>
<td>3.35 mmol(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>0.13</td>
<td>2.92 mmol g(^{-1})</td>
</tr>
<tr>
<td>GAC (Calgon F400)</td>
<td>15-150</td>
<td>11.8 mg(^{(1-1/n)}) L(^{1/n}) g(^{-1})</td>
<td>0.443</td>
<td>112.1 mg g(^{-1})</td>
</tr>
</tbody>
</table>
Thus the perfluorination or substitution of all of the organic hydrogens for fluorines in PFOS and PFOA renders these compounds inert to advanced oxidation techniques (Schroder et al., 2005). The addition of H$_2$O$_2$ is detrimental to the photolytic degradation of PFOA by competitively adsorbing photons (Hori et al., 2004). An upper limit for the second order rate constant of ‘HO$^-$ + PFOA’ has been determined to be $k_{\text{HO}^- + \text{PFOA}} \leq 10^5$ L.mol$^{-1}$.s$^{-1}$, which is multiple orders of magnitude slower than the reaction of hydroxyl radical with most hydrocarbons (Buxton et al., 1988). The futility of conventional advanced oxidation for the degradation of PFOS and PFOA is noted in the use of perfluorinated compounds to enhance advanced oxidation of other organics. PFOS is used as an additive to increase aqueous solubility of PAHs (An et al., 2009) enhancing their degradation by UV-H$_2$O$_2$. PFOS has also been utilized as a TiO$_2$ surface coating to increase adsorption of PCBs (Hung et al., 2000) and chlorinated aromatics (Yuan et al., 2001) leading to enhanced oxidation rates. Biphasic water-perfluorocarbon systems have been utilized to increase organic ozonation rates (Gromadzka et al., 2007) by increasing dissolved ozone concentrations. Conventional advanced oxidation methods utilizing oxygen-based radicals are not practical methods for the decomposition of perfluorochemicals.

2.7.4.2 Persulfate photolysis - Sulfate radical oxidation

Persulfate photolysis has been utilized for the oxidative degradation of a number of organics (Waldemar et al., 2007; Anipsitakis et al., 2004; Ball et al., 1956). Persulfate photolysis (Dogliott et al., 1967) or thermolysis (Kolthoff et al., 1956) generates two sulfate radicals, SO$_4^-$, (Eq. (1.4)). The sulfate radical is an oxidizing radical that reacts by a direct one-electron transfer to form sulfate (Eq. (1.5)). The sulfate radical has a one-electron reduction potential of 2.3 V (Wordman, 1989), making it a stronger direct electron transfer oxidant than the hydroxyl radical.

\[ \text{S}_2\text{O}_8^{2-} + h\nu \quad (270 \text{ nm})/\Delta \rightarrow \quad \text{SO}_4^- \]  \hspace{1cm} (2.4)

\[ \text{SO}_4^- + e^- \quad \rightarrow \quad \text{SO}_4^{2-} \]  \hspace{1cm} (2.5)

Persulfate photolysis has been utilized to degrade a number of perfluoroalkylcarboxylates of various chain lengths (Chen et al., 2006,b; Hori et
al., 2005 1-2; Kutsuna et al., 2007). PFOA degradation by sulfate radical oxidation has achieved minimum half-lives on the order of 1 hr with fluoride accounting for 15% of the total fluorine over the same period of time. The existence of total fluorine as fluoride can be used as a measure of PFOA mineralization. A reaction mechanism for the sulfate radical mediated degradation of perfluoroalkylicarboxylates was proposed by Kutsuna and Hori (Kutsuna et al., 2007). The initial degradation is postulated to occur through an electron transfer from the carboxylate terminal group to the sulfate radical (Eq. (1.6)). The oxidized PFOA subsequently decarboxylates to form a perfluoroheptyl radical (Eq. (1.7)) which reacts quantitatively with molecular oxygen to form a perfluoroheptylperoxy radical (Eq. (1.8)). The perfluoroheptylperoxy radical will react with another perfluoroheptylperoxy radical in solution since there are no reductants present to yield two perfluoroalkoxy radicals and molecular oxygen (Eq. (1.9)). The perfluoroheptyloxy has two branching pathways: unimolecular decomposition to yield the perfluorohexyl radical and carbonyl fluoride (Eq. (1.10)) or an H-atom abstraction from an acid such as HSO₄⁻ to yield perfluoroheptanol (Eq. (1.12)). The perfluorohexyl radical formed in Eq. (1.10) will react with O₂ (Eq. (1.9)) and resume the radical ‘unzipping’ cycle. The COF₂ will hydrolyze to yield CO₂ and two HF (Eq. (1.11)). The perfluoroheptanol from Eq. (1.12) will unimolecularly decompose to give the perfluoroheptylacyl fluoride and HF (Eq. (1.13)). Perfluoroheptyl acyl fluoride will hydrolyze to yield perfluoroheptanoate (Eq. (1.14)).

\[
\begin{align*}
\text{CF}_3(\text{CF}_2)_n\text{COO}^- + t\text{SO}_4^- &\rightarrow \text{CF}_3(\text{CF}_2)_n\text{COO}^- + \text{SO}_4^{2-} \quad (2.6) \\
\text{CF}_3(\text{CF}_2)_n\text{COO}^- &\rightarrow \text{CF}_3(\text{CF}_2)_n\text{CF}_2^- + \text{CO}_2 \quad (2.7) \\
\text{CF}_3(\text{CF}_2)_n\text{CF}_2^2 + \text{O}_2 &\rightarrow \text{CF}_3(\text{CF}_2)_n\text{CF}_2\text{OO}^- \quad (2.8) \\
\text{CF}_3(\text{CF}_2)_n\text{CF}_2\text{OO}^- + \text{R}_6\text{OO}^- &\rightarrow \text{CF}_3(\text{CF}_2)_n\text{CF}_2\text{O}^- + \text{R}_6\text{O} + \text{O}_2 \quad (2.9) \\
\text{CF}_3(\text{CF}_2)_n\text{CF}_2\text{O}^- &\rightarrow \text{CF}_3(\text{CF}_2)_n\text{CF}_2^- + \text{COF}_2 \quad (2.10) \\
\text{COF}_2 + \text{H}_2\text{O} &\rightarrow \text{CO}_2 + 2\text{HF} \quad (2.11) \\
\text{CF}_3(\text{CF}_2)_n\text{CF}_2\text{O}^- + \text{HSO}_4^- &\rightarrow \text{CF}_3(\text{CF}_2)_n\text{CF}_2\text{OH}^- + \text{HSO}_4^- \quad (2.12) \\
\text{CF}_3(\text{CF}_2)_n\text{CF}_2\text{OH}^- &\rightarrow \text{CF}_3(\text{CF}_2)_n\text{COF}^- + \text{HF} \quad (2.13) \\
\text{CF}_3(\text{CF}_2)_n\text{COF}^- + \text{H}_2\text{O} &\rightarrow \text{CF}_3(\text{CF}_2)_n\text{COO}^- + \text{HF} + \text{H}^+ \quad (2.14)
\end{align*}
\]

During photolysis, Kutsuna and Hori have observed the pH decline to lower than 3 due to HF production (Eq. (1.12)). The produced shorter chain carboxylates will be just as recalcitrant as PFOA. Persulfate photolysis in liquid carbon dioxide/water mixtures (Hori et al., 2005) has been reported to be a good medium for the degradation of longer chain carboxylic acids normally
insoluble in water. Through kinetic modeling of batch reactions the second order rate constants of the sulfate radical with various chain-length perfluorocarboxylates have been determined to be on the order of $10^4 \text{ L.mol}^{-1}.\text{s}^{-1}$ (Katsuna et al., 2007) consistent with a flash photolysis study (Maruthamuthu et al., 1995) which measured sulfate radical reaction with trifluoroacetate to be $1.6 \times 10^4 \text{ L.mol}^{-1}.\text{s}^{-1}$. A relatively slow rate when compared to second order rates of the sulfate radical with hydrocarbons; short-chain alcohols and carboxylic acids are at the lower end with reaction rates on the order of $10^6 \text{ L.mol}^{-1}.\text{s}^{-1}$ and aromatic organics are at the upper end with reaction rates being diffusion-controlled, $10^9$–$10^{10} \text{ L.mol}^{-1}.\text{s}^{-1}$ (Neta et al., 1998). The presence of any other dissolved organic species with aqueous PFOA will competitively inhibit degradation. Persulfate photolysis would be a practical technique for the degradation of ‘pure’ aqueous PFOA. When other organics are present, significant PFOA degradation will only occur when the PFOA concentration greatly exceeds the total organic concentration ([PFOA]/[Org]_{total} $>100$). Persulfate photolysis under the previously stated conditions would be a viable decomposition method for perfluoroalkyl carboxylates of all chain lengths, since they have similar second order kinetics with the sulfate radical (Katsuna et al., 2007).

### 2.7.4.3 Direct Ultra Violet (UV) photolysis

Photolysis is chemical bond-breaking driven by light. UV light adsorption yields an electronically excited molecule. An electronically excited molecule has a bonding (molecular) or non-bonding (atomic) electron promoted to an anti-bonding orbital. An electronically excited molecule is more susceptible to chemical reaction and may open new chemical reaction pathways unavailable to the ground state species. Terrestrial solar-driven photolytic processes require utilization of 290–600 nm photons due to atmospheric absorption of higher energy light. Organics with large chromophores can be directly photolyzed by solar irradiation (Schwarzenbach et al., 2003). Simulated sunlight applied to aqueous solutions of PFOS, and N-EtFOSE 30 days had no effect on their concentrations ([FC]$_i$ = 100 mmol.L$^{-1}$, $\lambda$ = 290–600 nm, 10W, 5W.L$^{-1}$). The 8:2 fluorotelomer alcohol have not significantly degraded under direct photolysis (Gauthier et al., 2005). Ultraviolet-C (UV-C, $\lambda$ <300 nm) and vacuum ultraviolet (VUV, $\lambda$ <200 nm) have been utilized for a number of disinfection and AOPs (Oppenlander, 2003). UV-C generated by a black or germicidal lamp ($\lambda$ = 250±10 nm) is primarily used for indirect photolyses (e.g., persulfate photolysis), disinfection and in some cases direct photolysis (Lee et al., 2005). VUV irradiation is of high enough energy to photodissociate water into an H-atom.
and HO (Eq. (1.15)) with a quantum yield of 0.3 at 185 nm (Getoff et al., 1968; Fricke et al., 1936).

\[
\text{H}_2\text{O} + \text{hv} (\lambda < 200 \text{ nm}) \rightarrow \text{H}^+ + \text{HO}^- \quad (2.15)
\]

Vacuum Ultra Violet (VUV) has a very short liquid penetration depth (<100 mm) due to the strong adsorption by water yielding a strongly oxidizing region near the lamp surface. Organic degradation during VUV photolysis is primarily via HO oxidation (Oppenlander et al., 2000; Jakob et al., 1993; Quici et al., 2008). Hori et al. (2004) reported on the photolytic degradation of PFOA which occurred with a half-life of 24 hrs ([PFOA]₀ = 1.35 mmol.L⁻¹, 200W Xe·Hg lamp, 22 mL, 4.8 atm of O₂). The primary photoproducts have been shorter chain carboxylic acids with fluoride accounting for 15% of the decomposed PFOA fluorine after 24 hrs. Aqueous PFOA VUV photolysis (Chen et al., 2006; Hori et al., 2008) proceeds at a faster rate having a photolysis half-life of 90 min ([PFOA]₀ = 100 mmol.L⁻¹, \( \lambda = 254 \text{ nm with minor } 185 \text{ nm}, 15W, 800 \text{ mL, pH } 3.7, 40^0\text{C, N}_2 \)) with fluoride accounting for 12% of the degraded PFOA fluorine. The gas-phase VUV photolysis of trifluoroacetic acid yields CO₂, CF₃, and H atom as predominant photoproducts (Eq. (16), Osborne et al., 1999). Aqueous PFOA will be dissociated into its ion products at pH 3.7 and direct photolysis will be of the PFOA anion (Eq. (1.17)), which may unimolecularly decompose to a perfluoroalkyl anion, CO₂, and an aqueous electron which will protonate under the experimental conditions (Eq. (1.18)).

\[
\text{CF}_3\text{COOH} + \text{hv} (\lambda = 172\text{nm}) \rightarrow \text{CF}_3\text{COOH}^+ \rightarrow \text{CF}_3^- + \text{CO}_2 + \text{H} \quad (2.16)
\]

\[
\text{CF}_3(\text{CF}_2)_6\text{COO}^- + \text{hv} (\lambda < 172\text{nm}) \rightarrow \text{CF}_3(\text{CF}_2)_6\text{COO}^+ \quad (2.17)
\]

\[
\text{CF}_3(\text{CF}_2)_6\text{COO}^+ + \text{H}^+ \rightarrow \text{CF}_3(\text{CF}_2)_6^- + \text{CO}_2 + \text{H} \quad (2.18)
\]

The hydroxyl radical concentration in the region near the VUV lamp surface may be great enough to also lead to PFOA oxidation and perfluoroalkyl radical formation. Since the photolysis conditions are anoxic (i.e., N₂ atmosphere) the perfluoroalkyl radical will react at diffusion-controlled rates with HO (Eq. (1.19)).

\[
\text{CF}_3(\text{CF}_2)_6^- + \text{HO}^- \rightarrow \text{CF}_3(\text{CF}_2)_6\text{OH} \quad (2.19)
\]

The overall degradation mechanism will occur through similar reactions as seen in persulfate photolysis (Eqs. (1.7)–(1.14)) to yield a perfluoroalkyl carboxylate (PFCA) one –CF₂– unit shorter than the initial species. The produced PFCA will undergo photolysis until the perfluorinated tail is completely unzipped. PFOS
photolytic degradation has also been reported (Yamamoto et al., 2007) and has a slower photolysis rate, half-life of 5.3 days, than PFOA under similar conditions ([PFOS] = 40 mmol.L⁻¹, λ = 254 nm, 32W, 750 mL, 36–40°C, N₂). Shorter chain perfluorocarboxylates and perfluoroalkyl alcohols were detected as reaction intermediates. After 50% of the PFOS is decomposed, fluoride accounts for 59% of the decomposed PFOS fluorine. The greater fluoride mass balance than observed with PFOA is likely due to faster photolysis rates of the PFCA intermediates than of the initial PFOS. Direct photolysis of PFOS and PFOA will be negligible under environmental conditions. Higher energy UV and VUV photolysis can degrade PFOX. Competitive UV light absorption by solvent and other matrix components will limit photolysis rates.

2.7.4.4 Phosphotungstic acid photocatalysis
Phosphotungstic acid, H₃PW₁₂O₄₀, is a heteropolyacid or polyoxometalate that has been utilized for photocatalytic degradation of contaminants (Ozer et al., 2001; Fox et al., 1987) and as an electron shuttle (Lee et al., 2007; Weinstock, 1998; Akid et al., 1985). PW₁₂O₄₀³⁻ is the predominant form when pH<2 and absorbs light with λ <390 nm. Upon light adsorption, PW₁₂O₄₀³⁻ enters a photoexcited state enhancing its oxidation strength (Eq. (1.20)).

\[ 
PW_{12}O_{40}^{3-} + \text{hv (} \lambda <390\text{nm)} \rightarrow PW_{12}O_{40}^{3-*} \]  

PFOA (Hori et al., 2004-a) and PFPA (Hori et al., 2004-b) have been reported to be decomposed by H₃PW₁₂O₄₀ photocatalysis. PFOA has half-life of 24 hrs during phosphotungstic acid photolysis ([PFOA] = 1.35 mmol.L⁻¹, [H₃PW₁₂O₄₀] = 6.7 mmol.L⁻¹, pH<2, 9000W.L⁻¹, Xe-Hg lamp, 4.8 atm of O₂). After 24 hrs of photolysis when 50% of the PFOA is degraded, fluoride accounts for 20% of the total fluorine. The extent of fluoride production is similar to that observed during persulfate photolysis suggesting a similar degradation mechanism where the carboxylate headgroup is oxidatively removed and a shorter-chain perfluoroalkylcarboxylate is formed.

Hori et al. (2004-b) proposed that PW₁₂O₄₀³⁻ photocatalytic PFOA decomposition involves a photo-Kolbe type mechanism. PFOA first complexes with PW₁₂O₄₀³⁻ (Eq. (1.21)), and upon photon adsorption an electron is directly transferred from PFOA to PW₁₂O₄₀³⁻ (Eq. (1.22) (Hori et al., 2004-b). Similar to the sulfate radical mechanism, PFOA will decarboxylate to form the perfluorohexyl radical. Oxygen is essential to the photocatalytic cycle in that it accepts an electron from the reduced phosphotungstic acid, PW₁₂O₄₀⁴⁻, (Eq. (1.23) returning it to its photoactive state.

23
\[ \text{CF}_3\text{(CF}_2\text{)}_n\text{COO}^- + \text{PW}_{12}\text{O}_{40}^{3-} \rightarrow \text{CF}_3\text{(CF}_2\text{)}_n\text{COO}^- \ldots \text{PW}_{12}\text{O}_{40}^{3-}. \] (2.21)
\[ \text{CF}_3\text{(CF}_2\text{)}_n\text{COO}^- \cdot \text{PW}_{12}\text{O}_{40}^{3-} + \text{hv} (\lambda < 390 \text{nm}) \rightarrow \text{CF}_3\text{(CF}_2\text{)}_n\text{CF}_2 + \text{CO}_2 + \text{PW}_{12}\text{O}_{40}^{4+}. \] (2.22)
\[ \text{PW}_{12}\text{O}_{40}^{4-} + \text{O}_2 \rightarrow \text{PW}_{12}\text{O}_{40}^{3-} + \text{O}_2^- . \] (2.23)

The superoxide produced in Eq. (1.13) will protonate when pH<2 to the hydroperoxy radical (Eq. (1.24)) which can act as a reductant for perfluoroalkylperoxy (Eq. (1.25)) and perfluoroalkoxy radicals (Eq. (1.26)). The perfluoroalkylhydroperoxide produced in Eq. (1.25) will likely photolyze to a perfluoroalkoxy radical and a hydroxyl radical (Eq. (1.27)).

Previous researches (yong qui 2007) in our group have found that the irradiation at UV\textsubscript{254} nm and UV\textsubscript{254+185} nm can both degrade PFCAs and stepwise decomposition mechanism of PFCAs was confirmed by mass spectra analysis, and \textit{consecutive} kinetics was proposed to simulate experimental data. It was also reported that PFASs can be degraded by UV\textsubscript{254+185} photolysis, although the products have not been identified yet. Coexisted NOMs have reduced performance of UV photolysis for PFCAs by competing for UV photons. Sample volume or irradiation intensity showed significant influence on degradation of PFCAs. They have recommended local river water polluted by PFOA to be cleaned up by UV\textsubscript{254+185} photolysis effectively. Ozone-related processes were also studied by them, but ineffective to degrade PFC molecules.
## Table 2.5 Summary of previous studies on PFOA degradation

<table>
<thead>
<tr>
<th>Technique</th>
<th>Condition</th>
<th>Power (W) &amp; Volume (mL)</th>
<th>k</th>
<th>product</th>
<th>Energy (kJ)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>UV direct photolysis</td>
<td>9.6 mg L&lt;sup&gt;-1&lt;/sup&gt; of PFOA</td>
<td>200</td>
<td>0.69 d&lt;sup&gt;-1&lt;/sup&gt;</td>
<td>33% F&lt;sup&gt;-&lt;/sup&gt;</td>
<td>792000</td>
<td>Hori et al. 2004</td>
</tr>
<tr>
<td></td>
<td>λ = 220–460 nm</td>
<td>22</td>
<td>τ&lt;sub&gt;1/2&lt;/sub&gt; = 1440 min</td>
<td>38% CO&lt;sub&gt;2&lt;/sub&gt;</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>1/2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>UV phosphotungstic photocatalysis</td>
<td>9.6 mg L&lt;sup&gt;-1&lt;/sup&gt; of PFOA</td>
<td>200</td>
<td>2.0 d&lt;sup&gt;-1&lt;/sup&gt;</td>
<td>30% F&lt;sup&gt;-&lt;/sup&gt;</td>
<td>276000</td>
<td>Hori et al. 2004</td>
</tr>
<tr>
<td></td>
<td>λ = 220–460 nm</td>
<td>22</td>
<td>τ&lt;sub&gt;1/2&lt;/sub&gt; = 500 min</td>
<td>25% CO&lt;sub&gt;2&lt;/sub&gt;</td>
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</tr>
<tr>
<td></td>
<td>0.48 MPa of O&lt;sub&gt;2&lt;/sub&gt;</td>
<td></td>
<td></td>
<td>1/2</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>6.6 mmol·L&lt;sup&gt;-1&lt;/sup&gt; of PTA</td>
<td></td>
<td></td>
<td>1/2</td>
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<td></td>
</tr>
<tr>
<td>TiO&lt;sub&gt;2&lt;/sub&gt; photocatalysis</td>
<td>414 mg L&lt;sup&gt;-1&lt;/sup&gt; of PFOA</td>
<td>75</td>
<td>0.69 d&lt;sup&gt;-1&lt;/sup&gt;</td>
<td>50% F&lt;sup&gt;-&lt;/sup&gt;</td>
<td>132000</td>
<td>Kutsuna et al.2006</td>
</tr>
<tr>
<td></td>
<td>λ = 310–400 nm</td>
<td>50</td>
<td>τ&lt;sub&gt;1/2&lt;/sub&gt; = 1440 min</td>
<td>50% CO&lt;sub&gt;2&lt;/sub&gt;</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>pH = 2–3, 0.1 g of TiO&lt;sub&gt;2&lt;/sub&gt;</td>
<td></td>
<td></td>
<td>1/2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>UV direct photolysis</td>
<td>20 g L&lt;sup&gt;-1&lt;/sup&gt; of PFOA</td>
<td>23</td>
<td>0.017 min&lt;sup&gt;-1&lt;/sup&gt;</td>
<td>10% F&lt;sup&gt;-&lt;/sup&gt;</td>
<td>49</td>
<td>Chen and Zhang 2006</td>
</tr>
<tr>
<td></td>
<td>λ = 185 nm</td>
<td>1000</td>
<td>τ&lt;sub&gt;1/2&lt;/sub&gt; = 41 min</td>
<td>90% PF&lt;sub&gt;acids&lt;/sub&gt;</td>
<td></td>
<td></td>
</tr>
<tr>
<td>UV persulfate photocatalysis</td>
<td>20 g L&lt;sup&gt;-1&lt;/sup&gt; of PFOA</td>
<td>23</td>
<td>0.012 min&lt;sup&gt;-1&lt;/sup&gt;</td>
<td>5% F&lt;sup&gt;-&lt;/sup&gt;</td>
<td>69</td>
<td>Chen and Zhang 2006</td>
</tr>
<tr>
<td></td>
<td>λ = 254 nm</td>
<td>1000</td>
<td>τ&lt;sub&gt;1/2&lt;/sub&gt; = 58 min</td>
<td>95% PF&lt;sub&gt;acids&lt;/sub&gt;</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>1.5 mmol·L&lt;sup&gt;-1&lt;/sup&gt; of S&lt;sub&gt;2&lt;/sub&gt;O&lt;sub&gt;8&lt;/sub&gt;&lt;sup&gt;2-&lt;/sup&gt;</td>
<td></td>
<td></td>
<td>1/2</td>
<td></td>
<td></td>
</tr>
<tr>
<td>UV persulfate photolysis</td>
<td>540 g L&lt;sup&gt;-1&lt;/sup&gt; of PFOA</td>
<td>200</td>
<td>0.69 h&lt;sup&gt;-1&lt;/sup&gt;</td>
<td>12% F&lt;sup&gt;-&lt;/sup&gt;</td>
<td>33600</td>
<td>Hori et al. 2005</td>
</tr>
<tr>
<td></td>
<td>λ = 220–460 nm, 0.48 MPa of O&lt;sub&gt;2&lt;/sub&gt;, 10 mmol·L&lt;sup&gt;-1&lt;/sup&gt; of S&lt;sub&gt;2&lt;/sub&gt;O&lt;sub&gt;8&lt;/sub&gt;&lt;sup&gt;2-&lt;/sup&gt;, pH = 2–3</td>
<td>22</td>
<td>τ&lt;sub&gt;1/2&lt;/sub&gt; = 58 min</td>
<td>85% PF&lt;sub&gt;acids&lt;/sub&gt;</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table 2.5 Summary of previous studies on PFOA degradation (Continued)

<table>
<thead>
<tr>
<th>Technique</th>
<th>Condition</th>
<th>Power (W) &amp; Volume (mL)</th>
<th>k</th>
<th>product</th>
<th>Energy (kJ)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>photocatalysis</td>
<td>20 g L(^{-1}) of PFOA</td>
<td>23</td>
<td>0.0077 min(^{-1})</td>
<td>10% F(^-)</td>
<td>500</td>
<td>Chen et al. 2006</td>
</tr>
<tr>
<td>TiO(_2)/Ni-Cu</td>
<td>(\lambda = 254) nm</td>
<td>250</td>
<td>(\tau^{1/2} = 90) min</td>
<td>90% PFacids</td>
<td></td>
<td></td>
</tr>
<tr>
<td>photoelectrocatalysis</td>
<td>20 g L(^{-1}) of PFOA</td>
<td>23</td>
<td>0.015 min(^{-1})</td>
<td>20% F(^-)</td>
<td>250</td>
<td>Chen et al. 2006</td>
</tr>
<tr>
<td></td>
<td>(\lambda = 254) nm</td>
<td>250</td>
<td>(\tau^{1/2} = 45) min</td>
<td>80% PFacids</td>
<td></td>
<td></td>
</tr>
<tr>
<td>persulfate photolysis</td>
<td>535 mg L(^{-1}) of PFBA</td>
<td>60</td>
<td>0.0096 min(^{-1})</td>
<td>n/a</td>
<td>1300</td>
<td>Kutsuna and Hori 2007</td>
</tr>
<tr>
<td></td>
<td>(\lambda = 254) nm</td>
<td>200</td>
<td>(\tau^{1/2} = 72) min</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>50 mmol·L(^{-1}) of S(_2)O(_8)(^{2-})</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>hydrogen peroxide photolysis</td>
<td>2.5 mmol·L(^{-1}) of PFBA</td>
<td>60</td>
<td>3.0e-5 min(^{-1})</td>
<td>n/a</td>
<td>420000</td>
<td>Kutsuna and Hori 2007</td>
</tr>
<tr>
<td></td>
<td>(\lambda = 254) nm</td>
<td>200</td>
<td>(\tau^{1/2} = 23100) min</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>250 mmol·L(^{-1}) of H(_2)O(_2)</td>
<td></td>
<td></td>
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<tr>
<td>sonolysis</td>
<td>8.2 L(^{-1}) of PFOA</td>
<td>150</td>
<td>0.018 min(^{-1})</td>
<td>95% F(^-)</td>
<td>670</td>
<td>Vecitis et al. 2008</td>
</tr>
<tr>
<td></td>
<td>(f = 354) kHz</td>
<td>600</td>
<td>(\tau^{1/2} = 39) min</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>sonolysis</td>
<td>82 (\mu)g L(^{-1}) of PFOA</td>
<td>150</td>
<td>0.047 min(^{-1})</td>
<td>95% F(^-)</td>
<td>260</td>
<td>Vecitis et al. 2008</td>
</tr>
<tr>
<td></td>
<td>(f = 354) kHz</td>
<td>600</td>
<td>(\tau^{1/2} = 15) min</td>
<td></td>
<td></td>
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</tr>
<tr>
<td>UV-KI photolysis</td>
<td>8.2 g L(^{-1}) of PFOA</td>
<td>1.5</td>
<td>0.0014 min(^{-1})</td>
<td>10% F(^-)</td>
<td>1500</td>
<td>Park et al. 2009</td>
</tr>
<tr>
<td></td>
<td>(\lambda = 254) nm</td>
<td>30</td>
<td>(\tau^{1/2} = 500) min</td>
<td>gaseous fluoroalkanes</td>
<td></td>
<td></td>
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Table 2.5 Summary of previous studies on PFOA degradation (Continued)

<table>
<thead>
<tr>
<th>Technique</th>
<th>Condition</th>
<th>Power (W) &amp; Volume(mL)</th>
<th>k</th>
<th>product</th>
<th>Energy (kJ)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>UV-KI photolysis</td>
<td>200 nmol·L⁻¹ of PFOA l = 254 nm</td>
<td>1.5</td>
<td>0.0025 min⁻¹</td>
<td>10% F⁻</td>
<td>820</td>
<td>Park et al. 2009</td>
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<tr>
<td></td>
<td>30</td>
<td>τ(^{1/2}) = 280 min</td>
<td></td>
<td>Gaseous fluoroalkanes</td>
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<td></td>
</tr>
<tr>
<td>Ferrophotolysis</td>
<td>14.3 gL⁻¹ of PFBA</td>
<td>200</td>
<td>0.028 h⁻¹</td>
<td>45% F⁻</td>
<td></td>
<td>Hori et al. 2007</td>
</tr>
<tr>
<td></td>
<td>2.5mmol.L⁻¹ of Fe(_2)(SO(_4))(_3)(^3)</td>
<td>105</td>
<td>τ(^{1/2}) = 1490 min</td>
<td>55% short chains</td>
<td>89400</td>
<td></td>
</tr>
<tr>
<td></td>
<td>λ = 220–460 nm</td>
<td></td>
<td></td>
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Table 2.6 Summary of previous studies on PFOS degradation

<table>
<thead>
<tr>
<th>Technique</th>
<th>Condition</th>
<th>Power (W) &amp; Volume(mL)</th>
<th>k</th>
<th>product</th>
<th>Energy (kJ)</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>sub-critical</td>
<td>185mgL⁻¹ of PFOS</td>
<td>0</td>
<td>0.013 min⁻¹</td>
<td>50% F⁻</td>
<td>2000</td>
<td>Hori et al. 2006</td>
</tr>
<tr>
<td>Fe(0)</td>
<td>0.5 g of Fe(0)</td>
<td>10</td>
<td>τ(^{1/2}) = 53 min</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>350°C, 20 MPa</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>UV direct</td>
<td>20mgL⁻¹ of PFOS</td>
<td>32</td>
<td>0.13 d⁻¹</td>
<td>71% F⁻</td>
<td>17000</td>
<td>Yamamoto et al. 2007</td>
</tr>
<tr>
<td>photolysis</td>
<td>λ = 254 nm</td>
<td>750</td>
<td>τ(^{1/2}) = 7700min</td>
<td>90% SO(_4)²⁻</td>
<td></td>
<td></td>
</tr>
<tr>
<td>UV alkaline</td>
<td>20mgL⁻¹ of PFOS</td>
<td>32</td>
<td>0.93 d⁻¹</td>
<td>NaF(_{(s)})</td>
<td>2500</td>
<td>Yamamoto et al. 2007</td>
</tr>
<tr>
<td>IPA photolysis</td>
<td>λ = 254 nm</td>
<td>750</td>
<td>τ(^{1/2}) = 1070min</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>sonolysis</td>
<td>10mgL⁻¹ of PFOS</td>
<td>150</td>
<td>0.011 min⁻¹</td>
<td>95% F⁻</td>
<td>945</td>
<td>Vecitis et al., 2008</td>
</tr>
<tr>
<td></td>
<td>f = 354 kHz</td>
<td>600</td>
<td>τ(^{1/2}) = 63 min</td>
<td>100% SO(_4)²⁻</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Technique</td>
<td>Condition</td>
<td>Power (W) &amp; Volume (mL)</td>
<td>k</td>
<td>product</td>
<td>Energy (kJ)</td>
<td>Reference</td>
</tr>
<tr>
<td>---------------</td>
<td>----------------------------------</td>
<td>-------------------------</td>
<td>-----------</td>
<td>--------------------</td>
<td>-------------</td>
<td>--------------------</td>
</tr>
<tr>
<td>UV-KI photolysis</td>
<td>10 mg L⁻¹ of PFOS (\lambda = 254 \text{ nm}) [KI] = 10 mmol L⁻¹</td>
<td>1.5 &amp; 30</td>
<td>0.002 min⁻¹ &amp; (\tau^{1/2} = 350 \text{ min})</td>
<td>50% F⁻</td>
<td>960</td>
<td>Park et al. 2009</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>50% fluoroalkanes</td>
<td></td>
<td></td>
</tr>
<tr>
<td>UV-KI photolysis</td>
<td>100 mg L⁻¹ of PFOS (\lambda = 254 \text{ nm}) [KI] = 10 mmol L⁻¹</td>
<td>1.5 &amp; 30</td>
<td>0.008 min⁻¹ &amp; (\tau^{1/2} = 87 \text{ min})</td>
<td>50% F⁻</td>
<td>260</td>
<td>Park et al. 2009</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>50% fluoroalkanes</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Sonolysis</td>
<td>100 mg L⁻¹ of PFOS (f = 354 \text{ kHz})</td>
<td>150 &amp; 600</td>
<td>0.023 min⁻¹ &amp; (\tau^{1/2} = 30 \text{ min})</td>
<td>95% F⁻</td>
<td>450</td>
<td>Vecitis et al. 2008</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>100% SO₄²⁻</td>
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</tbody>
</table>
Chapter 3  Ferrate oxidation of PFCs

3.1 Introduction

Oxidation of PFCs (if it is possible) is an important technique as a treatment process, because both elimination and degradation of PFCs can be achieved. According to the available literature, PFCs level in tap water has showed a linear correlation to surface water suggesting the incapability of conventional disinfection agents to oxidize PFCs. Advanced oxidation processes (AOPs) may be possible candidates to oxidize PFCs. Being involved in generation of hydroxyl radicals to enhance water treatment, this kind of process shows excellent performance to control micro pollutants in water and waste water. Most common processes of AOPs include O$_3$/H$_2$O$_2$, O$_3$/UV and UV/H$_2$O$_2$. UV/TiO$_2$ process and Fenton’s reagent have also shown efficiency on specific wastewater (Gottschalk, et al., 2000). However, previous researches have showed that AOPs including O$_3$, O$_3$/UV, O$_3$/H$_2$O$_2$ and Fenton process cannot degrade PFOS, but degrade PFOS precursors and partly fluorinated polymers effectively (Schröder and Meesters, 2005). Our research group too investigated on capabilities of AOPs to oxidize PFCs and deduced with four main conclusions (Yong 2007).

1. Mass spectra analysis on UV photolysis of PFCAs confirmed stepwise degradation mechanism of PFCAs. UV photolysis can also degrade PFAS, however, the velocity was very low and final products can not be identified yet.

2. Results of UV photolysis for PFCAs were successfully estimated by consecutive kinetics. Kinetic parameter for UV$_{254+185}$ irradiation on PFCAs was estimated as 2~6 hr$^{-1}$ in pure water. Background absorbance can significantly reduce degradation of PFOA in wastewater, by which $k$ was decreased from 2.5~3.5 hrs$^{-1}$ in pure water to 0.9~1 hr$^{-1}$ in sand filtration effluent.

3. PFOA polluted river water can be cleaned by UV photolysis process. UV$_{254+185}$ showed satisfactory performance to degrade more than 90% of 40 μg/L PFOA in 4 hrs, with $k$ value of 0.81 hr$^{-1}$. UV$_{254}$ was ineffective to degrade PFOA because of strong background absorbance. UV wave length and sample volume showed significant influences on degradation kinetics.

4. For ozone-related processes, mass spectra and batch experiments proved inefficiency of O$_3$/H$_2$O$_2$ to degrade PFCs. Semi-batch experiments showed obvious removal of PFCs in first order kinetics, which was attributed to air-floating effect by aeration.
3.1.1 Ferrate Oxidation

Although the ferrate(VI) species were discovered a century ago, a renaissance of interest in ferrate(VI) application began in the 1970s, when a number of researches were carried out for the degradation of various organic pollutants including the emerging micro-pollutants such as endocrine disrupt chemicals (EDCs) and pharmaceuticals. Extensive studies on the ferrate(VI) are also due to its unique oxidation/coagulation capacity in the environmental remediation and it is a green chemical. Ferrate(VI) is a strong oxidizing agent, which can be seen from the reduction potentials of reactions (3.1) and (3.2) in acidic and alkaline solutions, respectively (Wood, 1958).

\[
\text{FeO}_4^{2-} + 8H^+ + 3e^- \rightarrow \text{Fe}^{3+} + 4H_2O \quad E_\theta = +2.20V \quad (3.1)
\]

\[
\text{FeO}_4^{2-} + 4H^+ + 3e^- \rightarrow \text{Fe(OH)}_3 + 5OH^- \quad E_\theta = +0.72V \quad (3.2)
\]

The spontaneous reduction of ferrate (VI) in water forms non-toxic by-products, molecular oxygen and iron(III) (Eq. (3.3)) (Sharma, 2002a), suggesting that ferrate(VI) is an environment-friendly oxidant.

\[
2\text{FeO}_4^{2-} + 5H_2O \rightarrow 2\text{Fe(OH)}_3 + 3\text{O}_2 + 4OH^- \quad (3.3)
\]

Ferric(III) hydroxide, produced from ferrate(VI), acts as a coagulant for the removal of metals, non-metals, radionuclides, and organics (Potts and Churchwell, 1994: Sharma, 2002a). Ferrate(VI) has been proposed as an alternative to chlorine for the disinfection of water and wastewater (Sharma, 2002a, 2007b; Sharma et al., 2005a). Table 3.1 shows the oxidation potential of common disinfectants currently used in water and wastewater industry.

Fe(VI) solutions are generally unstable; their decomposition by reduction to Fe(III) species occurs rapidly at room temperature. The instability may be retarded but not stopped at low temperatures or with careful control of solution concentrations. Hence, without steps of refrigeration or high purification, the solutions cannot be stored for use in practice. Solid ferrate(VI) salts are stable, but they are costly as they require multiple chemical reagents and long synthesis time. This makes it difficult to be used in industry. In order to solve the problems of instability and the high cost of using ferrate(VI), some researchers have suggested to generate ferrate in situ and apply the generated ferrate(VI) directly for water and wastewater treatment.
3.1.2 Ferrate production

There are two basic methods for ferrate production: Chemical (Ockerman and Schereyer, 1951; Thompson et al., 1951) and electrochemical. The chemical methods are based on contacting iron compounds, such as iron(III) nitrate and iron oxide, with an oxidizing material in either an alkaline environment, the wet route, or under extreme temperatures in a controlled atmosphere, the dry route. The drawback of this method is that it requires additional processes and reagents to obtain high yield and purity which makes it expensive. On the other hand, the electrochemical method usually consists of a sacrificial iron anode in an electrolysis cell containing a strongly alkaline solution with an electric current serving to oxidize the iron to Fe(VI). The electrochemical method is more promising than the chemical for ferrate generation as it has a simpler process and does not require costly chemical reagents. The main elements that affect the electrochemical production of ferrate(VI) are the anode composition, the type and concentration of electrolyte, the current density and the cell design (Mácová et al., 2009).

3.1.3 Application of ferrate oxidation in water and wastewater treatment

It has been shown that ferrate(VI) has a high efficient oxidation performance for the degradation of a number of compounds that could contaminate water such as inorganic oxysulfur compounds (Read et al., 2001), thiourea and thioacetamide (Sharma, 2002), and H₂S could be completely converted to sulphate with the ratio 2.5:1 of ferrate or more to the total hydrogen sulfide (Simon et al., 2002), sulfide mine tailings where ferrate reduces the potential of acid production and enhance the potential of toxic metal species from tailing by oxidizing tailing bounded sulfide to

Table 3.1: Oxidation potentials of common oxidants

<table>
<thead>
<tr>
<th>Disinfectant</th>
<th>Oxidant</th>
<th>Reaction Eₒ, V</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chlorine</td>
<td>Cl₂(g)+2e ⇄ 2Cl⁻</td>
<td>1.358</td>
</tr>
<tr>
<td></td>
<td>ClO⁻+H₂O+2e ⇄ Cl⁻+2OH⁻</td>
<td>0.841</td>
</tr>
<tr>
<td>Hypochlorite</td>
<td>HClO+H⁺+2e ⇄ Cl⁻+H₂O</td>
<td>1.482</td>
</tr>
<tr>
<td>Chlorine dioxide</td>
<td>ClO₂(aq)+e ⇄ ClO₂</td>
<td>0.954</td>
</tr>
<tr>
<td>Perchlorate</td>
<td>ClO₄⁻+8H⁺+8e ⇄ Cl⁻+4H₂O</td>
<td>1.389</td>
</tr>
<tr>
<td>Ozone</td>
<td>O₃+2H⁺+2e ⇄ O₂+H₂O</td>
<td>2.076</td>
</tr>
<tr>
<td>Hydrogen peroxide</td>
<td>H₂O₂+2H⁺+2e ⇄ 2H₂O</td>
<td>1.776</td>
</tr>
<tr>
<td>Dissolved oxygen</td>
<td>O₂+4H⁺+4e ⇄ 2H₂O</td>
<td>1.229</td>
</tr>
<tr>
<td>Permanganate</td>
<td>MnO₄⁻+4H⁺+3e ⇄ MnO₂+2H₂O</td>
<td>1.679</td>
</tr>
<tr>
<td></td>
<td>MnO₄⁻+8H⁺+5e ⇄ Mn²⁺+4H₂O</td>
<td>1.507</td>
</tr>
<tr>
<td>Ferrate(VI)</td>
<td>FeO₄²⁻+8H⁺+3e ⇄ Fe³⁺+4H₂O⁻</td>
<td>2.20</td>
</tr>
</tbody>
</table>
sulphate (Mursheda et al., 2003), alachlor which could be totally eliminated from wastewater within 10 min under optimization conditions (Zhu et al., 2006), phenol and chlorophenol (Graham et al., 2004), endocrine disrupting chemicals (EDCs), which could be reduced to low levels ranging from 10 to 100 ng/L (Jiang et al., 2005), sulfamethoxazole (SMX) (Sharma et al., 2006) and cetylpyridinium chloride (CPC), which are mineralized after opening the pyridine ring by ferrate (Eng et al., 2006). Some organics such as methanol, ethanediol and phenol could be oxidized completely to CO2 and H2O even at room temperature (Denver and Pletcher, 1996 1, 2). Ethionine was oxidized by ferrate to sulfoxide within 500 s and thiourea was oxidized to urea within 10 s (Read et al., 2004). The oxidation of seleno-DL-methionine by potassium ferrate to the selenoxide is complete within 7.5 ms to 2 s (Read and Wyand, 1998). The half-lives for hydrogen sulfide, thiourea, thioacetamide, cyanide and thiocyanate are 3.0 ms, 0.6 s, 0.8 s, 9.3 s and 180 s, respectively. The oxidation of pollutants and amino acids with Fe(V), which is formed as a result of the use of ionizing radiation and photocatalytic techniques in the presence of Fe(VI), is 3–5 order of magnitude faster than Fe (VI) (Sharma, 2004). Offensive odor of the compounds generated during sewage treatment were reduced immediately and to an acceptable level by stabilization with Fe(VI) (Lucas et al., 1996). Most toxic ion cyanide in aqueous wastewater can be oxidized in few minutes (Tiwari et al., 2007). The disinfection performance of ferrate (VI) was also intensively studied by several researchers: removing more than 99.9% of total coliform was reported by Waite (1979), Kato and Kazama (1991) and Jiang et al. (2007). It was shown that ferrate can reduce 30% more COD and kill three log more bacteria than ferric sulphate and aluminum sulphate at a similar or even smaller dose (Jiang et al., 2006 2). It was reported that ferrate could rapidly inactivate virus at pHs 6–8 (Schink and Waite, 1980; Kazama, 1994; Kazama, 1995). Such high disinfection performance is of utmost importance for the water industry. It was also shown that ferrate could enhance the coagulation of algae (Lui and Ma, 2002; Ma and Liu, 2002 1, 2) and that microcystin was easily decomposed by ferrate depending on the dosage of ferrate, the pH, and the contact time (Yuan et al., 2002). On the other hand, the effect of ferrate on the enhancement of photocatalytic degradation of microcystin was studied by Yuan et al. (2006) and showed that above 81% degradation can be achieved at ferrate concentration of 0.08– 0.17 mmol/L within 10 min. In addition to the oxidation and disinfection effect, ferrate(VI) generates a coagulant in a single dosing and mixing unit process as a result of reduction of ferrate ions to Fe(III) during the process of oxidation/disinfection (Jiang et al., 2001). The research progress of using ferrate as a coagulant, a disinfectant and an oxidant has been reviewed by Jiang (Jiang, 2007). Table 3.2 shows a
summary of previous work on Ferrate oxidation of various organic compounds.

Table 3.2 Ferrate oxidation of various organic compounds

<table>
<thead>
<tr>
<th>Pollutants</th>
<th>pH</th>
<th>k (M$^{-1}$s$^{-1}$)</th>
<th>t$_{1/2}$</th>
<th>Reference</th>
</tr>
</thead>
<tbody>
<tr>
<td>Thioacetamide</td>
<td>9</td>
<td>5.6×10$^3$</td>
<td>0.36s</td>
<td>Sharma et al., 2000</td>
</tr>
<tr>
<td>Thiourea</td>
<td>9</td>
<td>3.4×10$^3$</td>
<td>0.59s</td>
<td>Sharma et al., 1999</td>
</tr>
<tr>
<td>p-Toluidine</td>
<td>9</td>
<td>1.3×10$^3$</td>
<td>1.5s</td>
<td>Sharma et al., 1995</td>
</tr>
<tr>
<td>Glyoxylic acid</td>
<td>8</td>
<td>7.0×10$^2$</td>
<td>2.9 s</td>
<td>Carr et al., 1985</td>
</tr>
<tr>
<td>Thiodiethanol</td>
<td>8</td>
<td>7.0×10$^2$</td>
<td>20.0s</td>
<td>Carr et al., 1985</td>
</tr>
<tr>
<td>Phenol</td>
<td>9</td>
<td>8.0×10$^1$</td>
<td>25.0s</td>
<td>Carr et al., 1985</td>
</tr>
<tr>
<td>p-Aminobenzoic acid</td>
<td>9</td>
<td>4.3×10$^1$</td>
<td>46.9s</td>
<td>Sharma et al., 1995</td>
</tr>
<tr>
<td>Methylamine</td>
<td>8</td>
<td>4.0×10$^1$</td>
<td>50.0s</td>
<td>Carr et al., 1985</td>
</tr>
<tr>
<td>Nitriloacetic acid</td>
<td>8</td>
<td>2.0×10</td>
<td>16.7min</td>
<td>Carr et al., 1985</td>
</tr>
<tr>
<td>Diethylamine</td>
<td>8</td>
<td>7.0×10$^{-1}$</td>
<td>47.6min</td>
<td>Carr et al., 1985</td>
</tr>
<tr>
<td>Neopentyl alcohol</td>
<td>8</td>
<td>1.0×10$^{-1}$</td>
<td>5.55h</td>
<td>Carr et al., 1985</td>
</tr>
<tr>
<td>Isopropyl alcohol</td>
<td>8</td>
<td>6.0×10$^{-2}$</td>
<td>9.26h</td>
<td>Carr et al., 1985</td>
</tr>
</tbody>
</table>

Note k = constant, $t_{1/2}$ = half time

3.1.4 Ferrate oxidation of PFCs

Even though we have realized the strength of C-F bond and the resistance of PFCs to oxidize by our previous work (Yong 2007), this experiment was carried out as ferrate (VI) technology has not been tested for PFCs oxidation including PFOS and PFOA.

3.2 Aims

Major aim of this chapter is to study the oxidation characteristics of PFCs by ferrate techniques. A series of batch type experiments were conducted at room temperature (24°C) and pressure to reach the objectives. A control test was done with Cetylpyridinium chloride (CPC) to ensure the accuracy of the methodology adapted. Objectives in detail are listed below.

(1) To conduct batch experiment with PFOA to identify the optimum Ferrate(VI):PFCs molar ratio for ferrate oxidation.

(2) To conduct batch experiment with PFOA to understand the kinetic behavior of ferrate oxidation of PFCs.
(3) To conduct batch experiment with a mixture of PFCs to understand the competitive ferrate oxidation of PFCs.

(4) To conduct batch experiment with CPC to ensure the accuracy of the methodology adapted.

3.3 Experimental method
3.3.1 Chemicals
All PFCs standards, sodium sulfite, sodium phosphate and 0.5 M borate buffer having purity more than 98% were obtained from WAKO pure chemical industries, Japan. Potassium ferrate(VI) and CPC salt (98+ % purity) were obtained from Aldrich Company. All chemicals were used without further purification. Solutions were prepared in ultra pure water purified with 18.2 MΩ Milli-Q (Millipore SAS 67120) water purification system. Ferrate(VI) solution was freshly prepared by dissolving a desired amount of ferrate(VI) salts into a 1.7 mM borate / 8 mM phosphate buffer solution. The phosphate served as a complexing agent for the Fe(III) produced, which otherwise could precipitate as a hydroxide and would accelerate the decomposition of ferrate(VI).

3.3.1.2 Analytical equipments and methods
All PFCs were directly analyzed and measured using LC/MS/MS. Extract 10μL was injected to a 2.1×100 mm (5 μm) Agilent Eclipse XDB-C18 column. Mobile phase consisted of (A) 5 mM ammonium acetate in ultrapure water (LC/MS grade) and (B) 100% Acetonitrile (LC/MS grade). At a flow rate of 0.25 mL/min, the mobile phase started with an initial condition of 30% (B), increased to 50% (B) at 16.5 min, then to 70% (B) at 16.6, 75 held at 70% (B) for 3.4 min, went up to 90% (B) at 21 min, kept at 90% (B) for 1 min, and then ramped down to 30% (B). The total running time was 34 min for each sample. For quantitative determination, the HPLC was interfaced with an Agilent 6400 Triple Quadrupole (Agilent, Japan) mass spectrometer (MS/MS). Mass spectrometer was operated with the electrospray ionization (ESI) negative mode. Analyte ion was monitored by using multiple reaction monitoring (MRM) mode. Both parent ion (499m/z) and daughter ion (80m/z) were monitored with the retention time of 13.8 min. A total organic carbon (TOC) analyzer, Shimadzu 5050 was used to measure the TOC content (in mg/L) of samples. The calibration curve of all PFCs gave good coefficient of determination (R^2>0.995).

3.3.1.3 Oxidation tests of PFC
Two kinds of batch type oxidation experiments were conducted for PFOA and a mixture of PFCs ((PFDA, PFOA, PFHpA, PFPeA, PFOS and PFHS).
In both cases, collected samples were centrifuged at 4000 rpm for 10 minutes to eliminate precipitant (Kubota ICE 61010-2-020). Then the samples were diluted by acetonitrile (LC/MS grade) to reach 40% acetonitrile level in the sample, which is required by LC/MS/MS. Figure 3.1 shows the flow chart of batch experiments; Figure 3.2 shows a schematic diagram of a batch experiment with the magnetic stirrer and Table 3.3 summarizes experimental conditions.

### 3.3.2 Determination of optimum Molar ratio

In this experiment, four parallel batch tests (250 mL each) were conducted with ferrate(VI) to PFOA molar ratios of 10, 100, 1000, and 10,000 at room temperature (25°C). The PFOA concentration was kept fixed at 16 µg/L, while concentration of ferrate(VI) was changed to increase the molar ratio. In each oxidation test, samples were constantly mixed by magnetic stirrer (Figure 3.2) and samples were taken (1 mL) at distinct time intervals up to 24 hrs (Table 3.3). Sodium sulfite solution was added immediately to each sample upon removal from the reactor in order to quench the ferrate and stop any further oxidation.

<table>
<thead>
<tr>
<th>Exp No.</th>
<th>Chemicals</th>
<th>Concentration</th>
<th>Chemicals, µg/L</th>
<th>Ferrate(VI), mg/L</th>
<th>Molar ratio</th>
<th>Contact time (hrs)</th>
<th>Measurement items</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>PFOA</td>
<td></td>
<td>16</td>
<td>0.071</td>
<td>10</td>
<td>1, 2, 3, 4, 5, 6, 7</td>
<td>Remaining PFOA</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>0.710</td>
<td>100</td>
<td>8, 9, 11, 14, 17, 24</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>7.100</td>
<td>1000</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>71.000</td>
<td>10000</td>
<td></td>
<td></td>
</tr>
<tr>
<td>2</td>
<td>Mixture of PFOS, PFOA, PFDA, PFHpA, PFPeA, PFHs</td>
<td>0.10, 0.030, 0.059, 0.089</td>
<td>0.050</td>
<td>0.150, 0.300, 44.000</td>
<td>100, 200, 300</td>
<td>24 (hrs)</td>
<td>Remaining PFCs</td>
</tr>
<tr>
<td></td>
<td>PFDA, PFHpA, PFPeA, PFHs</td>
<td>1.0, 0.300, 0.590, 0.890</td>
<td>5.0</td>
<td>1.480, 2.960, 4.440</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>PFHs</td>
<td>10</td>
<td>2.960, 5.920, 8.880</td>
<td>7.4100, 14.800, 22.200</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>CPC*</td>
<td>10,000</td>
<td>29.100</td>
<td>3</td>
<td>0, 5, 10, 20, 35, 50 (min)</td>
<td>TOC</td>
<td></td>
</tr>
<tr>
<td></td>
<td>PFOA</td>
<td>10,000</td>
<td>23.900</td>
<td>3</td>
<td>0, 5, 15, 35, 50 (min)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

* References: Yong et al., 2006
In this experiment, six parallel batch experiments (250 mL each) were conducted with different initial PFC concentrations (µg/L) of 0.10, 0.50, 1, 5, 10 and 25. The experiment was repeated for 3 ferrate(VI) to PFCs molar ratios of 100, 200 and 300 and the reaction time was 24 hrs. Separate experiment was conducted to determine the reduction of TOC equivalent to PFOA by ferrate oxidation and the results were compared with the results of the same experiment for CPC.
3.4 Results and discussion

3.4.1 Oxidation test of PFOA in a ferrate solution

Figure 3.3 shows results of the Experiment 1. Maximum PFOA reduction of 10.0 % was obtained at the molar ratio 100, followed by 6.0 % reduction by molar ratio 1000, 0.3 % reduction by molar ratio 10. It was noticed that PFOA concentration was kept unchanged at 10000 molar ratio. It was also noticed that the reaction with high ferrate(VI) to PFOA molar ratios (>1000) gave more fluctuating readings over the reaction time, whereas less molar ratio of 10 gave less fluctuation. One possible phenomenon to describe this fluctuation is adsorption. Temporary adsorption and desorption process of PFOA in to ferrate(VI) and ferrite(III) (solid particles) can cause fluctuation of the reading.

Previous studies have shown that five times higher concentration of ferrate(VI) than the concentration of target pollutant is good enough to oxidize most of other pollutants than PFCs, and observed that most pollutants are quick to react with ferrate. (Li et al., 2008; Yong et al., 1995; Sarma et al., 1995; Sharma et al., 2000; Carr et al., 1985). However, our results showed that the PFOA was not oxidized by ferrate(VI) even at very high molar ratio (10,000) and longer reaction time (24 hrs) (Figure 3.3).

![Fig. 3.3 Ferrate(VI) degradation of PFOA with time](image)

3.4.1.1 Stability of PFOA chain

As mentioned in the introduction of this chapter, PFOA is recalcitrant towards oxidation due to the complete substitution of fluorine (C-F bond) for hydrogen (C-H bond). Fluorine will retain its electrons (i.e., will resist oxidation) at any cost. Fluorine is nearly always found in the (−1) valence
state with the only exception being F₂ where its oxidation state is (0). The fluorine atom is the most powerful inorganic oxidant known with a reduction potential of 3.6 V (Eq. 3.4) (Wardman, 1989) and thus is thermodynamically unfavorable to create the fluorine atom with any other one-electron oxidant.

\[ \text{F} + e^- \rightarrow \text{F}^- \quad (E^o = 3.6 \text{V}) \]  

(3.4)

Perfluorination will also reduce the oxidizability of the ionic headgroup of CO₂⁻ for PFOA, since it inductively reduces headgroup electron density. The other important characteristic of PFOA is absence of C-H bonds which is important to initiate oxidation process. Particularly persistent organics such as PFOA hydroxyl radical normally reacts with saturated organics through an H-atom abstraction to form water (Eq. (3.6) and will react with unsaturated organics primarily via an addition reaction. The hydroxyl radical reacts with most aliphatic and aromatic organics at near diffusion-controlled rates (Buxton et al., 1988). At environmentally relevant pH, PFOA contains no hydrogens to abstract, thus the hydroxyl radical must act through a direct electron transfer to form the less thermodynamically favored hydroxyl ion (Eq. 3.6).

\[ \text{HO} + e^- \rightarrow \text{H}_2\text{O} \quad (E^o = 2.7 \text{ V}) \]  

(3.5)

\[ \text{HO} + e^- \rightarrow \text{HO}^- \quad (E^o = 1.9 \text{ V}) \]  

(3.6)

Thus the perfluorination or substitution of all of the organic hydrogens for fluorines in PFOA renders these compounds inert to advanced oxidation techniques (Schroder et al., 2005). Table 3.4 shows bond energy of each bond in the PFOA molecule.

3.4.1.2 Oxidation of PFOA by Ferrate

Oxidation can be defined as the loss of at least one electron when two or more substances interact. Carbon can take the oxidation numbers of -4, -3, -2, -1, +1, +2, +3 and +4. Fluorine has only one oxidation number of -1. Figure 3.4 shows oxidation numbers of each atoms of PFOA chain. According to this figure two carbon atoms at the end of the chain have same oxidation number of +3 and the interior carbon atoms have oxidation number of +2.
Table 3.4 Bond energies in PFOA molecule

<table>
<thead>
<tr>
<th>Bond no</th>
<th>Bond</th>
<th>Standard Bond Energy (kcal/mole)</th>
<th>Bond length (pm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>C-F</td>
<td>116</td>
<td>135</td>
</tr>
<tr>
<td>2</td>
<td>C-C</td>
<td>83</td>
<td>154</td>
</tr>
<tr>
<td>3</td>
<td>C=O</td>
<td>179</td>
<td>120</td>
</tr>
<tr>
<td>4</td>
<td>C-OH</td>
<td>110</td>
<td>143</td>
</tr>
<tr>
<td>5</td>
<td>O-H</td>
<td>111</td>
<td>96</td>
</tr>
</tbody>
</table>


Fig 3.4 Oxidation numbers in PFOA chain

It is obvious that the PFOA molecule has positive oxidation numbers except fluorine and oxygen. It has some possible oxidations such as Carbon in the PFOA chain in to +4 (CO₂). This reaction may take place at high temperature and pressure, but not by the strong oxidants at room temperature and pressure.

3.4.2 CPC oxidation by ferrate - reference study
Yong et al (2006) have studied the oxidation of CPC by ferrate. It was the first report to show that ferrate technique can open the pyridine ring and mineralize the aliphatic chain of the organic molecule resulting inorganic ions. They have demonstrated the unique property of ferrate(VI) to degrade almost completely the cationic surfactant, CPC. The decrease in total organic carbon (TOC) from CPC was more than 95%; suggesting mineralization of CPC to carbon dioxide. In order to check the reliability of the methodology adapted for this experiment, a part of ferrate oxidation test for CPC was imitated.
3.4.2.1 Kinetics of TOC reduction by ferrate oxidation.
In this experiment the ferrate to CPC molar ratio (beginning of the experiment) was kept at 3, which ensures the availability of excess ferrate for the reaction during entire reaction time. Yong et al (2006) has reported that the CPC can be completely oxidized by ferrate and this can be explained by the rate law as shown in Eq 3.7(Yong et al., 2006).

\[ d[\text{Fe(VI)}]/dt = k[\text{Fe(VI)}][\text{CPC}]^2 \]  

(3.7)

The reaction has taken only few minutes and molar consumption of ferrate(VI) has been nearly equal to the oxidized CPC. Also they have suspected the mineralization of CPC to carbon dioxide as they observed more than 95% TOC reduction in this experiment.

**Figure 3.5 a and b** show the rate of change of TOC with prevailing TOC concentration in the mixing flask. From our study the kinetic of TOC oxidation can be expressed as (Figure 3.5 a),

\[ d(\text{TOC})/dt = 0.1187x \quad R^2 = 0.8034 \]  

(3.8)

and according to the published data same oxidation can be written as (Figure 3.5 b)

\[ d(\text{TOC})/dt = 0.1249x \quad R^2 = 0.7732 \]  

(3.9)

![Fig 3.5a The changing rate of TOC with remaining TOC level. (This study)](image)

![Fig 3.5b The changing rate of TOC with remaining TOC level (Yong et al (2006))](image)

According to Eq 3.8 and 3.9 the kinetic constant for TOC reduction by ferrate for this study and the previous study were 0.1187 and 0.1249 respectively.
Our reading just deviated by 5% from the published reading and it is quite reasonable to conclude that the methodology adapted in this experiment is acceptable.

3.4.2.2 TOC reduction at the ferrate oxidation of PFOA and CPC

Figure 3.6 shows the percentage TOC reduction with time for PFOA and CPC. It was found that the TOC removal was 14% for PFOA, but for the control test with CPC the TOC reduction was more than 95% after one hour. This totally agrees with the previous results of this experiment, where PFOA was not oxidized by ferrate. Since process of mineralization was not started to reduce PFOA into CO₂, the levels of TOC remain unchanged. The main reason to reduce TOC by 14% in this experiment could be the temporary adsorption of PFCs onto ferrite (III). Since the concentrations used in this experiment was in mg/L level and ferrite to PFOA molar ratio was 3, there is a high possibility for the PFOA to attach with ferrite (III), which have been settled in the centrifuge process, reducing the TOC in the final analysis.

![Fig 3.6 Percentage TOC reduction with time for CPC and PFOA](image)

3.4.3 Ferrate oxidation of a mixture of PFCs

It is expected to understand the ability of ferrate to oxidize other PFCs with this experiment. Also the effect of initial concentration of PFCs on ferrate oxidation was studied in this experiment. Figure 3.7 shows the results of six individual batch tests of experiment No. 2 (table 3.3). It was observed that all PFCs were behaving in a similar pattern in each individual batch test for different PFCs concentrations. In the case of 25 µg/L batch experiment, molar ratio of 100 gave the best performance of about 15% reduction of overall PFCs and the performance was reduced as the molar ratio was...
increased. Similar pattern was noticed for the 10 µg/L batch experiment. For the 5µg/L batch experiment, ideal molar ratio was 200 which also gave about 15% overall PFCs reduction. For the concentrations of 0.5µg/L and 0.1µg/L, ideal molar ratio was 100 for one set of PFCs and 200 for another set of PFCs. High ferrate(VI) to PFC concentration seems to be discouraging PFCs removal. The reason may be coagulation effect of ferrate(VI), which may flocculate ferrite(III) particles reducing the adsorption process.

Fig 3.7  Ferrate(VI) degradation of PFCs with different initial concentrations and molar ratios

<table>
<thead>
<tr>
<th>Concentration (µg/L)</th>
<th>Ferrate(VI)/PFC Molar ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0</td>
</tr>
</tbody>
</table>

|$\Delta$PFPeA(C5)  □PFHpA(C7)  ◇PFOA(C8)  ○PFDA(C10)  ■PFHS(C6)  ●PFOS (C8)
Table 3.5 shows the summarized results of ferrate oxidation of PFCs. We tested different reaction combinations of initial PFCs concentrations and molar ferrate to PFCs molar ratios. The maximum PFCs reduction among all the combinations studied was 31.26%, which was given by PFHxA with molar ratio 100 and initial PFHxA concentration of 20 µg/L. It was observed in this experiment that the percentage reduction of PFCs with sulfonate functional group is lower than that of PFCs with acid functional group. Higher polarity of the acid functional group may be the reason for this phenomenon. It is suspected that the main removal mechanism is adsorption rather than oxidation which is encouraged by higher surface charges. This result totally agrees with the previous experiment done for PFCs oxidation with other oxidants. Yong Qiu et al. (2007) reported that PFCs cannot be decomposed by O₃/H₂O₂. Occurrence of PFCs in drinking water is an example of ineffectiveness for PFCs oxidation by the common oxidants used in water purification process.
3.5 Summary
Ferrate(VI) technique has been identified as an emerging water purification technique and has proved its effectiveness to treat many organic and inorganic pollutants even at trace level. We tested the ferrate technique to oxidize PFCs at trace concentrations (from 0.1 to 25µg/L).

1. The best ferrate to PFCs molar ratio to reach maximum PFCs reduction was identified as 100.

2. It was noticed that the average elimination of PFCs with sulfonane functional group is 18% and acid functional group is 24%.

3. PFCs reduction by ferrate(VI) technique might occur due to adsorption than oxidation. It was also found that high ferrate(VI) to PFC molar ratio discouraged PFC removal. Coagulation property of ferrate(VI), which flocculate ferrite(III) particles might be reducing the adsorption effect.

4. It was concluded that ferrate technique along is not sufficient to oxidize PFCs at environmental pressure and temperature.

5. Since the ferrate(VI) can oxidize many pollutants in water except PFCs, it might be useful in methodology development in PFCs determination, especially to degrade organic matrices in wastewater samples.
4.1 Introduction
In 1970s, synthetic resin sorbents became available as an alternative to GAC. One of the major advantages of these resins over GAC is their on-site regeneratability through steam stripping, solvent extraction, or microwave irradiation. The ability to regenerate resins on-site potentially offers economic advantages over GAC, which typically requires off-site high-temperature incineration for regeneration. In addition, because the manufacturing process of synthetic resins is a controlled process, resins can have specially designed functional groups and pore size ranges that can be manipulated to optimize their performance for specific applications. Currently, synthetic resin sorbents are used in a variety of industrial applications such as purification processes in the food and drug industry, odor control, and industrial wastewater treatment. They have also been used in a number of groundwater remediation applications (e.g., removal of halogenated organic compounds) and landfill leachate purification (Melin, 1999)

The higher unit costs of resins compared to the more traditional sorbent GAC has been mainly responsible for the limited applications of resin sorbents. The recent developments have suggested that synthetic resin sorbents may be economically competitive with other more established treatment technologies (AOPs, and GAC) for removing some organic compounds.

Improvements in resin regeneration processes such as steam regeneration, solvent regeneration, and microwave regeneration may make the life cycle cost of a resin system competitive or, perhaps, more economical than other options. Also the resins (unlike AOPs) do not produce oxidation by-products. The effectiveness of resin sorbents for PFCs removal is evaluated in this chapter.

4.1.1. Process principle
Synthetic resins, like the widely used GAC, rely on sorption processes to remove organic compounds from water. In the context of water treatment, the process of interest generally involves the sorption of contaminants, such as certain ions or organic compounds from water by porous solid or semi-solid sorbent particles. The sorption of compounds by such sorbents occurs because of two primary driving forces: the hydrophobic character of the sorbate and/or the high affinity of the sorbate for the sorbent. For the majority of systems encountered in water and wastewater treatment systems, sorption results from the net effect of the combined interaction of these two driving forces (Weber, 1972). The hydrophobicity of a compound is inversely proportional to its solubility in water. An extremely hydrophobic compound has a low aqueous solubility and, thus, may prefer to adsorb onto
a solid surface or into an amorphous matrix rather than remain surrounded by water molecules. On the other hand, hydrophilic compounds tend to be stable in aqueous solutions and will leave the solution only if the sorbent provides an attractive force sufficient to overcome the strong bonds between the compound and water molecules. This attractive force, or affinity of the compound for the sorbent, can result from physical or chemical mechanisms. Physical mechanisms include dipole-dipole interactions and Vander-Waals interactions. This intermolecular interaction results in a net attraction between the two molecules (Montgomery, 1985). When two neutral molecules that lack permanent dipoles approach each other, a weak polarization is induced in each because of quantum mechanical interactions between their charge distributions. As a general rule, Vander-Waals interactions increase with increasing size or surface area of the molecules involved (Schwarzenbach et al., 1993). Vander-Waals interactions are generally weaker than dipole-dipole interactions.

4.1.2 Synthetic Polymer Production and application

Non ion-exchange resins and ion-exchange resins are manufactured through very similar processes and are often created from the same base material or polymer backbone. The main difference between the two products is that ion-exchange resins contain charged functional groups, which can form chemical bonds with ions in the solution while non ion-exchange resins rely on physical or non-ionic interactions to remove contaminants from water. In addition to their chemical composition, resins are differentiated on the basis of their pore size distributions. As a matter of convention, micropores are defined as pores less than 20 angstroms (Å) (2x10⁻⁹ m) in diameter, mesopores are between 20 to 500 Å (2x10⁻⁹ to 5x10⁻⁸ m) in diameter, and macropores have diameters greater than 500 Å. Synthetic resins generally have a more controlled and even distribution of pore sizes than GAC. In order to be useful for sorptive applications in water treatment, resins have an extensive network of micropores, similar to GAC, which creates high surface areas and abundant sorption sites.

To produce superior kinetics over GAC, resins are also designed with a significant percentage of pores in the mesopore and macropore size range, which can provide access to the inner surfaces of resins (Melin, 1999). Similar to GAC, granular synthetic resins also can be used in fixed bed filters, particularly to eliminate organic pollutants. In the real applications, the process flow configuration of synthetic resin systems is very similar to that of GAC systems. The main difference between two systems is the provision for a regeneration process for resin systems as resins can be regenerated on-site (Melin, 1999). Sorbent columns can be operated in series, in parallel, or as a combination of the two configurations depending on a number of factors, including the need for
continuous operation, space constraints, effluent criteria, service cycle time constraints, operation logistics, and requirements for multi-barrier treatment. (Weston, 1995). However, the higher unit costs of resins compared to the more traditional sorbent GAC has been mainly responsible for the limited applications of resin sorbents in drinking water treatment scenarios (Melin, 1999).

4.1.3 Non ion-exchange Resins
Non ion-exchange polymeric resins are typically based on cross-linked polymers having polystyrene, phenolformaldehyde, or acrylate matrices (Faust and Aly, 1998). Most commercial macroporous polymeric sorbents are based on polystyrene-divinylbenzene copolymers (Neely, 1982) in which the divinylbenzene serves as a cross-linking agent that makes the styrene insoluble and confers physical strength to the resin (DeSilva, 1995). Although they can be based on the same matrices, polymeric resins differ from traditional ion-exchange resins in their lack of ionic functional groups.

4.1.4 Ion-exchange resins
4.1.4.1 Fundamentals of Ion-Exchange
Ion-exchange is the reversible interchange of ions between a solid (ion exchange material) and a liquid in which there is no permanent change in the structure of the solid. Ion-exchange is used in water treatment and also provides a method of separation in many non-water processes. It has special utility in chemical synthesis, medical research, food processing, mining, agriculture and a variety of other areas.

Ion-exchange occurs in a variety of substances and it has been used on an industrial basis since 1910 with the introduction of water softening using synthetic zeolites. Sulfonated coal, developed for industrial water treatment, was the first ion-exchange material that was stable at low pH. The introduction of synthetic organic ion-exchange resins in 1935 resulted from the synthesis (Adams et al., 1935) of phenolic condensation products containing either sulfonic or amine groups which could be used for the reversible exchange of cations or anions. A variety of functional groups have been added to the condensation or addition polymers used as the backbone structures. Porosity and particle size have been controlled by conditions of polymerization and uniform particle size manufacturing technology. Physical and chemical stability have been modified and improved. As a result of these advances, the inorganic exchangers (mineral, greensand and zeolites) have been almost completely displaced by the resinous types except for some analytical and specialized applications. Synthetic zeolites are still used as molecular sieves.
4.1.4.2 Cation Exchange Resins
Weak acid cation exchange resins are based primarily an acrylic or methacrylic acid that has been crosslinked with a di-functional monomer (usually divinylbenzene [DVB]). The manufacturing process may start with the ester of the acid in suspension polymerization followed by hydrolysis of the resulting product to produce the functional acid group. Weak acid resins have a high affinity for the hydrogen ion and are therefore easily regenerated with strong acids.

4.1.4.2 Anion Exchange Resins.
Strong base anion resins are classed as Type 1 and Type 2. Type 1 is the reaction of trimethylamine with the copolymer after chloromethylation. The Type 1 functional group is the most strongly basic functional group available and has the greatest affinity for the weak acids such as silicic acid and carbonic acid, which are commonly present during a water demineralization process. However, the efficiency of regeneration of the resin to the hydroxide form is somewhat lower, particularly when the resin is exhausted with monovalent anions, such as chloride and nitrate. The regeneration efficiency of a Type 2 resin is considerably greater than that of Type 1. Type 2 functionality is obtained by the reaction of the styrene-DVB copolymer with dimethylethanolamine. This quaternary amine has lower basicity than that of the Type 1 resin, yet it is high enough to remove the weak acid anions for most applications. The chemical stability of the Type 2 resins is not as good as that of the Type 1 resins, the Type 1 resins being favored for high temperature applications (Wheaton and Lefevre 2000).

4.1.4.3 Resins Capacity
Ion exchange capacity may be expressed in a number of ways. Total capacity, i.e., the total number of sites available for exchange, is normally determined after converting the resin by chemical regeneration techniques to a given ionic form. The ion is then chemically removed from a measured quantity of the resin and quantitatively determined in solution by conventional analytical methods. Total capacity is expressed on a dry weight, wet weight or wet volume. The ion exchange capacities relevant with the resins used in this study is tabulated in Table 4.2.

4.2 Objectives
The aim of this study is to investigate on different potential granular materials to identify optimum adsorbent to eliminate PFCs. Two ion-exchange resins and four non ion-exchange resins were tested. The specific objectives of the study explained in this chapter are listed below
1. To optimize the methodology for bottle point experiment.
2. To determine the PFCs sorption capacities of ion-exchange polymers, non ion-exchange polymers and activated carbon by fitting the bottle point experimental data into a suitable sorption model.
3. To determine the PFCs sorption kinetic characteristics of ion-exchange polymers, non ion-exchange polymers and activated carbon.
4. To check the suitability of new materials with actual wastewater

4.3 Experimental Description
4.3.1 Materials

Four kinds of non ion-exchange polymers, two kinds of ion-exchange polymers and one kind of GAC were studied in details. Some physical properties of these materials are shown in the Tables 4.1, 4.2 and 4.3. All granular materials were purchased from WAKO Company (Japan). Almost all the granular materials are specifically produced for water and wastewater treatment particularly to eliminate organic compounds in water.

<table>
<thead>
<tr>
<th>Table 4.1</th>
<th>Physical properties of non ion-exchange polymers</th>
</tr>
</thead>
<tbody>
<tr>
<td>Adsorbent</td>
<td>Matrix*</td>
</tr>
<tr>
<td>Dow V493</td>
<td>Styrene-DVB, macroporous</td>
</tr>
<tr>
<td></td>
<td>1025 (m²/g)</td>
</tr>
<tr>
<td>Dow L493</td>
<td>Styrene-DVB, macroporous</td>
</tr>
<tr>
<td></td>
<td>1100 (m²/g)</td>
</tr>
<tr>
<td>Dow V503</td>
<td>Styrene-DVB, macroporous</td>
</tr>
<tr>
<td></td>
<td>2003 (m²/g)</td>
</tr>
<tr>
<td>Amb XAD 4</td>
<td>Macroceticular crosslinked aromatic polymer</td>
</tr>
<tr>
<td></td>
<td>&gt;750 (m²/g)</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th>Adsorbent</th>
<th>Diameter (mm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dow V493</td>
<td>0.8</td>
</tr>
<tr>
<td>Dow L493</td>
<td>0.8</td>
</tr>
<tr>
<td>Dow V503</td>
<td>0.85–1.1</td>
</tr>
<tr>
<td>Amb XAD 4</td>
<td>0.35–1.15</td>
</tr>
</tbody>
</table>

*Provided by the supplier

<table>
<thead>
<tr>
<th>Table 4.2</th>
<th>Physical properties of ion-exchange polymers</th>
</tr>
</thead>
<tbody>
<tr>
<td>Adsorbent</td>
<td>Matrix</td>
</tr>
<tr>
<td>Amb IRA-400</td>
<td>Styrene-DVB</td>
</tr>
<tr>
<td>Dow MarathonA</td>
<td>Styrene-DVB</td>
</tr>
</tbody>
</table>
Table 4.3 Physical properties of GAC (Filtersorb 400, coal based)

<table>
<thead>
<tr>
<th>Property</th>
<th>Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total pore volume</td>
<td>0.61 cm$^3$/g</td>
</tr>
<tr>
<td>Macropores (&gt;500 A)</td>
<td>0.04 cm$^3$/g</td>
</tr>
<tr>
<td>Mesopores (20-500 A)</td>
<td>0.09 cm$^3$/g</td>
</tr>
<tr>
<td>Micropores (&lt;20A)</td>
<td>0.48 cm$^3$/g</td>
</tr>
<tr>
<td>Surface area</td>
<td>900-1100 m$^2$/g</td>
</tr>
<tr>
<td>Matrix</td>
<td>Stacked layers of fused hexagonal ring of C atoms</td>
</tr>
<tr>
<td>Diameter</td>
<td>0.25-0.50 mm</td>
</tr>
</tbody>
</table>

4.3.2 Material cleaning - Synthetic polymer materials.
Prior to the use in the sorption experiment, the resins were first washed in deionized water to remove dirt and then dried at 50°C (Qiang Yu, 2009). After draining, the materials were washed with MeOH (LC/MC grade) to ensure the materials are free of PFCs. Then the materials are again washed with deionized water and dried at 50°C till the weight is constant. Dried materials are stored in airtight container to prevent contamination of prepared material from external PFCs and moistures.

Above procedure was applied for both ion-exchange polymers of Dow MarathonA, Amb IRA-400 and non ion-exchange polymers of Dow V493, Dow L493 and Dow V503. The process was little changed for gel type non ion-exchange polymer of AMB XAD-4 as it was observed that the sorption capacity of this polymer is drastically reduced by drying at 50°C and the drying temperature was changed to 30°C. In order to make the correction for the moisture content at the material comparison, a separate experiment was carried out to determine the moisture content of AMB XAD4 at 50°C.

4.3.3 Material cleaning - Cleaning of GAC.
The GAC were first boiled in deionized water for one hour to remove fine particles and preloaded organics. Floating impurities and grease were readily removed from water surface during boiling. Right after boiling, GAC was flushed with abundant pure water to cool and clean carbons. After intensive washing, carbons were submerged in pure water and stored in room temperature overnight to equilibrate surface properties. The water was discarded and GAC was moved into an oven and dried up at 105°C for around two days to completely remove moisture inside pores (Kimura 2007). In order to obtain GAC in required diameters, dried GAC was firstly pulverized in a mortar by a pestle. Then the crushed GAC was separated by sieving in required ranges of diameters. The fine particles or powders attached on the surface of crushed GAC during pulverization should be
removed before experiments. Therefore, sieved GAC was washed again by deionized water, and used water with fine particles is poured out carefully. The washing step was repeated for several times until the water over GAC granules was clean and clear. The washed GAC was again dried in oven at 105°C for around two days and stored in airtight bottles. Benefited from former cleaning process, granular materials can be precisely weighed before experiments. After weighing, dried granular materials were moved into PP sampling valves (2mL) and soaked in small amount of pure water (1mL) to pre-wet inner pores by applying a vacuum for 24 hrs. This step is important and necessary because air inside pores might adversely reduce adsorption capacity and velocity (Nicholas and Paul 1993). After pre-wetted, excessive water was discarded by pipette and the materials are added to the bottles which were filled with PFCs solutions.

4.3.4 Preparation of PFCs solutions

PFCs solutions were prepared from stock solutions by diluting them in MilliQ water or wastewater matrix. Since the working maximum PFCs concentration was not exceeding 10μMol/L, pH was not significantly changed (6.4 – 6.9) and did not add any buffer solutions to control pH. Mainly two types of PFCs solutions were prepared for the batch experiment. First kind of PFCs solutions were prepared by dissolving individual PFCs in MilliQ water to determine isotherm and kinetic characteristics of each granular materials tested in this study. A series of individual PFCs solution with different concentration was prepared by adding required volume of stock solution in to MilliQ water and thoroughly mixing it (manual mixing and ultra-sonication).

The applicability of the real wastewater was examined in this study with batch test experiment with effluent wastewater from a PFCs related industry. The effluent wastewater was directly applied for the test without any pre preparation. The PFCs concentration of the wastewater was not changed by spiking PFCs solutions.

4.4 Experimental methods

4.4.1 Experimental apparatus

The batch test experiment was carried in a thermo-stat shaker with shaking speed of 140 rpm at 25°C. In the shaker, the prepared sample bottles were horizontally packed and clamped to the shaking basin. The arrangement is schematically shown in Figure 4.1. The volume of the sample bottle was 125mL and the screw type lid tightly closed the bottle. The temperature in the bottle was controlled by controlling the water bath temperature at 25°C.
4.4.2 Experimental conditions

Three series of batch experiments were carried out to meet the objectives with the apparatus shown in Figure 4.1. Details of each batch test are explained by following paragraphs.

![Diagram of batch test experiment](image)

Fig 4.1 Schematic diagram of batch test experiment to understand isotherm and kinetic characteristics of different granular materials

4.4.2.1 Isotherm experiments with different changing parameters

In the isotherm experiment sorption capacity was calculated with respect to different equilibrium concentrations. The shaking duration was 100 hrs and it was ensured that all the granular materials reached their equilibrium concentration within this shaking time. Different equilibrium concentrations could be reached by two methods, i.e. by changing initial sorbate concentration, while keeping the sorbent concentration constant and by changing the sorbent concentration, while keeping the sorbate concentration constant.

Sorption isotherm of GAC was tested by both methods and the experimental conditions are listed in Table 4.4.

4.4.2.2 Batch test to determine isotherm characteristics of individual PFCs

Adsorption isotherm characteristics of individual PFCs on different granular materials were studied by constant sorbent concentration method. For the known PFCs of PFOA, PFHxA and PFOS, ion-exchange, non ion-exchange and GAC were tested and other selected PFCs only non ion-exchange and GAC materials were tested. Table 4.5 shows the individual combinations studied in this experiment. The conditions of the experiment are shown in Table 4.6.
Table 4.4 Experiment condition for sorption isotherm experiment

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Constant sorbate concentration</th>
<th>Constant sorbent concentration</th>
</tr>
</thead>
<tbody>
<tr>
<td>Adsorbate concentration (μg/L)</td>
<td>2000</td>
<td>30, 60, 120, 250, 800, 2000, 5000</td>
</tr>
<tr>
<td>Selected adsorbent</td>
<td>F-400 (GAC)</td>
<td>F-400 (GAC)</td>
</tr>
<tr>
<td>Selected adsorbutes</td>
<td>PFOA, FFHpA</td>
<td>PFOA, FFHpA</td>
</tr>
<tr>
<td>Adsorbant concentration (g/L)</td>
<td>0.05, 0.1, 0.3, 0.8, 1.0, 1.5, 2.0</td>
<td>1± 0.05</td>
</tr>
<tr>
<td>Shaking time (hrs)</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>Volume (mL)</td>
<td>100</td>
<td>100</td>
</tr>
<tr>
<td>Temperature (°C)</td>
<td>25</td>
<td>25</td>
</tr>
<tr>
<td>Shaking speed (rpm)</td>
<td>150</td>
<td>150</td>
</tr>
<tr>
<td>pH</td>
<td>6.4&lt; pH &lt; 6.6</td>
<td></td>
</tr>
</tbody>
</table>

4.4.2.3 Batch test to examine the applicability of new materials to real wastewater

Treated wastewater was collected from the wastewater treatment plant in a PFCs related industry. This wastewater has been identified as one of a major PFCs polluter in Okayama prefecture (Okamoto, 2010). It was measured that the level of POFA in this wastewater is more than 4 μg/L. Since the level of PFCs in this wastewater was kept unchanged the sorbent concentration was changed in this experiment to get different equilibrium concentrations. The experimental conditions are tabulated in Table 4.7.

Table 4.5 Different sorbent-sorbate combinations considered for individual batch experiments to determine the characteristics of sorption isotherms kinetics

<table>
<thead>
<tr>
<th></th>
<th>PFBA</th>
<th>PFHxA</th>
<th>PFHpA</th>
<th>PFOA</th>
<th>PFDA</th>
<th>PFOS</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dow V493</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>Dow L493</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>Dow V503</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>Dow V503</td>
<td>√</td>
<td></td>
<td></td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>Amb XAD 4</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
<tr>
<td>Amb IRA-400</td>
<td>√</td>
<td></td>
<td></td>
<td>√</td>
<td></td>
<td>√</td>
</tr>
<tr>
<td>Dow MarathonA</td>
<td>√</td>
<td>√</td>
<td></td>
<td>√</td>
<td></td>
<td>√</td>
</tr>
<tr>
<td>F400(GAC)</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
<td>√</td>
</tr>
</tbody>
</table>
Table 4.6 Experiment condition for sorbent-sorbate combination (Table 4.4) for the sorption isotherm experiment-2

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Set value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Adsorbate concentration (μg/L)</td>
<td>10, 30, 60, 120, 250, 500, 1000, 2000, 5000</td>
</tr>
<tr>
<td>Shaking time (hrs)</td>
<td>100</td>
</tr>
<tr>
<td>Volume (mL)</td>
<td>100</td>
</tr>
<tr>
<td>Temperature (°C)</td>
<td>25</td>
</tr>
<tr>
<td>Shaking speed (rpm)</td>
<td>150</td>
</tr>
<tr>
<td>Adsorbant concentration (g/L)</td>
<td>1± 0.05</td>
</tr>
<tr>
<td>pH</td>
<td>6.4&lt; pH &lt; 6.9</td>
</tr>
<tr>
<td>Sampling</td>
<td></td>
</tr>
<tr>
<td>Before shaking (one sample from each bottle)</td>
<td>1*9</td>
</tr>
<tr>
<td>After shaking (two samples from each bottle)</td>
<td>2*9</td>
</tr>
</tbody>
</table>

Table 4.7 Experiment condition for sorption isotherm for industrial wastewater

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Set value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Adsorbate concentration (μg/L)</td>
<td>PFCs related industrial wastewater-PFOA is dominant (≈4 g/L)</td>
</tr>
<tr>
<td>Selected adsorbent</td>
<td>Dow V493, Dow L493, Amb XAD 4, Amb IRA-400, Dow MarathonA, F400(GAC)</td>
</tr>
<tr>
<td>Adsorbant concentration (g/L)</td>
<td>0.01, 0.02, 0.05, 0.08, 0.1</td>
</tr>
<tr>
<td>Shaking time (hrs)</td>
<td>100</td>
</tr>
<tr>
<td>Volume (mL)</td>
<td>100</td>
</tr>
<tr>
<td>Temperature (°C)</td>
<td>25</td>
</tr>
<tr>
<td>Shaking speed (rpm)</td>
<td>150</td>
</tr>
<tr>
<td>pH</td>
<td>7</td>
</tr>
<tr>
<td>Sampling</td>
<td></td>
</tr>
<tr>
<td>Before shaking</td>
<td>1<em>6</em>5</td>
</tr>
<tr>
<td>After shaking</td>
<td>2<em>6</em>5</td>
</tr>
</tbody>
</table>
4.4.2.4 Batch test to determine kinetic characteristics.
A series of experiments were carried out to determine kinetic characteristics for different sorbent-sorbate combinations. Similar to the second experiment of isotherm test, separate batch tests were carried out for each sorbent-sorbate combination as tabulated in Table 4.5. Contrasting to the sorption test, the sampling was done at different time intervals. The experiment conditions are summarized in Table 4.8.

Table 4.8 Experiment condition for sorbent-sorbate combinations (Table 4.5) for the sorption kinetic experiment

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Set value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Adsorbate concentration (µg/L)</td>
<td>500, 5000</td>
</tr>
<tr>
<td>Sampling intervals (hrs)</td>
<td>0, 1.5, 4.5, 21, 72, 100</td>
</tr>
<tr>
<td>Shaking time (hrs)</td>
<td>100</td>
</tr>
<tr>
<td>Volume (mL)</td>
<td>100</td>
</tr>
<tr>
<td>Temperature (°C)</td>
<td>25</td>
</tr>
<tr>
<td>pH</td>
<td>6.4-6.8</td>
</tr>
<tr>
<td>Shaking speed (rpm)</td>
<td>150</td>
</tr>
<tr>
<td>Adsorbant concentration (g/L)</td>
<td>1± 0.05</td>
</tr>
</tbody>
</table>

4.5 Modeling and simulation

4.5.1 Adsorption isotherms models
An adsorption isotherm describes the equilibrium of the sorption of a material at a surface (more general at a surface boundary) at a constant temperature. It represents the amount of material bound at the surface as a function of the material present in the solution. Sorption isotherms are often used as empirical models (Atkin 1998). Yong (2007) has summarized some of these equations as tabulated in Table 4.9.

Even though a number of isotherm equations are suggested based on different assumptions Freundlich equation and Langmuir equation are widely applied in water and wastewater treatment. Especially for the low equilibrium concentrations, Freundlich equation is the only possible isotherm equation to model the experimental data.
Table 4.9 summary of adsorption isotherm models developed with various assumptions

<table>
<thead>
<tr>
<th>Assumption</th>
<th>Name</th>
<th>Equations</th>
<th>Conditions</th>
<th>Comments</th>
</tr>
</thead>
<tbody>
<tr>
<td>Single layer</td>
<td>Generalized Langmuir</td>
<td>( \frac{q}{q_m} = \frac{(Kc)^n}{1 + (Kc)^n} )</td>
<td>( n=m=1 )</td>
<td>Langmuir</td>
</tr>
<tr>
<td></td>
<td>Local site</td>
<td>( \frac{q}{q_m} = \frac{(Kc)^n}{1 + (Kc)^n} )</td>
<td>( 0&lt;n&lt;m&lt;1 )</td>
<td>Langmuir-Freundlich</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>( m=1,0&lt;n&lt;1 )</td>
<td>General. Freundlich</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>( m=1,0&lt;n&lt;1 )</td>
<td>Tóth</td>
</tr>
<tr>
<td>Multilayer</td>
<td>B.E.T.</td>
<td>( \frac{q}{q_m} = 1 - \frac{Kx}{(1-x)} )</td>
<td>( n=1 )</td>
<td>Hütting</td>
</tr>
<tr>
<td></td>
<td>adsorption</td>
<td></td>
<td></td>
<td>Sircar</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Lopez-Gonzalez&amp;Dietz</td>
</tr>
<tr>
<td>Micropore</td>
<td>Dubinin-Astakhov</td>
<td>( q = \exp[-B(RT)^n \ln \frac{C_s}{C}] )</td>
<td>( n=1 )</td>
<td>Freundlich</td>
</tr>
<tr>
<td></td>
<td>filling</td>
<td></td>
<td></td>
<td>Dubinin-Raduskevich</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Exponential Jaroniec</td>
</tr>
<tr>
<td>Empirical</td>
<td>Radke-Prasunitz</td>
<td>( q = \frac{AKc}{1 + (Kc)^n} )</td>
<td></td>
<td>Combine Henry ((q=Kc)) and Freundlich ((q=Kc^m)), better performance than either</td>
</tr>
<tr>
<td></td>
<td>Jossens</td>
<td>( Kc = q \cdot \exp(B \cdot q^m) )</td>
<td>( n=1, \sum )</td>
<td>Exponentially depend on isosteric heat</td>
</tr>
<tr>
<td></td>
<td>Tiemkin</td>
<td>( q = A + B \log c )</td>
<td></td>
<td>Suitable for gas catalyst</td>
</tr>
</tbody>
</table>

Note: \( q \) is adsorbent loading rate, \( q_m \) is maximum loading rate, \( c \) is bulk concentration, \( c_s \) is saturated bulk concentration, \( K, A \) and \( B \) are adsorptive coefficients, \( n, m \) and \( R \) are constants, \( T \) is temperature.

Freundlich equation is an empirical relationship describing the adsorption of solutes from a liquid surface to a solid surface. It is widely applied to describe adsorption process for many compounds on to heterogeneous surfaces, including activated carbon, metals and polymers in dilute solutions (Bruce et al., 1997; Robert et al., 2000; Sirinivasan et al., 2008).

The Freundlich equation is defined by

\[ q_c = K_c c^{1/n} \]

and the equation can be rewritten as

4.1
Log \( q_e \) = Log\( K_f \) + \( \frac{1}{n} \) Log \( C_e \)  \( \text{(4.2)} \)

where \( q_e \) (\( \mu g/g \)) is the concentration in the solid phase, \( C_e \) (\( \mu g/L \)) is the equilibrium concentration of solute in solution, \( K_f \) (\( \mu g/g \)(\( \mu g/L \))\(^{-1/n} \)) is the Freundlich adsorption constant or capacity factor and \( n \) is the Freundlich exponent.

Parameters of \( K_f \) and \( n \) in Freundlich equation were determined by least square fitting of experiment data on \( q \) and \( C_e \), as shown in \textbf{Eq. (4.2)}.

According to the \textbf{Eq 4.1} adsorption capacity of a given sorbent (given sorbent concentration) is a constant for a given equilibrium concentration of a given sorbate (at given temperature).

The Freundlich isotherm is known to be operative only within certain concentration limits. This is because, given an exponential distribution of binding sites, the number of sites increases indefinitely with a decreasing association constant, implying that there are infinite number of sites. But the Freundlich isotherm will be a more accurate approximation at lower concentrations (Robert et al., 2001).

Most of the polymers tested in this study are widely used in water and wastewater treatment, particularly for organics. The adsorptive capacities of tested polymers at low concentrations (1 ng/L – 1 \( \mu g/L \)) were calculated with the Freundlich constants determined by the experiment.

In the case of unit equilibrium concentration (in this study \( C_e = 1 \mu g/L \)) the effect of power \( n \) can be neglected and the amount of PFCs adsorbed in to the polymer at this equilibrium concentration is given by \( K_f \).

### 4.5.2 Kinetics models

To further understand the sorption kinetics, the pseudo second-order model was selected to fit the kinetic data. The model assumes that the sorption rate is controlled by chemical sorption and the sorption capacity is proportional to the number of active sites on the sorbent (McKay 1999).

The concept is mathematically expressed in the \textbf{Eq 4.3}.

\[
\frac{dq_e}{dt} \propto (q_e - q_t)^2 \quad \text{(4.3)}
\]

or

\[
\frac{dq_e}{dt} = k_2 (q_e - q_t)^2 \quad \text{(4.4)}
\]

Where \( q_e \) and \( q_t \) are the sorption capacity at equilibrium and at time \( t \), respectively and \( k_2 \) is the rate constant of the pseudo-second order sorption.

For the boundary conditions \( t = 0 \) to \( t = t \) and \( q_t = 0 \) to \( q_t = q_t \), the integrated
form of Eq 4.4 becomes:

\[
\frac{1}{q_e - q_t} = \frac{1}{q_e} + k_2 t
\]

Which is the integrated rate law for a pseudo-second order reaction. Eq 4.5 can be rearranged to obtain

\[
q_t = \frac{1}{\frac{1}{k_2q_e^2} + \frac{1}{q_e} + \frac{1}{q_e}}
\]

Which has a linear form:

\[
\frac{1}{q_t} = \frac{1}{k_2q_e^2} + \frac{1}{q_e}
\]

4.6 Results and Discussion

4.6.1 Time required to reach the equilibrium concentration

In the sorption experiment, it is essential to reach all the sorbate in the bottles into the equilibrium concentration with the sorbent added in each bottle. Figure 4.2 shows the variations of material sorption capacities with time for PFOS and PFHxA. It was clearly identified that ion-exchange polymers and GAC reached their equilibrium concentrations much earlier than non-ion-exchange polymers. Since a comparative study was to be done with work, a common shaking duration of 100 hrs was selected. It was ensured with Figure 4.2 that all kinds of granular materials had reached their equilibrium concentrations during this shaking period.

4.6.2 Determination of isotherm characteristics with different methodologies

In order to determine the sorption isotherms, it is required to determine different sorption capacities at different equilibrium concentrations. In the experiment, actual equilibrium concentration was measured and based on which, material adsorption capacity was calculated with known values of initial sorbate and sorbent concentrations.
Different initial concentrations can be achieved by two methods, i.e. by changing initial sorbate concentration while keeping the sorbent concentration constant, or changing sorbent concentration while keeping the sorbate concentration constant. Isotherm characteristics of GAC were checked with both methods and the Freundlich isotherm curves drawn with different methods are shown in Figure 4.3. It was found that the Freundlich curves given by two methodologies were not same. The Freundlich coefficient given by constant sorbate method was 7.10 and same coefficient given by constant sorbent method was 24.51.

The experiment was repeated with different PFCs and found that constant sorbent method was better than constant sorbate method for isotherm experiments. It was noticed several times that unrealistic results were given by constant sorbate method. In this experiment, granular materials were shaken for 100 hrs in a 120 mL horizontally mounted PP bottles. It was assumed that the sorbate and sorbents are homogeneously mixed in the bottles during the shaking experiment. But in the real application it was noticed that heavy granular materials were confined into channels along with the hydraulic eddies, which are created by the constant shaking speed.

Fig 4.2 Time requirement for different granular materials to reach equilibrium concentration
4.6.3 Isotherm experiment for individual coagulants with individual PFCs

4.6.3.1 Non Ion-Exchange polymers
The values determined for the Freundlich constants are listed in Table 4.10. It was found that isotherms of all non ion-exchange granular medias were nicely fixed with Freundlich isotherm (R²= 0.76–0.99). Freundlich exponent (n) is an indicator of nonlinearity of Freundlich curve. Nonlinearity can occur due to many reasons including the sorption site heterogeneity and sorbate–sorbate interactions (Cheung, 2001). In this study GAC gave comparatively higher nonlinearity. It might be due to the sorption site heterogeneity of GAC is higher than that of synthetic polymers. Table 4.3 shows physical properties of GAC which could be calculated that more than 78% of total pore volume in GAC was occupied by micro pores, 14% meso pores and the rest was occupied by macro pores (Table 4.3). Pores in the GAC with different sizes occurred when it was prepared under controlled pressure and temperature.

Even though pore size distribution is comparatively uniform for synthetic non ion-exchange polymers there is a procedure to add some macro pores to enhance kinetic characteristics. To produce macroporous polymeric resins, the polymerization process was carried out in the presence of an inert material. Small amount of an inert material yields a non-macroporous,
three-dimensional network while a high inert material content leads to the formation of microstructures, or nuclei. As the polymerization progresses, the nuclei agglomerate and crosslink to form microspheres. Aggregates of microspheres form irregularly shaped particles, which constitute the resin beads (Malley et al., 1993). The pore size distribution of polymeric resins can also be controlled during their manufacture by varying the amount of extender used in the polymerization reaction: this governs the degree of cross-linking and the ultimate pore structure created (Weber and van Vliet, 1981).

In the case of unit equilibrium concentration (in this study $C_e = 1 \, \mu g/L$) the effect of power $n$ can be neglected and the amount of PFCs adsorbed into the polymer at this equilibrium concentration is given by $K_f$. It was noticed that $K_f$ was increased with the length of the carbon chain for all adsorbents suggesting longer chain PFCs can be easily adsorbed. Same results have been obtained in a previous research (Yong, 2007). However in actual situation PFCs in discharge water should be in ng/L level. Calculated $q_c$ values for 3 different equilibrium concentrations (1, 10 and 100 ng/L) with Freundlich equation are tabulated in Table 4.10. It was interesting to notice that synthetic polymer materials show better sorption capacities than GAC at low equilibrium concentrations. This might be the reason of failure for application of GAC to eliminate PFCs in German drinking water treatment plants (Schaefer, 2006).

Figure 4.4 shows the PFDA adsorption curves for synthetic non ion-exchange polymers and GAC. For the long chain PFCs of PFDA, highest $K_f$ was given by Amb XAD 4 (2965) followed by Dow L493 (434.51), Dow V493 (415.91) and GAC (121.89).
**Table 4.10** Freundlich constants for different sorbent/sorbate combinations and calculated $q_e$ for different $C_e$(ng/L) based on $K_f$ and n values

<table>
<thead>
<tr>
<th>Adsorbate</th>
<th>Adsorbent</th>
<th>Freundich Isotherm parameter</th>
<th>$q_e$&lt;sup&gt;2&lt;/sup&gt;</th>
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</thead>
<tbody>
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<td></td>
<td></td>
<td>$K_f$&lt;sup&gt;1&lt;/sup&gt;</td>
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</tr>
<tr>
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<td></td>
<td>Dowexoptopore V503</td>
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<td>Dowexoptopore L493</td>
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<tr>
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<td>Amberlite XAD 4</td>
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<td>Filtrasorb 400 (GAC)</td>
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<td>PFHxA</td>
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<td>176.36</td>
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<td>Amberlite XAD 4</td>
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<td></td>
<td>Filtrasorb 400 (GAC)</td>
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<td>PFHpA</td>
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<td>1.174</td>
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<tr>
<td></td>
<td>Dowexoptopore V503</td>
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</tr>
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<td>Dowexoptopore L493</td>
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</tr>
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</tr>
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<td></td>
<td>Filtrasorb 400 (GAC)</td>
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<td>1.924</td>
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<td>PFNA</td>
<td>Dowexoptopore V493</td>
<td>998.45</td>
<td>0.404</td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore L493</td>
<td>208.26</td>
<td>0.500</td>
</tr>
<tr>
<td></td>
<td>Amberlite XAD 4</td>
<td>554.50</td>
<td>0.541</td>
</tr>
<tr>
<td></td>
<td>Filtrasorb 400 (GAC)</td>
<td>602.42</td>
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</tr>
<tr>
<td>PFOA</td>
<td>Dowexoptopore V493</td>
<td>415.91</td>
<td>2.619</td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore V503</td>
<td>8.43</td>
<td>0.926</td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore L493</td>
<td>434.51</td>
<td>0.371</td>
</tr>
<tr>
<td></td>
<td>Amberlite XAD 4</td>
<td>2965.00</td>
<td>1.418</td>
</tr>
<tr>
<td></td>
<td>Filtrasorb 400 (GAC)</td>
<td>121.89</td>
<td>0.968</td>
</tr>
</tbody>
</table>

<sup>1</sup>Freundlich isotherm constants [(μg PFCs/g sorbent)(μg PFCs/L)<sup>n</sup>]

<sup>2</sup>PFCs adsorbed on the adsorbent at equilibrium at equilibrium concentrations of 1,10 and 100 ng/L (μg/g)

<sup>3</sup>Equilibrium concentration (ng/L)
PFDA adsorption by non ion-exchange polymers
But in further low concentrations of equilibrium concentration Dow L493 performed better than other polymers.

PFOA adsorption by non ion-exchange polymers
Three kinds of synthetic non ion-exchange resins (Dow V493, Dow L493, Amb XAD4 and F400 (GAC) were selected for this experiment. In the experiment, PFOA was identified as easily adsorbed PFCs like PFDA. Sorption capacity at unit equilibrium concentration of 1 μg/L, GAC showed higher sorption capacity of 602 μg/g which was the second best performance after Dow V493. But it was noticed that the sorption capacity at further low equilibrium concentrations of 100 ng/L and 10 ng/L of Amb XAD 4 showed better performances than GAC. **Figure 4.5** shows the variation of sorption capacity of each granular materials at equilibrium concentrations.
PFHpA adsorption by non ion-exchange polymers

PFHpA adsorption by four kind of synthetic non ion-exchange resins of Dow V493, Dow L493, Dow V503 and amb XAD4 and GAC were tested and Figure 4.6 shows the adsorption isotherms determined for them. With the comparison of capacity factors determined for PFDA and PFOA, clear reduction was observed. Amb XAD4 was identified as the best candidate material to eliminate PFHpA from water.
PFHxA adsorption by non ion-exchange polymers
PFHxA dominates as a problematic PFCs, because some industries use it as an alternative for PFOS/PFOA. Three kind of synthetic non ion-exchange resins of Dow V493, Dow L493, Amb XAD4 and F400 (GAC) were selected for this experiment. GAC clearly showed the best sorption capacity at unit equilibrium concentration of 1 µg/L, but it was noticed that the other non ion-exchange polymers of Dow V493 and Amb XAD4 showed better sorption capacities at further low equilibrium concentrations. Ion-exchange resins showed the best performance for PFHxA elimination and it will be discussed later in this chapter.

**Figure 4.7** shows the variation of sorption capacity of each granular material at equilibrium concentrations.

![Figure 4.7 Adsorption isotherm of PFHxA onto GAC(F400 • ▲), and non ion-exchange polymers (AmbXAD4 • ■, Dow L493 □, Dow V493 • ○)](image)

PFBA adsorption by non ion-exchange polymers
PFBA is the shortest PFCs tested in this chapter. It is said that the level of toxicity for PFBA is not high as other long chain and medium chain PFCs because the retention time of PFBA is much lower than that of long chain and medium chain PFCs (verbal communication with US EPA). For this batch experiment, four kind of synthetic non ion-exchange resins of Dow V493, Dow L493, Dow V503, Amb XAD4 and F400 (GAC) were selected. The sorption capacities determined by the results of the experiment clearly showed that the only possible non ion-exchange granular material to be
used for PFBA is GAC. This was valid even for the less equilibrium concentrations of 100ng/L, 10ng/L and 1 ng/L. **Figure 4.8** shows the variation of sorption capacities of each granular material at equilibrium concentrations.

**4.6.3.2 Adsorption by ion-exchange polymers**

**Figure 4.9** and **4.10** show the isotherm curves of Amb IRA 400 and Dow Marathon A drawn for PFHxA and PFOA. Same materials were tested also with PFOS and the results are discussed under PFOS adsorption. The results of the experiment were modeled with Freundlich isotherm and the determined coefficients are shown in **Table 4.11**.

Comparing the same coefficient determined for GAC and non ion-exchange polymers, ion-exchange polymers showed excellent adsorption capacities for both PFCs at any equilibrium considered in the experiment. For PFOA adsorption, both ion-exchanges showed over 5000 µg/g of sorption capacity and unit equilibrium concentration, which is ten times higher than that of non ion-exchange polymers including GAC. Dow Marathon A showed the best performance to eliminate PFHxA.
Table 4.11 Freundlich constants for different sorbent/ sorbate combinations and calculated $q_e$ for different $C_e$(ng/L) based on $K_f$ and n values

<table>
<thead>
<tr>
<th>Adsorbate</th>
<th>Adsorbent</th>
<th>Freundlich Isotherm parameter</th>
<th>$q_e$ $^2$</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>$K_f$$^1$ n $R^2$ $C_e$$^2$</td>
<td>$C_e=1$ $C_e=10$ $C_e=100$</td>
</tr>
<tr>
<td>PFH A</td>
<td>Amb IRA-400</td>
<td>340.96 0.836 0.94</td>
<td>1.059</td>
</tr>
<tr>
<td></td>
<td>Dow MarathonA</td>
<td>503.5 0.810 0.94</td>
<td>1.871</td>
</tr>
<tr>
<td>PFOA</td>
<td>Amb IRA-400</td>
<td>5154.23 0.712 0.73</td>
<td>37.633</td>
</tr>
<tr>
<td></td>
<td>Dow MarathonA</td>
<td>6356.00 1.487 0.99</td>
<td>0.219</td>
</tr>
</tbody>
</table>

$^1$Freundlich isotherm constants [(µg PFCs/g sorbent)(µg PFCs/L) $^n$]

$^2$PFCs adsorbed on the adsorbent at equilibrium at equilibrium concentrations of 1, 10 and 100 ng/L (µg/g)

$^3$Equilibrium concentration (ng/L)

Fig 4.9 Adsorption isotherm of PFOA onto ion-exchange polymers (Amb IRA400 · △, Dow MarathonA·○)
4.6.3.3 PFOS adsorption

PFOS is the latest POPs categorized by the Stockholm Convention in May 2009. Many researchers work on the treatment of PFOS with different strategies and adsorption has been identified as a possible method. Ion-exchange resins, non ion-exchange resins and GAC were tested for PFOS adsorption and the isotherm curves drawn onto various adsorbents separately. The determined Freundlich constants are listed in Table 4.12. The shaking duration of isotherm experiment is 100 hrs and which ensures that all filter materials have reached their equilibrium concentrations. It was found that isotherms of all granular medias were nicely fixed with Freundlich isotherm ($R^2=0.92 - 0.99$). Similar to the other PFCs in this chapter, Freundlich equation was used to calculate PFOS adsorption capacities onto various filter materials at further low equilibrium concentration of 100ng/L. Some possible errors can be expected with this calculation. Even though it is assumed in the calculation that all the materials follow Freundlich equation, practically all data points do not coincide with Freundlich curve ($R^2≠1$). Also it should be emphasized that the minimum equilibrium concentration received in this experiment is 150ng/L.

4.6.3.3.1 Adsorption of PFOS onto Granular Activated Carbon

Granular activated carbons are commonly used adsorbent materials to eliminate organic compounds in water. It has been identified that F400 (coal based) is the best type of GAC to eliminate PFOS (Valeria et al., 2008).
Same type of GAC was used in this experiment with the physical properties as tabulated in Table 4.3.

In this study too, GAC gave comparatively a higher Freundlich exponent \((n)\) than other polymer materials. \(n\) is an indicator of nonlinearity of Freundlich curve. Nonlinearity can occur due to many reasons including the sorption site heterogeneity and sorbate–sorbate interactions (Cheung, 2001). The reason for higher \(n\) for GAC may be the sorption site heterogeneity of GAC, which is higher than that of synthetic polymers. More than 78% total pore volume in GAC is occupied by micro pores, 14% meso pores and the rest is occupied by macro pores (Table 4.3).

With the isotherm experiment, it was determined that the PFOS adsorption of GAC (F400) at unit equilibrium concentration (1 \(\mu\)g/L) is 28.4 \(\mu\)g/g (Figure 4.11). Valeria et al. (2008) reported that the amount of PFOS adsorbed onto GAC (F400) at unit equilibrium concentration (mg/L) is 25.9 mg/g. According to the available literature data, 2.6 \(\mu\)g/g of Methyl Tertiary Butyl Ether (MTBE) detected in GRC-22 (coconut based GAC) at 1 \(\mu\)g/L of equilibrium concentration (Melin 1999). Hydrophobicity of PFOS than MTBE may be the reason to have higher \(K_f\) than MTBE.

Among the six materials tested in this study, GAC showed lowest sorption capacities at 1 \(\mu\)g/L (\(K_f\)) and 0.1 \(\mu\)g/L equilibrium concentrations suggesting its incapability at low equilibrium concentrations (< 1 \(\mu\)g/L). It has been reported that the application of GAC to eliminate PFOS in German drinking water treatment plants was not effective (Schaefer, 2006).
4.6.3.3.2 Adsorption of PFOS onto ion-exchange polymers

PFOS is a fully fluorinated compound, which commonly appear as a salt with a functional group of sulfonate attached at the end of the molecule. The carbon–fluorine bond length is shorter than any other carbon–halogen bond, and shorter than carbon–nitrogen and carbon–oxygen bonds. Short bond length and the high electronegativity of fluorine give the carbon–fluorine bond a significant polarity/dipole moment. The electron density is concentrated around the fluorine, leaving the carbon relatively electron poor. This introduces ionic character to the bond through partial charges (Cδ+—Fδ−) (O’Hagan, 2008). Negatively charged molecular saturate of PFOS suggests higher adsorption onto anion-exchange polymers.

Among the all polymers tested in this experiment, anion-exchange polymers of Amb IRA-400 and Dow MarathonA showed the highest Kᵢ of 108.8 and 95.9 (µg/g)(µg/L)^-n respectively (Figure 4.12). Some characteristics of these polymers are tabulated in Table 4.2. Quang et al. (2008) investigated Amb IRA 400 for PFOS adsorption and reported that Kᵢ is 250 for high equilibrium concentration of 1 mg/L.

![Adsorption isotherm of PFOS onto ion-exchange polymers(Amb IRA400 - △, Dow MarathonA-◇)](image)

Among the two ion exchange polymers tested, Dow MarathonA showed smaller n (1.68) giving flatter Frundich curve than Amb IRA-400 (n =2.8), which indicated better performance of Dow MarathonA at equilibrium concentrations in ng/L level. Calculated adsorption capacity for Dow Marathon400 is 2.01 µg/g whereas for Amb IRA-400 it is 0.9 µg/g.
4.6.3.3.3 Adsorption of PFOS onto non ion-exchange polymers

A significant aspect of the non ion-exchange polymer adsorption is that, the bonding forces between the adsorbent and the adsorbate are usually weaker than those encountered in activated carbon adsorption. Regeneration of the resin can be accomplished by simple, nondestructive means, such as solvent washing, thus providing the potential for solute recovery (Busca et al., 2008; Lin and Juang, 2009).

Dow L493, Dow V493 and Amb XAD 4 were tested for PFOS adsorption. Amb XAD4 and Dow V493 showed sorption capacities of 79.1 µg/g and 81.3 µg/g respectively at unit equilibrium concentration (1 µg/L) (Figure 4.13). These values are approximately in the range of 80% of sorption capacity determined for ion-exchange polymers and 250% of sorption capacity determined for GAC at 1 µg/L equilibrium concentration. Distribution of homogeneous pore size in the surface of synthetic non-ion exchange polymers may be the reason for higher Kf than GAC. Also it was noticed that the adsorption capacities of Dow V493 and Dow L493 at 100 ng/L (calculated values) were better than ion-exchange polymers. Comparing with ion-exchange polymers and GAC, adsorption capacity of non ion-exchange polymers showed a linear (comparatively) correlation to equilibrium concentration (n = 0.84, 1.61 and 0.94) (Table 4.12).

Fig 4.13 Adsorption isotherm of PFOS onto non ion-exchange polymers (AmbXAD4 • , Dow L493☐, Dow V493 ○)
Freundlich isotherm constants \[ \left( \frac{\mu g \text{ PF}_{Cs}}{g \text{ sorbent}} \right) \left( \frac{\mu g \text{ PF}_{Cs}}{L} \right)^{-n} \]

\[ q_{adsorbed \ on \ the \ adsorbent \ at \ equilibrium \ at \ equilibrium \ concentrations \ of \ 1,10 \ and \ 100 \ ng/L} \]

Table 4.12 Freundlich constants for PFOS with different adsorbents and calculated \( q_c \) for different \( C_e (ng/L) \) based on \( K_f \) and \( n \) values

<table>
<thead>
<tr>
<th>Adsorbate</th>
<th>Adsorbent</th>
<th>Freundlich Isotherm parameter</th>
<th>( q_c^{*2} )</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>( K_f )</td>
<td>( 1/n )</td>
<td>( R^2 )</td>
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<td>Dowexoptopore L493</td>
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<td>0.84</td>
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<td>Amberlite XAD 4</td>
<td>79.10</td>
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<td>Filtrasorb 400 (GAC)</td>
<td>28.40</td>
<td>2.20</td>
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<td></td>
<td>Amb IRA-400</td>
<td>95.90</td>
<td>1.68</td>
</tr>
<tr>
<td></td>
<td>Dow MarathonA</td>
<td>28.40</td>
<td>2.20</td>
</tr>
</tbody>
</table>

*1 Freundlich isotherm constants \([(\mu g \text{ PFCs/g sorbent})(\mu g \text{ PFCs/L})^{-n}]\)

*2 PFCs adsorbed on the adsorbent at equilibrium at equilibrium concentrations of 1,10 and 100 ng/L (\(\mu g/g\))

**4.6.3.3 Sorption kinetics of PFOS**

Figure 4.14 shows the amount of PFOS adsorption onto each adsorbent with time (with 5000\(\mu g/L\) initial concentration). Two ion-exchange polymers and GAC reached the equilibrium concentration within 4 hours.

Non ion-exchange polymer materials of Amb XAD4 reached it within 10 hours and Dow L493 and Dow V493 took 90 hours to reach the equilibrium concentration.

To further understand the sorption kinetics, the pseudo second-order model (Equation 4.7) was selected to fit the kinetic data. It was observed that our kinetic data of non ion-exchange polymers fixed well with the equation 2 \((R^2 = 0.99-1.00)\) suggesting the dominancy of the chemisorption at adsorption process.

Since ion-exchange polymers and GAC reached the equilibrium concentration within a very short time (< 4 h), the number of kinetic data was not sufficient to check with pseudo second-order kinetic model.
4.6.4 Adsorption kinetics of non ion-exchange polymers

Figure 4.15 shows the sorption kinetics for three PFCs on the selected adsorbents for two initial concentrations of 500 µg/L and 5,000 µg/L. We observed that our kinetic data fixed well with the pseudo second order kinetic model (Equation 4.7) (\(R^2=0.89\) to 1.00) suggesting the dominancy of the chemisorption at adsorption processes. Table 4.13 shows the \(k\) and the \(q_e\) values calculated with the initial concentrations of 5000 µg/L. Chemisorption is based on functional chemical group interactions. The principal difference between physical sorption and chemisorption is that the former is less specific with respect to which compounds sorb to which surface sites, has weaker forces and lower energies of bonding, and operates over longer distances between sorbate molecules and sorbent sites (Montgomery, 1985). In chemisorption, the attraction between sorbent and sorbate approaches by a covalent or electrostatic chemical bond between atoms, with shorter bond length and higher bond energy.

The sorption capacities observed in the kinetic experiment supported the results of isotherm experiment. The sorption capacities were increased with the length of the carbon chain of PFCs.
Sorbates bound by chemisorption to a surface generally cannot accumulate more than one molecular layer (monolayer) because of the specificity of the bond between the sorbate and the surface. The location of the sorption sites tends to be very specific since only certain functional groups on a sorbate molecule are able to form these chemical bonds. In general, synthetic resins designed as sorbents for organic compounds have lower densities and varieties of chemical functional groups than activated sorbents such as GAC, and sorbate interactions with their surfaces are, thus, primarily through physical, rather than chemical, mechanisms. Another important phenomenon observed in the kinetic experiment is the time required to reach equilibrium concentration for non ion-exchange polymers, which increased with the length of the carbon chain. This may be due to the lack of macro pores and meso pores in synthetic polymer surface which deaccelerates the access of long carbon PFCs in to inner microspores and ultimately it requires longer time to reach equilibrium concentration. Kinetic characteristics of PFCs on selected adsorbents were also examined with a low concentration of 500 μg/L. Medium chain (PFHpA and PFOS) and long chain (PFDA) PFCs well obeyed with the Eq 2 but for PFBA only GAC and Dow V493 behaved according to the kinetic model at low concentration of 500 μg/L. The graphs marked with b, d, and f of Figure 4.15 Show adsorption kinetics of PFCs with initial concentration of 500 μg/L and graphs marked with a, c and e show the results with 5,000 μg/L. It was obvious that an increase in initial PFCs concentration leads to an increase in adsorption capacity of PFCs on polymers. It was observed that GAC reached the equilibrium concentration within 4 hrs whereas Amb XAD 4 took less than 10 hrs for all PFCs except PFOS. Other polymers took more than 80 hrs to reach the equilibrium concentrations.
Fig. 4.15 Adsorption kinetics (for initial concentrations 500μg/L and 5000μg/L) for the PFCs of (a) PFBA 5000μg/L, (b) PFBA 500μg/L, (c) PFHpA 5000μg/L, (d) PFHpA 500μg/L, (e) PFDA 5000μg/L, (f) PFDA 500μg/L, and modeling using the pseudo second-order kinetic equation.
**Table 4.13** Pseudo-second-order kinetic parameters for different non ion-exchange sorbent/ sorbate combinations

<table>
<thead>
<tr>
<th>Adsorbate</th>
<th>Adsorbent</th>
<th>Pseudo-second-order kinetic parameter</th>
</tr>
</thead>
<tbody>
<tr>
<td>PFBA</td>
<td>Dowexoptopore V493</td>
<td>q_e^*2 13.7  k^*1 0.091  R^2 0.998</td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore V503</td>
<td>7.3 - 0.998</td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore L493</td>
<td>8.1 - 0.994</td>
</tr>
<tr>
<td></td>
<td>Amberlite XAD 4</td>
<td>10.4 1.024  0.998</td>
</tr>
<tr>
<td></td>
<td>Filtrasorb 400 (GAC)</td>
<td>27.0 1.369  1.000</td>
</tr>
<tr>
<td>PFHpA</td>
<td>Dowexoptopore V493</td>
<td>31.0 0.008  0.997</td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore V503</td>
<td>7.5 0.020  0.953</td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore L493</td>
<td>21.5 0.009  0.992</td>
</tr>
<tr>
<td></td>
<td>Amberlite XAD 4</td>
<td>19.7 0.214  1.000</td>
</tr>
<tr>
<td></td>
<td>Filtrasorb 400 (GAC)</td>
<td>14.37 0.457  0.999</td>
</tr>
<tr>
<td>PFDA</td>
<td>Dowexoptopore V493</td>
<td>66.6 0.001  0.983</td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore V503</td>
<td>30.3 0.001  0.885</td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore L493</td>
<td>41.6 0.003  0.995</td>
</tr>
<tr>
<td></td>
<td>Amberlite XAD 4</td>
<td>38.4 0.039  1.000</td>
</tr>
<tr>
<td></td>
<td>Filtrasorb 400 (GAC)</td>
<td>72.9 16.900  1.000</td>
</tr>
</tbody>
</table>

^*1Sorption rate constant \( [g \text{ sorbent}(mg \text{ PFCs})^{-1}(h)^{-1}] \)

^*2PFCs adsorbed on the adsorbent at equilibrium \( (mg/g) \)

---

![Solid phase concentration (mg PFCs/g sorbent)](image)

**Fig 4.16** Adsorption Kinetics of PFHxA and PFOA onto GAC(F400 - ▲), ion-exchange polymers(Amb IRA400 - △, Dow MarathonA- ◇) and non ion-exchange polymers (AmbXAD4 - ■, Dow L493-□, Dow V493 - ○ and pseudo second-order kinetic curve —
4.6.5 Adsorption kinetics of ion-exchange polymers

Figure 4.16 shows the variation of adsorbed PFCs concentration with time. Comparing with non ion-exchange polymers, adsorption kinetics for ion-exchange polymers and GAC were high. Among the materials tested AmbIRA 400 (ion-exchange resin) gave the highest sorption constant (k) for all three PFCs tested (Table 4.14). Both GAC and Dow MarathonA showed similar kinetic profiles.

Table 4.14 Pseudo-second-order kinetic parameters of PFHxA, PFOA and PFOS with different ion-exchange, non ion-exchange and GAC

<table>
<thead>
<tr>
<th>Adsorbate</th>
<th>Adsorbent</th>
<th>Pseudo-second-order kinetic parameter</th>
<th>q_e</th>
<th>k</th>
<th>R²</th>
</tr>
</thead>
<tbody>
<tr>
<td>PFHxA</td>
<td>Dowexoptopore V493</td>
<td>21.097</td>
<td>0.032</td>
<td>0.99</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore L493</td>
<td>13.831</td>
<td>0.014</td>
<td>0.98</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Amberlite XAD 4</td>
<td>16.340</td>
<td>0.052</td>
<td>0.99</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Filtrasorb 400 (GAC)</td>
<td>22.321</td>
<td>2.230</td>
<td>1.00</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Amb IRA-400</td>
<td>21.231</td>
<td>2.773</td>
<td>1.00</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Dow MarathonA</td>
<td>22.173</td>
<td>1.071</td>
<td>1.00</td>
<td></td>
</tr>
<tr>
<td>PFOA</td>
<td>Dowexoptopore V493</td>
<td>91.743</td>
<td>0.004</td>
<td>0.99</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore L493</td>
<td>74.074</td>
<td>0.012</td>
<td>0.99</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Amberlite XAD 4</td>
<td>88.495</td>
<td>0.008</td>
<td>0.99</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Filtrasorb 400 (GAC)</td>
<td>90.09</td>
<td>0.136</td>
<td>1.00</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Amb IRA-400</td>
<td>86.206</td>
<td>0.448</td>
<td>1.00</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Dow MarathonA</td>
<td>93.457</td>
<td>0.286</td>
<td>1.00</td>
<td></td>
</tr>
<tr>
<td>PFOS</td>
<td>Dowexoptopore V493</td>
<td>37.736</td>
<td>0.009</td>
<td>0.99</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore L493</td>
<td>38.462</td>
<td>0.010</td>
<td>0.99</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Amberlite XAD 4</td>
<td>41.322</td>
<td>0.063</td>
<td>1.00</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Filtrasorb 400 (GAC)</td>
<td>38.168</td>
<td>2.288</td>
<td>1.00</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Amb IRA-400</td>
<td>38.760</td>
<td>13.313</td>
<td>1.00</td>
<td></td>
</tr>
<tr>
<td></td>
<td>Dow MarathonA</td>
<td>39.216</td>
<td>8.128</td>
<td>1.00</td>
<td></td>
</tr>
</tbody>
</table>

*Sorption rate constant [g sorbent(mg PFCs)^(-1)(h)^(-1)]

*PFCs adsorbed on the adsorbent at equilibrium(mg/g)

4.6.6 Batch test with PFCs related industrial wastewater

Characteristics of wastewater

Collected wastewater from a discharge point of PFCs related industrial wastewater treatment plant was initially analyzed and found that pH was
6.9 and the TOC was 15.2 mg/L. The initial PFCs concentration of the wastewater was measured as shown in Table 4.15.

Table 4.15 Initial PFCs concentration of the wastewater

<table>
<thead>
<tr>
<th>Date</th>
<th>PFOA (µg/L)</th>
<th>PFOS (µg/L)</th>
<th>PFHxA (µg/L)</th>
<th>PFHpA (µg/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>2009-4</td>
<td>0.72</td>
<td>-</td>
<td>1.80</td>
<td>0.25</td>
</tr>
<tr>
<td>2009-5</td>
<td>2.60</td>
<td>-</td>
<td>4.20</td>
<td>1.40</td>
</tr>
<tr>
<td>2009-09</td>
<td>10.00</td>
<td>-</td>
<td>2.90</td>
<td>2.20</td>
</tr>
<tr>
<td>2010-02</td>
<td>4.35</td>
<td>0.02</td>
<td>0.75</td>
<td>0.33</td>
</tr>
</tbody>
</table>

Contrasting with individual PFCs in MilliQ water, it was difficult to get a good isotherm relationship with this experiment ($0.079 < R^2 < 0.99$) (Figure 4.17). The isotherm coefficient determined by the experiment and the calculated sorption capacities at further low concentrations of 1, 10 and 100 ng/L is tabulated in Table 4.16. It was observed in the experiment that AmbXAD4 can effectively remove PFOA, but negative isotherm coefficients were received.

According to the batch test results actual wastewater from PFCs related industry, GAC seems better for unit equilibrium concentration of 1 µg/L. For the low equilibrium concentrations (10 – 1 ng/L) synthetic resins seem to be more attractive than GAC in terms of sorption capacity. Especially DOW L493 gave best sorption capacity at ng/L level equilibrium concentrations.

The amount of research carried out with actual wastewater is not enough to come to a concrete solution, but the process of adsorption was identified as a possible method to eliminate PFCs in this wastewater. Long run column experiment is to be carried out with earmarked candidate materials and actual wastewater to understand the process clearly.
Fig 4.17 Adsorption isotherm of PFOA occurred in PFCs related industrial WW onto ion-exchange polymers, non ion-exchange polymers and GAC.
Table 4.16 adsorption characteristics of different filter materials with PFCs related industrial wastewater

<table>
<thead>
<tr>
<th>Adsorbate</th>
<th>Adsorbent</th>
<th>Freundich Isotherm parameter</th>
<th>$q_{c^2}$</th>
<th>$C_{e^3}$</th>
<th>$C_{e^3}$</th>
<th>$C_{e^3}$</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>$K_f^{*1}$</td>
<td>n</td>
<td>$R^2$</td>
<td>1</td>
<td>10</td>
</tr>
<tr>
<td>PFHxA</td>
<td>Dowexoptopore V493</td>
<td>9.795</td>
<td>0.809</td>
<td>0.616</td>
<td>0.037</td>
<td>0.236</td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore L493</td>
<td>22.999</td>
<td>3.225</td>
<td>0.817</td>
<td>0.000</td>
<td>0.000</td>
</tr>
<tr>
<td></td>
<td>Amberlite XAD 4</td>
<td>3.831</td>
<td>0.329</td>
<td>0.303</td>
<td>0.395</td>
<td>0.842</td>
</tr>
<tr>
<td></td>
<td>Filtrasorb 400 (GAC)</td>
<td>3147.748</td>
<td>1.266</td>
<td>0.838</td>
<td>0.503</td>
<td>9.264</td>
</tr>
<tr>
<td></td>
<td>Amb IRA-400</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Dow MarathonA</td>
<td>4.756</td>
<td>0.650</td>
<td>0.125</td>
<td>0.054</td>
<td>0.239</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>PFA</td>
<td>Dowexoptopore V493</td>
<td>7.658</td>
<td>0.371</td>
<td>0.970</td>
<td>0.591</td>
<td>1.388</td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore L493</td>
<td>5.829</td>
<td>0.285</td>
<td>0.658</td>
<td>0.815</td>
<td>1.570</td>
</tr>
<tr>
<td></td>
<td>Amberlite XAD 4</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Filtrasorb 400 (GAC)</td>
<td>204.644</td>
<td>0.842</td>
<td>0.787</td>
<td>0.608</td>
<td>4.229</td>
</tr>
<tr>
<td></td>
<td>Amb IRA-400</td>
<td>25.375</td>
<td>1.209</td>
<td>0.956</td>
<td>0.006</td>
<td>0.097</td>
</tr>
<tr>
<td></td>
<td>Dow MarathonA</td>
<td>15.470</td>
<td>0.927</td>
<td>0.724</td>
<td>0.026</td>
<td>0.216</td>
</tr>
<tr>
<td>PFOA</td>
<td>Dowexoptopore V493</td>
<td>38.265</td>
<td>0.227</td>
<td>0.900</td>
<td>7.976</td>
<td>13.452</td>
</tr>
<tr>
<td></td>
<td>Dowexoptopore L493</td>
<td>41.870</td>
<td>0.141</td>
<td>0.870</td>
<td>15.809</td>
<td>21.873</td>
</tr>
<tr>
<td></td>
<td>Amberlite XAD 4</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Filtrasorb 400 (GAC)</td>
<td>57.783</td>
<td>0.234</td>
<td>0.880</td>
<td>11.476</td>
<td>19.670</td>
</tr>
<tr>
<td></td>
<td>Amb IRA-400</td>
<td>7.204</td>
<td>1.124</td>
<td>0.990</td>
<td>0.003</td>
<td>0.041</td>
</tr>
<tr>
<td></td>
<td>Dow MarathonA</td>
<td>15.188</td>
<td>1.002</td>
<td>0.800</td>
<td>0.015</td>
<td>0.150</td>
</tr>
</tbody>
</table>

*1 Freundlich isotherm constants [(μg PFCs/g sorbent)(μg PFCs/L)]
*2 PFAs adsorbed on the adsorbent at equilibrium at equilibrium concentrations of 1, 10 and 100 ng/L (μg/g)
*3 Equilibrium concentration (ng/L)
4.7 Conclusion

1. Constant sorbent method was identified as the best method to determine sorption isotherm, where sorbent concentration is kept constant and sorbate concentration is changed.

2. Ion-exchange resins and GAC showed faster adsorption characteristics than non ion-exchange polymers. To reach the equilibrium concentration for all kind of granular materials, at least 100 hrs shaking is needed.

3. Synthetic polymer materials were identified as better filter materials (in terms of adsorption capacity) to eliminate PFCs in water at low concentration (1 μg/L). The magnitude of $K_f$ decreases in the following order for most of long chain and medium chain PFCs tested: Ion-exchange polymers > Non ion-exchange polymers > GAC, but some cases at further low equilibrium concentration (100 ng/L) non ion-exchange polymers showed higher adsorption capacity than other adsorbents.

4. Amb IRA-400 was identified as the best filter material to eliminate PFOS at equilibrium concentration > 1 μg/L. considering both adsorption isotherm and adsorption kinetics, Amb XAD 4 and Dow MarathonA can be recommended for eliminating PFOS at ng/L equilibrium concentrations.

5. Further studies are to be done to understand the sorption mechanisms of PFOS, but it can be suspected that the chemisorption is the dominant process.
Chapter 5 - PFCs coagulation

5.1 Introduction

Coagulation can be defined as the reactions and mechanisms involved in the chemical destabilization of particles in the formation of large particles through perikinetic flocculation. Coagulant is the chemical that is added to destabilize the colloidal particle in water so that floc formation can result. (Wastewater Engineering, Treatment and Reuser – Metcalf & Eddy). In the coagulation/flocculation process, very fine suspended particles are caused to come together to form larger particles that can be settled and filtered out of the water.

During coagulation, the coagulant is added to the water to be treated. The water is stirred vigorously in a fast mixer to assure quick, uniform dispersion of the coagulant. In the conventional treatment method, the coagulant may react rapidly with compounds in the water that contain carbonates, bicarbonates and hydroxides to produce a jelly-like substance that absorbs impurities. At the same time, coagulants can neutralize the charges on the particle surface.

5.1.1 Inorganic Coagulants

Most of the conventional inorganic coagulants are salts of aluminum or iron. These inorganic salts neutralize the charge on the particles causing raw water turbidity, and also hydrolyze to form insoluble precipitates, which entrap particles. Table 5.1 lists few common inorganic coagulants with some physical and chemical properties.

5.1.2 Organic Polymers

Organic Polymers are categorized as natural organic polymers and synthetic organic polymers. In this study, only synthetic polymers are considered. Organic coagulants are used in a wide variety of municipal and industrial applications to enhance the coagulation, flocculation and removal of impurities in water.

Synthetic long chain organic coagulants consist of simple monomers that are polymerized into high molecular weight substances. Depending on their charge, when placed in water, are negative, positive or neutral. These polymers are classified as anionic, cationic and nonionic respectively. The action of polymeric coagulants also classified as charge neutralization, polymer bridge formation and combine charge neutralization-polymer bridge formation process.
5.1.3 Uses and benefits of organic polymers in water Treatment

In general water soluble organic polymers are highly efficient contributors of cationic charge and molecular weight when applied in specific applications. The process of coagulation is made highly efficient by the proper application of chemistry and dosage. Sometimes organic coagulants are used most efficiently in combination with inorganic coagulants such as polyaluminum chloride and other aluminum and iron salts. The use of organic polymers offers several advantages over the use of inorganic coagulants:

1. Less sludge production.
2. The resulting sludge contains a less amount of chemically bound water and can be more easily dewatered.
3. Polymeric coagulants do not affect pH. Therefore, the need for supplemental alkalinity, such as lime, caustic, or soda ash, is reduced or eliminated.

5.1.4 Coagulation of anionic particles by cationic polymer coagulants

The basic reaction expected with anionic particles and cationic polymers is schematically shown in the Figure 5.1. The first expected reaction is the columbic attraction between opposite charge molecule and active site in the organic polymer. It is expected that this reaction is faster at the beginning of the experiment because of the availability of higher concentrations of opposite charges and it will decrease with time when
vacant sites and remaining anions decrease with the progression of the experiment. Also the coagulated polymers can combine with each other and make bigger particles. This bridging may be leading to floc formation.

Apart from the main two reactions mentioned above, there is a possibility for more reactions. Especially in this experiment, PFCs concentrations were in μg/L level, but the coagulant doze is in mg/L level. There is a chance for some coagulant polymers to not to have any PFCs during the coagulation process. In such case the polymer chain can be fold back and settle within the chain or can be attached with other coagulated polymers.

Fig 5.1 A diagram to explain conceptual mechanism of PFCs elimination by organic coagulants

5.2 Aims and Objectives

Major aim of this study is to identify possible candidate coagulants to eliminate PFCs for wastewater treatment process. Both inorganic coagulants and organic coagulants were tested by a series of jar test experiments. Coagulants were tested with both MilliQ water and effluent wastewaters from different WWTPs. The specific objectives of this chapter are listed below.

1. To determine the optimum coagulant doze to eliminate PFCs by jar tests.
2. To compare the performances of different coagulants (inorganic and organic) to coagulate PFCs by jar test.
3. To observe the individual coagulation of each PFCs when it appears as a mixture of PFCs in the initial solution.
4. To check the effect of other organic matters presence in water on PFCs coagulation.
5. To check the possibility of coagulation process to eliminate PFCs in the actual industrial water treatment process.
5.3 Materials and Method

5.3.1 Materials

All cationic organic coagulants (FL2250, FL2749, FL3050, FL4440, FL4620, FL4820) were received from SNS, Inc (Japanese agent). SNF, Inc. is one of the world's leading manufacturers of water-soluble polymeric organic coagulants. Inorganic coagulants (Iron(III) chloride, Sodium aluminate and Aluminum sulfate) were purchased from Wako chemicals (Japan). Some physico chemical properties of tested coagulants are tabulated in the Table 5.2 and Table 5.3

Table 5.2 Physico chemical properties of organic coagulants used in the experiment

<table>
<thead>
<tr>
<th>Coagulant</th>
<th>Specific gravity</th>
<th>Viscosity</th>
<th>Color</th>
<th>Mol Wt/1000</th>
<th>Remarks</th>
</tr>
</thead>
<tbody>
<tr>
<td>FL2250</td>
<td>1.12-1.16</td>
<td>25-35</td>
<td>Clear/ yellow</td>
<td>10</td>
<td>Low molecular weight, highly cationic</td>
</tr>
<tr>
<td>FL2749</td>
<td>-</td>
<td>-</td>
<td>Clear/ off white</td>
<td>120</td>
<td>Medium molecular weight, highly cationic</td>
</tr>
<tr>
<td>FL3050</td>
<td>1.12-1.16</td>
<td>1,000-4,000</td>
<td>Clear/ off white</td>
<td>500</td>
<td>High molecular weight, highly cationic</td>
</tr>
<tr>
<td>FL4440</td>
<td>1.05-1.10</td>
<td>1,000-3,000</td>
<td>Clear/ yellow</td>
<td>3,000</td>
<td>Medium molecular weight, homopolymer of</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>dallyldimethylammonium chloride.</td>
</tr>
<tr>
<td>FL4620</td>
<td>1.02-1.07</td>
<td>700-1,000</td>
<td>Clear/ off white</td>
<td>1,400</td>
<td>High molecular weight, homopolymer of</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>dallyldimethylammonium chloride.</td>
</tr>
<tr>
<td>FL4820</td>
<td>1.02-1.07</td>
<td>1000-3000</td>
<td>Clear/ off white</td>
<td>1,800</td>
<td>High molecular weight, homopolymer of</td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>dallyldimethylammonium chloride.</td>
</tr>
</tbody>
</table>
Table 5.3 Physico chemical properties of inorganic coagulants used in the experiment

<table>
<thead>
<tr>
<th>Coagulant</th>
<th>Mol wt</th>
<th>Molecular structure</th>
<th>Color</th>
<th>Physical form</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ferric Chloride</td>
<td>162.20</td>
<td>Fe(Cl)$_3$</td>
<td>Shining black</td>
<td>Granular</td>
</tr>
<tr>
<td>Sodium Aluminate</td>
<td>163.936 (without water)</td>
<td>Na$_2$Al$_2$O$_4$</td>
<td>White</td>
<td>Powder</td>
</tr>
<tr>
<td>Aluminium Sulfate</td>
<td>342.15 (without water)</td>
<td>Al$_2$(SO$_4$)$_3$.nH$_2$O (n=14-18)</td>
<td>White</td>
<td>Powder</td>
</tr>
</tbody>
</table>

5.3.2 Jar test

Coagulation and flocculation experiments were performed in a jar test apparatus (JMD6E-Osaka Miyamoto Riken Ind, Japan). Each sample (1.0 L) was rapidly mixed at 80 revolutions per minute (rpm) for 4 min, slowly mixed at 25 rpm for 30 min, and then settled (0 rpm) for 60 min. In the beginning of rapid mixing, inorganic and organic coagulants were added using the mechanism provided with the Jar test apparatus. For the jar test with MilliQ water, 1 mL sample was collected from each jar prior to adding coagulant and was analyzed to determine initial PFC$_S$ level. The dosage of organic polymer coagulant ranged from 0 to 1000 μL/L in the first experiment, which was carried out to decide the optimum coagulant dosage. 200 mg/L (for solid coagulant) or 200 μL/L (for liquid coagulant) was applied for other jar test experiments. Two duplicate jars were used for each condition. At the end of each jar test, the samples were filtered through membrane filters (GF/F; pore size. 0.45μm, and 0.2μm Millipore) and analyzed for remaining PFCs in the filtrate. Each steps involved with the jar test are schematically shown in Figure 5.2.

5.3.3 Methodology - Experiment 1 - Optimum doze

This experiment was carried out to determine the minimum effective organic coagulant doze to eliminate PFCs. One selected PFCs of PFOA was tested with different dozes of selected coagulant of FL2250 by a Jar test. PFOA stock solution was added to MilliQ water to yield the initial PFOA concentration of 10 μg/L and the prepared solution was filled in to 6 jars (1 L each). Five dozes of coagulants ranging from 0 μL/L to 1,000μL/L were added to five jars and the sixth one was kept with zero dose as the control. Then the jar test was conducted as described in Figure 5.2. In the jar test the coagulants were added after three seconds from starting the rapid
mixing by the facility provided by the jar test apparatus. Table 5.4 shows the summary of experimental conditions. Since the MilliQ water was used in this experiment, filtered samples after the jar test were directly analyzed with LC/MS/MS after diluting with acetonitrile to ensure 40% acetonitrile content in the analyzed samples.

Table 5.4 shows the summary of experimental conditions.

5.3.4 Methodology Experiment 2 – Comparison of coagulants

This experiment was designed to study the PFOA coagulation efficiencies by organic and inorganic coagulants. Three inorganic coagulants (Iron(III) chloride, Sodium aluminate and Aluminium sulfate) and six organic coagulants (FL2749, FL3050, FL2250, FL4620, FL4820 and FL4440) were tested with PFOA by a series of jar tests to determine the degree of coagulation. The optimum coagulant dose determined by the experiment 1 was applied in this experiment. The PFOA solution (10 µg/L) was prepared by adding PFOA stock solution into Milli Q to yield required concentration. Similar to the experiment 1, filtered samples were directly analyzed with LC/MS/MS after the dilution with acetonitrile. The experimental conditions of this test are summarized in the Table 5.4.

**Fig 5.2 Basic steps involved with Jar test**
5.3.5 Methodology Experiment 3 - Mixture of PFCs

This experiment was designed to investigate the PFCs coagulation as a mixture. A mixture of eight PFCs, which consist of long chain, medium chain, and short chain PFCs, and also different functional groups of acid and sulfonate was spiked into MilliQ water to prepare the initial solution for the jar test. The rest of experiment was similar to experiment 2. Filtered samples after the jar test were diluted with acetonitrile and analyzed with LC/MS/MS for remaining PFCs. The summary of the experimental conditions is shown in Table 5.4.

5.3.6 Methodology Experiment 4 – Wastewater spiked with PFCs

This experiment was designed to observe the effect of the presence of the organic matters in water on PFCs coagulation. Actual wastewater collected at a discharge point of a WWTP was spiked with a mixture of PFCs stock solution to prepare the initial solution for the jar test. The main steps of the jar test are shown in Figure 5.2. Since wastewater was used in this experiment, remaining organics other than PFCs had to be eliminated before analyze with LC/MS/MS. A measured volume of 50 mL from the filtered samples after jar test was sent through a Precip-C Agri cartridge by using the concentrator (flow rate 5mL/min). Then the cartridge was air dried for 1 hr using a vacuum manifold and then eluted it with organic solvents (methanol 3 mL + acetonitrile 3 mL). Eluted solution was completely dried by a flow of nitrogen gas at 60°C. then dried tube was reconstituted with 40% ACN. Final solution was analyzed and measured using LC/MS/MS.

5.3.7 Methodology Experiment 5 – Coagulation of industrial wastewater

This experiment was designed to check the applicability of inorganic and organic coagulants tested in previous experiments in the PFCs related industrial wastewater. Three inorganic coagulants and six organic coagulants shown in Table 5.3 were tested with the wastewater, which was collected at the discharge point of PFCs related industrial wastewater treatment plant. Collected wastewater samples were stored at 4°C and the temperature was adjusted to 25°C before the experiment.

The jar test was carried out according to the main steps shown in Figure 5.2 since the remaining organics other than PFCs in the wastewater used in this experiment had to be eliminated before analyze with LC/MS/MS. 100 mL of the filtrate from the filtered samples after the jar test was sent
through a Precip-C Agri cartridge by using the concentrator (flow rate 5mL/min). Then the cartridge was air dried for 1 hr using a vacuum manifold and was eluted with organic solvents (methanol 3 mL + acetonitrile 3 mL). Eluted solution was completely dried by a flow of nitrogen gas at 60º C. Then dried samples were reconstituted with 40% ACN solution. Final solution was analyzed and measured using HPLC/MS/MS.

Table 5.4 Summary of the experimental conditions

<table>
<thead>
<tr>
<th>Exp. No</th>
<th>PFCs</th>
<th>Coagulants</th>
<th>Type of water used</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>PFOA</td>
<td>FL2250</td>
<td>MilliQ</td>
</tr>
<tr>
<td>2</td>
<td>PFOA</td>
<td>Iron(III) chloride, Sodium aluminate, Aluminium sulfate FL2749, FL3050, FL2250, FL4620, FL4820 and FL4440.</td>
<td>MilliQ</td>
</tr>
<tr>
<td>3</td>
<td>PFPeA, PFHxA, PFOA, PFNA, PFDA, PFHS, PFOS</td>
<td>Iron(III) chloride, Sodium aluminate, Aluminium sulfate FL2749, FL3050, FL2250, FL4620, FL4820 and FL4440.</td>
<td>MilliQ</td>
</tr>
<tr>
<td>4</td>
<td>PFHxA, PFHpA, PFOA, PFNA, PFDA, PFHS, PFOS</td>
<td>Iron(III) chloride, Sodium aluminate, Aluminium sulfate FL2749, FL3050, FL2250, FL4620, FL4820 and FL4440.</td>
<td>Effluent wastewater</td>
</tr>
<tr>
<td>5</td>
<td>PFPeA, PFHxA, PFHpA, PFOA, PFNA, PFDA, PFOS</td>
<td>Iron(III) chloride, Sodium aluminate, Aluminium sulfate FL2749, FL3050, FL2250, Wastewater</td>
<td>Effluent wastewater</td>
</tr>
</tbody>
</table>

Note. The experiments were carried out at 20 ºC. pH was around 6.5 with the addition of PFCs (10µg/L) and it was not adjusted. The initial TOC level of wastewater was measured as 10.5 mg/L.

5.4 Results and discussion

5.4.1 Optimum dosage of organic coagulants

Figure 5.3 shows the percentage PFOA removal by coagulation plotted against the coagulation dosage. Since the concentration of the target compound (PFOA) is 10 µg/L, higher doses of coagulations were applied to get better results. This is a common situation at low concentration of target polar contaminant.
It was observed that the percentage PFOA removal was drastically increased up to 86% with the coagulant dosage. The corresponding coagulant dose at this point was 200µL/L. The additional dose after 200µL/L seems ineffective as there is no significant improvement in percentage PFOA removal. It was assumed that the effective coagulant dose is 200 µL/L and it was applied for the rest of the experiments in this chapter. Martial Pabon(2007) has reported that he could remove 81% PFOA by coagulation with same coagulant of FL2250 at a dosage of 1.4g/L, but, with different initial PFOA concentration of 280 mg/L.

5.4.2 Experiment 2- Performance of organic and inorganic coagulants

Experiment 2 was designed to understand the possibility of PFCs coagulation by inorganic and organic coagulant. Figure 5.4 shows the percentage removal of PFOA by each organic and inorganic coagulant. Initial PFOA concentration was 10µg/L and the optimum dosage of coagulant (200µL/L) determined by the experiment 1 was applied in this experiment.

Organic coagulants showed better performance than inorganic coagulants to coagulated PFOA. The average coagulation efficiency of organic coagulant was 78%, which is 40% higher than that of inorganic coagulants. Among the organic polymers, FL2749, FL3050 and FL2250 showed more than 80% coagulation efficiency. It was interesting to notice that the last
three organic coagulants with least performance were consisted of homopolymer of diallyldimethylammonium chloride suggesting a mixture of multipolymers are better coagulants for PFCs.

In the case of inorganic coagulants experiment, high coagulant dose of 200 mg/L was applied, which is comparable with the dosage of organic coagulants. It was noticed that Ferric Chloride showed almost 80% PFOA elimination, which dominated among all inorganic coagulants. In the process of coagulation by inorganic coagulants, it is difficult to differentiate coagulation and adsorption. Inorganic coagulants may not fully dissolve in the solution and some PFOA molecules can be adsorbed onto surface of the coagulants especially at the flocculation (slow mixing) process. Filtration was carried out (0.45µm and 0.2µm) to eliminate the particle remain in the solution before analysis for PFOA. Both adsorbed PFOA and coagulated PFOA were eliminated in this step. One explanation for higher performance of Ferric Chloride is that the PFOA component eliminated by adsorption is high, so that the overall percentage of PFOA removal reached 80%.

![Fig 5.4 Percentage PFOA removal with different type of coagulants](image-url)
5.4.3 Experiment 3 - PFCs coagulation as a mixture

Figure 5.5 shows the percentage removal of each PFCs in the mixture by various coagulants. Consolidating the results of experiment 2, organic coagulants showed better performances than conventional inorganic coagulants. Considering three inorganic coagulants and eight PFCs used in the experiment it can be figured out that an average PFCs removal by an inorganic coagulant is about 30% where as for organic coagulants same calculation gives 72%. In the first glance, it can be concluded that the PFCs coagulation efficiency by organic coagulants are double than that of inorganic coagulants for MilliQ water.

It was noticed for both organic and inorganic coagulants that there is no single coagulant to show the maximum performance for all PFCs. Different PFCs had different coagulants for maximum coagulation suggesting case by case consideration at the real application. Among the tested coagulants, FL 2749 was identified as the best material with average overall PFCs removal of 86%. For the inorganic coagulants, Aluminum sulfate was the best candidate with overall PFCs removal of 54% followed by Ferric Chloride with 36% overall removal.

It was clearly identified that short chain PFCs are difficult to coagulate than long chain PFCs. Figure 5.6 shows different PFCs (with different chain lengths) removal by organic and inorganic coagulants. Given removal percentages are average values of organic (six coagulants) and inorganic (two coagulants, Sodium aluminates was not considered because its elimination is almost zero) coagulants.

In this experiment, PFCs were spiked into MilliQ water and it can be assumed that the competition to attach with sites in the coagulant chain is only among PFCs molecules. Long chain PFCs seems to be more effective than short chain PFCs for attachment with active sites in the polymer chain. One possible reason for high efficiency is molecular charge. Longer the carbon chain, higher the number of Fluorine attached and which ultimately increase the negative charge in the molecule, which can beat the weakly charged short chain PFCs at the competition to attach with cationic coagulants. The second possible mechanism is enfishment, in which long chain PFCs can be trapped easily than smaller short chain PFCs.
Fig. 5.5 Percentage reduction of each PFCs by various organic and inorganic coagulants for MilliQ water spiked with a mixture of PFCs
5.4.3.1 PFCs coagulation by organic coagulants

This comparative study clearly showed that the organic coagulants are much effective than conventional inorganic coagulants to eliminate PFCs. Also it should be highlighted that this study was carried out for MilliQ water spiked with a mixture of PFCs solution. Especially for well known PFCs of PFOS and PFOA, some organic coagulants showed more than 98% removal efficiencies. Figure 5.7 shows the average PFCs removal for different molecular weights of coagulants. an organic coagulant (positively charged) with molecular weight around 100,000 seems to be ideal to eliminate PFCs of all chain lengths by coagulation/filtration process.

![Graph showing PFCs removal by coagulation](image)

**Fig. 5.6 PFCs with different chain lengths removal by coagulation**

□ organic coagulation △ inorganic coagulation
5.4.4 Experiment 4 – Coagulation of wastewater spike with a mixture of PFCs

Figure 5.8 shows the percentage removal of each PFC by various organic and inorganic coagulants in wastewater, which has been contaminated with a mixture of PFCs. In the same figure, the results of the experiment 3 have also been shown for the easy comparison. The TOC level is high in experiment 4 than experiment 3 because actual wastewater is used in this experiment. Measured TOC of the wastewater was 10.5 mg/L in this experiment whereas for MilliQ water measured TOC was zero.

It was clearly observed for all organic coagulants that the MilliQ spike with PFCs showed better removal percentage than that of wastewater spike with PFCs for short chain PFCs (PFHxA, PFHpA and PFHS). In case of medium chain and long chain PFCs, it was observed that a slight improvement of PFCs removal with the presence of other organics. These observations can be explained by charges of the PFCs molecules. For short chain PFCs, with less charge (negative) cannot compete with other organics to attach with coagulant polymer, so that their removal efficiencies were adversely affected. In the case of long chain PFCs (PFOS, PFOA and PFDA) with higher molecular charge, PFCs can compete with other organic matter and attach with the coagulant polymers. The process of coagulation for long chain PFCs seems to be accelerated by the presence of the other organic matter in the water. Some of the long chain PFCs may first attach...
with organic matter in the water and then get attached with the coagulant polymer improving the overall removal efficiency.

5.4.5 Experiment 5 – Experiment with real waste water

Actual discharge wastewater from PFCs related industry was tested with organic and inorganic coagulants in this experiment. Collected wastewater was initially analyzed. The pH was 6.9 and the TOC was 15.2 mg/L.

The initial PFCs concentration of the wastewater was measured as shown in Table 5.5.

Table 5.5 Levels of PFCs in the wastewater used in this study

<table>
<thead>
<tr>
<th>Month of analysis</th>
<th>PFOA μg/L</th>
<th>PFOS μg/L</th>
<th>PFHxA μg/L</th>
<th>PFHpA μg/L</th>
</tr>
</thead>
<tbody>
<tr>
<td>2009-4</td>
<td>0.72</td>
<td>-</td>
<td>1.80</td>
<td>0.25</td>
</tr>
<tr>
<td>2009-5</td>
<td>2.60</td>
<td>-</td>
<td>4.20</td>
<td>1.40</td>
</tr>
<tr>
<td>2009-09</td>
<td>10.00</td>
<td>-</td>
<td>2.90</td>
<td>2.20</td>
</tr>
<tr>
<td>2010-02 (This study)</td>
<td>4.35</td>
<td>0.02</td>
<td>0.75</td>
<td>0.33</td>
</tr>
</tbody>
</table>

Figure 5.9 shows removal percentage of each PFC by inorganic and organic coagulants. It should be emphasized that this wastewater is mainly polluted with PFOA (Table 5.5). Whatever the treatment technique applied in this wastewater must be essentially reduce the effluent PFOA level.

It was observed in the previous experiments in this chapter that the average PFCs removal by coagulation was more than 70%. Particularly in the experiments 3 and 4, the removal by some organic coagulants was more than 90%. But this experiment gave different results and none of the coagulants except ferric chloride reached 90% removal efficiency. Especially in PFOA removal, ferric chloride gave the best performance with 91% of removal efficiency followed by FL 2749, FL 3050 with removal efficiencies of 67% and 66% respectively. This observation clearly indicated that the behavior of PFCs in a solution of wastewater spiked with PFCs stock solution and the real wastewater from a PFCs related industry is not same. Another important observation is that the coagulation of short chain PFCs did not show much difference for MilliQ water spiked with PFCs, wastewater spiked with PFCs or actual wastewater from PFCs related industries.
PFCs, particularly long chain PFCs appeared in industrial wastewater seems to be attached to other polar particles. Most probably the negatively charged long chain PFCs may attach with positively charge matters appeared in the industrial wastewater as it has enough mixing and retention time before collecting wastewater sample. The story is different for wastewater spiked with PFCs stock solution, because the retention time is minimum and mixing is provided only to get homogenous solution. This explanation is further consolidated by the results of inorganic coagulants.

Fig. 5.8 Percentage reduction of each PFCs by various organic and inorganic coagulants for □ MilliQ water spiked with a mixture of PFCs ■ industrial wastewater ▼ spiked with a mixture of PFCs
Fig 5.9 Percentage reduction of each PFCs by various organic and inorganic coagulants for actual PFCs related industrial wastewater.
5.4.6 The role of filtration in the treatment process.
The filtration process was introduced after the Jar test experiment to simulate the actual treatment process for eliminating flocs and remaining coagulants to make sure the samples was free from suspended particles to analyze with LC/MS/MS. Each sample was filtered at least two times before analysis. First filtration was done at the end of the jar test (after 30 min of settling time) with 0.45μm syringe filters and the filtrate was again filtered by 0.2μm syringe filters. The control experiment carried out without coagulants showed that this pore size cannot filter PFCs. The results of experiment 3 and 4 of this chapter suggest that the positively charged organic coagulates drastically improve the efficiency of PFCs filtration by micro filtration (0.2μm). Further studies to be done to determine the optimum pore size to prevent PFCs by combine coagulation- micro filtration process.

5.4.7 Possible removal mechanism of organic coagulants for PFCs spiked waters
Once a polymeric coagulant was added to a solution the first expected process is mixing. In the mixing process the polymer becomes distributed evenly throughout the solution. The mixing was achieved in this experiment by rapid mixing at 80 rpm for 3 min. Comparatively low mixing speed was selected as there is a possibility to break some long chain polymers with a high degree of turbulence (Sikora et al., 1981). Since concentrated polymer solutions are quite viscous, mixing usually becomes easier, and flocculation more effective, with more dilute dosing solutions (Gregogy et al., 1991). Mixing effects are generally more important for more concentrated suspensions. Figure 5.10.a schematically shows mixing process.
The second possible process is the attachment of polymers with the charged particles in the solution. The rate of attachment mainly depends on their concentrations and it follows the Smoluchowski kinetics (Gregory et al., 1988). This process is schematically shown in Figure 5.10.b. According to the literature data, the dose of optimum polymeric coagulant and adsorption rate are proportional to particle concentration (Bolto et al., 2001). The PFCs concentration in the experiment was 10 μg/L and it was noticed that the longer slow mixing process showed better PFCs removal. Also we noticed that the optimum dose of polymeric coagulant for the elimination of PFCs at tracer level concentration is comparatively high (200 μL/L).
The third identified process is rearrangement of adsorbed chains (Figure 5.10.c). The polymer chain reaches its equilibrium adsorbed configuration with a characteristic distribution of loops, trains and tails. According to the literature data, high mw polymers take several seconds to reach equilibrium concentration (Pelssers et al., 1990), during which, the adsorbed polymers
have more extended configuration and form bridging contacts. Since the coagulant dose applied in this experiment is 250 μL/L, TOC level in wastewater was less than 10 mg/L and PFCs concentration was 10 μg/L, there was an unbalanced concentrations of opposite charges in the solution and total charge neutralization in polymer coagulants could not be achieved. Also there is a possibility for charge pockets or patch mechanism; it occurs when high charge density cationic polymers adsorb negative surfaces with a fairly low density of charged sites (Kasper, 1971). There is a possibility for some polymers to remain without attaching to a surface or a PFCs.

The final expected process is flocculation, mainly by bridging mechanism (Figure 5.10.d). Flocculation is a second order rate process, so that the rate depends on the square of the particle concentration. The mechanism of charge-neutralisation/ precipitation has been proposed for the removal of humic substances by cationic polymers. In many cases (Galser et al., 1979) it has been shown that mw has little or no effect, indicating that polymer bridging is not a significant mechanism. But in this experiment we identified the polymer bridging is important than charge neutralization and we identified the optimum mw is around 100,000 for best PFCs elimination.

![Fig. 5.10 Schematic diagram to explain possible PFCs coagulation mechanism. (a) Mixing (b) attachment (c) rearrangement (d) flocculation](image-url)
5.5 Conclusion

Following conclusions were derived from the results of this experiment.

1. The experiment with deionized water spiked with PFCs suggested that comparatively higher coagulant dose is required to get a good PFCs removal efficiency by organic coagulants.
2. The results of the experiments with deionized water spiked with PFCs and wastewater spike with PFCs showed that the efficiency of PFCs coagulation by organic coagulant is almost double than that of inorganic coagulants.
3. Among the organic coagulants tested, FL 2749 was identified as the best candidate to eliminate any PFCs.
4. The results of PFCs coagulation by different organic coagulants suggest that a mixture of multipolymers is better than single polymer coagulants to coagulate PFCs.
5. The results of the experiment with actual wastewater spiked with PFCs indicated that occurrence of other organic matters discourages coagulation of short chain PFCs, but it encourages the coagulation of long chain PFCs.
6. The results of the experiment with actual PFCs related industrial wastewater indicated that organic coagulants are not effective as it showed in wastewater spiked with PFCs to coagulate PFCs. The PFCs appear in real wastewater seems to be incorporated with other polar molecules in the wastewater.
7. Organic coagulation followed by microfiltration seems to be an effective combination to eliminate PFCs for some wastewaters, but more studies to be done on this topic.
6.1 Introduction

Four kinds of non-ion-exchange polymers, which were identified by the batch experiment in Chapter 4, were tested with a column experiment. This experiment setup is approximately simulated the real application of selected polymeric filter materials.

In the column experiment, the process of adsorption was accomplished by passing water through the porous filter medium for the removal of targeted PFCs (PFOS). We initiated this experiment with the latest POPs of PFOS and our research group will continue this experiment with other PFCs. Actually, the columns are designed for adsorbing soluble organic matters onto the media. Most filter columns are designed to operate in a down flow manner but up flow filters are occasionally used ahead of another filter in series.

The process of adsorption could be successfully simulated using the filter columns. However, special attention should be taken to minimize the flow ratio of the sidewall to cross section, which results in significant short circuiting and excessive bed expansion/compression (E.E Baruth, 1969). In addition it is essential to maintain a fixed flow rate.

6.1.1 Key variables for the design of synthetic resin sorbent systems

6.1.1.1 Types of synthetic resins

The results of isotherm experiments can be used to get an idea on candidate materials to eliminate target compound (in this study PFOS). Isotherm experiments and determined coefficients for seven granular materials are shown in Chapter 4. Although isotherm studies are useful for screening resins, it is important to note that these isotherm data are based on batch equilibrium sorption studies. That may not be directly representative of dynamic performance configurations. The actual mass loading on a resin may be significantly lower than that predicted from an isotherm study depending on numerous variables such as competitive sorption effects, background water quality and contact time. Tests performed by Calgon Carbon (Pittsburgh, PA) have shown that operating carbon usage rates, based on capacity at the time of breakthrough, can be estimated at 45 to 55 percent of the equilibrium capacity for VOCs (Stenzel and Merz, 1988). In order to determine a similar relationship for resins, it would be necessary to conduct dynamic column tests. For the usage in drinking-water applications, the synthetic resins have to be certified by the relevant authority to apply in portable water.
6.1.1.2 Background water quality
There is limited data currently available on the effects of background water quality on the PFOS removal efficiency by synthetic resins. One of our previous researchers has found that NOM can discourage the PFCs adsorption onto GAC by carbon foiling and competitive adsorption. Also he suggested that the effect of NOM on adsorption kinetics is minimum (Yong, 2007).

According to the available data on other target organics, the performance of Amberlite resins is unaffected by pH (6.5-8.5), temperature (10°C-25°C), oxidants (HOCl, H₂O₂, and O₃) and the presence of NOM. Amberlite resins have also been found to be unsusceptible to biofouling.

The issue of PFOS desorption would need to be addressed to determine the potential need for selection a mode of regeneration, multiple resin vessels in series, a greater number of sampling locations and a higher frequency of sampling, and a more frequent regeneration of resins.

To reduce the organic loading on the resin and to eliminate disinfection byproduct in tap water it may be prudent to use GAC columns at the front end of the process flow. GAC is cheaper on a per unit basis and is generally more effective for the removal of highly hydrophobic compounds.

6.1.1.3 Process flow configuration
In the real applications, the process flow configuration of synthetic resin systems is very similar to that of GAC systems. The main difference between a resin system and a GAC system is that the provision for a regeneration process. Sorption columns can be used in either down-flow or up-flow service mode. In general, the system configuration will be dependent on a number of factors, including the effluent standard, regeneration technique, and vessel design constraints. In situations where low effluent standards must be met and, thus, low leakage levels are allowed, a resin system works best under countercurrent operation and regeneration, with operation in the up-flow mode and regeneration in the down-flow mode (Rohm and Haas, 1992).

Using the down-flow mode for regeneration has been found to have better removal efficiencies and lower leakage levels once the column is returned to service after regeneration (Rohm and Haas, 1992). Sorbent columns can be operated in series, in parallel (also referred to as carousel), or as a combination of the two configurations depending on a number of factors, including the need for continuous operation, space constraints, effluent criteria, service cycle time constraints, operation logistics, and requirements for multi-barrier treatment.

Many field systems do not exhibit ideal, narrow breakthrough curves, but show elongated curves with tailing on the front and back end (Suffet, 1999 and Sun, 1999). This would suggest that the in-series operation offers an
economic advantage over the carousel operation. Table 6.1 shows the design flow rates of some GAC filters available in the market.

Table 6.1 Design flow rates for some GAC filters available in the market

<table>
<thead>
<tr>
<th>Brand</th>
<th>Model</th>
<th>Maximum Dimensions</th>
<th>Initial ΔP (psi) @ Flow Rate (gpm)</th>
<th>Chlorine Taste &amp; Odor Reduction @ Flow Rate (gpm)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pentec</td>
<td>GAC-5</td>
<td>2-7/8” x 4-7/8” (73 x 124 mm)</td>
<td>3.0 psi @ 0.5 gpm (0.2 bar @ 1.9 L/min)</td>
<td>250 gallons @ 0.5 gpm (900 liters @ 1.9 L/min)</td>
</tr>
<tr>
<td>filtration</td>
<td>GAC-10</td>
<td>2-7/8” x 9-3/4” (73 x 248 mm)</td>
<td>7.0 psi @ 1.0 gpm (0.5 bar @ 3.8 L/min)</td>
<td>5,000 gallons @ 1.0 gpm (18,900 liters @ 3.8 L/min)</td>
</tr>
<tr>
<td></td>
<td>GAC-20</td>
<td>2-7/8” x 20” (73 x 508 mm)</td>
<td>16 psi @ 2.0 gpm (1.1 bar @ 7.6 L/min)</td>
<td>10,000 gallons @ 2.0 gpm (37,800 liters @ 7.6 L/min)</td>
</tr>
<tr>
<td></td>
<td>GAC-BB</td>
<td>4-1/2” x 9-3/4” (114x 248 mm)</td>
<td>6.0 psi @ 2.0 gpm (0.4 bar @ 7.6 L/min)</td>
<td>12,500 gallons @ 2.0 gpm (47,000 liters @ 7.6 L/min)</td>
</tr>
<tr>
<td></td>
<td>GAC-20BB</td>
<td>4.1” x 20” (114x 508 mm)</td>
<td>5.0 psi @ 4.0 gpm (0.3 bar @ 15 L/min)</td>
<td>25,000 gallons @ 4.0 gpm (95,000 liters @ 15 L/min)</td>
</tr>
<tr>
<td>Siemens Water Tec.</td>
<td>2.875&quot; (7.3 cm)</td>
<td>1.0 gpm (3.8 L/min)</td>
<td>9.75&quot; (24.8 cm)</td>
<td>2.875&quot; (7.3 cm)</td>
</tr>
</tbody>
</table>

6.1.2 Regeneration of granular materials

All the granular materials used as adsorbents in filters must be regenerated. In the case of GAC, normally regeneration is done at a separate plant. Table 6.2 shows the main steps involved with this process (for GAC). Regeneration allows recovery of approximately 70% of the original carbon with this process. The re-activated carbon can be mixed with a portion of new carbon for higher effectiveness and is then returned to its place in the plant process (Clark, 1989). Off site regeneration of adsorbant materials is not cost effective. Also it requires time to install and uninstall filter materials.

Table 6.2 The basic steps involve with GAC regeneration

<table>
<thead>
<tr>
<th>Stage</th>
<th>Temperature (degrees C)</th>
<th>Action</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Drying</td>
<td>&lt; 100</td>
</tr>
<tr>
<td>2</td>
<td>Desorption</td>
<td>100 - 649</td>
</tr>
<tr>
<td>3</td>
<td>Pyrolysis</td>
<td>100 - 649</td>
</tr>
<tr>
<td>4</td>
<td>Gasification</td>
<td>&gt;&gt;649 and &gt;&gt;1038</td>
</tr>
</tbody>
</table>
In the case of synthetic resins, there is a possibility to regenerate the materials on site, which is one of the main advantages over conventional GAC filters. Several alternatives are available for regeneration including steam regeneration, solvent regeneration, and microwave regeneration.

6.1.2.1 Steam Regeneration
In steam regeneration, saturated steam is passed through a loaded/saturated resin bed, condensed, and collected. The steam is used to desorb organic compounds from the resin and to transport them away from the column. The condensed steam is typically discarded or treated through “superloading.” In superloading, the condensate is passed through a sorbent column just prior to the column’s regeneration cycle, taking advantage of the additional capacity of the sorbent at higher concentrations. The use of steam has been demonstrated to be an effective means of regenerating Ambersorb 563 resins loaded with a variety of organic compounds such as trihalomethanes (THMs) (Vandiver and Isacoff, 1994) and TCE (Parker and Bortko, 1991). In these cases, steam regeneration was able to fully restore the resin’s sorption capacity for these compounds. Suri et al. (1999) confirmed these findings, which showed that six regeneration cycles each with 28 to 40 bed volumes (BV) of steam (160°C) effectively regenerated resins saturated with p-PCB, perchloroethylene (PCE), and CCl4. In the case of o-PCB, there was a 20 percent loss in capacity observed after the first regeneration cycle. However, subsequent cycles did not result in further loss of sorption capacity (Suri et al., 1999).

6.1.2.2 Solvent Regeneration
In solvent regeneration, a solvent in which the adsorbate is highly soluble is passed through the saturated bed. In this experiment, organic regeneration was carried out for regeneration of several non ion-exchange polymers which have adsorbed PFOS.

6.1.2.3 Microwave Regeneration
Most studies on the use of microwave regeneration of sorbents examined vapor-phase applications in which the sorbents are used to remove volatile organic compounds (VOCs) from gaseous emission streams. However, the use of microwave regeneration for a liquid phase sorbent system is likely to be a similar process. Unlike steam regeneration, which uses steam to heat up the sorbent system, microwave irradiation generates heat directly in the sorbent bed by exciting sorbent and sorbate molecules. Contaminants adsorbed onto a resin column are volatilized and subsequently extracted through an induced vacuum (AmeriPure, Inc., 1999). Microwave heating has been shown to effectively eliminate the heat and mass transfer
resistances that limit the rate of regeneration in conventional steam systems (Price and Schmidt, 1997).

6.2 Aim and objectives
Major aim of this chapter is to study selected non ion-exchange polymer materials with a column experiment. Only non ion-exchange polymers were specifically selected for the column test mainly for two reasons. First reason is that it was found in the isotherm experiment that non ion-exchange polymers are possible candidates to adsorb PFOS; prior studies on this are rarely available. The second reason is easy regeneration of materials than other materials tested. The tested polymer materials were selected by the batch experiment conducted in Chapter 4. The results of the column experiment must be explained by the results of the batch test, which was conducted to determine adsorption isotherms. Also the results of this experiment are useful to determine some design and operational parameters in the real application in the field. The objectives in details are listed as below.

6.2.1 Objectives
1. To obtain column breakthrough curves up to 90% of PFOS removal for selected non ion-exchange filter materials.
2. To develop a mathematical model to explain the behaviors of the selected filter materials in the column.
3. To check the regenerability of non ion-exchange filter materials by washing of organic solvent.

6.3 Methodology
6.3.1 Adsorbent Pretreatment
Prior to the usage in the sorption experiment, Dowex polymers were first washed in ultrapure water to remove dirt and then dried at 50°C until reached a constant weight. Amberlite was also washed with deionized water first and then dried at 30°C. Once the polymers reached a constant weight, each column was filled with a required volume of each polymer. Similarly, the coal-based activated carbon of Filtrasorb 400 was first rinsed with deionized water for several times and then washed in 80°C deionized water for 2 hrs to remove the impurities. After being dried in an oven at 105°C for 48 hrs, they were crushed by a mortar and passed through 0.25 -0.5 mm sieve. After selecting the GAC with required size, again it was washed by deionized water to remove PAC particles attached with GAC. Washed GAC was dried and measured the required volume and installed into the column.
6.3.2 Analytical Equipments and Methods for PFOS Determination
At each sampling event, about 500mL of sample was collected from 1 column for the analysis of TOC and remaining PFOS. For the analysis of PFOS, 100mL was measured from the collected sample and sent through a Precip-C agri cartridge by using a concentrator (flow rate 5mL/min). Then, the cartridge was air dried for 1 hr using a vacuum manifold and then eluted it with methanol 3 mL. Eluted methanol was completely dried by flow of nitrogen gas at 60°C. Then, dried tube was reconstituted with 40% ACN solution and at the same time the concentration was adjusted such it did not exceed 10 μg/L. Final solution was analyzed and measured using HPLC/MS/MS (Agilent, Japan) and final concentration was calculated. In Agilent 1200SL HPLC, 5mM ammonium acetate and acetonitrile were used as mobile phases. Agilent 6400 triple quadrupole MS/MS was used in multiple reaction monitoring (MRM) at negative ionization mode for the detection of m/z of parent ion (499) and daughter ion (80) of PFOS.

6.3.3 Column Experiment
The column experiment setup is schematically shown in Figure 6.1. PP columns with dimensions of 30 cm length and 2 cm internal diameter were used to contain four types of filter materials as fixed-bed adsorbers. One column was run without a filter material as a control. The columns were first filled with measured pre treated material volume of 20 cm³. Then the columns were filled with MilliQ water and applied a vacuum for 24 hrs, which ensured all entrapped air bubbles in material surfaces were released. Tap water filtered with an activated carbon filter (to eliminate residual chlorine) was mixed with PFOS stock solution to adjust the feed PFOS concentration to 10 μg/L. The mixed PFOS solution was fed through five columns in down-flow mode and channeling of bed materials were avoided by mesh fixed at the bottom of each filter beds (Figure 6.1). Peristaltic pumps were used to control the flow rates at the inlets of each column and the mixing tank. The effluent of each column were collected periodically and analyzed for the remaining PFOS concentration (C) using LC/MS/MS. The desired break through concentration (C_b) was determined at 10% of the inlet feed concentration (C_0), which is 0.1 C_0 or 1 μg/L. The flows through the tested columns were continued until the PFOS concentration of all columns effluent approached 0.2 C_0 which took about 60 days.

6.3.4 PFOS regeneration
All filter materials of each column were completely removed after operation and dried in 60°C for 48 hrs. The dried materials were thoroughly mixed to make it homogenous. For the regeneration experiment, 0.02 g of each dry material were added to 1L of LC/MS grade methanol and shaken at control
temperature of 25°C. Sampling was done at different time intervals of 0, 20, 80 and 270 min. 2 mL of each collected samples were dried with nitrogen gas at 55°C and then reconstituted with 40% acetonitril (4mL). After filtration (0.2μm), the final samples were analyzed with LC/MS/MS for recovered PFOS.

Fig 6.1 Column experiment setup for PFOS adsorption onto different filter materials

6.4 Results and discussion

6.4.1 Forces effect on adsorption

Previous studies have demonstrated that anionic surfactants uptake by a adsorbent surface is strongly dependent on electrostatic interactions between a surfactant molecule and the surface (Dobson et al., 2000; Johnson et al., 2007). In addition, the Columbic repulsion between adjacent PFOS molecules is likely to play an important role. Other non electrostatic interactions, such as hydrophobic interaction (Dobson et al., 2000; Torn et al., 2003), may also contribute to surfactant adsorption onto mineral surfaces. Thus, the change in total free energy ((ΔG) adsorption) associated with PFOS adsorption can breakdown into three components:

\[ \Delta G_{\text{adsorption}} = (\text{PFOS} - \text{surface electrostatic interaction}) + (\text{PFOS} - \text{PFOS electrostatic interaction}) + (\text{non} - \text{electrostatic interactions}) \]
6.4.1.1 PFOS-surface electrostatic interaction
PFOS, being a strong acid with an estimated pKa of around -3 (OECD, 2002), carries a negatively charged site at its sulfonate head group under all environmentally relevant pH. In general, a mineral surface becomes more positively charged (or less negatively charged) at lower pH. This results in an enhanced electrostatic attraction force (or reduced electrostatic repulsion force) between the negatively charged PFOS molecules and the surface, which is expected to promote PFOS adsorption at lower pH. Another important parameter that affects the electrostatic interaction between PFOS molecules and the adsorbent surface is ionic strength. The electrostatic interaction can be significantly weakened at higher ionic strength due to the double layer compression effect. For a positively charged mineral surface, the amount of adsorbed PFOS tends to be reduced due to the weaker electrostatic attraction (Chuyang et al., 2010).

6.4.1.2 PFOS-PFOS electrostatic repulsion.
In addition to the PFOS surface electrostatic interaction, two adjacent PFOS molecules on a surface will also repel each other due to their negatively charged sulfonate head groups. A strong PFOS-PFOS repulsion tends to prevent these molecules getting close to each other. Thus, a solution with high ionic strength has a tendency to promote PFOS adsorption as a result of the suppressed electrostatic repulsive force. In contrast, the PFOS-PFOS electrostatic repulsion is not directly affected by pH since these molecules carry a constant charge over all environmentally relevant pH values (Chuyang et al., 2010).

6.4.1.3 Other non-electrostatic interactions.
Non-electrostatic interactions such as hydrophobic interaction (Higgins and Luthy, 2006; Ochoa-Herrera and Sierra-Alvarez, 2008; Torn et al., 2003) may also contribute to PFOS adsorption, given the strong hydrophobic nature of its perfluoroalkyl chain. For example, hydrophobic interaction may arise between the hydrophobic chain of a PFOS molecule and the hydrophobic moiety on a mineral surface or between the hydrophobic chains of different PFOS molecules. Such non-electrostatic interaction has much weak or little dependence on the solution chemistry compared to the PFOS-surface and PFOS-PFOS electrostatic interactions.

6.4.1.4 Effect of pH
In general, lowering pH makes a surface more positively charged, which promotes PFOS adsorption (enhanced PFOS-surface interaction). However, the effect of pH diminishes at high ionic strength, as the PFOS-surface electrostatic interaction becomes screened by counter ions as a result of
electrical double layer compression (Chuyang et al., 2010).

6.4.1.5 Effect of ionic strength
Increasing ionic strength tends to suppress the PFOS-surface as well as PFOS-PFOS electrostatic interactions. The net result on PFOS adsorption depends on the competition between the two types of interactions. For a weakly charged mineral surface (pH~$H_{pzc}$), the PFOS-surface electrostatic interaction is negligible compared with the PFOS-PFOS interaction. Consequently, increasing ionic strength promotes PFOS adsorption as a result of reduced repulsive force between adjacent PFOS molecules. In contrast, the PFOS-surface interaction dominates for a highly charged surface.

6.4.2 Overall Performance of the columns with time
We applied comparatively higher flow rate of 15 mL/min which ensures at least 1.3 min retention time and inflow PFOS concentration was kept constant at 10 $\mu$g/L. Figure 6.2 a and b show the percentage removal of PFOS in each column over the time (60 days) and the variation of effluent PFOS concentration with operation time and filtered bed volume. It was noticed for all polymers that the percentage removal efficiency for first 5000 bed volume is more than 99%. Amb XAD showed excellent performance by removing more than 99.99% PFOS from first 23,000 bed volumes passing through it. The column break through point was assumed at 90% removal because the PFOS concentration in environmental water is in ng/L level.

Fig. 6.2(a) Variation of percentage PFOS removal with time (60 days) for different filter materials in the column experiment
6.4.2.1 Dowex L493

The PFOS removal efficiency of the column with Dow L493 was more than 99% for first 7 days. It took 30 days to reduce the removal efficiency from 100% to 90%, another 18 days to reduce it from 90% to 80% and 9 days to reduce from 80% to 70%. Figure 6.3 shows the percentage relative abundance with time.

Fig 6.3 Fractional effluent PFOS concentration in the column with DOW L493
6.4.2.2 Amberlite XAD 4

Amb XAD 4 was the best candidate among the tested filter materials. The effluent PFOS concentration of AmbXAD4 was less than detectable level for first 20 days (Figure 6.4(a)).

The removal efficiency was decreased to 96% during next 20 days and showing a sudden efficiency drop from 40 days removal efficiency was 80% after 60 days. This behavior emphasizes the superior performance of AmbXAD at low PFOS concentrations.

Amberlite XAD 4 is a non-ionic cross linked polymer which derives its adsorptive properties from its macroreticular structure (containing both continuous polymer phase and continuous pore phase), high surface area and the aromatic nature of its surface (Figure 6.4 (b)). This structure gives Amb XAD polymer adsorbent an excellent physical, chemical and thermal stability. Average surface area of other granular materials tested was around 1,000 m²/g. The surface area of AmbXAD4 is unknown, but it is said by the manufactures that the minimum surface area is 750 m²/g. it can be reasonably assumed that the actual surface are of Amb XAD4 is more than other materials tested.

6.4.2.3 Granular Activated carbon

GAC showed more than 99.95% of PFOS removal for first 7 days and then started to reduce the performance. Unlike other granular materials tested, it was noticed that the relative abundance of PFOS is linearly increased.
with time (filtered volume) (Figure 6.5) for this material. GAC column took about 22 days to reduce the removal efficiency by 10% (from 100% to 90%) and 40 days to reduce it by 20%.

![Graph](image1)

Fig 6.5 Fractional effluent PFOS concentration in the column with F400

**6.4.2.4 Dowex V493**
Among all the granular materials tested, DowV493 showed the minimum performance to remove PFOS. The performance of the column was reduced by 10% within 12 days and by 20% within 35 days (Figure 6.6).

![Graph](image2)

Fig 6.6 Fractional effluent PFOS concentration in the column with DOW V493

**6.4.3 Column test results and Batch test results**
The results of column test experiment can be explained by the results of batch test and kinetic test experiments. The PFOS concentration of the column outflow is decided by two main factors. First factor is the amount of
vacant sites of the filter media which can attach PFOS molecules and the second factor is the time requirement to make the bond between sorbent and sorbate. Isotherm experiment gives an idea on available vacant sites and kinetic experiment implies the speed of attachment. Amb XAD4 showed higher sorption rate constant \((k = 0.48)\) and Freundlich isotherm constant \((K_f = 79.1)\) and it was the best material even in the column test. GAC also showed higher sorption rate constant \((k = 80.28)\) but lower Freundlich isotherm constant \((K_f = 28.4)\) whereas for Dow L493 and Dow V493 showed higher Freundlich isotherm constants \((K_f = 54.6\, \text{and}\, 81.3)\) but lower sorption rate constants \((k = 0.07\, \text{and}\, 0.08)\).

### 6.4.4 Mathematical modeling

Lin and Huang (2007) proposed a mathematical model for the change of sorbate concentration in a column (with short column operation time of 1-5 min). Using the same assumption they made, we come up with a modeling equation as derived below.

![Fig 6.7 Schematic representative of a column for modeling](image)

Fraction of adsorption

\[
A = \frac{C_0 - C_1}{C_0}
\]  

6.1

Fraction of discharge

\[
P = \frac{C_1}{C_0}
\]  

6.2

\[A + P = 1\]  

6.3

The rate of change of fraction of adsorption

\[
\frac{dA}{dt} = \frac{A}{dt} = \frac{d[(C_0 - C_1)/C_0]}{dt}
\]  

6.4

According to Lin and Huang (1999) it can be assumed that the rate of change of adsorption fraction is linearly promotional to the fraction of adsorption \((A)\) itself.
and the fraction of discharge \((P)\).

With this assumption it can be written as

\[
\frac{dA}{dA} \propto AP
\]

6.5

Same expression can be rewritten with a constant as

\[
\frac{dA}{dt} = kAP
\]

6.6

From equations 6.3 and 6.6

\[
\frac{dA}{dt} = kA(1:A)
\]

6.7

With the initial conditions of \(A = A_a\) and \(t = t_a\), the equation can be integrated as

\[
\int \frac{dA}{A(1:A)} = -\int_{t_a}^{t} kdt
\]

6.8

\[
\ln \frac{A(1:A_a)}{A(1:A)} = k(t_a-t)
\]

6.9

Which is the same as:

\[
\ln \left[ \frac{P_a(1:A)}{P_a(1:A)} \right] = k(t_a-t)
\]

6.10

It the half removal time is denied by \(\tau\), \(P\) (remaining fraction of PFOS) is 0.5

at \(t = \tau\)

\[
P = \frac{1}{1 + \exp \left[ k \left( \tau \cdot t \right) \right]}
\]

6.11

Or

\[
t = \tau + \frac{1}{k} \ln \left[ \frac{P}{(1:P)} \right]
\]

6.12

Since the inflow and the outflow of the column is same, the remaining fraction of PFOS at the outflow can be represented by the concentration ratio, \(C/C_0\). Where \(C\) is the PFOS concentration at the outflow of the column and \(C_0\) is the inflow PFOS concentration.

Then, the equation 6.12 can be rewritten as

\[
t = \tau + \frac{1}{k} \ln \left[ \frac{C}{(C_0 \cdot C)} \right]
\]

6.13
According to equation 6.13, a linear plot of adsorption time (t) vs \( \ln[C/(C_0 - C)] \) can be used to determine the half saturation time (\( \tau \)) and column constant k.

**Figure 6.8.a** shows such a plot for PFOS adsorption on to different filter materials and **Table 6.3** shows the calculated \( \tau \) and k. **Figure 6.8b** shows the theoretical curves which have been drawn with calculated \( \tau \) and k and actual data point for first 60 days.

Table 6.3. parameters of the theoretical model proposed by Lin and Huang (1999) for a column experiment for PFOS adsorption onto different filter materials.

<table>
<thead>
<tr>
<th>Adsorbent</th>
<th>( \tau ) (days)</th>
<th>( k ) (1/days)</th>
<th>( R^2 )</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dow V493</td>
<td>64.5</td>
<td>0.049</td>
<td>0.93</td>
</tr>
<tr>
<td>Amb XAD 4</td>
<td>70.7</td>
<td>0.117</td>
<td>0.97</td>
</tr>
<tr>
<td>Dow L493</td>
<td>67.4</td>
<td>0.069</td>
<td>0.98</td>
</tr>
<tr>
<td>Filtrasorb 400</td>
<td>65.5</td>
<td>0.060</td>
<td>0.94</td>
</tr>
</tbody>
</table>

**Note:** Half saturation time (\( \tau \)) and column constant k with 10 \( \mu \)g/L inflow concentration and 0.75 bed volume/min flow rate

### 6.4.5 Material regeneration by organic solvents

In the real application of solvent regeneration, a solvent in which the adsorbate is highly soluble is passed through the saturated bed. Studies by Rohm and Haas (1992) have demonstrated that the use of solvents such as methanol or acetone at a flow rate of two bed volumes per hour can successfully regenerate resin columns. In one study, methanol was found to extract more than 99% of 280 mg of trichloroethylene adsorbed per gram of Amb 563 (Parker and Bortko, 1991). A second study showed that 4 to 5 BVs of methanol at a flow rate of 1 bed volume per hour can remove more than 95% of 1,2-dichloroethane loaded on an Amb 563 column (Isacoff et al., 1992). Rinsing with water or steam is typically performed after regeneration to remove any residual regenerant prior to the next sorption cycle. In another experiment by Malley et al. (1993), methanol was demonstrated to be ineffective in regenerating some synthetic resins. The published data on the effectiveness of solvent regeneration specifically for PFCs-saturated columns is rarely available.
The accumulation of PFOS over the operation of column test of 60 days is shown in Figure 6.9. The cumulative PFOS inflow into the column is also shown in the same figure. We only measured the PFOS concentration at in-flow and out-flow at different times (days) and the PFOS accumulation in the granular materials were calculated considering the difference of concentrations, sampling intervals and flow rates.
Figure 6.10 shows the variation of percentage PFOS recoveries of each granular material with time. As reported by many researchers in previous studies (Rohm and Haas, 1992, Parker and Bortko, 1991, Isacoff et al., 1992) for other organic compounds, methanol has been identified as an ideal organic solvent to regenerate PFOS adsorbed synthetic resins. For the synthetic polymers, a sudden recovery was observed for first 30 min and they reached the maximum recovery of 100% within 80 min. For GAC it was observed that the percentage recovery just reached 40% after 270 min of shaking time. One possible reason for faster desorption kinetics of synthetic resins than GAC is the strength of sorbent - sorbate. There is a physico-chemical bond between the internal surface of adsorbent and adsorbate at any adsorption process (Montgomery, 2008). The bond between GAC surface and PFOS seems to be stronger than that of synthetic resins and PFOS. The second possible explanation is pore diffusion. PFOS seems to be diffused into deep pores in the active GAC surface and it takes time to dissolve PFOS back in organic solvent. In the case of synthetic resins, PFOS attachment to the outer surface is dominant suggesting minimum pore diffusion. This may the reason for higher PFOS recovery from synthetic resins than from GAC mainly during the first few minutes. The results suggest onsite regeneration of synthetic resins by organic solvents is much more feasible than regeneration of GAC. PFOS might be easily recovered from methanol by evaporation and it can be reused.
6.5 Summary
In this chapter, non ion exchange polymer materials, which were selected by the batch experiment in Chapter 4 was tested with a continues column experiment. Three synthetic granular materials and one GAC material were tested by the column experiment, which was continuously run for 60 days. The conclusions of the experiment are summarized below.

1. Amb XAD 4 was recognized as the best candidate among the tested four filter materials to eliminate PFOS.

2. It was observed that the results of column experiment cannot be explained by individual results of isotherm experiment or kinetic experiment suggesting the importance of both isotherm (available vacant sites in the materials to adsorb) and kinetics (the speed of attachment) in the real application.

3. It was interesting to notice in the column experiment that Amb XAD 4 removed 99.99% PFOS up to 23,000 bed volumes pass through with the condition of 10 μg/L inflow concentration and flow rate of 15 ml/min (0.75 bed volumes/min).

4. Within 80 min, all most 100% PFOS was recovered from synthetic resins in the regeneration experiment suggesting that synthetic polymers can be effectively regenerated by an organic solvent.
Chapter 7 - Combined treatment of PFOS

7.1 Introduction
The combined process of coagulation and filtration is not a new concept. Coagulation, flocculation and clarification, followed by rapid gravity sand filtration, are the key steps in conventional water treatment systems. This is a well-proven technology for the significant removal of color and particulate matter including protozoa (e.g. Cryptosporidium oocysts and Giardia cysts), viruses, bacteria, and other micro-organisms. Iron, manganese, tastes and odors may also be removed from the water by these processes (Mercalf & Eddy, 2003; Fiksdal et al., 2006, Dugan et al, 2001). This chapter combines the two unit operations of coagulation, adsorption and filtration, studied under Chapter 5 and 6 to remove PFOS from water. Conventional treatment (coagulation, sedimentation and sand filtration), has several distinct stages. A coagulant is added to neutralize the natural electrical charges on the colloidal particles that prevent them from agglomerating, and is rapidly mixed into the water to be treated. The processed water will then enter a flocculation chamber, where further chemicals may be added. Gentle mixing during this stage allows particles to agglomerate and form settleable flocs. We tried to simulate the process in the lab scale with polymer type organic coagulants and non ion exchange granular materials. It was assumed that the attachment of PFCs into long chain organic coagulant can be further enhanced the adsorption process and can be achieved superior performance than the individual coagulation and adsorption process.

7.2 Aim and objectives
The overall objective of this experiment was to examine the improvement of adsorption process by prior coagulation by organic coagulants. Non ion-exchange polymer adsorbents tested in Chapter 4 and the organic coagulants tested in Chapter 5 were used in this experiment. Coagulation and flocculation processes were simulated by lab scale unit and same column setup used in Chapter 6 was used to treat coagulated water. The specific objectives of the experiment are as follows.
1. To observe the PFOS adsorption characteristics of non ion-exchange polymers for coagulated water by obtaining the column break through curves.
2. To determine the effect of TOC on PFOS adsorption.
3. To check the effectiveness of micro-filtration to eliminate PFOS from coagulated water.
7.3 Methodology

7.3.1 Adsorbent Pretreatment

Pretreatment of granular materials were similar to that of column experiment at Chapter 6. Prior to the use in the sorption experiment, Dowex polymers were first washed in ultrapure water to remove dirt and then dried at 50°C until reached a constant weight. Amberlite also washed with ultrapure water first then dried at 30°C. Once the polymers reach the constant weight, each column was filled with the required volume of each polymer. Similarly, the coal-based activated carbon of Filtrasorb 400 was first rinsed with deionized water for several times and then washed in 80°C deionized water for 2 hrs to remove the impurities. After being dried in an oven at 105°C for 48 hrs, they were crushed by a mortar and passed through 0.25-0.5 mm sieve. After selecting the GAC with required size, again it was washed by deionized water to remove PAC particles attached with GAC. Washed GAC was dried and measured the required volume and installed into the column.

7.3.2 Analytical Equipments and Methods for PFOS Determination

At one sampling, about 1 L of sample was collected from one column. 100 mL was measured from the collected sample and first it was filtered with 0.2mm syringe filters. The main purpose of this step is to eliminate all the remaining coagulants in the sample as they can interfere with LC/MS/MS analysis. The filtrate was sent through a Presep-C agri cartridge by using the concentrator (flow rate 5mL/min). Then, the cartridge was air dried for 1 hr using a vacuum manifold and then eluted it with methanol 2 mL and Acetonitrile 2mL. Eluted methanol was completely dried by a flow of nitrogen gas at 55°C. Then, dried tube was reconstituted with 40% ACN solution and at the same time the concentration was adjusted such that it did not exceed 10 μg/L. Final solution was analyzed and measured by using LC/MS/MS and final concentration in the initial solution was calculated. In Agilent 1200SL HPLC, 5mM ammonium acetate and acetonitrile were used as mobile phases. Agilent 6400 triple quadrupole MS/MS was used in multiple reaction monitoring (MRM) at negative ionization mode for the detection of m/z of parent ion (499) and daughter ion (80) of PFOS.

7.3.3 Column Experiment

The column experiment setup is schematically shown in Figure 7.1. Similar kind of PP columns used in the column test in Chapter 6 with dimensions of 30 cm length and 2 cm internal diameter were used. Four non ion-exchange filter materials already tested in Chapter 4 and Chapter 6 were placed in the columns. Fifth column was run without a filter material and only coagulated water was collected at the effluent of column 5. The
columns were first filled with measured pre treated material volume of 20 cm³. Then, the columns were filled with deionized water and applied a vacuum for 24 hrs which ensured all entrapped air bubbles in material surfaces were released.

As an addition to the previous column test, rapid mixing tank was introduced for this experiment. Tap water filtered with an activated carbon filter (to eliminate residual chlorine) was mixed with PFOS stock solution (to adjust the feed concentration to 10 μg/L) and coagulation stock solution at the rapid mixing tank. The coagulated PFOS solution was sent to a slow mixing tank at which flocculation process is enhanced. Then, the solution was fed through five columns in down-flow mode and channeling of bed materials were avoided by mesh fixed at the bottom of each filter beds (Figure 7.1).

Fig 7.1 Schematic diagram of the experimental setup for combined treatment processes

Peristaltic pumps were used to control the flow rates at the inlets of each column and the mixing tanks. The effluent of each column were collected periodically and analyzed for the remaining PFOS concentration using LCMS/MS as described above. The conditions of the experiment are shown in Table 7.1.
Table 7.1 Experimental conditions for combined treatment process

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Set value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Feed concentration - PFOS</td>
<td>10μg/L</td>
</tr>
<tr>
<td>Feed concentration - Coagulant</td>
<td>6.5μL/L</td>
</tr>
<tr>
<td>Retention time – rapid mixing tank</td>
<td>4 min</td>
</tr>
<tr>
<td>Retention time – slow mixing tank</td>
<td>5 hrs</td>
</tr>
<tr>
<td>Flow rate in columns</td>
<td>15 mL/min</td>
</tr>
<tr>
<td>Retention time - column</td>
<td>1.3 min</td>
</tr>
</tbody>
</table>

### 7.4 Results and Discussion

**Figure 7.2** compares the percentage removal of PFOS at each column (with different granular materials) for previous column experiment (Chapter 6) and this column experiment.

First observation in Figure 7.2 is the overall improvement of columns by organic coagulation to remove PFOS. The main reason for the improvement is the combined effect of coagulation and adsorption process. As schematically shown in **Figure 7.3**, coagulated water was sent through the adsorption column in this experiment whereas in the previous experiment only adsorption columns were used.

![Graph showing PFOS removal over time](image)

**Fig. 7.2** Percentage PFOS removal with time by adsorption process and combined coagulation and adsorption process for non ion exchange filter materials.
Since an organic coagulant was added in the experiment three possible mechanisms can be expected. First obvious phenomenon is pure coagulation, which did not appear in the first column test discussed in Chapter 6. The coagulated PFOS can be settled in slow mixing tank and filter columns. The second PFOS removal mechanism in this column test is adsorption. Similar to the first column experiment, non coagulated PFOS molecules can be adsorbed onto the granular materials packed in the column. But in the second column experiment some organic coagulants also occupied the filter bed. Most of the time these coagulants negatively affect material adsorption capacity as they can block the macrospores of granular materials preventing the access into inner micro pores. The effect is more serious as the retention time of the column is just 1.5 min. The retaining coagulants in the granular materials also have a positive effect on PFOS removal as it can add some active sites to the filter materials. Unfilled active sites of long chained polymeric coagulates attached to the filter media can attach some free PFOS molecules as it passes through the columns. It can be reasonably assumed that the real performance of the column is the net effect of above two phenomena (one negative and one positive).
The third PFOS removal mechanism is filtration. Filtration (0.2μm syringe filters) was done as an essential step in LC/MS analysis for both column tests before analysis. It was tested and observed that the PFOS removal by filtration (0.2μm syringe filters) is almost zero for non coagulated samples. As discussed in the Chapter 5 we observed that micro filtration is very effective to eliminate PFOS in coagulated water.

7.4.1 Reduction of PFOS by coagulation and filtration process in the combine treatment process

PFOS reduction by the coagulation and filtration process is determined by measuring the remaining PFOS after filtration in the effluent of control column. The control column was kept empty and the PFOS reduction was only by coagulation and filtration processes. Figure 7.4 shows the percentage PFOS removal by coagulation with time. For the first month a long range polymer coagulant of Fl4820 was used as the coagulant. It was observed in the experiment that the coagulation was increased for the first week from 65% to 85%. After that the performances of coagulation was deteriorated with time from 85% to 70% within the next 3 weeks. From the second month onward, another long range coagulant of FL 2250 was applied and found that the percentage PFOS removal was around 95% and it was stable with time.

Fig. 7.4 Percentage PFOS removal with time by coagulation and filtration processes
7.4.2 Reduction of PFOS by adsorption process in the combined treatment process

PFOS reduction by adsorption process was calculated with overall PFOS reduction and the PFOS reduction by coagulation and filtration. It was assumed in the calculation that the adsorption process in the column was independent and it did not affect with coagulation and filtration process and vice versa. Figure 7.5 shows the percentage PFOS removal by adsorption. Compared with the previous column experiment (Chapter 6) it was noticed that the column capacities did not reduce smoothly with time. It was also noticed in the first month that the removal efficiencies of the columns for all filter materials were less than that of first column experiment (Chapter 6). With the introduction of new organic coagulant (FL 2250) some performance improvement was observed for some polymers, particularly for AMB XAD 4. Disturbances by coagulants on adsorption process at the filter bed may be the main reason for the fluctuation and reduction (than previous column test) of column efficiencies. Also it was interesting to notice that the properties of coagulants affect the adsorption capacity in the column.

![Graph showing PFOS removal percentage over time](image)

Fig. 7.5 Percentage PFOS removal by adsorption (with and without coagulation process)

7.4.3 TOC and PFOS removal

Figure 7.6 shows the variation of TOC with time. Tap water was used in this experiment as the inflow source where average TOC was measured as 2.5 mg/L. PFOS stock solution and organic coagulant solution were mixed with
tap water before feeding in to the column to adjust PFOS concentration to 10μg/L and coagulant concentration 6 mg/L. After all the additions, TOC should be around 8 mg/L but it was noticed that measured TOC was around 1 mg/L. The TOC could have been reduced in the process at several places. One place for the reduction of the TOC concentration is by the carbon filter installed to eliminate residual chlorine. The TOC added with the coagulant could have been reduced at the slow mixing tank by settling it, at filtration column and final filtration process before analysis. One of the main disadvantages with this experimental set up was that there was no proper provision to flush out settled coagulants in slow mixing tank and filter beds. Due to this fault, sudden TOC peaks were observed and it was overcome by cleaning the tanks manually and flushing the columns with tap water (free of residual chlorine). First such peak was observed after 60 days of operation as shown in Figure 7.6.

It was noticed that there was a reduction of TOC with the introduction of different organic coagulants, which may be the reason for the improvement of adsorption capacity in some granular materials. It can be deduced with this observation that the filtration capacity of granular materials tested
were reduced by coagulants (may be due to material fouling) but the degree of effect was minimum for some granular materials such as AMB XAD 4.

7.5 Economic Analysis

7.5.1 Objective and Background

The primary purpose this basic economical analysis was to determine the potential cost saving granular material for treating water contaminated with PFOS. Since the column experiment and the material regeneration experiment were limited to non ion-exchange polymers, cost analysis also limited to the same granular materials.

7.5.2 Assumptions

This analysis was a basic cost comparison considering only two parameters of adsorption capacity and material regeneration. Following assumptions were made to figure out some parameters to be used in the calculation as described below.

1. Unit price of each polymer material and GAC were assumed to be same as the supply price to the laboratory. Table 7.3 shows the unit price of each granular material.

2. The performance of the column experiment can be applied to the real scale applications.

3. Three times bed volumes of organic solvent is sufficient to regenerate the synthetic polymer materials.

4. Cost of organic solvent (Methanol) is same to the supply price to the laboratory.

5. 60% of material cost of GAC is required for regenerating them (for one time).

6. Handling of recovered PFOS is a separate business and will not involve with regeneration process.

7. The time required to reduce the column efficiency by given valve will not change with initial PFOS concentration for a given flow rate.

8. The column efficiency curve will not change for further low inflow concentration.
Table 7.2 Cost of Granular Materials

<table>
<thead>
<tr>
<th>Polymer brand</th>
<th>Producer</th>
<th>Supplier</th>
<th>Amount purchased (g)</th>
<th>Bill value (Yen)</th>
<th>Unit cost (Yen/kg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dow V493</td>
<td>Dow Chemical</td>
<td></td>
<td>100</td>
<td>9000</td>
<td>90000</td>
</tr>
<tr>
<td>Dow L493</td>
<td>Dow Chemical</td>
<td></td>
<td>500</td>
<td>18000</td>
<td>36000</td>
</tr>
<tr>
<td>Amb XAD 4</td>
<td>Rohm and Haas</td>
<td>Sigma-Aldrich</td>
<td>500</td>
<td>21000</td>
<td>42000</td>
</tr>
<tr>
<td>F400 (GAC)</td>
<td>Calgon Company</td>
<td>Japan</td>
<td>500</td>
<td>8500</td>
<td>17000</td>
</tr>
<tr>
<td>Amb IRA-400</td>
<td>Rohm and Haas</td>
<td></td>
<td>500</td>
<td>16700</td>
<td>33400</td>
</tr>
<tr>
<td>Dow Marathon</td>
<td>Dow Chemical</td>
<td></td>
<td>250</td>
<td>11000</td>
<td>44000</td>
</tr>
</tbody>
</table>

### 7.5.3 Cost Scenarios

For this economic analysis, some common factors were not considered because the purpose of this analysis was to compare the materials, but not compare the PFOS treatment techniques. For example, minimum by-product formation in the treatment process was not considered for the analysis because it is common for all granular materials. Two main factors considered for economical analysis were adsorption capacities of each material and their cost of regeneration. Some data related to the calculation was not available in the experiment and also in the literature data. For example, it is unknown how many times non-ion-exchange polymers can be regenerated while keeping the acceptable removal efficiency. Also, it is unknown the amount of organic solvent required for material regenerations. The economic analysis was done for three scenarios, for different inflow PFOS concentrations, different outflow PFOS concentrations and different times of regeneration. For the inflow PFOS concentrations, 10,000 ng/L, 4,000 ng/L, and 1000 ng/L were considered and for outflow PFOS concentrations 100 ng/L and 10 ng/L were considered. The cost implications were also calculated for 10, 5 and 2 times of materials regenerations before replacing them with new materials.

### 7.5.4 Methodology

This economic analysis was done by comparing the material installation cost and the operational cost for four filters containing the target non-ion-exchange filter materials. It was assumed that the filter bed volume is 1 L for each filter. For easy comparison, material installation cost and operational cost were calculated for the daily basis. Each component of the cost was calculated as described below.

### 7.5.4.1 Material installation cost

For this comparative study, it was assumed that the material supply price
remained as same as the laboratory supply price (Table 7.3). However, in the real application there is a significant price reduction for bulk purchasing.

7.5.4.2 Material regeneration interval
Material regeneration interval of each material was determined by considering the results of column experiment in Chapter 6 based on the required removal efficiencies of each scenario. Table 7.4 shows the drop of removal efficiencies of each material with time (based on the results of Chapter 6).

7.5.4.3 Material regeneration cost
It was assumed that three times bed volumes of methanol (3 L) is sufficient to regenerate synthetic polymers and since the methanol cost is dominant other costs can be negligible. Based on the literature data 60% of material cost was allocated for the offsite regeneration of GAC.

Table 7.3. Drop of PFOS removal efficiencies with time

<table>
<thead>
<tr>
<th>Days</th>
<th>No of bed volumes</th>
<th>DOWV493</th>
<th>DOW L493</th>
<th>Amb XAD 4</th>
<th>F400(GAC)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0</td>
<td>99.95</td>
<td>100.00</td>
<td>99.95</td>
<td>99.98</td>
</tr>
<tr>
<td>1</td>
<td>1080</td>
<td>99.90</td>
<td>100.00</td>
<td>99.95</td>
<td>99.96</td>
</tr>
<tr>
<td>6</td>
<td>6480</td>
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<td>100.00</td>
<td>99.41</td>
<td>100.00</td>
</tr>
<tr>
<td>7</td>
<td>7560</td>
<td>96.27</td>
<td>100.00</td>
<td>99.95</td>
<td>100.00</td>
</tr>
<tr>
<td>10</td>
<td>10800</td>
<td>92.72</td>
<td>100.00</td>
<td>98.37</td>
<td>99.44</td>
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<tr>
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<td>12960</td>
<td>93.74</td>
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<td>100.00</td>
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<td>96.56</td>
</tr>
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<td>99.64</td>
<td>94.48</td>
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</tr>
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<td>98.02</td>
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Table 7.4 Results of economic analysis for three times material regenerations

<table>
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<th>cost factors</th>
<th>Total</th>
</tr>
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<tbody>
<tr>
<td></td>
<td>IF</td>
<td>OF</td>
<td>MRI</td>
</tr>
<tr>
<td>Dow V493</td>
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</tr>
<tr>
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<td>15429</td>
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<td>4846</td>
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<tr>
<td>F400(GAC)</td>
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<td>7286</td>
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</table>

Note:

IF- Inflow PFOS concentration (ng/L), OF- Targeted outflow PFOS concentration (ng/L), MRI-Material regeneration interval (days), RV-PFOS removal efficiency, C_{mi}-Material installation cost, C_{mr}- Material regeneration cost

Assumptions: 1. three times bed volume of organic solvent is needed for material regeneration 2. 60% of materials cost (GAC) is needed for one time GAC regeneration 3. Cost of organic solvent is 1200 JPY/L
Table 7.5 Results of economic analysis for five times material regenerations

<table>
<thead>
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<th>Operation parameters</th>
<th>cost factors</th>
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<tr>
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<td>MRI</td>
</tr>
<tr>
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<td>150000</td>
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<tr>
<td>F400(GAC)</td>
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<table>
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<th>Total</th>
</tr>
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<tbody>
<tr>
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<td>5625</td>
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<table>
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<th>Material</th>
<th>Operation parameters</th>
<th>cost factors</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dow V493</td>
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<td>99.75</td>
<td>150000</td>
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<tr>
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</tr>
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<td>F400(GAC)</td>
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<td>99.75</td>
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</tr>
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</table>

Note

IF- Inflow PFOS concentration (ng/L), OF- Targeted outflow PFOS concentration (ng/L), MRI-Material regeneration interval (days), RV-PFOS removal efficiency, C_{mi}-Material installation cost, C_{mr}- Material regeneration cost
Assumptions: 1. three times bed volume of organic solvent is needed for material regeneration 2. 60% of materials cost (GAC) is needed for one time GAC regeneration 3. Cost of organic solvent is 1200 JPY/L
Table 7.6 Results of economic analysis for eight times material regenerations

<table>
<thead>
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<th>Material</th>
<th>Operation parameters</th>
<th>cost factors</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
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<td>IF</td>
<td>OF</td>
<td>MRI</td>
</tr>
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<td>Dow V493</td>
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<td></td>
</tr>
<tr>
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</tr>
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<td>Amb XAD 4</td>
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</tr>
<tr>
<td>Dow V493</td>
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<tr>
<td>Dow L493</td>
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<td></td>
</tr>
<tr>
<td>Amb XAD 4</td>
<td>1,000</td>
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</tr>
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<td>Dow V493</td>
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</tr>
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</tr>
<tr>
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<tr>
<td>F400(GAC)</td>
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<td></td>
</tr>
<tr>
<td>Amb XAD 4</td>
<td>26</td>
<td>99.75</td>
<td></td>
</tr>
<tr>
<td>F400(GAC)</td>
<td>7</td>
<td>99.75</td>
<td></td>
</tr>
</tbody>
</table>

Note

IF- Inflow PFOS concentration (ng/L), OF- Targeted outflow PFOS concentration (ng/L), MRI-Material regeneration interval (days), RV-PFOS removal efficiency, \(C_{mi}\)-Material installation cost, \(C_{mr}\)- Material regeneration cost

Assumptions: 1. three times bed volume of organic solvent is needed for material regeneration 2. 60% of materials cost (GAC) is needed for one time GAC regeneration 3. Cost of organic solvent is 1200 JPY/L
7.5.5 Results
Nine different combinations of removal efficiencies and material regenerations were compared in terms of financial aspects. It was found in each calculation that Dow V493 was the least cost effective followed by Dow L493. Amb XAD 4 and GAC showed similar range of cost effectiveness for 90% and 99% PFOS removals. For the higher removal of 99.75%, Amb XAD4 showed the best cost effectiveness.

7.5.5.1 Factors affecting cost effectiveness

Higher sorption capacities
Since the environmental PFOS concentration is in ng/L level, it is essential to meet high removal efficiencies at the unit operation to treat PFOS, especially at industrial wastewater treatment. It was observed in this experiment that non ion-exchange polymers, especially Amb XAD4 showed an excellent performance for PFOS removal. Higher sorption capacity at lower equilibrium concentrations (outflow concentration) increases the duration of column operation between material regeneration which ultimately reduces the operation cost.

Material regeneration by organic solvent
Despite the high initial capital investment in resins compared to GAC, resins can be regenerated and reused on-site whereas carbon must be taken off-site, reactivated, and replaced. Consequently, if resin regeneration costs are sufficiently low and resin usage rates are less than carbon usage rates, the lifecycle costs for a resin system can be less than those for a carbon system.

Hydraulic retention time.
The hydraulic retention time applied in the experiment was 1.35 min, which is less than conventional GAC treatment units. It was observed that a comparatively better performance was given by non ion-exchange polymers than GAC with this retention time. Similar performance can be expected from the GAC column with lower flow rate. Consequently, resin vessels can sustain a much higher flow-through than equivalently sized carbon vessels, resulting in either a reduction in the total number of required resin vessels or a reduction in the size of the resin vessels. The smaller volume of resins required may lead to lower costs.
Minimal interference with other organics.
The results of combine treatment of coagulation and adsorption suggest the interferences for adsorption by other organics are comparatively less for some non ion-exchange polymers than GAC. This observation indicates the direct applicability of non ion-exchange polymers especially in the industrial wastewater.

7.6 Summary

1. Combined treatment process of coagulation (by organic coagulants) followed by adsorption and filtration was identified as an excellent method to treat PFCs in water. The combination with some adsorbents, more than 99% removal was obtained even after 100 days of continuous run.

2. Removal of PFCs by coagulation was higher than that determined by the jar test in Chapter 5. This may be due to the higher retention time at slow mixing tank.

3. Addition of coagulants negatively affected the adsorption of most of the adsorbents, but for AmbXAD4, the effect was positive.

4. Economical analysis with different scenarios for the level of treatment and regenerations identified AmbXAD as the cheapest material for PFOS adsorption.
8.1 Conclusions

Perfluorinated compounds are anthropogenic organic pollutants with the properties of persistence, bio-accumulation and toxicity. Many researchers have reported that conventional water and wastewater treatment methods cannot eliminate PFCs. Fully saturated C-F bonds give the stability and polarity to the PFCs molecules. This study tried to eliminate PFCs by using the property of polarity. Main removal mechanisms tested were adsorption and coagulation. The conclusions are drawn by chapters as follows:

In Chapter 2, available literature was summarized, basically on PFCs treatment. It was noticed that PFCs oxidation is extremely difficult at environmental temperature and pressure. Adsorptions by GAC and membrane filtration were identified as attractive methods to eliminate PFCs.

Chapter 3 tested the ferrate technique to oxidize PFCs at trace concentrations (from 0.1 to 25 µg/L). Ferrate(VI) technique has been identified as an emerging water purification technique and proved its effectiveness to treat many organic and inorganic pollutants. Conclusions of this chapter are shown below

1. The best ferrate to PFCs molar ratio to reach maximum PFCs reduction was identified as 100.
2. It was noticed that the average elimination of PFCs with sulfonane functional group was 18% and acid functional group was 24%.
3. PFCs reduction by ferrate(VI) might occur due to the adsorption than oxidation. It was also found that high ferrate(VI) to PFC molar ratio discouraged PFC removal. Coagulation property of ferrate(VI), which flocculate ferrite(III) particles might be reducing the adsorption effect.
4. It was concluded that ferrate technique alone is not sufficient to oxidize PFCs at environmental pressure and temperature. The oxidation numbers of carbon atoms in the PFCs chain are +2 and +3.
5. Since the ferrate(VI) can oxidize many pollutants in water except PFCs, it might be useful in methodology development in PFCs determination, especially to degrade organic matrices in wastewater samples.
Chapter 4 studied the PFCs adsorption by batch experiments. Ion-exchange polymers, non ion-exchange polymers and GAC were studied in detail for kinetic and isotherm characteristics. Conclusions of this chapter were shown as follows.

1. Constant sorbent method was identified as the best method to determine sorption isotherm, where sorbent concentration was kept constant and sorbate concentration was changed.

2. Ion-exchange resins and GAC showed faster adsorption characteristics than non ion-exchange polymers. To reach the equilibrium concentration for all kind of granular materials, at least 100 hrs shaking was needed.

3. Synthetic polymer materials were identified as better filter materials (in terms of adsorption capacity) to eliminate PFCs in water at low concentration (1 μg/L). The magnitude of Freundlich isotherm constants (Kf) decreases in the following order for most of the long chain and medium chain PFCs tested: Ion-exchange polymers > Non ion-exchange polymers > GAC, but in some cases at further low equilibrium concentrations (100 ng/L) non ion-exchange polymers showed higher adsorption capacity than other adsorbents.

4. Amb IRA·400 was identified as the best filter material to eliminate PFOS at equilibrium concentrations > 1 μg/L. Considering both adsorption isotherm and adsorption kinetics, Amb XAD 4 and Dow MarathonA can be recommended for eliminating PFOS at ng/L equilibrium concentrations.

5. Further studies are to be done to understand the sorption mechanisms of PFOS, but it can be suspected that the chemisorption must be the dominant process.

Chapter 5 studied PFCs coagulation by a series of jar tests. Long chain polymer type cationic coagulants and conventional inorganic coagulants were comparatively studied for PFCs coagulation. Conclusions are listed in detail as follows.

1. The experiment with deionized water spiked with PFCs suggested that comparatively higher coagulant dose is required to get a good PFCs removal efficiency by organic coagulants.
2. The results of the experiments with deionized water spiked with PFCs and wastewater spiked with PFCs showed that the efficiency of PFCs coagulation by organic coagulants is almost double than that of inorganic coagulants.

3. Among the organic coagulants tested, FL 2749 was identified as the best candidate to eliminate any PFCs.

4. The results of PFCs coagulation by different organic coagulants suggest that a mixture of multipolymers is better than single polymer coagulant to coagulate PFCs.

5. The results of the experiment from actual wastewater spiked with PFCs indicated that existence of other organic matter discourages coagulation of short chain PFCs, but it encourages the coagulation of long chain PFCs.

6. The results of the experiment from actual industrial wastewater related to PFCs industry indicated that organic coagulants are not effective as it showed in wastewater spiked with PFCs to coagulate PFCs. The PFCs that appear in real wastewater seems to be incorporated with other polar molecules in the wastewater.

7. Organic coagulation followed by microfiltration seems to be an effective combination to eliminate PFCs for some wastewaters, but more studies to be done on this topic.

Chapter 6 studied the PFOS adsorption by column experiment. Non ion-exchange synthetic polymers identified in chapter 4 were tested with long run column experiment (60 days) for PFOS adsorption. At the end of the experiment, the granular materials were tested for PFOS recovery. The conclusions of the chapter are summarized as following.

1. Amb XAD 4 was recognized as the best candidate among the tested four filter materials to eliminate PFOS.

2. It was observed that the results of column experiment cannot be explained by individual results of the isotherm experiment or the kinetic experiment suggesting the importance of both isotherm (available vacant sites in the materials to adsorb) and kinetics (the speed of attachment) in the real application.

3. It was interesting to notice that in the column experiment with Amb XAD 4 removed 99.99% PFOS up to 23,000 bed volumes pass through with the condition of 10 μg/L inflow concentration and flow rate of 15 ml/min (0.75 bed volumes/min).
4. Within 80 min, all most 100% PFOS was recovered from synthetic resins in the regeneration experiment suggesting that synthetic polymers can be effectively regenerated by an organic solvent.

Chapter 7 studied the combined treatment process of adsorption (studied in Chapter 4 and Chapter 6) and coagulation (studied in Chapter 5) for PFOS elimination. This study was done in a continuous run column experiment (>100 day). Later part of Chapter 7 calculated the cost effectiveness of each polymer material. The conclusions of this chapter are summarized as following.

1. Combined treatment process of coagulation (by organic coagulants) followed by adsorption and filtration was identified as an excellent method to treat PFCs in water. Some polymers showed more than 99% removal even after 100 days of continuous run.

2. The higher removal of PFCs by coagulation than that was determined by the jar test at Chapter 5 may be due to the higher retention time at slow mixing tank.

3. Addition of coagulants negatively affected the adsorption of most of the adsorbents, but for AmbXAD4, the effect was positive.

4. Economical analysis with different scenarios for the level of treatment and regeneration identified AmbXAD 4 as the cheapest material for PFOS adsorption.

8.2 Recommendations

The results of this study suggest the following recommendations. A limited number of synthetic materials were tested in this experiment. Four synthetic resins out of six (two ion-exchange resins and four non ion-exchange resins) studied in this experiment showed better performance than conventional GAC for PFCs adsorption. It is highly recommended to test more synthetic materials for material optimization. Also the dominancy of chemisorption was observed and further studies are recommended for complete understanding of PFCs adsorption process. Medium molecular weight (around 100,000Da) cationic organic coagulants showed the best PFCs coagulation and further studied to be done to confirm it. Organic coagulation followed by micro filtration seems to be an effective combination for tap water treatment and further research on this is recommended. Among the inorganic coagulants, ferric chloride showed better performance for PFOA coagulation and it can be recommended for WWTPs. In
the column experiment, best performance was given by Amb XAD4. It was found that this material can be regenerated on site (solvent regeneration) and economically viable than other granular materials tested. It is highly recommended to install a filter column with Amb XAD4 in a PFCs related industry to study the performance in the real field conditions.
References


27 Chuyang Y. Tanga, Q. Shiang Fub, c, d, Dawen Gao, e, Craig S. Criddled and James O. Leckied, 2010, Effect of solution chemistry on the adsorption of perfluorooctane sulfonate onto mineral surfaces, Water Research 44(8), 2654-2662


38 Environmental health perspectives National Institutes of Health U.S. Department of Health and Human Services

39 Environmental Science and Technology 31 (9): 2445-2454.


54 Hatfield, T. L. (2001a), Hydrolysis reactions of perfluorooctanoic acid (PFOA), 3M Environmental Laboratory, USEPA docket: EPA-HQ-OPPT-2002-0051-0013.

55 Hatfield, T. L. (2001b), Screening studies on the aqueous photolytic degradation of perfluorooctanoic acid (PFOA), 3M Environmental Laboratory, USEPA docket: EPA-HQ-OPPT-2002-0051-0023.


60 Higgins C.P and Luthy R.G, 2006, Sorption of perfluorinated surfactants on sediments, Environmental Science and Technology 40 (23), 7251–7256


128 Ma, J., Liu, W., 2002-1. Effectiveness of ferrate (VI) peroxidation in enhancing the coagulation of surface waters. Water Res. 36, 4959–4962.


137 Melzer, D., Rice, N., Depledge, H.M., Henley, e.W., Tamara, Galloway, S.  2010. Association Between Serum Perfluorooctanoic Acid (PFOA) and Thyroid Disease in the NHANES Study. Env. Health perspective


Technol. 39(9), 3388-3392.


146 Ochoa-Herrera V and Sierra-Alvarezb R 2008, Removal of perfluorinated surfactants by sorption onto granular activated carbon, zeolite and sludge, Chemosphere 72 (10), 1588–1593


150 Oppenlander T, Gliese S. Mineralization of organic micropollutants (homologous alcohols and phenols) in water by vacuum-UV oxidation (H2O-VUV) with an incoherent xenon-excimer lamp at 172 nm. Chemosphere, 2000, 40(1): 15–21

151 Osborne M C, Li Q, Smith I W M. Products of the ultraviolet photodissociation of trifluoroacetic acid and acrylic acid. Physical Chemistry Chemical Physics, 1999, 1(7): 1447–1454


153 Parker G.R. and Bortko S, 1991.“Groundwater Remediation Using Ambersorb Adsorbents.” Presented at the Florida Environmental Chemistry Conference. October 30 - November 1,


156 Prevedouros, K., Cousins, I.T., Buck, R.C., Korzeniowski, S.H., 2006. Sources, fate and


pregnancy in rat and mouse. I: Maternal and prenatal evaluations, Toxicol. Sci., 74 (2), 369-38


201 US EPA (2002), Hazard assessment of perfluorooctanoic acid and its salts, 107 pp, Office of Pollution Prevention and Toxics - Risk Assessment Division, US.

202 US EPA (2003), Preliminary risk assessment of the developmental toxicity associated with exposure to perfluorooctanoic acid and its salts, 64 pp, Office of Pollution Prevention and Toxics - Risk Assessment Division, US


204 Vandiver M. and Isacoff E.G., 1994 “THM Reductions with Ambersorb 563 Adsorbent.” Presented at the Society of Soft Drink Technologists 41st Annual Conference, Albuquerque, NM. April


156
218 Yong, Q., Fujii, S., Tanaka, S. and Koizumi, A. 2006 Preliminary study on the treatment of Perfluorinated chemicals (PFCs) by Advanced Oxidation Process (AOPs). The 12th Seminar of JSPS-MOE Core University Program on Urban Environment, Kyoto, Japan