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1 **Cellulase Activity in Meiobenthos in Wetlands**

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Abstract

To validate the involvement of meiobenthos in cellulose breakdown in wetlands, meiobenthos were collected from the sediments of Lake Furen and the Biwase River in Hokkaido prefecture, the Kako River in Hyogo prefecture, and the Chinai River in Shiga prefecture. Cellulase activities of the meiobenthos were measured by cellulose zymographic analysis using SDS-PAGE gels containing 0.5% carboxymethyl cellulose. The results showed that most of the Turbellaria, Nematoda, Harpacticoida and Oligochaeta species exhibited cellulase activity. The molecular sizes of the cellulase-active bands of the sediments in Lake Furen, the Biwase River, and the Chinai River coincided with those of meiobenthos. The findings suggest that meiobenthos might play a major function in cellulose breakdown in these wetlands. This paper is the first to report cellulase activity in meiobenthos and that they are possibly involved in the breakdown of cellulose in wetlands.

KEY WORDS: cellulase • cellulose • meiobenthos • sediment • wetland

Introduction

31 Cellulose, the most abundant organic matter on earth, is a high molecular weight
32 substance consisting of glucose residues bound by β -1,4 linkages, unlike starch, another
33 glucan consisting of a α -1,4 linked glucose residues. Cellulose is resistant to enzyme
34 degradation [1, 2]. However, cellulose can be degraded by specific enzymes collectively
35 named cellulases [3].

36 Fungi and bacteria as well as symbiotic protozoa in herbivorous animals and
37 termites have been studied as known consumers of cellulose [4]. Recently, an intrinsic
38 cellulase gene was isolated from termite [5], and a variety of intrinsic cellulase genes
39 were identified from various animals, including beetle [6], nematode [7, 8], abalone [9],
40 mussel [10], sea urchin [11], and brackish-water clam [12].

41 Recent stable isotopic analysis showed that a brackish water clam *Corbicula*
42 *japonica* consumes land-derived organic materials mainly composed of cellulose [13].
43 Identification of the intrinsic cellulase gene and immunological detection of the enzyme
44 protein in *C. japonica* strongly suggest that *C. japonica* plays an important role in the
45 process of degradation of cellulose in rivers [14].

46 Besides macrobenthos such as *C. japonica*, a group of small animals called
47 meiobenthos also inhabits the sediments of aquatic areas. Meiobenthos are defined as

animals that pass through a 1-mm mesh filter and are known to be composed of a variety of fauna corresponding to 22 phyla [15].

In the present study, we attempted to validate the role of meiobenthos in the process of breakdown of cellulose in wetlands. We chose Lake Furen and the Biwase River located in the subarctic area, since they were reported to be the wetlands demonstrating the highest cellulase activities in Japan [16]. On the other hand, we chose the Kako River and the Chinai River as typical rivers in temperate area. We also expected the difference in distribution of meiobenthos between the Kako River and the Chinai River, because the Chinai River is a fresh water river. We report that meiobenthos have cellulase activity and possibly play substantially important roles in the cellulose degradation process in the sediments of some wetlands.

Materials and Methods

Materials

We collected sediments from Lake Furen and the Biwase River in Hokkaido prefecture, the Kako River in Hyogo prefecture, and the Chinai River in Shiga prefecture. River

66 sediments from all rivers were collected in the wetland within 50 m from the river
67 mouth. From each site, we collected sediment samples from a 5-cm depth at low tide
68 from May 2006 to October 2007. The sediment samples were transported to the
69 laboratory in Kyoto University at 4°C, and meiobenthos were recovered in the fraction
70 that included material small enough to pass through a 1mm-mesh filter but too large to
71 pass through a 40 µm-mesh filter. These meiobenthos were classified under microscopic
72 observation according to Higgins and Thiel [15]. About 500 g of the sediments were
73 filtered through 1 mm-mesh. Sediment samples that were not filtered were designated
74 “total sediment fraction,” while sediments less than 1 mm and larger than 40 µm were
75 collected and designated “meiobenthos fraction.”

76

77 Measurement of cellulase activity

78

79 Meiobenthos were separated from the sediments by using a pair of tweezers under the
80 microscope (Olympus, S2X12, Tokyo), and each single meiobenthos was homogenized
81 with 20 µl of phosphate-buffered saline (PBS) containing 140 mM NaCl, 2.7 mM KCl,
82 8 mM Na₂HPO₄, and 1.5 mM KH₂PO₄ (pH 7.4) to prepare a meiobenthos extract. The
83 total sediment and meiobenthos fractions were homogenized with 1.5-fold volume of

PBS, and the supernatants were obtained by centrifugation at $10,000 \times g$ for 10 min. Ten microgram of the supernatant was applied on cellulase zymographic analysis. Cellulase zymographic analysis was performed as described previously by using 7.5% or 10% SDS-PAGE gel containing 0.5% carboxymethyl cellulose (CMC, Sigma, St Louis, MO, US). After electrophoresis, the gels were soaked in 10 mM acetate buffer (pH 5.5) containing 0.1% TritonX-100 for 30 min to remove SDS from the gels. The gels were transferred to 10 mM acetate buffer (pH 5.5), incubated at 37°C overnight, and then stained with 0.1% Congo Red. The gels were destained using 1 M NaCl. The active bands were detected as non-stained bands. Unless otherwise specified, special grades of reagents were commercially obtained from Nacalai Tesque (Kyoto). Protein concentration was measured according to the method of Bradford (17).

Results

Distribution of meiobenthos

Oligochaeta species were the dominant meiobenthos in Lake Furen, where the sediments are mainly composed of sand. In addition, a variety of Turbellaria, Nematoda,

102 Harpacticoida species were also observed. Oligochaeta species were also dominant in
103 the Chinai River, where the sediments were, like in Lake Furen, mainly composed of
104 sand.

105 In the Biwase and Kako Rivers, where the sediments were mainly composed of
106 clay, Nematoda and Harpacticoida species were dominantly observed.

107

108 Cellulase activity of sediments and meiobenthos

109

110 Cellulase activity in meiobenthos extracts, total sediment fractions, and meiobenthos
111 fractions were measured by cellulose zymography.

112 Cellulase activity bands of the total sediment fraction were detected between

Fig.1

113 32.5 kDa and 47.5 kDa in samples from Lake Furen, which coincided with those of the

114 meiobenthos fraction (lanes 1 and 2 in Fig. 1). Oligochaeta species assayed by using a

115 single animal showed an active band corresponding to 32.5 kDa, which is the band size

116 observed for the total sediment fraction and the meiobenthos fraction (lane 3 in Fig. 1).

117 Different species of Nematoda showed active bands of different molecular sizes, as

118 shown in lanes 4–7 of Fig. 1. Interestingly, common active bands at 25 kDa were

119 detected for all the Nematoda species. Various active bands were also detected for

120 Turbellaria species (lane 8 in Fig. 1), but their sizes differed from those of Nematoda
121 species.

122 Figure 2 shows the cellulase activity in sediment samples corresponding to
123 approximately 5 mg per one lane from the Biwase River. Intensive active bands were
124 observed, especially above 47.3 kDa in the total sediment fraction and the meiobenthos
125 fraction (lanes 1 and 2 in Fig. 2), and the active band patterns were nearly identical.
126 Harpacticoida species, the dominant organisms of meiobenthos in the Biwase River,
127 also exhibited remarkably intensive activity bands of above 47.3 kDa (lane 3 in Fig. 2).

Fig.2

128 Figure 3 shows the active bands of the samples from the Chinai River. Faint
129 active bands were observed at 25 kDa and between 47.5 kDa and 62 kDa in the total
130 sediment fraction and the meiobenthos fraction (lanes 1 and 2 in Fig. 3). Bands with
131 nearly the same activity were observed for Oligochaeta species, as shown in lane 3.

Fig.3

132 Figure 4 shows the active bands in the Kako River samples. In the total
133 sediment fraction and the meiobenthos fraction, faint bands of less than 25 kDa were
134 observed (lanes 1 and 2 in Fig. 4). In lane 3, active bands of Harpacticoida species were
135 observed at approximately 47.5 kDa, which did not coincide with the active bands in the
136 total sediment fraction and the meiobenthos fraction.

Fig.4

137

138 **Discussion**

139 Cellulase activities were detected in the extracts of the meiobenthos collected from all
140 the sampling sites examined, suggesting that meiobenthos may be involved in the
141 breakdown of cellulose in the sediments. Interestingly, 25 kDa active bands were
142 commonly detected in the extracts of morphologically distinct Nematoda species
143 collected from Lake Furen (lanes 4–7 in Fig. 1), suggesting that Nematoda species
144 possibly share a related cellulase gene. On the other hand, Oligochaeta species collected
145 from Lake Furen and the Chinai River were morphologically distinct and the band sizes
146 of the cellulase were also different (compare lane 3 in Fig. 1 and lane 3 in Fig. 3).

147 The International Collaborative Research on the Management of Wetland
148 Ecosystem of the National Institute for Environmental Studies [16] reported the
149 outstanding strong cellulase activities of sediments collected from Lake Furen and the
150 Biwase River in Hokkaido among many Japanese wetlands tested. The report attributed
151 the strong cellulase activity in the sediments of Lake Furen and the Biwase River to
152 microorganisms including bacteria and fungus. However, the active bands in cellulose
153 zymographic analyses showed that the position of the active bands coincided with the
154 sediment fractions and the extracts of meiobenthos (Figs. 1 and 2), which supported the

155 hypothesis that meiobenthos might be involved in the breakdown of cellulose in Lake
156 Furen and the Biwase River.

157 Because recent molecular biological studies suggest the endogenous origin of
158 the cellulase genes in aquatic invertebrates [9, 11, 12], cellulase genes of meiobenthos
159 could be encoded in the DNA of meiobenthos themselves. We are now trying to clone
160 the cellulase genes of meiobenthos to validate the possibility of its endogenous origin.

161 As shown in Fig. 3, active bands of the total sediment fraction and the
162 meiobenthos fraction coincided with those of Oligochaeta species from the Chinai River.
163 On the other hand, active bands of the total sediment fraction and the meiobenthos
164 fraction did not coincide with those of the Harpacticoida species from the Kako River,
165 as shown in Fig. 4. Thus, the origin of cellulase could not be concluded to be the
166 meiobenthos in the case of Kako River sediment. Further studies are required to
167 evaluate the contribution of meiobenthos to the breakdown of cellulose in wetlands in
168 the temperate area.

169 The contribution of termites to the breakdown of cellulose in the forests of
170 tropical zones is assumed to correspond to 80% the total cellulose breakdown in this
171 area [18]. Like termites, meiobenthos could be major consumers of cellulose, especially
172 in some wetlands in Hokkaido, because cellulase activity of meiobenthos in Lake Furen

173 and the Biwase River were detected at 4°C (data not shown), which is a temperature at
174 which the growth of bacteria and fungi would be suppressed. Therefore, it seems
175 probable that meiobenthos would play important roles in cellulose degradation
176 especially in low temperature environments like wetlands in Hokkaido.

177

178

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182

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- 231

232 Figure legends

233

234 **Fig. 1.** Cellulase activities in Lake Furen. Activities were detected in 10% SDS-PAGE
235 gel containing 0.5% carboxymethyl cellulose. The positions for molecular mass marker
236 proteins are shown by arrows. Lanes: 1, total sediment fraction; 2, meiobenthos
237 fraction; 3, Oligochaeta species; 4–7, morphologically distinct species of Nematode; 8,
238 Turbellaria species.

239

240 **Fig. 2.** Cellulase activities in the Biwase River. Ten percent SDS-PAGE gel containing
241 0.5% carboxymethyl cellulose was used for the detection of cellulase bands of the
242 sediments, while 7.5% gel was used for Harpacticoida species. The positions for
243 molecular mass marker proteins are shown by arrows. Lanes: 1, total sediment fraction;
244 2, meiobenthos fraction; 3, Harpacticoida species.

245

246 **Fig. 3.** Cellulase activities in the Chinai River. Activities were detected in 10%
247 SDS-PAGE gel containing 0.5% carboxymethyl cellulose. The positions for molecular
248 mass marker proteins are shown by arrows. Lanes: 1, total sediment fraction; 2,
249 meiobenthos fraction; 3, Oligochaeta species.

250

251 **Fig. 4.** Cellulase activities in the Kako River. Activities were detected in 10%

252 SDS-PAGE gel containing 0.5% carboxymethyl cellulose. The positions for molecular

253 mass marker proteins are shown by arrows. Lanes: 1, total sediment fraction; 2,

254 meiobenthos fraction; 3, Harpacticoida species.

255

(Figure 1)

(Figure 2)

(Figure 3)

(Figure 4)

和文要旨

湿地帯に生息するメイオベントスのセルラーゼ活性

豊原治彦，朴 煥華，土屋佳奈子，劉 文（京大院農）

湿地帯におけるセルロース分解に関わるメイオベントスの役割を明らかにする

目的で，琵琶瀬川と風連湖（北海道），知内川（滋賀県），加古川（兵庫県）の

底泥に生息するメイオベントスのセルラーゼ活性を測定し，ほとんどのメイオ

ベントスに活性を認めた。とくに風連湖，琵琶瀬川および知内川ではザイモグ

ラフィー分析により，底泥とメイオベントスの活性バンドのサイズが一致した

ことから，これらの湿地帯においてはセルロース分解にメイオベントスが主要

な役割を果たしていると考えられた。