

1 Mucosal IgA induction in calves

2 **Effects of Feeding Whey Protein on Growth Rate and mucosal IgA**
3 **Induction in Japanese Black Calves**

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19 **ABSTRACT:** Data from 63 Japanese Black calves were collected to clarify the effects of
20 feeding whey protein on the growth rate and mucosal IgA induction in calves. Dietary
21 treatments in milk replacers were 1) 26% CP as in skim milk (control), 2) 26% CP as whey

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22 and skim milk and 3) 26% CP as whey. Diets were offered from 3 to 63 days of age in calves.
23 Feeding whey protein had no effects on growth rate, fecal consistency and fecal water in
24 calves. Compared with 2 days of age, fecal IgA concentration in calves decreased at 14 days
25 of age, while fecal water increased. Feeding whey protein increased fecal IgA in calves after
26 14 days of age, which was thought to be the increased mucosal IgA induction in the gut.
27 Serum cholesterol concentration tended to be lower in calves fed whey than in control group,
28 but feeding whey protein had no clear effects on serum glucose, NEFA, total protein and
29 urea-N concentrations. These results suggest that feeding whey protein enhances mucosal IgA
30 induction in calves, but feeding whey protein has little effect on growth rate and fecal
31 consistency in calves.

32

33 **Key Words:** neonatal calves, mucosal IgA induction, whey, milk replacer

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35 **1. Introduction**

36 Whey protein concentrate has an adequate amino acid profile than that in dried skim
37 milk and casein, and higher proportion of whey protein concentrate in milk replacers
38 improved calf performance when only milk replacer was fed (Lammers et al., 1998). The
39 absorbed and retained N in dairy calves fed 16.1, 18.5, 22.9 and 25.8% of CP from whey
40 protein sources increased linearly as dietary CP increased (Blome et al., 2003). In the
41 previous study (Nishiyama et al., 2011b), daily gains of calves fed whey protein or skim milk
42 at 26% CP were very similar, because the appropriate supply of CP in the diets maintained
43 normal growth rate of calves.

44 Mortality and morbidity of neonates continue to be major problems in calves, and their
45 most common disease is diarrhea, which can cause growth retardation and death of calves.
46 Successful neonatal health depends on many factors related to management and nutrition, but

47 the improvement of the immune system is required for preventing diarrhea. Whey protein
48 concentrate contains antiviral and immunomodulatory components, and supplemental whey
49 protein concentrate reduces rotavirus-induced disease symptoms in suckling mice (Wolber et
50 al., 2005) and enhances mucosal innate immunity during early life in suckling rats
51 (Perez-Cano et al., 2007).

52 Passive immunity is critical to the survival and health of neonates, and colostrum or milk
53 is a source of nutrients and immune components for neonatal calves (Blum, 2006). IgA is the
54 most abundant Ig isotype in mucosal secretions and provides protection against microbial
55 antigens at mucosal surfaces (Fagarasan and Honjo, 2003; Mora and von Andrian, 2009).
56 Most IgA antibody secreting cells (ASC) express chemokine receptor CCR10, but IgA ASC
57 from CCR10-deficient mice do not efficiently accumulate in the lactating mammary gland
58 and lead to a significant decrease in milk IgA and fecal IgA of neonatal mice (Morteau et al.
59 2008). Additionally, the mucosal immune induction is also needed in neonatal calves, because
60 the disease resistance acquired from colostrum Ig is only temporary (Quigley and Drewry,
61 1998). In the previous studies (Nishiyama et al., 2011a, 2011b), supplemental β -carotene with
62 whey to maternal mice during pregnancy and lactation is useful to increase IgA transfer from
63 maternal milk to neonatal mice, while supplemental β -carotene with whey may have little
64 effect on mucosal IgA induction in neonatal mice and calves. However, supplemental whey
65 protein has been expected to enhance mucosal IgA induction in neonatal calves owing to the
66 high level of fecal IgA at 14 days of age (Nishiyama et al., 2011b).

67 The objective of this study was to clarify the effects of feeding whey protein on daily
68 gains, fecal consistency and levels of fecal IgA in Japanese Black calves in order to evaluate
69 the role of whey protein on the growth rate and mucosal IgA induction in calves.

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71 **2. Materials and methods**

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73 *2.1. Experimental design*

74

75 This research was approved by the guide for the care and use of animal in Shiga Prefectural
76 Livestock Technology Promotion Center (Hino, Japan), Northern Center of Agricultural
77 Technology (Asago, Japan), Nara Prefectural Livestock Technology Center (Mitsue, Japan)
78 and the Livestock Technology Center Department of The Kyoto Prefectural Agriculture,
79 Forestry & Fisheries Technology Center (Ayabe, Japan). Sixty three Japanese Black calves
80 born in their centers were used, and calves consisted of 43 males and 20 females.

81 Dietary treatments in milk replacers were 1) commercial milk replacer which contained
82 26% CP as in skim milk (control group, 14 males and 7 females), 2) experimental milk
83 replacer which contained 26% CP as whey and skim milk (whey plus skim milk group, 14
84 males and 7 females) and 3) experimental milk replacer which contained 26% CP as whey
85 (whey group, 15 males and 6 females). These milk replacers were provided by Chubu Shiryo
86 Co. Ltd (Ohbu, Japan), and dietary ratio of protein source and chemical composition in milk
87 replacers are shown in Table 1.

88 Calves lived with their dams after birth and received only their dam's colostrum, but
89 colostrum IgA was not determined. At 3 days of age, calves were separated from their dams
90 and housed in individual pens. From 3 to 63 days of age, calves received appropriate amounts
91 of milk replacers and calf starter pellets to meet recommendations (Agriculture, Forestry, and
92 Fisheries Research Council Secretariat, 2000) for TDN, protein and minerals of calves. The
93 amounts of milk replacers offered to calves were increased from 0.5 to 0.9 kg/d during 3 to 15
94 days of age, maintained at 1.0 to 1.3 kg/d (Mean \pm SD, 1.05 \pm 0.05 kg/d) during 16 to 50
95 days of age and decreased by 0.25 kg/d during 51 to 63 days of age. Milk replacers were

96 diluted with warm water at 40°C and offered twice a day throughout the experiment. Calf
97 starter pellets (TDN, 75%; CP, 20%) were offered from 7 days of age, and the amounts of calf
98 starter were gradually increased by 63 days of age, according to the pellet refusals of calves.
99 Intake of milk replacers and calf starter pellets were measured every day, and their data were
100 averaged by each week. Additionally, the calves were given free access to timothy hay from
101 20 days of age.

102

103 *2.2. Sample collection and analyses of serum components and fecal IgA*

104

105 Body weights of calves were measured on day 0, 7, 14, 21, 28, 42, 56 and 63 after birth.
106 Fecal consistency of calves was observed every day throughout the experiment. Fecal scores
107 were measured on a scale of 1 to 3 (1= firm, normal; 2=soft, 3=watery), and their data were
108 averaged by each week. Blood and fecal grab samples were collected at 13:00 hour on day 2,
109 14, 28, 42 and 56 after birth. Blood was sampled by a jugular vein puncture into vacuum
110 tubes, left to stand at room temperature for 1 hour and centrifuged at 3,000 × g for 15 min.
111 Serum glucose, total protein, nonesterified fatty acid (NEFA), triglyceride, urea N and
112 cholesterol were determined by an Automatic analyzer (7600, Hitachi, Tokyo, Japan). Fecal
113 water and fecal IgA were determined as previously described (Nishiyama et al., 2011b).

114

115 *2.3. Statistics*

116

117 Data of body weight, daily gain, feed intake, fecal score and components of serum and
118 feces were analyzed by least squares ANOVA using the general linear model procedure of
119 SAS (1997). The model was as follows;

$$120 \quad Y_{ijk} = u + D_i + E_j + C_{(ij)k} + T_1 + DT_{il} + e_{ijkl}$$

121 where u is the overall mean, D_i is the effect of diet, E_j is the effect of the experimental
122 center, $C_{(ij)k}$ is the random variable of calves nested in diet and experimental center, T_1 is the
123 effect of time, DT_{ij} is the interactions, and e_{ijkl} is the residuals. Data obtained from serum and
124 feces at 14, 28, 42 and 56 days of age were used for this model. In addition, the general linear
125 model procedure of SAS (1997) was used to analyze the effect of time on fecal content and
126 fecal IgA at 2, 14, 28, 42 and 56 days of age.

127 An ANOVA was performed, and the differences were tested by Tukey-Kramer's multiple
128 comparisons. Significance was declared at $P < 0.05$.

129

130 **3. Results**

131 Body weights and daily gains of calves were not affected by treatment. Body weights
132 increased from 31.2 kg at birth to 77.0 kg at 63 days of age, and weight gains in control group
133 was slightly high level (Table 1). Calves were fed almost all the milk replacers, and calf
134 starter intake was not affected by treatment. Calf starter intake increased from 41g/d at 2
135 weeks of age to 285g/d at 6 weeks of age and reached at 906g/d at 9 weeks of age.

136 Fecal scores of calves were almost similar in the control, whey plus skim milk and whey
137 groups, and fecal water was not affected by treatment (Table 2). Compared with 2 days of age,
138 fecal water increased ($P < 0.001$) at 14 days of age, while fecal IgA concentration decreased
139 ($P < 0.001$) (Fig.1). Fecal IgA concentration in calves after 14 days of age was significantly
140 higher in whey group than in control ($P < 0.001$) and whey plus skim milk ($P < 0.05$) groups.
141 Compared with control group, fecal IgA concentration in calves fed whey plus skim milk
142 group was slightly high level.

143 Serum glucose and total protein concentrations in calves were not affected by treatment
144 (Table 3). Serum cholesterol concentration tended to be lower ($P < 0.10$) in whey group than in
145 control group, and serum triglyceride concentration in whey group was slightly high level.

146 Serum NEFA concentration at 28 days of age and serum urea-N concentration at 14 days of
147 age were higher ($P<0.05$) in whey plus skim milk group than in control group.

148

149 **4. Discussion**

150

151 *4.1. Effects of feeding whey protein on growth rate in calves*

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153 Whey protein concentrate is useful for calf growth (Lammers et al., 1998; Blome et al.,
154 2003), and the lean tissue gain of calves continued to increase with dietary CP up to 26%
155 when calves were fed at 1.75% of BW daily (Bartlett et al., 2006). Terosky et al. (1997)
156 reported that diets varying in the ratio of dried skim milk to whey protein concentrates at 20.6
157 to 21.1% CP had no effects on health, growth, apparent digestibility and blood measurements
158 including glucose, total protein, NEFA, triglyceride and urea-N in Holstein calves up to 8
159 weeks of age. In the previous (Nishiyama et al., 2011b) and present studies, feeding whey
160 protein had no effects on the growth rates and serum glucose, total protein, NEFA,
161 triglyceride and urea-N in Japanese Black calves. The lower serum cholesterol in calves fed
162 whey protein agreed with the previous reports (Nagaoka et al., 1992; Sautier et al., 1983),
163 which showed that serum cholesterol was lower in the rats fed whey protein and whey protein
164 exhibited a greater hypocholesterolemic effect in comparison with casein or soybean protein.

165 Severe diarrhetic feces of calves contain more than 85 % moisture, while feces that contain
166 less than 80% moisture are considered as normal (Abe et al., 1999). In the present study, the
167 average fecal water was below 80% in the 3 groups and feeding whey protein had no effects
168 on fecal consistency and fecal water in calves. These results indicate that feeding whey
169 protein at 26% CP may have little effect on the growth rate and fecal consistency in calves.

170

171 *4.2. Effects of feeding whey protein on mucosal IgA induction in calves*

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173 The increased IgA transfer from maternal milk to neonates is important for maintaining
174 normal calf health, because IgA antibodies are specific for antigens of the intestinal
175 microflora and act to limit penetration of commensal intestinal bacteria through the neonatal
176 intestinal epithelium (Harris et al., 2006; Roux et al., 1977). Fecal IgA in calves at 2 days of
177 age was relatively high level, but varied from 0.004 to 59.3 mg/g in the present study. The
178 lower fecal IgA may be inappropriate for newborn calves, but their values of fecal water were
179 almost below 80%.

180 The gut-associated lymphoid tissue is the largest immunologic tissue in the body, and the
181 mucosal immune induction of the newborn gastrointestinal tract is dependent on an active
182 process of IgA ASC accumulation in the gut (Nishiyama et al., 2011a). IgA antibodies
183 produced from IgA ASC in the guts are secreted mainly as dimmers after incorporation of the
184 J chain and association with a transmembrane epithelial glycoprotein known as polymeric-Ig
185 receptor (Fagarasan and Honjo, 2003). In the present study, feeding whey protein increased
186 fecal IgA in calves after 14 days of age, which was thought to be the increased mucosal IgA
187 induction in the gut. In addition, compared with the skim milk feeding, fecal IgA in calves fed
188 whey plus skim milk was slightly high level after 14 days of age.

189 Whey protein concentrate promoted the expansion of cell subsets involved in innate and
190 mucosal immune response in suckling rats (Perez-Cano et al., 2007). The globulin fraction of
191 whey was shown to contain a nondialyzable factor that is chemotactic for IgA-positive
192 lymphocytes (Czinn and Lamm, 1986). In addition, the beta-lactoglobulin in whey protein has
193 more resistance to pepsin degradation than casein, and the undigested beta-lactoglobulin
194 activates IgA production (Takasugi et al., 2001; Wong et al., 1998). These results suggest that
195 feeding whey protein is useful to enhance mucosal IgA induction in calves, and this effect

196 may be partly due to the globulin-mediated activation on IgA ASC accumulation in the gut.
197 However, further studies are needed to evaluate the role of whey protein on the immune
198 system in neonatal calves.

199

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204

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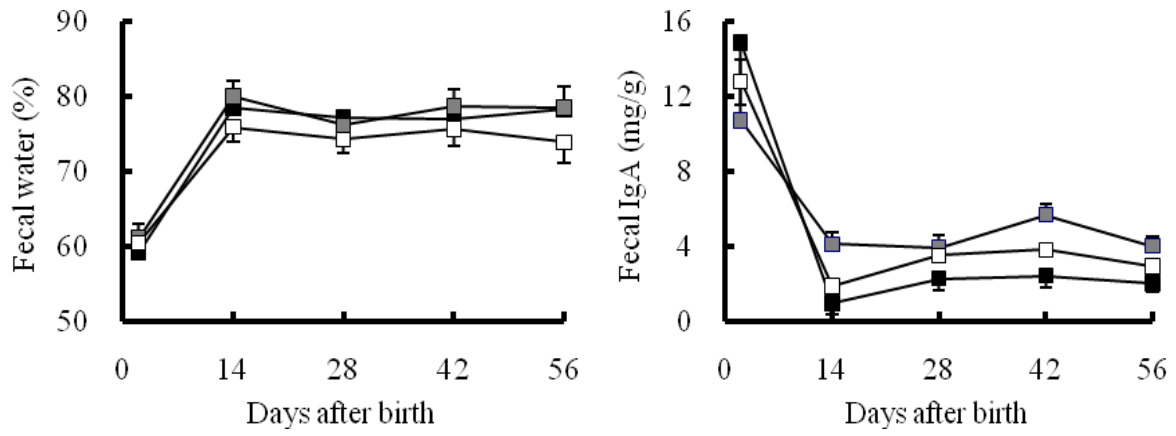
272 Figure 1. Fecal water content and fecal IgA concentration (Mean \pm SE) of calves in control
273 (■), whey plus skim milk (□) and whey (■) groups. Fecal IgA concentration was
274 expressed on a fresh matter basis, and fecal samples were obtained at 2, 14, 28, 42
275 and 56 days of age.

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281 Figure 1.

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Table 1

Dietary ratio of protein source and chemical composition in milk replacers for control, whey plus skim milk (WS) and whey groups in calves.

	Control	WS	Whey
Ingredient (%)			
Skim milk	66.3	28.5	0
Dried whey	3.1	2.8	17.5
Whey protein concentrate	7.4	45.0	56.5
Soybean protein concentrate	2.0	2.0	0
Composition (as-fed basis)			
CP, %	26.3	26.4	26.1
Crude fat, %	17.2	17.3	17.3

Table 2

Daily gain, calf starter intake, fecal score and fecal components for control, whey plus skim milk (WS) and whey groups in calves

	Control	WC	Whey	SE	<i>P</i>		
					Diet	Time	Diet×Time
Daily gain, kg/d	0.771	0.734	0.722	0.022	NS	***	NS
Starter intake, g/d	304	217	263	31	NS	***	NS
Fecal score	1.31	1.36	1.35	0.04	NS	***	NS
Fecal water ¹ , %	77.5	75.3	78.1	2.2	NS	NS	NS
Fecal IgA ¹ , mg/g	1.86 ^B	2.91 ^b	4.67 ^{A,a}	0.45	***	**	NS

*** $P < 0.001$, ** $P < 0.01$.

^{A,B} $P < 0.001$, ^{a,b} $P < 0.05$.

¹ Collected at 14, 28, 42 and 56 days of age.

Table 3

Serum components for control, whey plus skim milk (WC) and whey groups in calves at 14, 28, 42 and 56 days of age

	Control	WC	Whey	SE	<i>P</i>		
					Diet	Time	Diet×Time
Glucose, mg/dl	107.0	110.6	102.0	2.4	NS	NS	NS
Cholesterol, mg/dl	112.1 ^a	103.5 ^{ab}	90.7 ^b	6.5	**	***	NS
NEFA, μ Eq/l	268.7	309.9	280.6	23.0	NS	NS	*
Triglyceride, mg/dl	14.4	14.8	17.1	1.0	*	NS	NS
Total protein, g/dl	5.5	5.6	5.5	0.1	NS	***	NS
Urea-N, mg/dl	11.0	11.5	11.6	0.4	NS	**	*

*** $P < 0.001$, ** $P < 0.01$, * $P < 0.05$.

^{a,b} $P < 0.10$.