<table>
<thead>
<tr>
<th>Title</th>
<th>Design and synthesis of amidine-type peptide bond isosteres: application of nitrile oxide derivatives as active ester equivalents in peptide and peptidomimetics synthesis.</th>
</tr>
</thead>
<tbody>
<tr>
<td>Author(s)</td>
<td>Inokuchi, Eriko; Yamada, Ai; Hozumi, Kentaro; Tomita, Kenji; Oishi, Shinya; Ohno, Hiroaki; Nomizu, Motoyoshi; Fujii, Nobutaka</td>
</tr>
<tr>
<td>Citation</td>
<td>Organic &amp; biomolecular chemistry (2011), 9(9): 3421-3427</td>
</tr>
<tr>
<td>Issue Date</td>
<td>2011-05-07</td>
</tr>
<tr>
<td>URL</td>
<td><a href="http://hdl.handle.net/2433/156967">http://hdl.handle.net/2433/156967</a></td>
</tr>
<tr>
<td>Right</td>
<td>© The Royal Society of Chemistry 2011; This is not the published version. Please cite only the published version.; この論文は出版社版でありません。引用の際には出版社版をご確認ご利用ください。</td>
</tr>
<tr>
<td>Type</td>
<td>Journal Article</td>
</tr>
<tr>
<td>Textversion</td>
<td>author</td>
</tr>
<tr>
<td>School</td>
<td>Kyoto University</td>
</tr>
</tbody>
</table>
Design and Synthesis of Amidine-type Peptide Bond Isostere: Application of Nitrile Oxide Derivatives as Active Ester Equivalents to Peptide and Peptidomimetics Synthesis

Eriko Inokuchi, Ai Yamada, Kentaro Hozumi, Kenji Tomita, Shinya Oishi, Hiroaki Ohno, Motoyoshi Nomizu, and Nobutaka Fujii*

*Graduate School of Pharmaceutical Sciences, Kyoto University, Sakyo-ku, Kyoto 606-8501, Japan

School of Pharmacy, Tokyo University of Pharmacy and Life Sciences, 1432-1 Horinouchi, Hachioji, Tokyo 192-0392, Japan

Corresponding Author:
Nobutaka Fujii, Ph.D.
Graduate School of Pharmaceutical Sciences
Kyoto University
Sakyo-ku, Kyoto 606-8501, Japan
Tel: +81-75-753-4551; Fax: +81-75-753-4570
E-mail: nfujii@pharm.kyoto-u.ac.jp
Abstract: Amidine-type peptide bond isosteres were designed based on the substitution of the peptide bond carbonyl (C=O) group with an imino (C=NH) group. The positively charged property of the isosteric part resembles the reduced amide-type peptidomimetic. The peptidyl amidine units were synthesized by the reduction of a key amidoxime (N-hydroxyamidine) precursor, which was prepared from nitrile oxide components as an aminoacyl or peptidyl equivalent. This nitrile oxide-mediated C–N bond formation was also used for peptide macrocyclization, in which the amidoxime group was converted to peptide bonds under mild acidic conditions. Syntheses of the cyclic RGD peptide and a peptidomimetic using both approaches and the inhibitory activity against integrin-mediated cell attachment are presented.

Introduction

The backbone modification of amide bonds 1 in bioactive peptides is one of the most promising approaches for improving the resistance toward degradation by peptidases. A number of peptide bond isosteres that reproduce the electrostatic properties and secondary structure conformations have been reported. Reduced amide bonds (-CH₂-NH-) 2 with a positively charged secondary amine provide a flexible and hydrogen bond-donating substructure (Figure 1). The success of this substructure was exemplified by several enzyme inhibitors of HIV-1 protease and neuronal nitric oxide synthase. Alkene dipeptide isosteres (-CR=CH-, R = H, F or Me) also represent steady-state peptide bond mimetics. This motif has been employed for the preparation of functional probes to identify indispensable peptide bonds. During the course of our medicinal chemistry studies using these isosteres, it was demonstrated that a heavy atom corresponding to the carbonyl oxygen in peptide bonds favorably modulates the local and global peptide conformations.

The uncharged form of amidines 4 resembles the peptide bond structures 1, in which both imino- and amino- functional groups share an sp²-carbon. Under physiological conditions, amidines are protonated and the positive charge of the conjugated acid is delocalized over two nitrogens. The characteristic substructure 4* can be viewed as a modified motif of the peptide bond and/or reduced
amide structure. However, there are few reports on amidine-type peptide bond isosteres, while acyclic amidines and cyclic amidines have been utilized as an equivalent of the basic guanidino group for several bioactive molecules.

Whereas amidines have been synthesized directly by the Pinner reaction or coupling of imidyl chlorides with amines, these reactions are not applicable to peptidyl amidine synthesis because of the harsh reaction conditions or arduous substrate preparation. We postulated that amidoximes (N-hydroxyamidines) represents an appropriate key precursor for peptidyl amidine synthesis, which is obtained by coupling of nitrile oxides with nucleophilic amines (Scheme 1). Reduction or hydrolysis under mild acidic condition of the key amidoximes would provide the target peptidyl amidines or the parent peptide bonds, respectively. It was also expected that the highly reactive nitrile oxides derived from peptide aldoximes could be exploited as active ester equivalents for fragment condensation to prepare various protected peptides and peptidomimetics. Herein we describe a novel approach to the synthesis of peptides and amidine-type peptidomimetics via peptide amidoximes.

Results and Discussion

Preparation of Amino Acid-derived Nitrile Oxide and the Application to the Synthesis of the Peptide Bond and Amidine-type Peptidomimetics

Nitrile oxides are useful reactive species, which can be formed from aldoximes by treatment with a chlorinating agent and a weak base. There have been a number of reports on 1,3-dipole cycloaddition of a nitrile oxide with olefin to produce isoxazoline derivatives, whereas examples to utilize nitrile oxides as active ester equivalents are limited. We expected that α-aminoaldoximes and peptide aldoximes serve as useful precursors of reactive nitrile oxide components for peptide and peptidomimetic synthesis.

Initially, we optimized the coupling conditions of α-aminoaldoxime and α-amino ester. This consists of a two-step process including chlorination of aldoxime and the subsequent
nucleophilic attack of an amino ester 12 onto the nitrile oxide 11, which is derived from 10 under basic conditions (Table 1). The major isomer of N-Boc-valine aldoxime 9a reacted with a NaOCl solution followed by workup and treatment with the amino ester 12 to give the desired amidoxime (N-hydroxyamidine) product 13 in 77% yield (entry 1), while the minor isomer 9b produced a complex mixture of unidentified products by the same reagent (entry 2). This was presumably due to the concomitant formation of unstable nitrile oxide 11 under basic conditions of the first chlorination step. Treatment of both aldoxime isomers 9a,b with N-chlorosuccinimide (NCS) in DMF without base provided the same product 13 in satisfying yields (entries 3 and 4). Of note, the chlorination of 9a with NCS in CHCl3 did not work, resulting in the recovery of the starting material. As such, a facile protocol to prepare amidoximes from the both isomers of amino acid-derived aldoximes was established.

Conversion processes from amidoxime 13 were next investigated. Hydrogenation of 13 with Raney Ni cleaved the N–O bond to afford the expected amidine-type isosteric unit 14 in 67% yield (Scheme 2). Alternatively, hydrolysis of 13 under mild acidic conditions containing NaNO2 gave the parent dipeptide unit 15 in 62% yield. No epimerization of the amidoximes occurred during the coupling process, which was verified by comparing 15 with two authentic diastereomers prepared from the standard protocol for peptide synthesis.

**Solid-phase Synthesis of Peptide Aldoxime and the Application of Nitrile Oxide-mediated Coupling to Cyclic Peptide Synthesis**

The nitrile oxide-mediated synthesis of peptides and peptidomimetics was applied to the solid-phase approach. We chose a cyclic RGD peptide 16, *cyclo*(-Arg-Gly-Asp-D-Phe-Val-), which is a highly potent integrin αvβ3 antagonist that includes two reactive side-chains (Arg and Asp), and the mimetic 17 *cyclo*(-Arg-Gly-ψ[C(=NH)-NH]-Asp-D-Phe-Val-) as target peptides. We planned to synthesize the RGD peptide 16 and peptidomimetic 17 by nitrile oxide-mediated cyclization followed by hydrolysis and hydrogenolysis, respectively. For application to solid-phase synthesis, the
preparations of aldoxime resin 19 were investigated. The direct attachment of Fmoc-protected aminoaldoximes such as Fmoc-NH-CH$_2$-CH=NH-OH onto the (2-Cl)trityl chloride resin failed to afford the expected resin under any conditions. In contrast, the resin 18 was prepared from the (2-Cl)trityl chloride resin and Fmoc-protected hydroxyamine (89% loading) followed by piperidine treatment. The reaction of Fmoc-protected $\alpha$-aminoaldehyde with the aminooxy (2-Cl)trityl resin 18 gave the desired aldoxime resin 19 (83% loading, Table 2, entry 1). An acidic additive improved the reactivity and the reaction proceeded smoothly at 60 °C within 2 h to give the aldoxime resin 19 in 99% yields (entry 5).\textsuperscript{16}

Peptide elongation was performed by the standard Fmoc-based solid-phase synthesis approach using N,N$'$-diisopropylcarbodiimide (DIC)/HOBt in DMF to give the peptide aldoxime resin 20 (Scheme 3). During the solid-phase process, the oxime-ether linker was inert even by treatment with 20% piperidine in DMF for Fmoc removal. For peptide cleavage from the solid-support, the standard condition [30% 1,1,1,3,3,3-hexafluoropropan-2-ol (HFIP) in CH$_2$Cl$_2$, rt, 2 h\textsuperscript{17}] was ineffective to the aldoxime resin 20, indicating that the oxime-ether linkage is less acid-labile compared with the peptide acids and peptide alcohols. Treatment of resin 20 in TFA/triisopropylsilane (TIS)/CH$_2$Cl$_2$ (0.5/0.1/99.4) provided the linear peptide aldoxime 21 in a quantitative yield.

Cyclization of acyclic peptide aldoxime 21 by treatment with NCS followed by Et$_3$N gave the amidoxime-containing peptide 22 in a moderate yield (36%, Scheme 4). The yield of aldoxime-mediated cyclization is comparable with approaches using the azide method or DPPA-mediated cyclizations (11–52% cyclization yields for the RGD peptide 16 and the derivatives).\textsuperscript{15a} Subsequently, amidoxime 22 was converted smoothly to amide 23a and amidine 23b in 46% and 95% yields by NaNO$_2$-mediated acidic hydrolysis and Raney Ni-mediated hydrogenation, respectively. The protecting groups for Arg and Asp were cleaved off using a cocktail of 1 M TMSBr–thioanisole/TFA in the presence of $m$-cresol and 1,2-ethanedithiol (EDT) in a short time, providing the desired parent RGD peptide 16 and the peptidomimetic 17 in 87% and 73% yields, respectively. It is of note that no hydrolyzed product 16 was observed during the deprotection
treatment of 23b and subsequent HPLC purification process.

**Biological Activity of the Cyclic RGD Peptide with an Amidine-type Isosteric Unit for the Gly-Asp Dipeptide.**

The resulting cyclic RGD peptidomimetics 17 was evaluated for its inhibitory effect of integrin-mediated cell attachment (Figure 2). Peptide 17 with an amidine moiety showed moderate inhibitory activity (IC$_{50}$ = 4.77 μM) compared with the original peptide 16 (peptide 16, IC$_{50}$ = 0.157 μM). The X-ray crystal structure of the α$_v$β$_3$ integrin-cyclic RGD peptide complex indicated that the uncharged amide NH of Gly-Asp is located proximal to the integrin residue Arg216, which is likely to be involved in the interactions.$^{18}$ These results suggest that substitution of the Gly-Asp peptide bond with the positively charged amidine unit partially eliminated the highly potent binding affinity towards the α$_v$β$_3$ integrin.

**Conclusion**

In conclusion, we have established a novel approach to synthesize acyclic amidine and amide units via a key amidoxime (N-hydroxyamidine) precursor, which was prepared from nitrile oxide component as an active ester equivalent. This method was used for Fmoc-based solid-phase synthesis of peptides and peptidomimetics containing an amidine-type isostere. The peptide aldoxime represented a functional precursor for a protected cyclic peptide and peptidomimetic, suggesting that the nitrile oxide-mediated coupling reaction should serve as an alternative method for peptide macrocyclizations. Further studies on the scope and limitations of this approach as well as applications for structure-activity relationship studies of bioactive peptides are currently in progress.

**Experimental Section**

**Synthesis**

*tert*-Butyl [(S)-1-(hydroxyiminomethyl)-2-methylpropyl]carbamate (9). To a solution of
Boc-Val-NMe(OMe) (5.00 g, 19.2 mmol) in Et₂O (60 cm³) was added dropwise a solution of LiAlH₄ (1.02 g, 27.0 mmol) in Et₂O (20 cm³) at –40 °C and the mixture was stirred for 40 min. The reaction was quenched at –40 °C by addition of Na₂SO₄ solution. The reaction mixture was washed with saturated aqueous NaHCO₃ and brine, and dried over Na₂SO₄. Concentration under reduced pressure gave Boc-valinal. To a solution of NH₂OH·HCl (1.66 g, 23.9 mmol) and AcONa (1.96 g, 23.9 mmol) in EtOH (50 cm³) was added the solution of the aldehyde in EtOH (15 cm³). The reaction mixture was stirred at 80 °C for 15 min. The mixture was concentrated under reduced pressure. The residue was extracted with CH₂Cl₂, and the extract was washed with H₂O and dried over Na₂SO₄. Concentration under reduced pressure followed by flash chromatography over silica gel with n-hexane/EtOAc (3/1) gave the title compounds 9a and 9b (3.44 g, 82% yield, 9a/9b = 58/42) both as a white solid. Compound 9a: mp 35.5–36.5 °C; [α]²⁶D +11.0 (c 0.58, CHCl₃); δ₁H (500 MHz, DMSO, Me₄Si) 0.82 (6H, dd, J 13.7 and 6.9), 1.37 (9H, s), 1.75 (1H, td, J 13.7 and 6.9), 3.72–3.78 (1H, m), 6.96 (1H, d, J 8.8), 7.14 (1H, d, J 7.3) and 10.63 (1H, s); δ₁C (125 MHz, DMSO-d₆, Me₄Si) 18.6, 18.8, 28.2 (3C), 30.9, 55.4, 77.7, 149.0 and 155.1. Anal. Calcd for C₁₀H₂₀N₂O₃: C, 55.53; H, 9.32; N, 12.95. Found: C, 55.29; H, 9.17; N, 12.81. Compound 9b: mp 114.0–115.0 °C; [α]²⁶D +50.0 (c 0.18, CHCl₃); δ₁H (500 MHz, DMSO-d₆, Me₄Si) 0.81 (6H, t, J 7.2), 1.37 (9H, s), 1.76–1.85 (1H, m), 4.54 (1H, dd, J 15.7 and 7.1), 6.51 (1H, d, J 7.1), 6.95 (1H, d, J 8.9) and 10.86 (1H, s); δ₁C (125 MHz, DMSO, Me₄Si) 18.3, 18.7, 28.2 (3C), 30.7, 50.2, 77.7, 149.9 and 155.2. Anal. Calcd for C₁₀H₂₀N₂O₃: C, 55.53; H, 9.32; N, 12.95. Found: C, 55.25; H, 9.32; N, 12.71.

**tert-Butyl**

(S)-2-{{[(S)-2-tert-butoxycarbonylamino-N-hydroxy-3-methylbutanimidoyl]amino}-3-phenylpropionate (13). To a solution of aldoxime 9b (30.0 mg, 0.140 mmol) in DMF (0.6 cm³) was added N-chlorosuccinimide (20.0 mg, 0.150 mmol) and the mixture was stirred at room temperature for 4 h. The reaction mixture was extracted with EtOAc and the extract was washed with a solution of H₂O/brine (1/1), and dried over Na₂SO₄. After concentration under reduced pressure, the residue was
dissolved in Et₂O (5 cm³). To the solution were added Et₃N (210 mm³, 0.150 mmol) and H-Phe-O'Bu 12 (30.0 mg, 0.140 mmol) and the mixture was stirred at room temperature overnight. The reaction mixture was washed with brine and dried over Na₂SO₄. Concentration under reduced pressure followed by flash chromatography over silica gel with n-hexane/EtOAc (3/1) gave the title compound 13 (50.0 mg, 81% yield, inseparable mixture of major/minor = 97/3) as a colorless oil: [α]²⁶ D −19.2 (c 0.73, CHCl₃); δ_H (500 MHz, DMSO-d₆, Me₄Si) 0.63 (3H, d, J 6.6), 0.73 (3H, d, J 6.6), 1.32 (9H, s), 1.37 (9H, s), 1.79 (1H, dt, J 21.7 and 6.6), 2.84–2.94 (2H, m), 3.70 (1H, t, J 9.0), 4.44–4.52 (1H, m), 5.38 (1H, d, J 10.5), 6.66 (1H, d, J 9.5), 7.19–7.29 (5H, m) and 10.86 (1H, s); δ_C (125 MHz, DMSO-d₆, Me₄Si) 18.3, 19.8, 27.5 (3C), 28.2 (3C), 29.8, 55.0, 56.1, 77.8, 80.6, 126.5, 128.0 (3C), 129.5 (2C), 137.0, 150.5, 155.3 and 171.4; HRMS (FAB) m/z calcd for C₂₃H₃₈N₃O₅ ([M+H]+) 436.2811, found 436.2808.

**tert-Butyl**

((S)-2-{{((S)-2-tert-butoxycarbonylamino-3-methylbutanimidoyl}amino}-3-phenylpropionate (14). To a solution of amidoxime 13 (29.1 mg, 0.0670 mmol) in MeOH (1 cm³) and AcOH (0.011 cm³) was added Raney Ni (0.85 cm³, slurry in H₂O) and the mixture was stirred under atmospheres of hydrogen at room temperature for 1 h. The mixture was filtered through celite. Concentration under reduced pressure followed by flash chromatography over silica gel with n-hexane/EtOAc (3/1) gave the title compound 14 (18.9 mg, 67% yield) as a yellow oil: [α]²⁶ D +7.53 (c 0.46, CHCl₃); δ_H (500 MHz, DMSO-d₆, Me₄Si) 0.76 (6H, dd, J 13.5 and 6.7), 1.28 (9H, s), 1.38 (9H, s), 1.80–1.88 (1H, m), 2.86 (1H, br s), 2.92 (1H, dd, J 13.5 and 6.9), 3.77 (1H, br s), 4.26 (1H, br s), 4.99 (1H, d, J 9.5), 6.16 (1H, br s), 6.96 (1H, d, J 9.5) and 7.14–7.26 (5H, m); δ_C (125 MHz, DMSO-d₆, Me₄Si) 18.0 (2C), 19.3, 27.5 (3C), 28.2 (3C), 31.0, 37.8, 59.5, 77.8, 78.9, 126.1, 127.9 (2C), 127.9, 129.2 (2C), 138.1, 155.2 and 171.2; HRMS (FAB) m/z calcd for C₂₃H₃₈N₃O₄ ([M+H]+) 420.2862, found 420.2864.
**tert-Butyl (S)-2-[(S)-2-tert-butoxycarbonylamino-3-methylbutrylamino]-3-phenylpropionate (15).** To a solution of amidoxime 13 (35.3 mg, 0.0810 mmol) in MeOH (0.8 cm³) and H₂O (0.8 cm³) were added AcOH (0.00800 cm³, 0.120 mmol) and NaNO₂ (8.30 mg, 0.120 mmol). The mixture was stirred at room temperature overnight. The mixture was concentrated under reduced pressure. The residue was extracted with CH₂Cl₂, and the extract was washed with H₂O and dried over Na₂SO₄. Concentration under reduced pressure followed by flash chromatography over silica gel with n-hexane/AcOEt (3/1) gave the title compound 15 (21.0 mg, 62% yield) as a white solid: mp 115.0–116.0 °C; [α]$_{24}^D$ +60.0 (c 0.87, CHCl₃); δ$_{H}$ (500 MHz, DMSO-$_d$₆, Me$_4$Si) 0.87 (3H, d, J 5.6), 0.93 (3H, d, J 6.8), 1.38 (9H, s), 1.45 (9H, s), 2.04–2.14 (1H, m), 3.04–3.11 (2H, m), 3.91 (1H, t, J 5.6), 4.74 (1H, dd, J 13.8 and 6.2), 5.16 (1H, d, J 6.6), 6.30 (1H, d, J 6.2) and 7.14–7.31 (5H, m); δ$_{C}$ (125 MHz, DMSO-$_d$₆, Me$_4$Si) 17.7, 19.2, 27.9 (3C), 28.3 (3C), 38.2, 52.2, 53.6, 55.1, 82.2, 82.3, 126.9, 127.0 (2C), 128.4 (2C), 129.5, 136.0, 170.3 and 171.0; HRMS (FAB) m/z calcd for C$_{23}$H$_{37}$N$_2$O$_5$ ([M+H]$^+$) 421.2702, found 421.2702.

**H$_2$N-O-(2-Cl)Trt resin (18).** 2-Chlorotrityl resin chloride (loading: 1.31 mmol g$^{-1}$, 76.3 mg) was reacted with Fmoc-NHOH (128 mg, 0.500 mmol) and pyridine (0.0810 cm³, 1.00 mmol) in THF (0.8 cm³) at 60 °C for 6 h. The solution was removed by decantation and the resulting resin was washed with the solution of DMF/(iPr)$_2$NEt/MeOH (17/2/1). The Fmoc-protecting group was removed by treating the resin with a DMF/piperidine solution (80/20, v/v). The loading was determined by measuring at 290 nm UV absorption of the piperidine-treated sample: 0.900 mmol g$^{-1}$, 89%.

**H-Asp(O'Bu)-d-Phe-Val-Arg(Pbf)-Gly-aldoxime-(2-Cl)Trt resin (20).** The solid supported hydroxyamine 18 (loading: 0.900 mmol g$^{-1}$, 91.6 mg, 0.0820 mmol) was reacted with Fmoc-glycinal (0.500 mmol) in dichloroethane (0.7 cm³), HC(OMe)$_3$ (0.5 cm³) and AcOH (0.001 cm³) at 60 °C for 2 h. The solution was removed by decantation and the resulting resin was washed with DMF to afford resin 19. The peptide-resin 20 was manually constructed using Fmoc-based solid-phase
synthesis on resin 19. The Fmoc-protecting group was removed by treating the resin with a DMF/piperidine solution (80/20, v/v). Fmoc-protected amino acid (0.500 mmol, 6.1 equiv) was successively condensed using 1,3-diisopropylcarbodiimide (0.0770 cm$^3$, 0.500 mmol, 6.1 equiv) in the presence of $N$-hydroxybenzotriazole (77 mg, 0.500 mmol, 6.1 equiv) to give resin 20. $t$Bu ester for Asp and 2,2,4,6,7-pentamethyldihydrobenzofuran-5-sulfonyl (Pbf) for Arg were employed for side-chain protection.

**H-Asp(O$t$Bu)-D-Phe-Val-Arg(Pbf)-Gly-aldoxime (21).** Resin 20 was treated with TFA/TIS/CH$_2$Cl$_2$ (20 cm$^3$, 0.5/0.1/99.4) at room temperature for 1.5 h. After removal of the resin by filtration, the filtrate was concentrated under reduced pressure to give off a crude peptide aldoxime 21 as a yellow oil (74.0 mg, quant. from resin 18). The crude product was used without further purification.

**Cyclo[-Arg(Pbf)-Gly-ψ[C(=NOH)NH]-Asp(O$t$Bu)-D-Phe-Val-] (22).** To a solution of peptide aldoxime 21 (74.0 mg) in DMF (1 cm$^3$) was added $N$-chlorosuccinimide (14.7 mg, 0.100 mmol). The solution was stirred at room temperature overnight, and then DMF (40 cm$^3$) and Et$_3$N (0.4 cm$^3$) were added. The mixture was stirred at room temperature overnight, and was then concentrated under reduced pressure. The residue was extracted with EtOAc and the extract was washed with brine. The organic layer was dried over Na$_2$SO$_4$, and concentrated under reduced pressure to give a yellow oil, which was purified by column chromatography over silica gel with CH$_2$Cl$_2$/MeOH (95/5) to give 22 (26.9 mg, 36% yield, major/minor = 79/21) as a yellow solid: mp 168.0–169.0 °C; $[\alpha]_{25}^{25}$D $-$52.7 (c 0.28, CHCl$_3$); $\delta$ (500 MHz, DMSO-$d_6$, Me$_4$Si) 0.85 (major, 3H, d, J 6.9), 0.68 (minor, 3H, t, J 6.4), 0.73 (major, 3H, d, J 6.7), 0.72–0.76 (minor, 3H, m), 1.25–1.50 (2H, m), 1.34 (minor, 9H, s), 1.37 (major, 9H, s), 1.36 (minor, 6H, s), 1.41 (major, 6H, s), 1.74–1.76 (2H, m), 2.00 (3H, s), 2.41 (3H, s), 2.47 (3H, s), 2.30–2.50 (2H, m), 2.59 (1H, dd, J 15.9 and 5.9), 2.82–2.91 (2H, m), 2.96 (2H, s), 3.79 (major, 1H, t, J 7.0), 3.82–3.88 (minor, 1H, m), 3.98 (major, 1H, dd, J 14.7 and 7.7), 4.02–4.08
(minor, 1H, m), 4.10–4.14 (minor, 1H, m), 4.16–4.25 (major, 1H, m), 4.50 (major, 1H, dd, J 14.6 and 8.4), 4.35–4.45 (minor, 1H, m), 4.54–4.65 (major, 1H, m), 4.60–4.75 (minor, 1H, m), 5.17 (minor, 1H, d, J 9.6), 5.27 (major, 1H, d, J 10.6), 6.37 (major, 1H, br s), 6.70 (minor, 1H, br s), 7.12–7.33 (5H, m), 7.42–7.53 (1H, m), 8.05–8.14 (2H, m), 8.32 (minor, 1H, d, J 7.3), 8.45 (major, 1H, d, J 5.9), 9.17 (minor, 1H, s) and 9.63 (major, 1H, s); δC (125 MHz, DMSO-d$_6$, Me$_4$Si) 12.1, 12.3, 17.3 (minor), 17.6 (major), 17.8 (major, 2C), 17.9 (minor, 2C), 18.9, 19.0 (major), 19.1 (minor), 21.1, 27.7 (3C), 27.7, 28.3 (major, 2C), 28.8 (minor, 2C), 36.3, 42.5 (2C), 52.0, 52.6, 55.0, 59.8 (minor), 60.3 (major), 62.8, 79.7 (minor), 80.3 (major), 86.3, 116.3, 124.3, 126.5, 128.1 (minor, 2C), 128.2 (major, 2C), 129.1 (major, 2C), 129.3 (minor, 2C), 131.4, 134.2, 137.0, 137.3, 148.9, 156.0, 157.5, 169.3 (major), 169.5 (minor), 170.7, 171.2, 172.2 and 172.4; HRMS (FAB) m/z calcd for C$_{43}$H$_{64}$N$_9$O$_{10}$S ([M+H]$^+$) 898.4497, found 898.4502.

_Cyclo[-Arg(Pbf)-Gly-Asp(O'Bu)-d-Phe-Val-] (23a)._ To a solution of amidoxime 22 (20.0 mg, 0.0220 mmol) in MeOH (0.5 cm$^3$) and H$_2$O (0.2 cm$^3$) were added AcOH (0.00500 cm$^3$) and NaNO$_2$ (4.60 mg, 0.0660 mmol). The mixture was stirred at room temperature overnight. The mixture was concentrated under reduced pressure. The residue was extracted with EtOAc, and the extract was washed with H$_2$O and dried over MgSO$_4$. Concentration under reduced pressure followed by PTLC purification with CH$_2$Cl$_2$/MeOH (95/5) gave the title compound 23a (8.90 mg, 46% yield) as a white solid: mp 247.0–248.0 °C; [α]$^2$$_D$ –32.3 (c 0.27, MeOH); δ$_\text{H}$ (500 MHz, DMSO-d$_6$, Me$_4$Si) 0.70 (6H, dd, J 20.7 and 6.7), 1.18–1.50 (2H, m), 1.34 (9H, s), 1.41 (6H, s), 1.65–1.72 (1H, m), 1.80–1.88 (1H, m), 2.01 (3H, s), 2.36 (1H, dd, J 15.7 and 8.9), 2.41 (3H, s), 2.46 (3H, s), 2.80 (1H, dd, J 13.7 and 6.6), 2.91–3.06 (2H, m), 2.96 (2H, s), 3.28 (2H, s), 3.82 (1H, t, J 7.6), 4.00–4.10 (2H, m), 4.54–4.62 (2H, m), 6.35 (1H, br s), 6.70–6.80 (1H, m), 7.13–7.28 (5H, m), 7.42–7.50 (5H, m), 7.74 (2H, dd, J 11.5 and 8.3), 7.95 (1H, d, J 8.3), 8.06 (1H, d, J 7.6) and 8.36 (1H, dd, J 7.3 and 4.4); δ$_\text{C}$ (125 MHz, DMSO-d$_6$, Me$_4$Si) 12.3, 17.6, 18.2, 18.9, 19.2, 25.8, 27.6 (3C), 28.3 (2C), 28.4, 29.7, 36.4, 37.1, 39.8, 42.5, 43.1, 48.9, 52.2, 53.9, 60.1, 80.0, 86.3, 116.3, 119.7, 124.3, 126.2, 128.1 (2C), 129.0 (2C),
130.3, 131.4, 137.3, 156.0, 157.4, 169.1, 169.4, 169.9, 170.8, 171.0 and 171.1; HRMS (FAB) m/z calcd for C_{43}H_{63}N_{8}O_{10}S ([M+H]^+) 883.4388, found 883.4397.

_Cyclo[-Arg(Pbf)-Gly-ψ(C(=NH)NH)-Asp(O^tBu)-D-Phe-Val-] (23b)._ To a solution of amidoxime 22 (30.0 mg, 0.0330 mmol) in MeOH (0.6 cm^3) and AcOH (0.006 cm^3) was added Raney Ni (0.440 cm^3, slurry in H_{2}O) and the mixture was stirred under H_{2} atmospheres at room temperature for 2 h. The mixture was filtered through celite. Concentration under reduced pressure followed by flash chromatography over silica gel with CH_{2}Cl_{2}/MeOH (95/5) gave the title compound 23b (27.4 mg, 95% yield) as a colorless oil: [α]^{26}_{D} –53.3 (c 0.14, CHCl_{3}); δ_{H} (500 MHz, CD_{3}OD, Me_{4}Si) 0.74 (6H, dd, J 14.7 and 6.9), 1.43 (9H, s), 1.45 (6H, s), 1.45–1.52 (1H, m), 1.55–1.60 (1H, m), 1.82–1.89 (1H, m), 1.95–2.00 (1H, m), 2.07 (3H, s), 2.56 (3H, s), 2.59 (1H, d, J 6.6), 2.77 (1H, dd, J 16.5 and 6.9), 2.94 (1H, dd, J 13.3 and 6.7), 2.99 (2H, s), 3.05 (1H, dd, J 13.2 and 9.0), 3.11–3.18 (1H, m), 3.53 (1H, d, J 15.2), 3.87 (1H, d, J 6.9), 4.28 (1H, d, J 15.2), 4.34–4.37 (1H, m), 4.39–4.45 (1H, m), 4.68 (1H, dd, J 9.0 and 6.9) and 7.15–7.29 (5H, m); δ_{C} (125 MHz, CD_{3}OD, Me_{4}Si) 12.5, 18.4, 18.7, 19.6, 19.7, 28.4, 28.4, 28.4 (3C), 29.6 (2C), 30.9, 37.9, 38.4, 44.0, 49.5, 49.7, 54.0, 56.4, 62.5, 82.6, 87.7, 118.5, 126.0, 127.9, 129.6 (2C), 130.4 (2C), 132.4, 133.5, 134.4, 138.0, 139.4, 158.1, 160.0, 172.0, 173.3, 173.6, 173.9, 174.2 and 174.3; HRMS (FAB) m/z calcd for C_{43}H_{62}N_{8}O_{9}S ([M–H]^-) 880.4397, found 880.4395.

_Cyclo[-Arg-Gly-ψ(C(=NH)NH)-Asp-D-Phe-Val-] (17)._ The protected amidine 23b (7.90 mg, 0.00900 mmol) was treated with 1M TMSBr-thioanisole in TFA (10 cm^3) in the presence of m-cresol (0.1 cm^3) and 1,2-ethanedithiol (0.5 cm^3) at 4 °C for 15 min. The mixture was poured into ice-cold dry Et_{2}O (50 cm^3). The resulting powder was collected by centrifugation and the washed three times with ice-cold dry Et_{2}O. The crude product was purified by preparative HPLC to afford the expected peptide 17 as a white powder (5.30 mg, 0.00660 mmol, 73% yield): [α]^{25}_{D} –129.2 (c 0.17, MeOH); δ_{H} (500 MHz, DMSO-d_{6}, Me_{4}Si) 0.70 (3H, d, J 6.6), 0.74 (3H, d, J 6.6), 1.32–1.60 (3H, m),
1.73–1.84 (1H, m), 1.88–1.98 (1H, m), 2.59 (1H, dd, J 17.0 and 5.7), 2.78 (1H, dd, J 13.5 and 6.5),
2.84 (1H, dd, J 17.2 and 8.2), 3.00 (1H, dd, J 13.0 and 8.4), 3.04–3.13 (2H, m), 3.72–3.78 (2H, m),
3.90–3.98 (1H, m), 4.23 (1H, dd, J 13.5 and 8.2), 4.43 (1H, t, J 16.2 and 7.0), 4.53–4.60 (1H, m),
4.62–4.68 (1H, m), 6.80–7.40 (2H, br s), 7.16–7.28 (5H, m), 7.72 (1H, t, J 5.7), 7.93 (1H, dd, J 11.3
and 8.4), 8.12 (1H, d, J 7.7), 8.28–8.32 (1H, m), 8.53 (1H, d, J 7.7), 8.92–8.98 (1H, m), 9.10–9.20
(1H, m) and 9.64 (1H, s); δC (125 MHz, DMSO-d6, Me4Si) 17.9, 25.3, 28.2, 29.6, 34.2, 37.0, 37.1,
40.2, 51.7, 51.9, 54.2, 59.9, 126.4, 128.2 (2C), 129.1 (2C), 137.2, 156.8, 158.4, 164.8, 166.8, 170.7,
171.2, 171.3 and 171.7; HRMS (FAB) m/z calcd for C26H40N9O6 ([M+H]+) 574.3102, found 574.3101.

Cyclo(-Arg-Gly-Asp-D-Phe-Val-) (16). By the identical procedure as described for the
preparation of 17, 23a (8.00 mg, 0.00900 mmol) was converted into the cyclic RGD peptide 16
(0.00790 mmol, 87% yield). All characterization data were in agreement with the date of control
peptide which was synthesized using Fmoc-based solid-phase synthesis. [α]25D –21.6 (c 0.27,
MeOH); δH (500 MHz, DMSO-d6, Me4Si) 0.68 (3H, d, J 6.7), 0.75 (3H, d, J 6.7), 1.32–1.45 (2H, m),
1.45–1.55 (1H, m), 1.69–1.80 (1H, m), 1.80–1.90 (1H, m), 2.38 (1H, dd, J 16.4 and 5.5), 2.72 (1H,
dd, J 16.4 and 8.9), 2.81 (1H, dd, J 13.5 and 6.1), 2.94 (1H, dd, J 13.5 and 8.0), 3.05–3.14 (2H, m),
3.26 (1H, dd, J 15.2 and 4.2), 3.82 (1H, t, J 7.4), 4.04 (1H, dd, J 15.2 and 7.7), 4.08–4.16 (1H, m),
4.55 (1H, dd, J 14.2 and 7.2), 4.60–4.68 (1H, m), 6.58–7.11 (1H, br s), 7.15–7.25 (5H, m), 7.58 (1H,
t, J 5.7), 7.78 (1H, d, J 7.4), 7.87 (1H, d, J 8.0), 8.00 (1H, d, J 7.4), 8.08 (1H, d, J 8.6), 8.36 (1H, dd,
J 7.4 and 4.2) and 12.3 (1H, s); δC (125 MHz, DMSO-d6, Me4Si) 18.1, 19.1, 25.3, 28.2, 29.5, 34.8,
37.1, 40.2, 43.0, 48.8, 52.0, 53.9, 60.1, 126.1, 128.0 (2C), 129.0 (2C), 137.3, 156.6, 158.3, 169.4,
169.8, 170.6, 171.1 and 171.6; HRMS (FAB) m/z calcd for C26H36N8O7 ([M+H]+) 575.2942, found 575.2952.

Evaluation of Inhibitory Activity against Integrin-mediated Cell Attachment. Human dermal
fibroblasts (HDFs; AGC Techno Glass, Chiba, Japan) were maintained in DMEM containing 10% FBS, 100 U cm$^{-3}$ penicillin, and 100 μg cm$^{-3}$ streptomycin (Invitrogen, Carlsbad, CA). Human plasma vitronectin (0.1 μg in 0.050 cm$^3$ well$^{-1}$; EMD Chemicals Inc., Gibbstown, NJ) were added to 96-well plates (Nalge Nunc, Rochester, NY) and incubated for 1 h at 37 °C. The plates were washed and blocked with 1% bovine serum albumin (BSA; Sigma-Aldrich, St. Louis, MO) in DMEM. HDFs were incubated at room temperature for 15 min in the various concentrations of peptides (0.001–200 μM in 1% DMSO). Then 0.100 cm$^3$ HDFs (2 x 10$^4$ cells) in DMEM containing 0.1% BSA were added to each well and incubated at 37 °C for 30 min in 5% CO$_2$. The attached cells were stained with 0.2% crystal violet aqueous solution in 20% MeOH (0.150 cm$^3$) for 15 min. After washing with Milli-Q water, the plates were dried overnight at room temperature and dissolved by 0.150 cm$^3$ of 1% SDS solution. The absorbance at 570 nm was measured. Each sample was assayed in triplicate, and cells attached to the BSA were subtracted from all measurements. 1% DMSO did not have any effect on HDF attachment to vitronectin.

**Acknowledgement**

This work was supported by Grants-in-Aid for Scientific Research and Targeted Protein Research Program from the Ministry of Education, Culture, Sports, Science, and Technology of Japan. E.I. and K.T. are grateful for Research Fellowships from the JSPS for Young Scientists.

**Supplementary data**

Supplementary data associated with this article can be found, in the online version, at doi:XXXXX/XXXXX.
References and notes


7 For acyclic amidine-type peptide isostere, see: H. Moser, A. Fliri, A. Steiger, G. Costello, J.


Racemization (20%) was observed when Fmoc-d-Phe-H was utilized for on-resin aldoxime formation under the conditions of entry 5.

Table 1. Optimization of the Aldoxime–Amino Acid Coupling Conditions.

<table>
<thead>
<tr>
<th>entry</th>
<th>substrate</th>
<th>step a</th>
<th>step b</th>
<th>yield (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>9a</td>
<td>NaOCl (3.0 equiv), Et$_3$N (3.0 equiv)$^a$</td>
<td>Et$_3$N (6.0 equiv)/ CH$_2$Cl$_2$</td>
<td>77</td>
</tr>
<tr>
<td>2</td>
<td>9b</td>
<td>NaOCl (3.0 equiv), Et$_3$N (3.0 equiv)</td>
<td>Et$_3$N (6.0 equiv)/ decomp</td>
<td>71$^c$</td>
</tr>
<tr>
<td>3</td>
<td>9a</td>
<td>NCS (1.4 equiv)</td>
<td>Et$_3$N (4.0 equiv)/ Et$_2$O</td>
<td>71$^c$</td>
</tr>
<tr>
<td>4</td>
<td>9b</td>
<td>NCS (1.4 equiv)</td>
<td>Et$_3$N (4.0 equiv)/ Et$_2$O</td>
<td>81</td>
</tr>
</tbody>
</table>

$^a$ Substrates 9 were prepared from Boc-valinal according to the literature procedure.$^{14}$ $^b$ 30% aqueous solution. $^c$ When CHCl$_3$ was used as the reaction solvent in step a, the starting material 9a was recovered.

Table 2. Preparation of the Aldoxime Resin 19.

<table>
<thead>
<tr>
<th>entry</th>
<th>additive (1.2 equiv.)</th>
<th>conditions$^a$</th>
<th>loading (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>-</td>
<td>rt, o/n</td>
<td>83</td>
</tr>
<tr>
<td>2</td>
<td>Et$_3$N</td>
<td>rt, o/n</td>
<td>56</td>
</tr>
<tr>
<td>3</td>
<td>AcOH</td>
<td>rt, o/n</td>
<td>90</td>
</tr>
<tr>
<td>4</td>
<td>AcOH</td>
<td>60 °C, o/n</td>
<td>99</td>
</tr>
<tr>
<td>5</td>
<td>AcOH</td>
<td>60 °C, 2 h</td>
<td>99</td>
</tr>
</tbody>
</table>

$^a$ Dichloroethane (0.15M), HC(OMe)$_3$ (0.2M).
**Scheme 1.** Synthetic Scheme for Amidine-type Peptide Bond Isosteres 4 and Native Peptide Bonds 1 Using Nitrile Oxides 6 as the Reactive Acyl Equivalents.

**Scheme 2.** Conversion of N-Hydroxyamidine 13 to Amidine 14 and Peptide Bond 15.

**Scheme 3.** Preparation of the Peptide Aldoxime 21.
Scheme 4. Synthesis of the Cyclic RGD Peptide 16 and the Amidine-type Isosteric Congener 17.
**Figure 1.** Structures of the Peptide Bond and the Mimetics.

native peptide bonds 1

alkene isosteres 3
(R = H, F or Me)

reduced amide bonds 2

amidine-type isosteres 4

---

**Figure 2.** Inhibitory effect of cyclic RGD peptides on HDF attachment to vitronectin. HDFs were allowed to attach to human vitronectin in the presence of various concentrations of cyclic RGD peptides. Peptides were added to the cell suspension and the cells were plated. After a 30 min-incubation period, the attached cells were stained with crystal violet and dissolved in a 1% SDS solution. The absorbance at 570 nm was measured. Triplicate experiments gave similar results.