

Global meta-analysis for controlling factors on carbon stable isotope ratios of lotic periphyton

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Abstract

Carbon stable isotope ratios ($\delta^{13}\text{C}$) are widely used to trace resource transfer pathway, yet $\delta^{13}\text{C}$ variation of freshwater autotrophs has not been fully understood. We analyzed data from 42 published studies supplemented with some unpublished data to show the determinants of lotic periphyton $\delta^{13}\text{C}$. At large spatial scales, we observed broad differences in periphyton $\delta^{13}\text{C}$ among biomes and consistent longitudinal variation related to watershed area.

Longitudinal increases in $\delta^{13}\text{C}$ indicated the importance of in-stream processes on lotic carbon cycles and autotroph $\delta^{13}\text{C}$ variation. At local spatial scales, periphyton $\delta^{13}\text{C}$ was related negatively to canopy cover and water current velocity and positively to chlorophyll *a* density. Autotroph $\delta^{13}\text{C}$ varied among taxonomic groups. Cyanobacteria and red algae had significantly higher and lower $\delta^{13}\text{C}$ than other taxa, respectively. A hierarchical model across spatial scales showed that local controls for periphyton $\delta^{13}\text{C}$ were nested by regional controls, which suggested that productivity and CO_2 availability determine $\delta^{13}\text{C}$. Overall our results revealed general patterns of periphyton $\delta^{13}\text{C}$ and provided improved information for study design and use of $\delta^{13}\text{C}$ in isotopic mixing models in lotic food web studies.

Key words: $\delta^{13}\text{C}$ variation; regional control; local control; structural equation modeling; lotic ecosystem

Introduction

Most ecosystems receive resource subsidization from other systems. Estimating contributions of spatial subsidies to organisms provides information of food web production and trophic structure via consideration of resource movement across ecosystem boundaries (Polis et al. 1997). Lotic food webs are a good example of subsidized systems, supported by two carbon sources: autochthonous (i.e., periphytic algae, mosses and other aquatic plants attaching to a substrate) and allochthonous (i.e., terrestrial litter and invertebrates, Nakano and Murakami 2001) resources. Distinguishing the contributions of these two sources to lotic food webs is challenging because direct observations of food sources are difficult. Carbon stable isotope ratios ($\delta^{13}\text{C}$) are widely used to discriminate between periphytic and terrestrial production in lotic ecosystems (e.g., Fry 1991; France 1995; Finlay 2001).

While the use of $\delta^{13}\text{C}$ has many advantages as a natural tracer, growing awareness of the large spatial and temporal variability in $\delta^{13}\text{C}$ values of lotic periphyton suggest limitation of $\delta^{13}\text{C}$ applications under some circumstances (e.g., Finlay et al. 1999; Zah et al. 2001). Large variability in $\delta^{13}\text{C}$ values of periphyton can make the isotopic baseline of food webs uncertain and consequently influence the accuracy of food web analysis (McCutchan and Lewis 2001; Zah et al. 2001). Since two-source mixing models require isotopic separation between potential sources, the variability in $\delta^{13}\text{C}$ values of periphyton can compromise the precise estimation of the relative contribution of carbon sources to food webs. For example, Finlay et al. (1999) show large variations in $\delta^{13}\text{C}$ values of periphyton and the herbivores over small spatial scales (1-10m) in a stream related to *in-situ* water current velocity. Such effects greatly complicate mixing model applications.

Many previous studies have pointed out the importance of understanding the pattern of variability in $\delta^{13}\text{C}$ values of lotic periphyton for the purpose of precise food web analysis

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(MacLeod and Barton 1998; Finlay et al. 1999; Trudeau and Rasmussen 2003; Singer et al. 2005; Doi et al. 2007; Hill et al. 2008). Although the importance of understanding the variability in periphyton $\delta^{13}\text{C}$ is well recognized, the factors that determine $\delta^{13}\text{C}$ values of periphyton are complex due to influences of spatial heterogeneity at multiple spatial scales such as local habitat, reach, and watershed (Doi et al. 2007). To understand the factors controlling periphyton $\delta^{13}\text{C}$ values, both regional controls (e.g., biome or river size) and local controls (e.g., light/flow conditions, taxonomic composition) must be considered. These diverse controls over periphyton $\delta^{13}\text{C}$ have not yet been adequately classified and brought into a hierarchically structured and causally plausible framework.

In this study, we conduct a global meta-analysis to identify the factors that determine the $\delta^{13}\text{C}$ value of lotic periphyton. For this meta-analysis, we use a dataset including a total of 765 records from 42 references and some unpublished data. From the dataset, regional and local drivers hypothesized to affect periphyton $\delta^{13}\text{C}$ are analyzed. Particularly we focus on biome, season, and watershed area as regional controls because they strongly affect periphytic production and dissolved inorganic carbon (DIC) availability in lotic ecosystems. Canopy cover, water current velocity, chlorophyll *a* density, and dominant taxa in periphyton are used as local controls. Canopy cover regulates light intensity, chlorophyll *a*, and dominant taxa in periphyton, which controls DIC demand. Water current velocity regulates DIC availability for periphyton. Finally we build a hierarchical model and synthesize controlling pathways on periphyton $\delta^{13}\text{C}$.

Materials and methods

Data sources

We searched for data for carbon stable isotope values of lotic periphyton from published

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sources using ISI Web of Science (<http://www.isiknowledge.com>). Search terms included ‘periphyt*’, ‘algae’, ‘isotope’, and ‘stream’. The search was conducted on 24-Feb-2009 and returned 192 studies. We also added studies found in the journals Canadian Journal of Aquatic and Fisheries Science, Ecology, Freshwater Biology, Hydrobiologia, Journal of the North American Benthological Society, Limnology and Oceanography, and Marine and Freshwater Research that were not detected in our Web of Science search. Also, we included unpublished data from our own studies. For this analysis, we defined periphyton as predominantly algal material attached to solid surfaces. Attached autotrophic algae described by authors as benthic algae, epilithic algae, algal-dominated epilithon, filamentous algae, littoral algae, micro algae, epiphyton, periphytic biofilms, phototrophic biofilms, and phytomicrobenthos were regarded as periphyton in this meta-analysis. Data were used for epilithon only if authors specified the dominance of epilithic algae in the epilithic material. Neither periphyton nor epilithon are pure algae, and the terminology is not necessarily consistent among researchers. Some studies have defined epilithon as a mixed assemblage of algae, fungi, and bacteria (e.g., England and Rosemond 2004), and others regard epilithon as a surrogate of autochthonous materials (e.g., Watanabe et al. 2008) because they are highly influenced by polysaccharides derived from microalgal production (Lock 1993). Data for periphyton was often a mix of microalgae but information for specific taxonomic groups was extracted where possible. Data for macrophytes and aquatic bryophytes were also gathered.

We adopted the data that met following conditions:

- i) The study was conducted in a field setting
- ii) The study provided periphyton $\delta^{13}\text{C}$ and environmental drivers
- iii) The study did *not* use ^{13}C tracer additions

When the data were presented in figures, we extracted data using graph digitizing software

PlotDigitizer X ver. 2.0.1

(<http://www.surf.nuqe.nagoya-u.ac.jp/~nakahara/Software/PlotDigitizerX/index-e.html>).

Finally, we selected 42 papers and used 765 data points.

Explanatory variables

Explanatory variables for periphyton $\delta^{13}\text{C}$ were extracted from individual studies or, for a small number of cases, provided directly by authors. Biome (alpine; arctic; boreal; temperate; tropical), sampling season, and watershed area (km^2 ; hereafter, WA) of the study sites were used as “regional controls”. We categorized biome of the study sites from the location of rivers studied. Sampling periods were categorized into groups approximating four seasons. The periods March-May, June-August, September-November, December-February corresponded to spring, summer, autumn, and winter for the Northern hemisphere while to autumn, winter, spring, and summer for the Southern hemisphere. The effects of seasonality on periphyton $\delta^{13}\text{C}$ were categorized as a regional control because of strong relationship between season and productivity. Canopy cover percentage (%; hereafter, CC), water current velocity (cm s^{-1} ; hereafter, WCV), chlorophyll *a* density (mg m^{-2} ; hereafter, CHLA), and dominant taxa (bryophytes; cyanobacteria; diatoms; green algae; macrophytes; red algae; multiple taxa mixtures) were used as “local controls”.

All numerical variables, excluding percent or categorical variables, were analyzed after transformation by $\log_{10} x$ for WA and CHLA and by $\log_{10} (x + 1)$ for WCV because there were zero values in the WCV (Table 1), except for missing values. In the meta-analysis dataset, we did not have enough data for a statistically robust sample size for aqueous CO_2 concentration and $\delta^{13}\text{C}$ signatures, although these factors are important for determining $\delta^{13}\text{C}$

Ishikawa et al. Meta-analyses of lotic periphyton $\delta^{13}\text{C}$ of periphyton (Finlay 2004).

Linear and non-linear modeling procedures

We tested the relationships between periphyton $\delta^{13}\text{C}$ and the potential explanatory variables. We analyzed the effects of “regional” and “local” controls separately by building two general linear models (GLMs) to test the relative contributions of potential explanatory variables to periphyton $\delta^{13}\text{C}$ as follows:

Full model (regional controls):

$$\text{Periphyton } \delta^{13}\text{C} = \text{Biome} + \text{Season} + \text{WA}$$

Full model (local controls):

$$\text{Periphyton } \delta^{13}\text{C} = \text{CC} + \text{WCV} + \text{CHLA} + \text{Dominant taxa}$$

We selected the best GLM by downward stepwise selection according to the Akaike Information Criterion (AIC, Akaike 1974). We also calculated AIC differences (Δ_i) and Akaike weight (ω_i), which is considered as the weight of evidence in favor of a candidate model being the best model out of the set of models considered (Burnham and Anderson 2002). For the individual relationship between each explanatory variable and periphyton $\delta^{13}\text{C}$, we compared both regressions of single linear and non-linear models with generalized cross validation (GCV) score. The smaller GCV score indicates higher fitness to the models (Wood 2008). Differences in periphyton $\delta^{13}\text{C}$ among biomes, season, and dominant taxa were tested by post-hoc multiple comparison (Holm’s test). For rivers that reported velocity data, the taxa-specific or river-specific relationships between periphyton $\delta^{13}\text{C}$ and the water current velocity were analyzed using ANCOVA (dominant taxa and rivers as the covariance of each

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ANCOVA model; $n = 283$).

Structural equation modeling procedure

To consider hierarchical orders among regional and local controls, we used structural equation modeling (SEM). The dataset ($n = 765$) included four numerical variables (WA; CC; WCV; CHLA), and periphyton $\delta^{13}\text{C}$. The SEM procedure for ecological studies is described in detail elsewhere (e.g., Shipley 2000; Grace 2006). Briefly, observed correlation matrix among the numerical variables was set to calculate a path-coefficient between two variables (see Appendix). Biome, season, and dominant taxa could not be included in the SEM because they were the nominal variables, which did not set reasonable correlation matrices.

To build a hierarchical model to explain periphyton $\delta^{13}\text{C}$, we used a three step analyses using a correlation matrix among causal and response variables. First, non-hierarchical models that assumed all possible correlations between variables were examined. Second, all reasonable pathways among variables were included *a priori* in the model. Third, we performed downward stepwise selection of the model judged by whether a path-coefficient was significant ($p < 0.05$) or not ($p > 0.05$) to arrive at the final model. The model fit to the data was assessed with the χ^2 test (Grace 2006). The SEM approach logically and statistically takes account of multiple co-linearity among the numerical variables, and thus allows more direct identification of causal associations.

The significance level of all statistical tests was assessed at $\alpha = 0.05$. We performed all statistical analyses and graphics using R ver. 2.13.1 software (R Development Core Team 2011) with DAAG (Maindonald and Braun 2011), MASS (Venables and Ripley 2002), mgcv (Wood 2011), and sem (Fox et al. 2010) packages in the library.

Results

Variation in $\delta^{13}\text{C}$ signatures of periphyton

Periphyton $\delta^{13}\text{C}$ ranged from -47.3‰ to -9.3‰ (Fig. 1). The mean periphyton $\delta^{13}\text{C}$ in the whole dataset was $-25.7 \pm 6.8\text{‰}$ (Mean \pm 1 SD, $n = 765$). The most frequent values of periphyton $\delta^{13}\text{C}$ ($-28 \sim -26\text{‰}$) overlapped with typical value of terrestrial C3 plants (Fig. 1); about 7% of the data fell into this range.

Regional patterns and controls

The model that had the lowest AIC value for regional controls included all explanatory variables (Table 2). We adopted the non-linear model as the best-fit model for the relationship between WA and periphyton $\delta^{13}\text{C}$ (Fig. 2a) because it explained more variation than the linear model (GCV: 44.3 for non-linear, 47.5 for linear). There was a significant relationship between periphyton $\delta^{13}\text{C}$ and WA (Df of smooth term = 5.69, $R^2 = 0.18$, $p < 0.001$, $n = 269$; Fig. 2a). Arctic and boreal rivers had significantly lower periphyton $\delta^{13}\text{C}$ than other biomes (Holm's test: $p < 0.05$; Fig. 3a). Periphyton $\delta^{13}\text{C}$ in winter was significantly higher than in spring and summer (Holm's test: $p < 0.001$; Fig. 3b).

Local patterns and controls

The model that had the lowest AIC value for local controls included canopy cover and dominant taxa, but Δ_i and ω_i values of the model were similar to those of the full model (Table 2). We again adopted the non-linear model as the best-fit model for the relationship between CC and periphyton $\delta^{13}\text{C}$ (Fig. 2b) because it explained more variation than the linear model (GCV: 28.8 for non-linear, 39.1 for linear). There was a significant relationship between periphyton $\delta^{13}\text{C}$ and CC (Df of smooth term = 6.33, $R^2 = 0.40$, $p < 0.001$, $n = 244$; Fig. 2b). Cyanobacteria (e.g., *Nostoc*) had significantly higher $\delta^{13}\text{C}$ than other groups (Holm's test: $p <$

0.001; Fig. 3c). Red algae (e.g., *Batrachospermum* and *Lemanea*) had significantly lower $\delta^{13}\text{C}$ than the other taxa except bryophytes and macrophytes (Holm's test: $p < 0.001$; Fig. 3c).

The effect of WCV on periphyton $\delta^{13}\text{C}$ was significantly different among dominant taxa and rivers (ANCOVA; dominant taxa: $p < 0.001$, rivers studied: $p < 0.001$, the interaction: WCV*dominant taxa: $p = 0.59$, WCV*rivers: $p = 0.048$). The $\delta^{13}\text{C}$ values of cyanobacteria and bulk periphyton (multiple taxa mixtures) had significantly negative relationships with WCV (cyanobacteria: slope = -4.2 , $R^2 = 0.26$, $p = 0.01$, $n = 22$; multiple taxa mixtures: slope = -1.8 , $R^2 = 0.02$, $p = 0.04$, $n = 200$), while other taxa did not (diatoms: $p = 0.92$, $n = 19$; green algae: $p = 0.09$, $n = 42$; bryophyte, macrophyte, red algae: no data) (Fig. 2c). The relationship between CHLA and periphyton $\delta^{13}\text{C}$ was significantly positive (slope = 3.7 , $R^2 = 0.08$, $p < 0.001$, $n = 204$; Fig. 2d).

Structural equation modeling

The SEM returned three models, though we presented only the final reduced model that contained only significant path-coefficients. Non-hierarchical null model indicated that three variables (CC, WCV, and CHLA) significantly influenced periphyton $\delta^{13}\text{C}$. The path-coefficient between WA and periphyton $\delta^{13}\text{C}$ was not significantly different from zero ($p = 0.45$). All explanatory variables were significantly correlated with each other, indicating that they were not completely independent. Since GLM results showed that WA was a strong regional control, the null model suggested that WA affected periphyton $\delta^{13}\text{C}$ through regulating other local controls.

A hierarchical model that contained all reasonable pathways was saturated (i.e., $df = 0$). The model indicated that WA occupied the top level of the hierarchy, controlling all other variables but a direct path from WA to periphyton $\delta^{13}\text{C}$ was not significantly different from zero ($p = 0.45$). Path-coefficients from WA to CC, WCV, and CHLA were all significantly

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different from zero ($p < 0.001$). CC and WCV occupied the same layer, both significantly controlling CHLA. There was no correlation between CC and WCV ($p = 0.36$). CHLA occupied the lowest level of the hierarchy. CC, WCV, and CHLA had significant paths to periphyton $\delta^{13}\text{C}$ ($p < 0.001$).

Downward stepwise selection based on p values deleted insignificant paths in saturated model. Direct regression from WA to periphyton $\delta^{13}\text{C}$ was not significant, but WA strongly regulated local CC and CHLA, which controlled periphyton $\delta^{13}\text{C}$ (Fig. 4). Correlation between CC and WCV was also deleted from the model. Insignificant χ^2 ($p > 0.05$) indicated that the model provided an acceptable fit to the data (Grace 2006).

Discussion

The mean $\delta^{13}\text{C}$ value of periphyton in various lotic ecosystems is close to or overlaps that of terrestrial litter for C3 plants. However, large variations in periphyton $\delta^{13}\text{C}$ are evident such that periphyton $\delta^{13}\text{C}$ are distinct from terrestrial sources in many streams and rivers. Below, we integrate understanding of regional and local controls of periphyton $\delta^{13}\text{C}$ into a hierarchical framework, which is useful in designing studies with carbon stable isotopic techniques.

Regional patterns and controls

Significant $\delta^{13}\text{C}$ difference among biomes suggests that variation in stream productivity controls, at least in part, fractionation of ^{13}C in photosynthesis. Although often unshaded by terrestrial vegetation, arctic and boreal rivers have low water temperatures and are nutrient poor, resulting in very low rates of primary productivity (Peterson et al. 1986). Low primary production promotes discrimination against $^{13}\text{CO}_2$ during photosynthesis (Finlay 2001).

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Isotopic fractionation is thus likely larger, resulting in lower periphyton $\delta^{13}\text{C}$ in arctic and boreal rivers than in other biomes. Alpine rivers often have little canopy shading and show higher $\delta^{13}\text{C}$ than rivers in arctic and boreal biomes, probably owing to higher $\delta^{13}\text{C}$ of DIC and/or higher primary production (e.g., McCutchan and Lewis 2001; Zah et al. 2001).

An influence of primary production is also likely to underlie the positive relationship between watershed area and periphyton $\delta^{13}\text{C}$. Gross primary production increases with watershed area due to channel widening, nutrient loading, and increased light availability (Battin et al. 2008; Bernot et al. 2010; Finlay 2011). Thus, photosynthetic activity generally increases with river size, while CO_2 concentrations tend to decrease due to uptake and degassing (Finlay 2003; Butman and Raymond 2011). These changes increase carbon limitation effects on isotopic fractionation in productive rivers, leading to the pattern of increasing periphyton $\delta^{13}\text{C}$ with river size (Finlay 2001).

Primary production increases in deciduous forest rivers during winter, because seasonal defoliation reduces canopy cover (Roberts et al. 2007). Canopy opening increases primary production, resulting in smaller fractionation and higher $\delta^{13}\text{C}$. Seasonality in periphyton $\delta^{13}\text{C}$ has been reported for studies conducted in alpine and temperate rivers (e.g., McCutchan and Lewis 2001; Finlay 2004). Since our dataset was biased towards temperate rivers, the seasonal influence we observed is most relevant to temperate biomes. We did not have enough data to address the mechanisms driving relationships with seasonality. The patterns shown here indicate that regional influences on in-stream productivity and possibly watershed biogeochemistry are controlling periphyton $\delta^{13}\text{C}$ variation.

Local patterns and controls

Periphyton $\delta^{13}\text{C}$ was lower under highly shaded, low light conditions. Moreover, a positive relationship was observed between chlorophyll *a* density and periphyton $\delta^{13}\text{C}$. Both results

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support the hypothesis that algal photosynthetic activity determines isotopic fractionation between DIC and algal cells. Photosynthetic activity decreases with increased canopy cover because of low light availability and colder temperature for primary producers, leading to greater isotopic fractionation (Behmer and Hawkins 1986; Lamberti and Steinman 1997; Finlay 2011). Thus decreasing periphyton $\delta^{13}\text{C}$ with increased canopy cover indicates that there is considerable isotopic discrimination against $^{13}\text{CO}_2$ between aqueous CO_2 and algae during photosynthesis, or possibly less HCO_3^- use by algae in shaded habitats (Doi et al. 2007; Finlay 2004; Hill and Middleton 2006; Hill et al. 2008).

The negative relationship between water current velocity and periphyton $\delta^{13}\text{C}$ values suggests that the increased CO_2 supply to algal cells with high water current velocity allows more selective $^{12}\text{CO}_2$ uptake by algae, intensifying the isotopic fractionation between aqueous CO_2 and periphyton (MacLeod and Barton 1998; Finlay et al. 1999; Trudeau and Rasmussen 2003; Singer et al. 2005). Although field and laboratory studies have often observed that faster current velocity strongly decreases periphyton $\delta^{13}\text{C}$, the general influence of water velocity on periphyton $\delta^{13}\text{C}$ was weaker in the meta-analyses than for individual studies. This difference may exist because the flow history, rather than the instantaneous current velocity, is more influential to $\delta^{13}\text{C}$ at sites with variable hydrograph conditions (Singer et al. 2005). Overall, water current velocity effects on periphyton $\delta^{13}\text{C}$ values must be considered with other environmental variables.

Our analyses show that variation in physiology among taxa exerts a strong influence on patterns of ^{13}C fractionation. Cyanobacteria typically concentrate inorganic carbon more efficiently than any other group, and are thus able to use HCO_3^- under low water current velocity (Merz-Preiß and Riding 1999; Badger 2003). The $\delta^{13}\text{C}$ value of HCO_3^- is higher than that of aqueous CO_2 (Mook et al. 1974) explaining the higher $\delta^{13}\text{C}$ of cyanobacteria compared to all other taxa examined. In contrast, red algae and bryophytes had

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lower $\delta^{13}\text{C}$ than other taxa. For red algae, isotopic discrimination during inner-cell carbon transportation is large, as most red algae cannot concentrate inorganic carbon within their cells (Raven et al. 2005). Furthermore, most bryophytes cannot use HCO_3^- as photosynthetic substrate (Prins and Elzenga 1989). Significant $\delta^{13}\text{C}$ difference among taxa suggests that the researchers should pay attention to the community structure in freshwater autotroph.

Relationships between regional and local controls

Our hierarchical model shows that multilevel-controls determine periphyton $\delta^{13}\text{C}$. The effect of watershed area on periphyton $\delta^{13}\text{C}$ was indirect in our model. Increasing watershed area is associated with higher discharge, channel widening, and reduced canopy shading above the water surface. Thus watershed area controls canopy cover and water current velocity on average, which locally control periphyton $\delta^{13}\text{C}$. The significant direct paths from watershed area to chlorophyll *a* density suggests that large rivers can support higher biomass of periphyton, probably because of greater light and nutrient availability.

The model is closely related to the structure of geomorphology and productivity in lotic ecosystems (e.g., Vannote et al. 1980), indicating that periphyton $\delta^{13}\text{C}$ changes along with river continuum through several different pathways. In a food web context, a recent study showed that $\delta^{13}\text{C}$ gradients of macroinvertebrates from upland to lowland rivers reflect longitudinal patterns of their resource base (Kobayashi et al. 2011). Variability in periphyton $\delta^{13}\text{C}$ across multiple spatial scales is clearly transferred to the lotic food webs, showing the importance of multiscale analyses for understanding food web dynamics using $\delta^{13}\text{C}$ signatures in fluvial networks.

In the dataset, however, the low R^2 values of both non-hierarchical and hierarchical models suggest that unexplained $\delta^{13}\text{C}$ variation in lotic periphyton remains. Lack of data for dissolved CO_2 concentration and/or $\delta^{13}\text{C}$ values of DIC in study sites might limit our

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modeling approach for prediction of periphyton $\delta^{13}\text{C}$. For example, the positive relationship between watershed area and periphyton $\delta^{13}\text{C}$ may reflect lower CO_2 concentration in large rivers (Finlay 2003; Butman and Raymond 2011). DIC $\delta^{13}\text{C}$ variation may also have been important since values typically range between -13‰ to -9‰ across most rivers (Finlay 2003).

Since periphyton carbon has longer turnover time than aqueous CO_2 and HCO_3^- , $\delta^{13}\text{C}$ of periphyton integrates short-term fluctuation in DIC $\delta^{13}\text{C}$ caused by variation in discharge (Finlay 2003; Doctor et al. 2008). Such fluctuations are about 2-4‰, much smaller than observed variation in periphyton $\delta^{13}\text{C}$ (i.e., $\sim 15\text{‰}$) at small scales. In addition to the effect of DIC, short-term variations in water current velocity and primary production may also be important for periphyton $\delta^{13}\text{C}$ (France and Cattaneo 1998; Singer et al. 2005). Finer resolution studies, rather than global meta-analysis, are thus required to address temporal variation in periphyton $\delta^{13}\text{C}$.

Perspectives for food web studies

While a global model to predict specific $\delta^{13}\text{C}$ values of periphyton was beyond the scope of this study, our analyses provides information on the likelihood of overlap or isotopic separation of organic matter sources. In settings where $\delta^{13}\text{C}$ value of periphyton are most likely to overlap with that of terrestrial litter, alternative methods can be used to avoid expense associated with extensive sampling for $\delta^{13}\text{C}$. We suggest use of alternative isotopes such as carbon-14 natural abundance ($\Delta^{14}\text{C}$), and sulfur and hydrogen stable isotopes to identify the food web structure and energy sources (Doi et al. 2006; Finlay et al. 2010; Ishikawa et al. 2010). For example, the $\Delta^{14}\text{C}$ value of periphyton is corrected by $\delta^{13}\text{C}$ and therefore is not affected by isotopic fractionation (Ishikawa et al. 2012). These isotope tracers may in some situations provide better estimation of food web attributes than $\delta^{13}\text{C}$. The

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combined use of such alternative isotopes with $\delta^{13}\text{C}$ analysis will be helpful for better understanding of carbon source contribution to freshwater food webs.

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2 Table 1:

3 Summary of variables with abbreviation, type of data, units, category, and statistical transformation for modeling.

Control	Variable	Abbreviation	Type of data	Unit	Category	Statistical transformation
Regional	Biome		Categorical		Alpine; Arctic; Boreal; Temperate; Tropical	
	Season		Categorical		Spring; Summer; Autumn; Winter	
	Watershed area	WA	Numerical	km ²		log ₁₀ x
Local	Canopy cover	CC	Numerical	%		NA
	Water current velocity	WCV	Numerical	cm s ⁻¹		log ₁₀ (x+1)
	Chlorophyll <i>a</i> density	CHLA	Numerical	mg m ⁻²		log ₁₀ x
	Dominant taxa		Categorical		Bryophytes; Cyanobacteria; Diatoms; Green algae; Macrophytes; Red algae; Multiple taxa mixtures	

4

5 Table 2: Stepwise tests for regional and local controls to explain periphyton $\delta^{13}\text{C}$. Δ_i and ω_i
 6 were AIC differences and Akaike weights, respectively.

	Df	Sum of sq	RSS	AIC	Δ_i	ω_i
Regional controls						
Full model			8997	964.2		0.999
+ WA + Biome	4	897.2	9894	981.8	17.6	<0.001
+ Season + Biome	1	1957.1	10954	1015.1	50.9	<0.001
+ WA + Season	4	2810	11807	1029.3	65.1	<0.001
Local controls						
+ Dominant taxa + WCV + CC	1	5.1	2447	365.8		0.48
+ Dominant taxa + CHL + CC	1	21.6	2463	366.6	0.8	0.32
Full model			2441	367.5	1.7	0.20
+ CC + WCV + CHLA	1	270.6	2712	377.9	12.1	<0.001
+ Dominant taxa + WCV + CHLA	1	666.2	3108	394.0	28.2	<0.001

7

8

9 **Figure legends**

10

11 Figure 1:

12 Frequency of $\delta^{13}\text{C}$ of lotic periphyton (‰, mean: -25.7‰ , SD: 6.8, $n = 765$). The black bar
13 indicates overlap with typical values of terrestrial C3 plants ($-28 \sim -26 \text{‰}$)

14

15 Figure 2:

16 Relationships between periphyton $\delta^{13}\text{C}$ (‰) and a) watershed area (WA: km^2 , $n = 269$), b)
17 canopy cover (CC: %, $n = 244$), c) water current velocity (WCV: cm s^{-1} , $n = 283$), and d)
18 chlorophyll *a* density (CHLA: mg m^{-2} , $n = 204$). The lines of a) and b) are given by smooth
19 spline regression. The purple dashed line of c) is given by linear regression for $\delta^{13}\text{C}$ of
20 cyanobacteria. The black lines of c) and d) are given by linear regression for $\delta^{13}\text{C}$ of multiple
21 taxa mixtures. Color version available online

22

23 Figure 3:

24 Box plot for periphyton $\delta^{13}\text{C}$ (‰) and a) biome (Alpine: $n = 30$, Arctic: $n = 19$, Boreal: $n =$
25 104, Temperate: $n = 570$, Tropical: $n = 42$), b) season (Spring: $n = 87$, Summer: $n = 329$,
26 Autumn: $n = 95$, Winter: $n = 114$), and c) dominant taxa (Bryophytes: $n = 16$, Cyanobacteria:
27 $n = 46$, Diatoms: $n = 52$, Green algae: $n = 94$, Macrophytes: $n = 8$, Red algae: $n = 22$, Multiple
28 taxa mixtures: $n = 527$). The box and bar depict inter-quartile (Q1 and 3) and median,
29 respectively. The whisker represents the most extreme data point that is no more than 1.5
30 times the inter-quartile. Outliers are shown where applicable

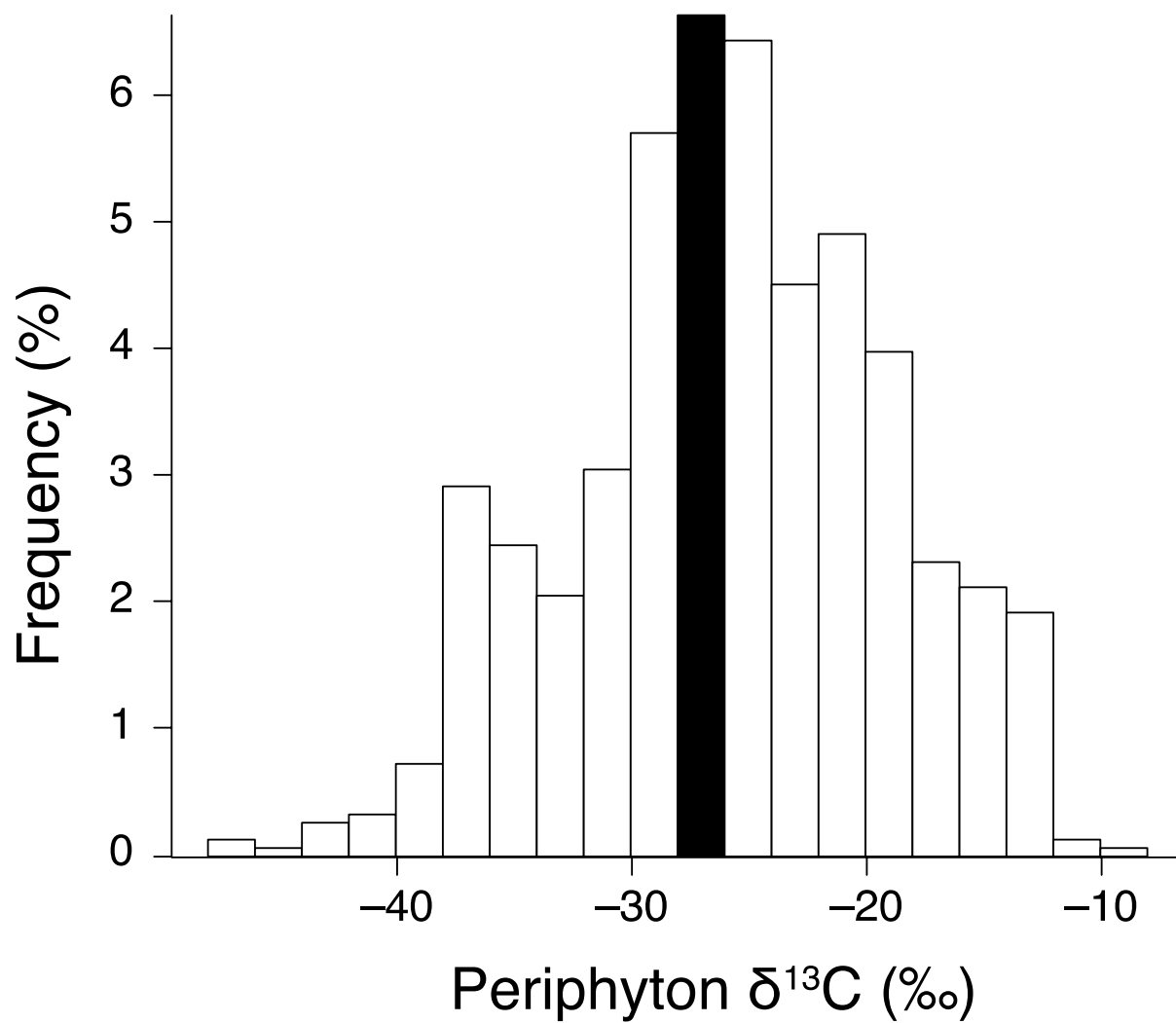
31

32 Figure 4:

33 Path-diagram containing only significant paths. For the non-hierarchical null model

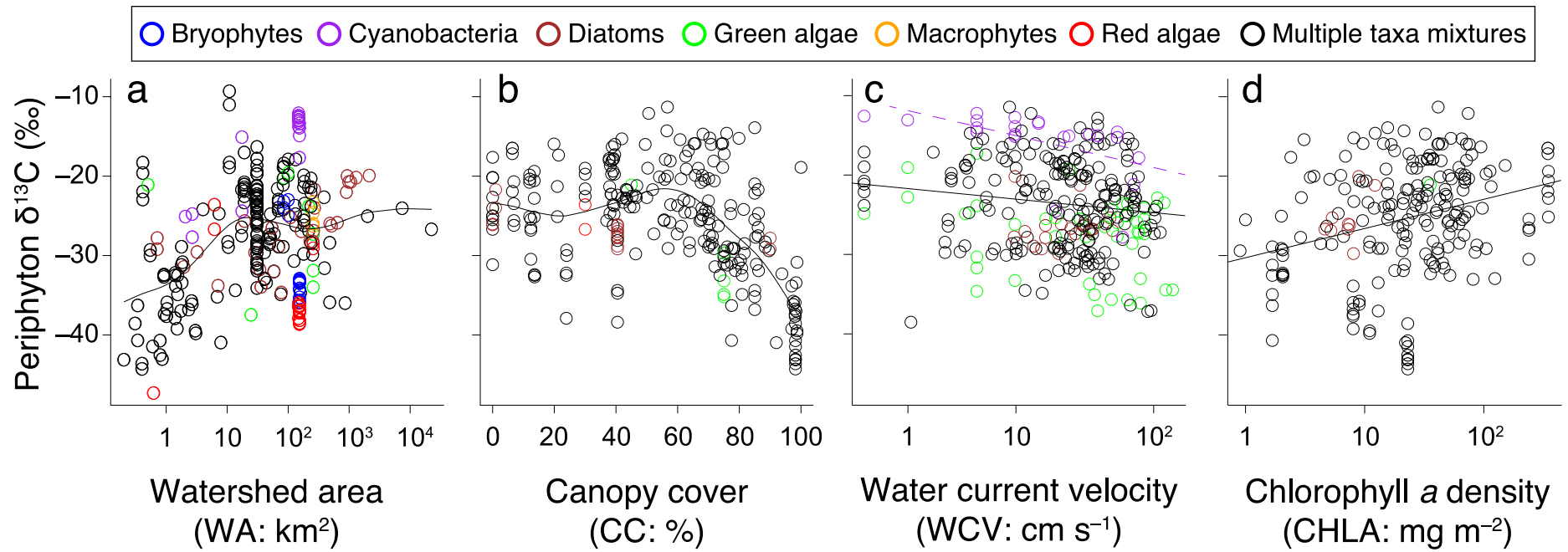
34 (including multiple linear regression model), and for the full model with all reasonable
35 combinations of paths, see Appendix. The numbers near each arrow indicate the standardized
36 path-coefficient. One-sided arrows indicate causality. Solid and dashed arrows indicate
37 positive and negative relationships, respectively, and arrow thickness is scaled to the
38 path-coefficient value. “e” represents unexplained variance of dependent variables.
39 Insignificant χ^2 of 1.40 (df = 2, n = 765, p = 0.50) indicates that the model provides an
40 acceptable fit to the data (Grace 2006)

41



42

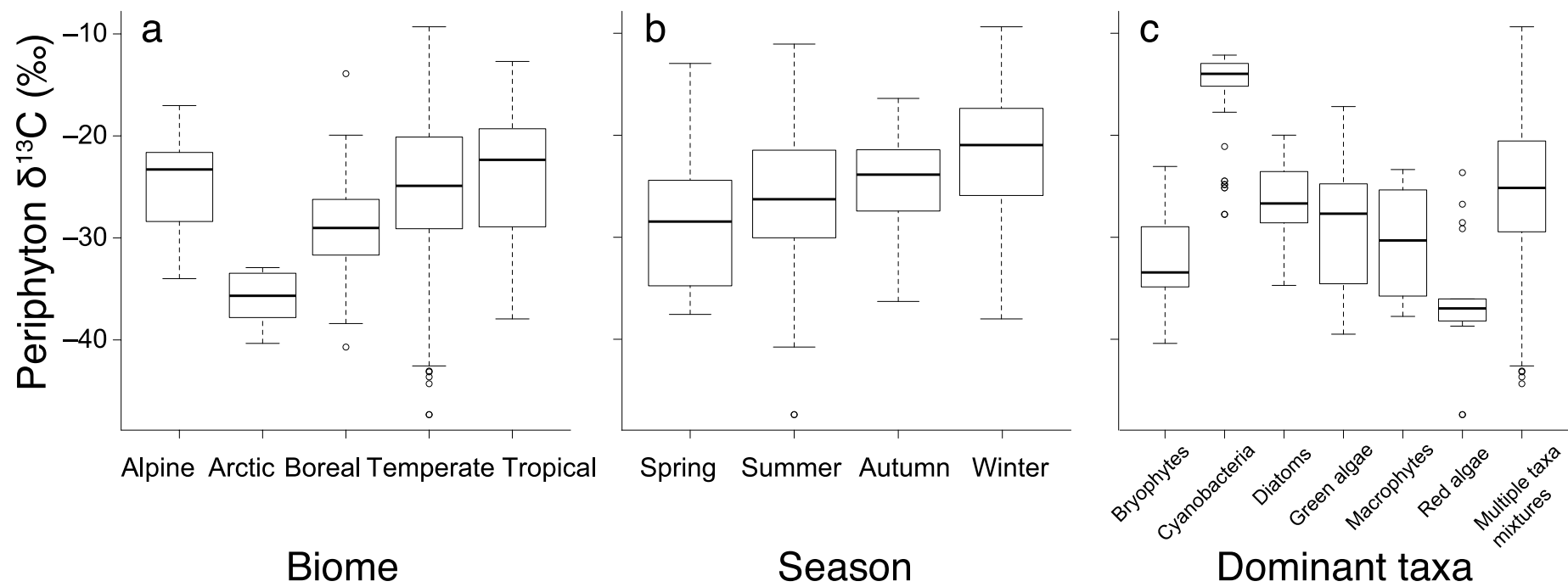
43 Figure 1



44

45 Figure 2

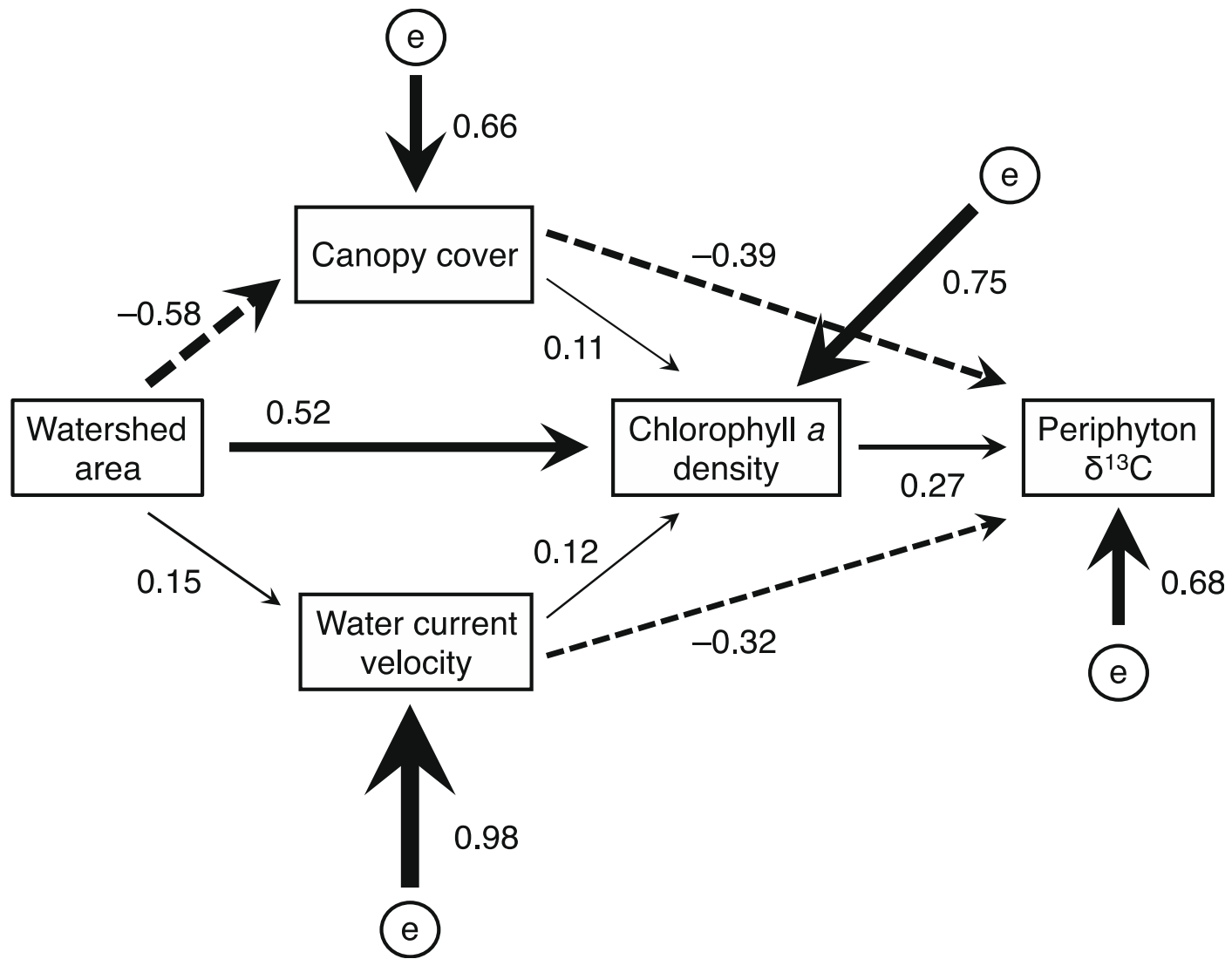
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48 Figure 3

49



50

51 Figure 4

Appendix 1: List of the papers analyzed in this study. “#” represents paper ID used throughout the Appendices

#	Author(s)	Year	Title	Journal	Volume	Page(s)
1	Bergfur J, Johnson RK, Sandin L, Goedkoop W	(2009)	Effects of nutrient enrichment on C and N stable isotope ratios of invertebrates, fish and their food resources in boreal streams	Hydrobiologia	628	67–79
2	Bunn SE, Davis PM, Winning M	(2003)	Sources of organic carbon supporting the food web of an arid zone floodplain river	Freshw Biol	48	619–635
3	Chessman BC, Westhorpe DP, Mitrovic SM, Hardwick L	(2009)	Trophic linkages between periphyton and grazing macroinvertebrates in rivers with different levels of catchment development	Hydrobiologia	625	135–150
4	Dekar MP, Magoulick DD, Huxel GR	(2009)	Shifts in the trophic base of intermittent stream food webs	Hydrobiologia	635	263–277
5	Doi H, Takemon Y, Ohta T, Ishida Y, Kikuchi E	(2007)	Effects of reach-scale canopy cover on trophic pathways of caddisfly larvae in a Japanese mountain stream	Mar Freshw Res	58	811–817
6	England LE, Rosemond AD	(2004)	Small reductions in forest cover weaken terrestrial-aquatic linkages in headwater streams	Freshw Biol	49	721–734
7	Evans-White M, Dodds WK, Gray LJ, Fritz KM	(2001)	A comparison of the trophic ecology of the crayfishes (<i>Orconectes nais</i> (Faxon) and <i>Orconectes neglectus</i> (Faxon)) and the central stoneroller minnow (<i>Campostoma anomalum</i> (Rafinesque)): omnivory in a tallgrass prairie stream	Hydrobiologia	462	131–144
8	Finlay JC	(2001)	Stable-carbon-isotope ratios of river biota: implications for energy flow in lotic food webs	Ecology	82	1052–1064
9	Finlay JC	(2004)	Patterns and controls of lotic algal stable carbon isotope ratios	Limnol Oceanogr	49	850–861
10	Finlay JC, Power ME, Cabana G	(1999)	Effects of water velocity on algal carbon isotope ratios: implications for river food web studies	Limnol Oceanogr	44	1198–1203
11	France R, Cattaneo A	(1998)	$\delta^{13}\text{C}$ variability of benthic algae: effects of water colour via modulation by stream current	Freshw Biol	39	617–622
12	Fuentes Brito E, Moulton TP, De Souza M, Bunn SE	(2006)	Stable isotope analysis indicates microalgae as the predominant food source of fauna in a coastal forest stream, south-east Brazil	Austral Ecol	31	623–633
13	Füreder L, Welter C, Jackson JK	(2003)	Dietary and stable isotope ($\delta^{13}\text{C}$, $\delta^{15}\text{N}$) analyses in alpine stream insects	Int Rev Hydrobiol	88	314–331
14	Godwin CM, Arthur MA, Carrick HJ	(2009)	Periphyton nutrient status in a temperate stream with mixed land-uses: implications for watershed nitrogen storage	Hydrobiologia	623	141–152
15	Göthe E, Lepori F, Malmqvist B	(2009)	Forestry affects food webs in northern Swedish coastal streams	Fund Appl Limnol	175	281–294

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Continued

#	Author(s)	Year	Title	Journal	Volume	Page(s)
16	Hamilton SK, Sippel, SJ, Bunn SE	(2005)	Separation of algae from detritus for stable isotope or ecological stoichiometry studies using density fractionation in colloidal silica	Limnol Oceanogr Methods	3	149–157
17	Hury AD, Riley RH, Young RG, Arbuckle CJ, Peacock K	(2002)	Natural-abundance stable C and N isotopes indicate weak upstream-downstream linkage of food webs in a grassland river	Arch Hydrobiol	153	177–196
18	Ishikawa NF, Uchida M, Shibata Y, Tayasu I	(2012)	Natural C-14 provides new data for stream food-web studies: a comparison with C-13 in multiple stream habitats	Mar Freshw Res	63	210–217
19	Junger M, Planas D	(1994)	Quantitative use of stable carbon-isotope analysis to determine the trophic base of invertebrate communities in a boreal forest lotic system	Can J Fish Aquat Sci	51	52–61
20	Lau DCP, Leung KMY, Dudgeon D	(2009)	Evidence of rapid shifts in the trophic base of lotic predators using experimental dietary manipulations and assimilation-based analyses	Oecologia	159	767–776
21	Lau DCP, Leung KMY, Dudgeon D	(2009)	What does stable isotope analysis reveal about trophic relationships and the relative importance of allochthonous and autochthonous resources in tropical streams? A synthetic study from Hong Kong	Freshw Biol	54	127–141
22	Li AOY, Dudgeon D	(2008)	Food resources of shredders and other benthic macroinvertebrates in relation to shading conditions in tropical Hong Kong streams	Freshw Biol	53	2011–2025
23	Manetta GI, Benedito-Cecilio E, Martinelli M	(2003)	Carbon sources and trophic position of the main species of fishes of Baía River, Paraná River floodplain, Brazil	Braz J Biol	63	283–290
24	McCutchan JH, Lewis WM	(2001)	Seasonal variation in stable isotope ratios of stream algae	Verh Internat Verein Limnol	27	3304–3307
25	Parkyn SM, Collier KJ, Hicks BJ	(2001)	New Zealand stream crayfish: functional omnivores but trophic predators?	Freshw Biol	46	641–652
26	Pereira AL, Benedito E, Sakuragui CM	(2007)	Spatial variation in the stable isotopes of ^{13}C and ^{15}N and trophic position of <i>Leporinus friderici</i> (Characiformes, Anostomidae) in Corumbá Reservoir, Brazil	An Acad Bras Cienc	79	41–49
27	Perry RW, Bradford MJ, Grout JA	(2003)	Effects of disturbance on contribution of energy sources to growth of juvenile chinook salmon (<i>Oncorhynchus tshawytscha</i>) in boreal streams	Can J Fish Aquat Sci	60	390–400
28	Primavera JH	(1996)	Stable carbon and nitrogen isotope ratios of Penaeid juveniles and primary producers in a riverine mangrove in Guimaras, Philippines	B Mar Sci	58	675–683
29	Rasmussen JB, Trudeau V	(2007)	Influence of velocity and chlorophyll standing stock on periphyton $\delta^{13}\text{C}$ and $\delta^{15}\text{N}$ in the Ste. Marguerite River system, Quebec	Can J Fish Aquat Sci	64	1370–1381
30	Rasmussen JB, Trudeau V	(2010)	How well are velocity effects on $\delta^{13}\text{C}$ signatures transmitted up the food web from algae to fish?	Freshw Biol	55	1303–1314

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Continued

#	Author(s)	Year	Title	Journal	Volume	Page(s)
31	Rasmussen JB, Trudeau V, Morinville G	(2009)	Estimating the scale of fish feeding movements in rivers using $\delta^{13}\text{C}$ signature gradients	J Anim Ecol	78	674–685
32	Robinson CT, Schmid D, Svoboda M, Bernasconi SM	(2008)	Functional measures and food webs of high elevation springs in the Swiss alps	Aquat Sci	70	432–445
33	Singer GA, Panzenböck M, Weigelhofer G, Marchesani C, Waringer J, Wanek W, Battin TJ	(2005)	Flow history explains temporal and spatial variation of carbon fractionation in stream periphyton	Limnol Oceanogr	50	706–712
34	Spencer CN, Gabel KO, Hauer FR	(2003)	Wildfire effects on stream food webs and nutrient dynamics in Glacier National Park, USA	Forest Ecol Manag	178	141–153
35	Thorp JH, DeLong MD, Greenwood KA, Casper AF	(1998)	Isotopic analysis of three food web theories in constricted and floodplain regions of a large river	Oecologia	117	551–563
36	Trimmer M, Hildrew AG, Jackson MC, Pretty JL, Grey J	(2009)	Evidence for the role of methane-derived carbon in a free-flowing, lowland river food web	Limnol Oceanogr	54	1541–1547
37	Verburg P, Kilham SS, Pringle CM, Lips KR, Drake DL	(2007)	A stable isotope study of a neotropical stream food web prior to the extirpation of its large amphibian community	J Trop Ecol	23	643–651
38	Walters AW, Barnes RT, Post DM	(2009)	Anadromous alewives (<i>Alosa pseudoharengus</i>) contribute marine-derived nutrients to coastal stream food webs	Can J Fish Aquat Sci	66	439–448
39	Watanabe K, Monaghan MT, Takemon Y, Omura T	(2008)	Biodilution of heavy metals in a stream macroinvertebrate food web: Evidence from stable isotope analysis	Sci Total Environ	394	57–67
40	Winemiller KO, Hoeninghaus DJ, Pease AA, Esselman PC, Honeycutt RL, Gbanaador D, Carrera, Payne J	(2011)	Stable isotope analysis reveals food web structure and watershed impacts along the fluvial gradient of a Mesoamerican coastal river	River Res Appl	27	791–803
41	Zah R, Burgherr P, Bernasconi SM, Uehlinger U	(2001)	Stable isotope analysis of macroinvertebrates and their food sources in a glacier stream	Freshw Biol	46	871–882
42	Zeug SC, Winemiller KO	(2008)	Evidence supporting the importance of terrestrial carbon in a large-river food web	Ecology	89	1733–1743
43	Unpublished data from JC Finlay	-	-	-	-	-
44	Unpublished data from H Doi	-	-	-	-	-

Total**42 published & 2 unpublished sources**

Appendix 2: Study site, Sample size, periphyton $\delta^{13}\text{C}$, and explanatory (numerical) variables collected from each paper. When a paper has multiple data, mean and ± 1 SD are shown

#	Site	n	Periphyton $\delta^{13}\text{C}$ (‰)	Watershed area (km^2)	Canopy cover (%)	Water current velocity (cm s^{-1})	Chlorophyll <i>a</i> density (mg m^{-2})
1	Algangsán & others	6	-31.6 \pm 3.4	299.8 \pm 290.7			
2	Mayfield & others	13	-19.6 \pm 3.0				
3	Murrumbidgee River	43	-26.6 \pm 1.6				
4	Arkansas River	15	-23.4 \pm 3.5	163.0 \pm 64.5			
5	Kamo River	8	-17.4 \pm 2.0		48.6 \pm 22.6	64.9 \pm 9.5	52.5 \pm 14.2
6	Chattahoochee River	2	-24.5 \pm 0.4	5.5 \pm 2.3	79.3 \pm 3.0		
7	Kings Creek	4	-14.4 \pm 4.9	10.6 \pm 0.0			
8	Eel River	6	-32.3 \pm 5.1	79.4 \pm 148.3	51.0 \pm 46.6		
9	Eel River	33	-34.4 \pm 6.9	971.1 \pm 4001.7	77.6 \pm 35.8		15.2 \pm 6.6
10	Eel River	6	-23.3 \pm 3.7	108.9 \pm 74.8	44.0 \pm 35.6		21.9 \pm 10.7
11	Streams in Laurentian mountains	15	-28.8 \pm 4.1		40.5 \pm 0.0	21.0 \pm 11.9	
12	Córrego da Andorinha	5	-22.5 \pm 2.8		27.8 \pm 6.8		
13	River Inn	2	-26.3 \pm 2.1	50.0 \pm 0.0			
14	Spring Creek	20	-21.9 \pm 4.2		25.9 \pm 30.4		203.3 \pm 141.8
15	Swedish streams	14	-33.1 \pm 3.3	1.1 \pm 0.5	45.0 \pm 30.1		1.9 \pm 0.2

Continued

#	Site	n	Periphyton $\delta^{13}\text{C}$ (‰)	Watershed area (km^2)	Canopy cover (%)	Water current velocity (cm s^{-1})	Chlorophyll <i>a</i> density (mg m^{-2})
16	Several sites in Queensland	20	-25.5 ± 4.7				
17	Taieri River	22	-24.2 ± 4.3	173.1 ± 78.5			
18	Seri River	48	-24.5 ± 3.9	30	56.1 ± 16.0	37.3 ± 18.0	53.5 ± 29.7
19	Montmorency River	6	-25.5 ± 2.9	90.6 ± 135.1	40.3 ± 40.6		
20	Pak Ngau Shek Stream	1	-14.2				
21	Several streams in Hong Kong	4	-22.7 ± 2.3		50.0 ± 23.1		
22	Hong Kong stream	9	-18.0 ± 4.3			37.8 ± 9.7	
23	Parana River	1	-28.6				5.3
24	St. Vrain Creek	14	-19.1 ± 2.1				
25	Mangaotama Stream & Firewood Creek	1	-21.1	0.5	45.0		35.5
26	Corumba River	1	-21.6	65.0			6.8
27	Yukon River	6	-32.4 ± 3.2	195.2 ± 214.2			
28	Sibunag River & others	5	-19.8 ± 3.9				
29	St. Marguerite River	9	-25.4 ± 3.0		0.0 ± 0.0	32.0 ± 13.2	7.4 ± 2.3
30	St. Marguerite River & others	15	-29.8 ± 0.7			21.0 ± 17.0	

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Continued

#	Site	n	Periphyton $\delta^{13}\text{C}$ (‰)	Watershed area (km^2)	Canopy cover (%)	Water current velocity (cm s^{-1})	Chlorophyll <i>a</i> density (mg m^{-2})
31	St. Marguerite River & others	25	-27.0 ± 4.6	406.3 ± 525.8			
32	Several springs in Switzerland	6	-30.6 ± 1.8				
33	Kleine Erlauf	3	-31.8 ± 1.9	40.0 ± 0.0		44.7 ± 3.4	96.5 ± 23.7
34	Akokala & Bowman creeks	1	-21.4				
35	Ohio River	3	-21.9 ± 1.1				
36	River Lambourn	4	-35.8 ± 1.4				
37	Rio Guabal	8	-31.7 ± 2.4		75.0 ± 0.0		
38	Several streams in Connecticut	34	-28.8 ± 2.4				
39	Several streams in Japan	4	-21.7 ± 2.7		37.5 ± 47.9		
40	Monkey River	9	-25.2 ± 6.7				
41	Roseg River	22	-22.9 ± 3.0	31.8 ± 24.8			
42	Brazos River	3	-21.1 ± 4.0		0.0 ± 0.0		2.3 ± 1.3
43	Several streams in the US	229	-26.1 ± 8.3	127.7 ± 50.2		40.2 ± 35.3	
44	Kurama River	60	-21.8 ± 5.7		65.5 ± 12.1	18.4 ± 12.1	26.5 ± 20.8
Total		765	-25.7 ± 6.8	272.2 ± 1569.5	54.8 ± 28.6	33.5 ± 27.2	47.5 ± 72.3

Appendix 3: Explanatory (categorical) variables and topographical information (altitude, slope, and width of site) collected from each paper.

When a paper has multiple data, all attributes are listed and/or mean and ± 1 SD are shown

#	Biome	Season	Dominant taxa	Altitude (m a.s.l.)	Slope (%)	Width (m)
1	Boreal	Autumn	Multiple taxa mixtures	66.6 \pm 68.7		7.8 \pm 2.6
2	Temperate	Spring; Autumn	Multiple taxa mixtures			70.4 \pm 51.6
3	Temperate	Annual	Multiple taxa mixtures			
4	Temperate	Spring; Summer; Winter	Multiple taxa mixtures			28.5 \pm 3.5
5	Temperate	Winter	Multiple taxa mixtures	110.0		27.0 \pm 0.0
6	Temperate	Autumn	Multiple taxa mixtures	465.0 \pm 48.1	1.1 \pm 0.6	4.2 \pm 0.8
7	Temperate	Summer; Winter	Multiple taxa mixtures			
8	Temperate	Summer	Multiple taxa mixtures			
9	Temperate	Spring; Summer	Multiple taxa mixtures			
10	Temperate	Summer	Multiple taxa mixtures			
11	Boreal	Summer	Diatoms			
12	Tropical	Summer	Multiple taxa mixtures	60.8 \pm 16.5		14.1 \pm 3.6
13	Alpine		Multiple taxa mixtures	1900.0		
14	Boreal	Annual	Multiple taxa mixtures			
15	Boreal	Summer	Multiple taxa mixtures	30.8 \pm 23.6	8.2 \pm 5.3	1.2 \pm 0.4

Continued

#	Biome	Season	Dominant taxa	Altitude (m a.s.l.)	Slope (%)	Width (m)
16	Temperate		Multiple taxa mixtures			
17	Temperate	Summer; Autumn	Bryophytes; Green algae; Red algae; Macrophytes; Multiple taxa mixtures	669.5 \pm 117.5	0.3 \pm 0.3	15.8 \pm 3.9
18	Temperate	Winter	Multiple taxa mixtures			
19	Boreal		Diatoms; Red algae	642.3 \pm 104.5	4.5 \pm 3.8	19.8 \pm 26.5
20	Tropical		Multiple taxa mixtures			
21	Temperate		Multiple taxa mixtures			
22	Tropical	Winter	Multiple taxa mixtures	150.0 \pm 96.8		3.8 \pm 0.9
23	Temperate		Multiple taxa mixtures			
24	Boreal	Annual	Multiple taxa mixtures			
25	Temperate	Summer; Winter	Green algae	81.1 \pm 25.2	2.5 \pm 1.2	
26	Tropical		Multiple taxa mixtures			
27	Boreal	Summer	Multiple taxa mixtures			
28	Tropical	Winter	Multiple taxa mixtures ; Green algae			
29	Boreal	Summer	Diatoms			18.0 \pm 15.8
30	Boreal	Summer	Multiple taxa mixtures			

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Continued

#	Biome	Season	Dominant taxa	Altitude (m a.s.l.)	Slope (%)	Width (m)
31	Boreal		Diatoms			
32	Alpine		Multiple taxa mixtures	2008.7 ± 277.2		
33	Boreal		Multiple taxa mixtures			
34	Boreal	Summer	Diatoms			
35	Temperate	Summer; Winter	Diatoms			920.0
36	Temperate		Macrophytes			
37	Tropical	Summer	Multiple taxa mixtures ; Green algae	750.0		
38	Temperate	Spring; Summer	Multiple taxa mixtures			
39	Temperate		Multiple taxa mixtures	206.0 ± 91.2		4.8 ± 2.3
40	Tropical	Winter	Multiple taxa mixtures			
41	Alpine		Multiple taxa mixtures	1833.8 ± 431.3		
42	Temperate		Multiple taxa mixtures			
43	Arctic; Boreal; Temperate		Bryophytes; Cyanobacteria; Diatoms; Green algae; Red algae; Macrophytes; Multiple taxa mixtures			
44	Temperate	Winter	Multiple taxa mixtures			
Total				747.8 ± 747.3	3.4 ± 4.7	29.2 ± 89.2

Appendix 4: Physicochemical (nutrient concentration, water temperature, pH, geology, and vegetation of site) information collected from each paper. When a paper has multiple data, all attributes are listed and/or mean and ± 1 SD are shown

#	Phosphorous (ng $\text{PO}_4^{3-} \text{L}^{-1}$)	Nitrate+Nitrite (($\mu\text{g NO}_2^- + \mu\text{g NO}_3^-$) L^{-1})	Ammonium (ng $\text{NH}_4^+ \text{L}^{-1}$)	Water temperature ($^\circ\text{C}$)	pH	Geology	Vegetation
1		0.488 \pm 0.705			7.1 \pm 0.5		Coniferous forest
2							
3	276.3 \pm 65.0	0.559 \pm 0.247	399.9 \pm 112.6				
4		0.010 \pm 0.000					Upland hardwood species mixed with shortleaf pine
5							Oak; Bamboo
6							
7							Grasses; Shrubs
8							
9							
10				20.9 \pm 3.3	8.4 \pm 0.3		
11				25.0	6.9		
12					6.6	Granite	
13							
14	4.8 \pm 1.6	2.590 \pm 1.497		9.4 \pm 1.0	7.7 \pm 0.6		Several vegetations included
15					6.7 \pm 0.5	Granite; Diabase	<i>Picea abies</i> ; <i>Alnus incana</i>

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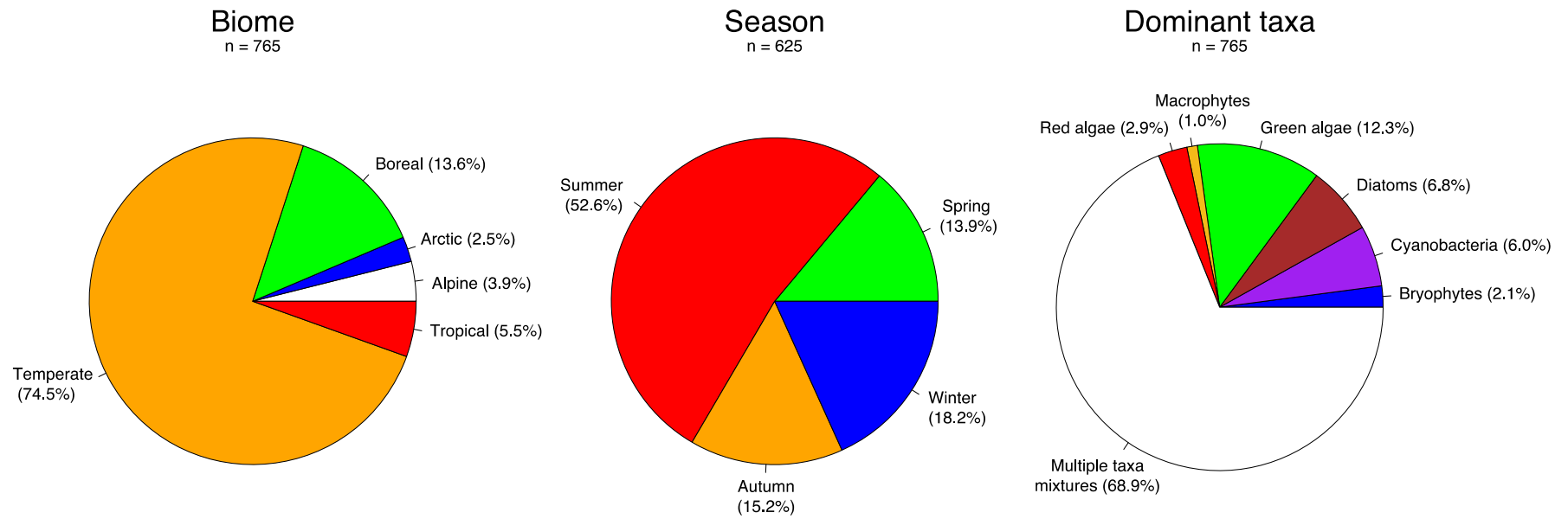
Continued

#	Phosphorous (ng $\text{PO}_4^{3-} \text{L}^{-1}$)	Nitrate+Nitrite (($\mu\text{g NO}_2^- + \mu\text{g NO}_3^-$) L^{-1})	Ammonium (ng $\text{NH}_4^+ \text{L}^{-1}$)	Water temperature ($^\circ\text{C}$)	pH	Geology	Vegetation
16							
17	3.6 \pm 0.4	0.002 \pm 0.001		13.4 \pm 0.7			Native tussock grasses; Exotic pasture grasses
18						Limestone	Fagaceae; Taxodiaceae
19							
20							
21							
22	20.3 \pm 14.4	0.252 \pm 0.349	0.1 \pm 0.2	14.7 \pm 2.5	6.7 \pm 0.1		Several vegetations included
23							Pastureland
24							Pine; Spruce-fir forest
25							Pasture; Native forest
26							
27				5.4 \pm 0.8	7.9 \pm 0.1		
28							Mangrove
29	40.3 \pm 5.7	3.208 \pm 0.352	21.1 \pm 5.5		7.2		Several vegetations included
30							

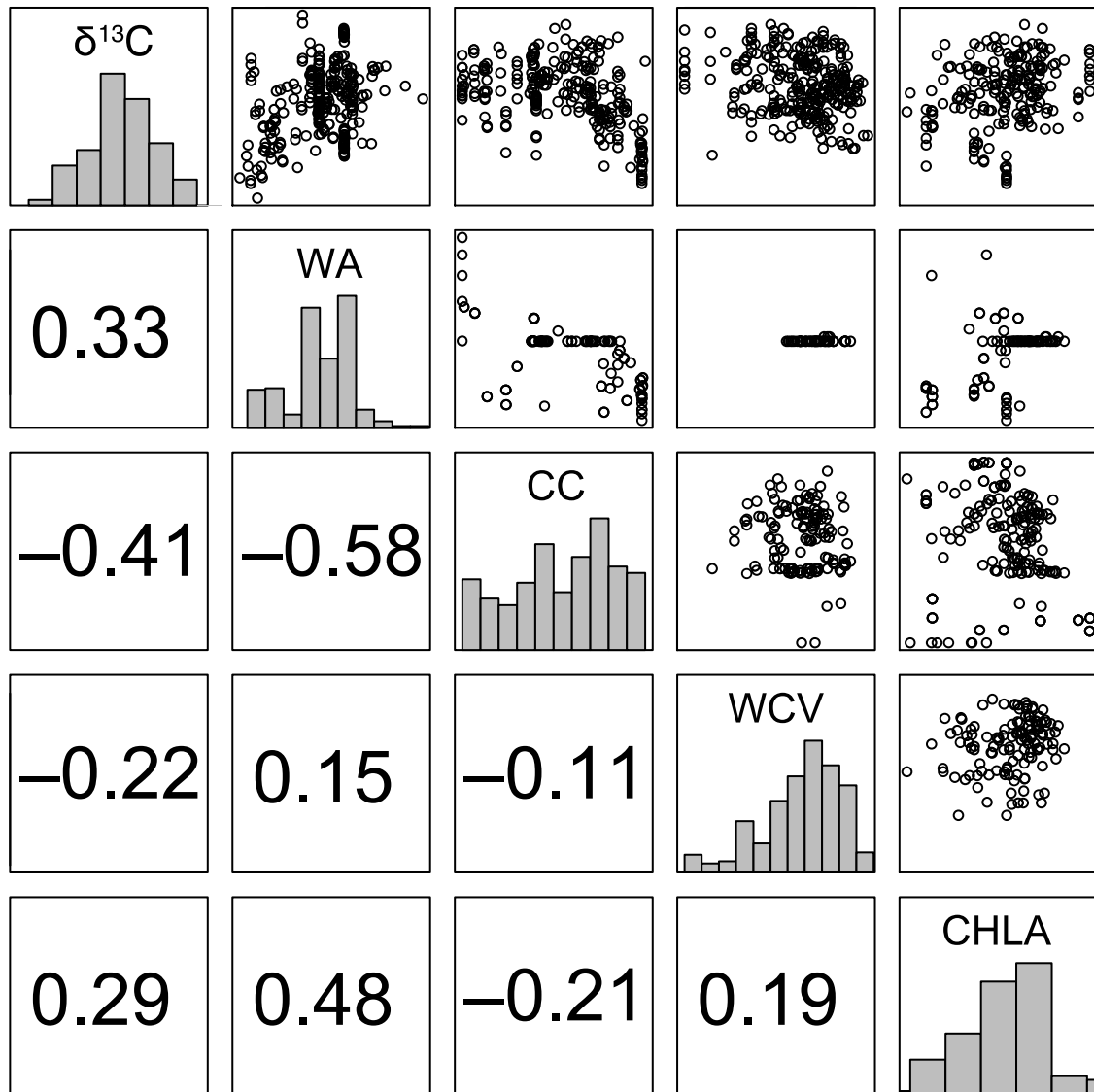
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#	Phosphorous (ng $\text{PO}_4^{3-} \text{L}^{-1}$)	Nitrate+Nitrite ($(\mu\text{g NO}_2^- + \mu\text{g NO}_3^-) \text{L}^{-1}$)	Ammonium (ng $\text{NH}_4^+ \text{L}^{-1}$)	Water temperature ($^\circ\text{C}$)	pH	Geology	Vegetation
31							
32	1.7 \pm 0.9	0.154 \pm 0.038		5.5 \pm 1.0	7.5 \pm 0.6		
33	36.0 \pm 24.2	1.420 \pm 0.277	70.0 \pm 52.0	9.3 \pm 0.1	8.3	Limestone	Agriculture; Forestry
34							
35						Limestone	Deciduous trees
36							
37				23.0			
38		0.163 \pm 0.199	18.2 \pm 5.4				
39		0.413 \pm 0.219					Several vegetations included
40							
41				3.2 \pm 0.3			Grasses; Shrubs
42							<i>Salix nigra</i> ; <i>Platanus occidentalis</i>
43	14.7 \pm 7.2	0.022 \pm 0.013	26.5 \pm 20.1				
44							
Total	76.6 \pm 117.9	0.491 \pm 0.965	235.1 \pm 208.5	12.5 \pm 7.5	7.3 \pm 0.6		

Appendix 5: Distribution of data among biomes, seasons and dominant taxa



- 1 Appendix 6: Histograms, scatter plots, and correlation matrices of variables used for structural
- 2 equation modeling to explain periphyton $\delta^{13}\text{C}$. WA, CC, WCV, and CHLA are watershed area,
- 3 canopy cover, water current velocity, and Chlorophyll *a* density, respectively



4

5

- 6 Appendix 7: Non-hierarchical null model and hierarchically saturated model. Single asterisks
 7 indicate that standardized path-coefficients are significantly different from zero ($\alpha = 0.05$)

