Factors that differentiate the H-bond strengths of water near the Schiff bases in bacteriorhodopsin and *Anabaena* sensory rhodopsin

Keisuke Saito 1, Hideki Kandori2, and Hiroshi Ishikita 1,3*

Running Title: H-bond strength of water in BR and ASR

To whom correspondence should be addressed: Hiroshi Ishikita, 202 Building E, Career-Path Promotion Unit for Young Life Scientists, Graduate School of Medicine, Kyoto University, Yoshida-Konoe-cho, Sakyo-ku, Kyoto 606-8501, Japan, Tel. +81-75-753-9286, Fax. +81-75-753-9286, E-mail: hiro@cp.kyoto-u.ac.jp

Keywords: bacteriorhodopsin, *Anabaena* sensory rhodopsin, Fourier transform infrared spectroscopy, hydrogen bond, pKa, proton transfer

Background: Bacteriorhodopsin (BR) functions as a proton pump whereas Anabaena sensory rhodopsin (ASR) function as a photosensor.

Results: pK_a of the conserved Asp residues near the Schiff base significantly differ between BR and ASR.

Conclusion: The O-D stretching frequencies for D_2O is correlated with $pK_a(Asp)$.

Significance: The presence of a strongly H-bonded water results from the proton-pumping activity in BR.

SUMMARY

Bacteriorhodopsin (BR) functions as a lightdriven proton pump, whereas Anabaena sensory rhodopsin (ASR) is believed to function as a photosensor, despite the high similarity in their protein sequences. In Fourier transform infrared (FTIR) spectroscopic studies, the lowest O-D stretch for D₂O was observed at ~2200 cm⁻¹ in BR but was significantly higher in ASR (>2500 cm⁻¹), which was previously attributed to a water molecule near the Schiff base (W402) that is Hbonded to Asp85 in BR and Asp75 in ASR. We investigated the factors that differentiate the lowest O-D stretches of W402 in BR and ASR. Quantum mechanical mechanical/molecular (QM/MM)

calculations reproduced the H-bond geometries of the crystal structures and the calculated O-D stretching frequencies were corroborated by the FTIR band assignments. The potential-energy profiles indicate that the smaller O-D stretching frequency in BR originates from the significantly higher $pK_a(Asp85)$ in BR relative to the $pK_a(Asp75)$ in ASR, which were calculated to be 1.5 and -5.1, respectively. The difference is mostly due to the influences of Ala53, Arg82, Glu194-Glu204, and Asp212 on pK_a (Asp85) in BR, and the corresponding residues Ser47, Arg72, Ser188-Asp198, and Pro206 on $pK_a(Asp75)$ in ASR. Because these residues participate in proton transfer pathways in BR but not in ASR, the presence of a strongly H-bonded water molecule near the Schiff base ultimately results from the proton-pumping activity in BR.

Bacteriorhodopsin (BR) functions as a light-driven proton pump and the driving force for its action is provided by photoisomerization of the all-*trans* retinal chromophore, which is covalently attached to Lys216 via the Schiff base, to 13-*cis*. This leads to proton transfer pathways that proceed toward the extracellular side via Asp96, the Schiff base, Asp85, and Glu204 (1-3). Asp 85 is located near the Schiff

¹ From 202 Building E, Career-Path Promotion Unit for Young Life Scientists, Graduate School of Medicine, Kyoto University, Yoshida-Konoe-cho, Sakyo-ku, Kyoto 606-8501, Japan

² Department of Frontier Materials, Nagoya Institute of Technology, Showa-ku, Nagoya 466-8555, Japan

³ Japan Science and Technology Agency (JST), PRESTO, 4-1-8 Honcho Kawaguchi, Saitama 332-0012, Japan

base and serves as the counter ion. The presence of water molecules in this region had been suggested by Fourier transform infrared (FTIR) studies (4,5), and these were later confirmed as W401, W402, and W406 in the high-resolution crystal structure of the BR ground state (PDB code 1C3W; Figure 1) (6). The three water molecules form an H-bond network with Asp85, Arg82, Asp212, and the Schiff base. Among the three water molecules, the chemical properties of W402 are of particular interest because the water molecule is located at H-bond distances of 2.63 Å and 2.87 Å from the carboxyl O atom of Asp85 and the Schiff base N atom, respectively (Table 1). From FTIR analysis of various mutants in D₂O, the lowest O-D stretching frequency of 2171 cm⁻¹ in BR was assigned to the O_{W402}–H...O_{Asp85} bond, implying that this is the strongest H-bond in the network near the Schiff base (7). The band assignment presented in FTIR studies was also supported by previous computational studies by Hayashi and coworkers (8,9).

Water molecule W402 was also found in the crystal structure of *Anabaena* sensory rhodopsin (ASR) (PDB code 1XIO; Figure 1) (10). In contrast to the proton pumping of BR, ASR is believed to function as a photosensor. Despite the high similarity of their protein sequences, some key residues that are functionally important in the proton-pumping event in BR are not conserved in ASR: Asp96 in BR, which serves as a proton donor to the Schiff base, is replaced with Ser86 in ASR; Glu194 and Glu204 in BR, which is located in the terminal region of the proton transfer pathway, are replaced with Ser188 and Asp198, respectively. Although W402 is present in both BR (6) and ASR (10), Asp212 in BR is replaced with Pro206 in ASR (Figure 1). The absence of W401 and W406 in the corresponding H-bond network of ASR may be associated with the absence of an acidic residue corresponding to Asp212 in BR. FTIR studies have suggested that the O-D stretch in water molecules was only observed at >2500 cm⁻¹ in ASR (11), which is $\sim 300 \text{ cm}^{-1}$ higher than the lowest O-D stretching frequency of 2171 cm⁻¹ in BR (7).

The significant difference in the lowest O-D stretching frequency implies that the H-bond properties of W402 are significantly different in BR and ASR. It has been established by FTIR that the lowest O-D stretching frequency of water can be found at less than 2400 cm⁻¹ in a number of proton-pumping rhodopsins (12). The same tendency also holds true for BR and ASR.

In the BR and ASR crystal structures, the angle defined by the Schiff base N, W402 O, and Asp85 O atoms $(N_{Lys}...O_{W402}...O_{Asp})$ differs significantly: the angle is 106° in BR (6) and 83° in ASR (10). Thus, it was proposed that the angle difference may

differentiate the H-bond strength of the water molecules in BR and ASR (11). On the other hand, it also appears that the O_{W402} –H... O_{Asp} angle is more crucial than the N_{Lys} ... O_{W402} ... O_{Asp} angle to the energetics of the H bond since it involves a H atom. In general, the O_{donor} –H... $O_{acceptor}$ angle strongly depends on the O_{donor} – $O_{acceptor}$ distance, i.e., the O_{donor} –H... $O_{acceptor}$ angle is more linear when the O_{donor} – $O_{acceptor}$ bond is shorter (13).

In addition, the $N_{Lys}...O_{W402}...O_{Asp}$ angle in the crystal structure can be directly altered in response to changes in the O_{W402} – $H...O_{Asp}$ distance. Because of the nature of H bonds (14-17), the O_{W402} – $H...O_{Asp85}$ distance could also be predominantly determined by the pK_a difference between the H-bond donor (W402) and acceptor (Asp) moieties. In general, a smaller pK_a difference yields a short, symmetrical H bond (14-17), and the H atom of O_{W402} –H migrates toward the acceptor moiety, Asp. Unfortunately, the energetics of the O_{W402} – $H...O_{Asp}$ bond, particularly the pK_a difference between W402 and the Asp residue, remain unclear. Thus, it is essential to clarify how the O_{W402} – $H...O_{Asp}$ distance is energetically determined in the BR and ASR crystal structures.

Here, we present calculated O-D stretching frequencies of D₂O near W402 in BR and ASR in the quantum state using a large-scale ground mechanical/molecular mechanical (OM/MM) approach. We also present the potential-energy profiles of the H bond between W402 and Asp85 in BR, and W402 and Asp75 in ASR, and demonstrate that the frequencies and H-bond properties are ultimately determined predominantly by the pK_a values of H-bond acceptors Asp85 and Asp75. By calculating p K_a (Asp85) in BR and p K_a (Asp75) in ASR through solving the linear Poisson-Boltzmann equation with explicit consideration of the protonation states for all titratable residues, we are finally able to pinpoint the factors that cause a difference of ~300 cm⁻¹ between the O-D stretching frequencies of W402 in BR and ASR.

EXPERIMENTAL PROCEDURES

As demonstrated in our previous work with photoactive yellow protein (18), we employed the following systematic modeling procedure.

First, we constructed initial molecular models of BR and ASR using their crystal structures and adding hydrogen atoms. Second, to gain better understanding of the electronic structure of W402 and the associated H-bond network, we performed large-scale QM/MM calculations for the entire BR and ASR proteins. To

gain insight into the QM/MM potential-energy profiles of the H bonds, O_{W402} –H... O_{Asp85} in BR and O_{W402} –H... O_{Asp75} in ASR, we calculated p K_a (Asp85) in BR and p K_a (Asp75) in ASR by simultaneously titrating all titratable residues in BR and ASR. Technical details of each modeling procedure are summarized below.

Atomic coordinates and charges. As a basis for the computations, the crystal structures of BR (PDB codes 1C3W (6) and 2NTU (19)) and ASR (PDB code 1XIO (10)) were used. To generate the initial geometries, the positions of the H atoms were energetically optimized with CHARMM (20) using the CHARMM22 force field. During this procedure, the positions of all non-H atoms were fixed and the standard charge states of all the titratable groups were maintained (i.e., basic and acidic groups were considered to be protonated and deprotonated, respectively). The Schiff base was considered to be protonated. Atomic partial charges of the amino acids and the Schiff base were adopted from the all-atom CHARMM22 (20) parameter set.

Protonation pattern and pK_a . The present computation is based on the electrostatic continuum model created by solving the linear Poisson-Boltzmann equation with the MEAD program (21). To facilitate a direct comparison with previous computational results (e.g., Refs. (22,23)), identical computational conditions and parameters, such as atomic partial charges and dielectric constants, were used. To obtain absolute pK_a values of target sites (e.g., $pK_a(Asp85)$ of BR), we calculated the difference in electrostatic energy between the two protonation states, protonated and deprotonated, in a reference model system using a known experimentally measured pK_a value (e.g., 4.0 for Asp (24)). The difference in the pK_a value of the protein relative to the reference system was added to the known reference pK_a value. The experimentally measured pK_a values employed as references were 7.2 for the Schiff base (25,26), 12.0 for Arg, 4.0 for Asp, 9.5 for Cys, 4.4 for Glu, 10.4 for Lys, 9.6 for Tyr (24), and 7.0 and 6.6 for the Nε and Nδ atoms of His, respectively (27-29). All other titratable sites were fully equilibrated to the protonation state of the target site during the titration. The ensemble of the protonation patterns was sampled by a Monte Carlo (MC) method with Karlsberg (30). The dielectric constants were set to $\varepsilon_p = 4$ inside the protein and $\varepsilon_w =$ 80 for water. All computations were performed at 300 K, pH 7.0, and an ionic strength of 100 mM. The linear Poisson-Boltzmann equation was solved using a three-step grid-focusing procedure at resolutions of 2.5 Å, 1.0 Å, and 0.3 Å. The MC sampling yielded the probabilities [protonated] and [deprotonated] of the

two protonation states of the molecule. The pK_a value was evaluated using the Henderson-Hasselbalch equation. A bias potential was applied to obtain an equal amount of both protonation states ([protonated] = [deprotonated]), yielding the pK_a value as the resulting bias potential.

OM/MM calculations. We employed electrostatic embedding QM/MM scheme and used the Osite (31) program code, as performed in previous studies (32). We employed the restricted DFT method with B3LYP functional and LACVP**+ basis sets. The geometries were refined using a constrained QM/MM optimization, whereby the coordinates of the heavy atoms in the surrounding MM region were fixed to the original X-ray coordinates, whereas those of the H atoms in the MM region were optimized with the OPLS-2005 force field. To investigate the energetics of the entire H-bond network, the QM region was defined as follows (Figure 2): (a) W402 and the residues/groups in the same H-bond network, i.e., W401, W406, Tyr57, Arg82, Asp85, Ser85, Thr89, Tyr185, Asp212, Lys216 (the Schiff base), and retinal for BR; and (b) W402, and the residues/groups in the same H-bond network, i.e., Tyr11, Tyr51, Asp75, Trp76, Thr79, Ser47, the backbone of Phe202, Lys210 (the Schiff base), and retinal for ASR. Other residues that are not in the H-bond network of W402 (e.g., Trp86 in BR is not involved in the same H-bond network, even though the residue corresponds to Trp76 in ASR) were approximated by the MM force field. The resulting OM/MM optimized geometries are listed in Table S1 in the SI. The potential-energy profile of the H bond was obtained in the following manner: first, we optimized the geometry without constraints using QM/MM and used the resulting geometry as the initial geometry for the subsequent steps. Next, we moved the H atom from the H-bond donor atom (O_{donor}) to the acceptor atom (O_{acceptor}) by 0.05 Å, optimized the geometry by constraining the $O_{donor} ext{-H}$ and $O_{acceptor} ext{...} ext{H}$ distances, and then calculated the energy of the resulting geometry. This procedure was repeated until the H atom reached the O_{acceptor} atom. After obtaining the stable geometry of the QM fragment, we calculated the O-D stretching frequencies of W401, W402, and W406 in BR, and of W402 in ASR. The frequency calculations were performed using a numerical differentiation method at the same level as the QM/MM geometry optimization. The calculated frequencies were scaled using a standard factor of 0.9614 for B3LYP (33).

Calculation of the O-D stretching frequencies of D_2O from an empirical equation. The O-D stretching frequencies of D_2O were also evaluated on the basis of the resultant QM/MM optimized geometries by using the correlation of stretching frequency with respect to bond distance of the O_{donor} -

D...O_{acceptor} bond, as previously proposed by Mikenda (34). This correlation was empirically expressed with the O-D stretching frequency, v_{O-D} , and the O_{donor}-O_{acceptor} bond distance, r_{O-O} , by

$$v_{O-D}$$
 [cm⁻¹] = 2727 – $A \exp(-B r_{O-O} [Å]),$ (eq. 1)

where $A = 2.11 \times 10^6$ and B = 3.23 for the O_{water} - D_{water} ... $O_{acceptor}$ bond (34). Note that these parameters were derived from spectroscopic and X-ray diffraction data for 61 different solid deuterates, e.g., $CaSO_4 \cdot 2D_2O$ and $MnCl_2 \cdot 6D_2O$ (34).

To predict the frequencies, eq. 1 requires only the O_{donor} — $O_{acceptor}$ distance, which can be preferentially taken from the QM/MM optimized geometry. Remarkably, the O-D stretching frequencies calculated by a QM/MM numerical differentiation method were reasonably well-reproduced using eq. 1 solely from the optimized O_{donor} — $O_{acceptor}$ distance, particularly for frequencies <2500 cm⁻¹ (Table 2b).

Notably, eq. 1 indicates that the O-D stretching frequency of 2500 cm⁻¹ corresponds to the $O_{acceptor}$ – O_{donor} distance of ~2.8 Å. The deviation from the values calculated with the QM/MM numerical differentiation method appears to be more pronounced for frequencies >2500 cm⁻¹ (Figure 3). Because the original parameters were derived from solid hydrates (34), the larger deviation for weak H bonds at these frequencies may be due to H-bond patterns or mode couplings specific to the protein environments.

RESULTS

H-bond geometries near W402 in BR. The O-D stretching frequencies of D_2O predominantly depend on the H-bond geometry, particularly the H bond donor-acceptor distance $(O_{donor}-O_{acceptor})$ (34). If the assumed H-bond pattern is not correct, the resulting QM/MM optimized geometries significantly deviate from the original crystal structure. Thus, although the QM/MM optimized geometries (especially the heavy atom positions) are not always consistent with the crystal structures, it is necessary to carefully evaluate the H-bond pattern with respect to the atomic coordinates of the crystal structures prior to frequency calculations.

In BR, two H-bond patterns are geometrically possible for the H-bond network of W402 (Figure 4). A model previously presented from FTIR studies (7) suggests that an H atom of W401 has no H-bond acceptor (model 1), while an alternative model suggests that an H atom of W406 has no H-bond acceptor (model 2). The QM/MM optimized geometries of the two models appear to represent the

actual H bond patterns of the crystal structures because of the low RMSD (0.17–0.19 Å, Table 1), implying that the two models probably coexist as tautomers in the actual BR protein environment.

Model 1 resembles the QM/MM geometry reported by Hayashi and Ohmine (8). However, in their QM/MM geometry, the O_{W402} – O_{Asp212} distance was significantly elongated to 4.07 Å and the H bond is absent (8). This varies significantly from the crystal structures and the present QM/MM geometries (O_{W402} – $O_{Asp212} = 2.85$ –3.01 Å, Table 2). In the reported QM/MM geometry, W402 forms a new H bond with another carboxyl O atom of Asp212 (Figure 4). Hence, it is to be considered that the band assignment reported by Hayashi and Ohmine (8) refers to another possible H-bond geometry that differs from the crystal structures.

The QM/MM geometries imply that model 1 can explain the O_{donor}-O_{acceptor} distances in the crystal structure provided by Luecke et al. (PDB code 1C3W) (6) more reasonably than model 2 (Table 1). On the other hand, model 2 appears to explain the O_{donor}-Oacceptor distances in the crystal structure determined by Lanyi et al. (PDB code 2NTU) (19) more reasonably, namely in terms of O_{W402}-O_{Asp85}, O_{W402}- $O_{W401} - O_{Asp85}$, and O_{Arg82} – O_{W406} O_{Asp212} , distances(Table 1). However, the small discrepancies could also be due to being artifacts of the crystallization process. Because the resolutions of the two crystal structures are 1.53 and 1.55 Å (which are essentially the same), both of the H-bond tautomers are likely to contribute to the O-D stretching frequencies measured by FTIR. If both tautomers are present, this might corroborate the rapid H/D exchange observed in this region.

Calculated O-D stretching frequencies in BR. In FTIR studies (7), the three H bonds O_{W401} –H... O_{Asp85} , O_{W402} –H... O_{Asp85} , and O_{W406} –H... O_{Asp212} were classified as being strong H bonds because of the assignment of low frequencies (Figure 4).

a) Model 1. In FTIR studies (7), the lowest frequency of 2171 cm $^{-1}$ was assigned to O_{W402} – $H...O_{Asp85}$. However, in model 1, O_{W402} – $H...O_{Asp85}$ was longer than O_{W401} – $H...O_{Asp85}$ (Table 1), leading to a larger calculated O-D stretching frequency for O_{W402} – $H...O_{Asp85}$ than O_{W401} – $H...O_{Asp85}$ (Table 2a), as predicted from the empirical equation by Mikenda (eq. 1) (34).

In model 1, two mode couplings between O_{W401} – $H...O_{Asp85}$ and O_{W401} – $H...O_{Asp85}$, and between O_{W406} – $H...O_{Asp212}$ and O_{W406} – $H...O_{W401}$ were observed. However, the latter coupling is not consistent with the FTIR assignments. In FTIR studies, H bonds O_{W406} –

 $H...O_{Asp212}$ and O_{W406} – $H...O_{W401}$ were assigned as strong (2292 cm⁻¹) and weak (2599 cm⁻¹), respectively (Table 2a) (7). Because mode couplings are unlikely to occur between a strong and weak H bond, model 1 fails to sufficiently explain the FTIR assignments.

b) Model 2. In contrast to model 1, model 2 can reasonably explain the strong H bond (2292 cm⁻¹) of O_{W406}–H...O_{Asp212} and the weak H bond (2636 cm⁻¹) of O_{W406}–H...O_{W401} that were assigned by FTIR studies (Table 2b). Furthermore, the lowest calculated frequency was observed for O_{W402}–H...O_{Asp85}, which is also consistent with FTIR studies (7). Significantly high correlations of the frequencies predicted from eq. 1 suggest that assignment of the O-D stretching frequencies on the basis of model 2 is also justified by the H-bond geometry. Note that no remarkable mode couplings were observed for model 2 in the QM/MM calculations.

In the following section, further analysis of BR will be discussed on the basis of model 2 using the crystal structure by Luecke et al. (PDB code 1C3W) (6) unless otherwise specified.

H-bond geometries near W402 and O-D stretching frequencies in ASR. In contrast to BR, the ASR crystal structure only possesses a single water molecule, W402, near the Schiff base (Figure 4). The H-bond geometry of the crystal structure was reasonably reproduced by the QM/MM geometry optimization (Table 3). In the QM/MM calculations, the lowest O-D stretching frequency of 2376 cm⁻¹ in ASR, which was found for O_{W402} – $H...O_{Asp75}$ (Table 4), was significantly larger (by ~300 cm⁻¹) than that for the corresponding O_{W402} –H... O_{Asp85} in BR (2078 cm⁻¹, Table 2b). In FTIR studies, the O-D stretching of D₂O was only observed in the >2500 cm⁻¹ region for ASR (11), which was also $\sim 300 \text{ cm}^{-1}$ higher than the lowest frequency of 2171 cm⁻¹ in BR (7). Since the main purpose of the present study is to understand the reason for the significant discrepancy between the lowest frequencies of D₂O (assumed as W402 (11)) of BR and ASR, these computational results are sufficiently accurate for our purpose to describe the chemical properties of W402.

DISCUSSION

Understanding frequencies from energetics.

Significant differences between the H-bond energy profiles of BR and ASR. To identify the origin of the difference in O-D stretching frequencies of W402, we analyzed the potential-energy profiles of H-bonds O_{W402} –H... O_{Asp85} in BR and O_{W402} –H... O_{Asp75} in ASR by altering the H atom position

along the O_{W402} – $O_{Asp85/75}$ bond (Figure 5). In both H bonds, the energy minimum was found at the W402 moiety rather than at the Asp moiety, suggesting that in the ground state, W402 exists as H_2O in the presence of deprotonated Asp in BR and ASR.

On the other hand, the potential-energy profiles of the Asp moiety were significantly different for BR and ASR. For an H atom along the O_{W402}-O_{Asp85/75} bond, the energy of the Asp85 moiety of BR is significantly lower than that of the Asp75 moiety of ASR. Thus, it is more favorable for an H atom of O_{W402}–H to be at the Asp moiety in BR than in ASR, indicating that $pK_a(Asp)$ is significantly higher for BR than ASR (arrow 1 in Figure 5). As clearly shown in Figure 5, the lower energy near the Asp85 moiety in BR leads to greater broadening of the potential-well width near W402 toward Asp85, which corresponds to an elongation of O_{W402}–H. Thus, the H atom of O_{W402}-H migrates toward the H-bond acceptor Asp more in BR than in ASR (arrow 2 in Figure 5). It should be noted that the longer O_{W402}–H corresponds to a smaller O-D stretching frequency. It is also obvious from the width of the entire well of the potential-energy profiles that the O_{W402}-O_{Asp85} length in BR (~2.6 Å, Table 1), which is shorter than the O_{W402} – O_{Asp75} length in ASR (~2.7 Å, Table 3), is a result of the smaller pK_a difference between W402 and the Asp residue in BR relative to the corresponding difference in ASR (arrow 3 in Figure 5).

Thus, from the potential-energy profiles (Figure 5), the difference of $\sim 300~\text{cm}^{-1}$ in the O-D stretching frequency is mainly due to the difference between the p K_a values for the Asp residues in BR and ASR.

$pK_a(Asp85)$ in BR and $pK_a(Asp75)$ in ASR

Factors that differentiate $pK_a(Asp)$ of BR and ASR. By solving the linear Poisson–Boltzmann equation, we calculated $pK_a(Asp85)$ in BR and $pK_a(Asp75)$ in ASR to be 1.5 and –5.1, respectively, in the presence of the protonated Schiff base (Table 5). The calculated $pK_a(Asp85)$ of 1.5 is consistent with an experimentally measured $pK_a(Asp85)$ of 2.2–2.6 in the presence of the protonated Schiff base (2,35). Although $pK_a(Asp75)$ is not known, the significantly low $pK_a(Asp85)$ in BR relative to $pK_a(Asp75)$ in ASR is consistent with the QM/MM-calculated potential-energy profiles (Figure 5). The significantly low $pK_a(Asp75)$ in ASR appears to be consistent with the fact that Asp75 remains deprotonated throughout the photocycle (36).

a) Asp212 in BR. The most significant difference observed between $pK_a(Asp85)$ and $pK_a(Asp75)$ was induced by the substitution of Asp212 in BR for Pro206 in ASR (Figure 6), which is responsible for the pK_a difference of ~6 (Table 5). Because Asp212 is an H-bond acceptor for O_{W402} —H and is thus far the most proximal negative charge to Asp85 (5.1 Å), Asp212 upshifts $pK_a(Asp85)$ by the largest amount among all residues in BR (Table 5).

It is widely known that both Asp85 and Asp212 are ionized in the ground state (2,37,38). However, the deprotonated state of Asp212 is more energetically stable than the deprotonated state of Asp85, as seen in the lower $pK_a(Asp212)$ of -2.0 (Table 6); this is consistent with previous pK_a assignments of <1 for Asp212 (39,40) or $pK_a(Asp85) \le pK_a(Asp212)$ (41,42). The deprotonated state of Asp212 was facilitated by from H-bond donations Tyr57 (decreasing $pK_a(Asp212)$ by 3.4) and Tyr185 (decreasing $pK_a(Asp212)$ by 3.3). In addition, Arg82 is closer to Asp212 (3.8 Å) than Asp85 (6.6 Å) (Figure 6), stabilizing the ionized state of Asp212 more effectively than Asp85 (increasing pK_a by ~ 6 , Table 6).

Although the most significant difference between $pK_a(Asp85)$ and $pK_a(Asp75)$ induced by the substitution of Asp212 in BR for Pro206 in ASR is remarkable, this is not sufficient to entirely explain the difference between $pK_a(Asp85)$ and $pK_a(Asp75)$ (see below), which is in agreement with mutational studies of ASR mutant P206D (43).

b) Ser47 in ASR. Ser47 donates an H bond to Asp75 in ASR, decreasing $pK_a(Asp75)$ by 3.2, whereas the corresponding residue in BR is the non-polar Ala53 (Table 5). This was responsible for a pK_a difference of 2.7 between Asp85 and Asp75 (Table 5).

c) Glu194-Glu204 pair in BR. Another remarkable $pK_a(Asp85)$ upshift in BR originates from the Glu194-Glu204 moiety near the terminal region of the proton transfer pathway (2,3) (Figure 6). It is likely that the two acidic residues share a proton. The corresponding acidic residue pair is absent in ASR, thereby contributing slightly to the larger $pK_a(Asp)$ in BR relative to that in ASR (~0.5 pK_a units, Table 5). In the present study, Glu204 is protonated and Glu194 is deprotonated when Ser193 donates an H bond to Glu194, which is consistent with the previously reported $pK_a(Glu204)$ of ~9 (35,44,45).

Nevertheless, the geometry of the crystal structure also suggests that protonated Glu194 and deprotonated Glu204 are almost equally possible when Ser193 donates an H bond to Glu204, as suggested by Gerwert et al. (46). Thus, it would be

more plausible to consider the two acidic residues as a pair that possesses $\sim 1~\text{H}^+$ as concluded in previous QM/MM simulations (47). This resembles a pair of acidic residues, Glu-L212 and Asp-L213, in the proton transfer pathway of photosynthetic reaction centers, which has been interpreted as sharing $\sim 1~\text{H}^+$ (48-50). Deprotonated Glu194 contributes to an increase in p K_a (Asp85) of ~ 1 (Table 5); this therefore corroborates the linkage between p K_a (Asp85) and p K_a (Glu204) previously reported in mutant BR studies (35) if we consider Glu194-Glu204 as a pair of acidic residues sharing $\sim 1~\text{H}^+$.

d) Orientation of Arg82 in BR and Arg72 ASR. Arg82 was found to contribute to an increase in $pK_a(Asp85)$ of 3.1 in BR (Table 5), which is in agreement with a similar increase (~4.5) reported for BR mutants R82Q and R82A (51,52). Irrespective of the conservation of Arg82/Arg72 in BR/ASR, their $pK_a(Asp85)$ influence on and $pK_a(Asp75)$ significantly differ by ~2 because the side chain of Arg72 is oriented away from the Schiff base and toward the extracellular side in ASR (O_{Asp75} - N_{Arg72} = 9.6 Å) (10), which is the opposite of the orientation of Arg82 in BR $(O_{Asp85}-N_{Arg82} = 6.6 \text{ Å})$ (6) (Figure 6). Thus, the differing orientations of the Arg side chains decrease the $pK_a(Asp)$ difference between BR and ASR (Table 6).

In summary, the significantly large $pK_a(Asp85)$ in BR relative to $pK_a(Asp75)$ in ASR is largely due to the presence of Asp212 near Asp85 in BR and the absence of the corresponding acidic residue in ASR. In addition, the donation of an H-bond from Ser47 to Asp75 lowers the $pK_a(Asp75)$ in ASR by facilitating deprotonation. The corresponding polar residue is absent in BR; this upshifts $pK_a(Asp85)$ in BR relative to $pK_a(Asp75)$ in ASR. The Glu194-Glu204 pair is located near the terminal region of the proton transfer pathway in BR, whereas only a single acidic residue is located near the corresponding position in ASR. This also upshifts $pK_a(Asp85)$ in BR relative to $pK_a(Asp75)$ in ASR.

CONCLUSIONS

QM/MM calculations revealed that the heavy atom positions in the BR crystal structures could be explained by one of two models. Model 2 can reasonably explain the strong H bond (2292 cm⁻¹) of O_{W406} –H... O_{Asp212} and the weak H bond (2636 cm⁻¹) of O_{W406} –H... O_{W401} , as previously assigned by FTIR (Table 2b). In model 2, the lowest calculated frequency was observed at O_{W402} –H... O_{Asp85} , which is also consistent with FTIR studies. Essentially the same frequencies were also predicted solely from the

 O_{donor} — $O_{acceptor}$ distances using eq. 1 proposed by Mikenda (34). This suggests that eq. 1 can be used to support the frequency assignments made from spectroscopic studies. In ASR, the lowest calculated O-D stretching frequency is observed at O_{W402} — $H...O_{Asp75}$, which is ~300 cm⁻¹ higher than that of BR (Tables 2b and 4), as previously reported in FTIR studies of BR (7) and ASR (11).

The potential-energy profiles of two H bonds, O_{W402} –H... O_{Asp85} in BR and O_{W402} –H... O_{Asp75} in ASR, demonstrate that the significant p K_a difference between Asp85 in BR and Asp75 in ASR is ultimately responsible for the difference of ~300 cm⁻¹ in the O-D

stretching frequency of W402 between BR and ASR (Figure 5). Electrostatic calculations resulted in a p K_a (Asp85) of 1.5 in BR and a p K_a (Asp75) of -5.1 in ASR (Table 5). The p K_a difference is mainly due to differences in the influences of Ala53, Arg82, Glu194, and Asp212 on Asp85 in BR, and the corresponding residues Ser47, Arg72, Ser188, and Pro206 on Asp75 in ASR, indicating the presence of long-range electrostatic interactions along the proton transfer pathways (53). A strongly H-bonded water near the Schiff base, which results from electrostatic interactions between Asp85 and these acidic residues in BR, could play a key role in proton transfer.

REFERENCES

- 1. Lanyi, J. K. (1998) Understanding structure and function in the light-driven proton pump bacteriorhodopsin. *J Struct Biol* **124**, 164-178
- 2. Balashov, S. P. (2000) Protonation reactions and their coupling in bacteriorhodopsin. *Biochim Biophys Acta* **1460**, 75-94
- 3. Kandori, H. (2000) Role of internal water molecules in bacteriorhodopsin. *Biochim Biophys Acta* **1460**, 177-191
- 4. Maeda, A., Sasaki, J., Yamazaki, Y., Needleman, R., and Lanyi, J. K. (1994) Interaction of aspartate-85 with a water molecule and the protonated Schiff base in the L intermediate of bacteriorhodopsin: a Fourier-transform infrared spectroscopic study. *Biochemistry* 33, 1713-1717
- 5. Kandori, H., Yamazaki, Y., Sasaki, J., Needleman, R., Lanyi, J. K., and Maeda, A. (1995) Water-mediated proton transfer in proteins: an FTIR study of bacteriorhodopsin. *J Am Chem Soc* **117**, 2118-2119
- 6. Luecke, H., Schobert, B., Richter, H. T., Cartailler, J. P., and Lanyi, J. K. (1999) Structure of bacteriorhodopsin at 1.55 A resolution. *J Mol Biol* **291**, 899-911
- 7. Shibata, M., and Kandori, H. (2005) FTIR studies of internal water molecules in the Schiff base region of bacteriorhodopsin. *Biochemistry* **44**, 7406-7413
- 8. Hayashi, S., and Ohmine, I. (2000) Proton transfer in bacteriorhodopsin: structure, excitation, IR spectra, and potential energy surface analyses by an ab initio QM/MM method. *J Phys Chem B* **104**, 10678-10691
- 9. Hayashi, S., Tajkhorshid, E., Kandori, H., and Schulten, K. (2004) Role of hydrogen-bond network in energy storage of bacteriorhodopsin's light-driven proton pump revealed by ab initio normal-mode analysis. *J Am Chem Soc* **126**, 10516-10517
- 10. Vogeley, L., Sineshchekov, O. A., Trivedi, V. D., Sasaki, J., Spudich, J. L., and Luecke, H. (2004) Anabaena sensory rhodopsin: a photochromic color sensor at 2.0 A. *Science* **306**, 1390-1393
- 11. Furutani, Y., Kawanabe, A., Jung, K. H., and Kandori, H. (2005) FTIR spectroscopy of the all-trans form of Anabaena sensory rhodopsin at 77 K: hydrogen bond of a water between the Schiff base and Asp75. *Biochemistry* **44**, 12287-12296
- 12. Kandori, H. (2010) in Hydrogen bonds of protein-bound water molecules in rhodopsins, In Hydrogen bonding and transfer in the excited state (Han, K.-L. and Zhao, G.-J., Eds). pp 377-391, John-Wiley & Sons Ltd., West Sussex
- 13. Frey, P. A. (2006) in Isotope Effects in Chemistry and Biology (Kohen, A. and Limbach, H.-H., Eds.), pp 975-993, CRC press, Boca Raton, FL
- 14. Cleland, W. W., and Kreevoy, M. M. (1994) Low-barrier hydrogen bonds and enzymic catalysis. *Science* **264**, 1887-1890

- 15. Frey, P. A., Whitt, S. A., and Tobin, J. B. (1994) A low-barrier hydrogen bond in the catalytic triad of serine proteases. *Science* **264**, 1927-1930
- 16. Perrin, C. L., and Nielson, J. B. (1997) "Strong" hydrogen bonds in chemistry and biology. *Annu Rev Phys Chem* **48**, 511-544
- 17. Schutz, C. N., and Warshel, A. (2004) The low barrier hydrogen bond (LBHB) proposal revisited: the case of the Asp... His pair in serine proteases. *Proteins* **55**, 711-723
- 18. Saito, K., and Ishikita, H. (2012) Energetics of short hydrogen bonds in photoactive yellow protein. *Proc Natl Acad Sci U S A* **109**, 167-172
- 19. Lanyi, J. K., and Schobert, B. (2007) Structural changes in the L photointermediate of bacteriorhodopsin. *J Mol Biol* **365**, 1379-1392
- 20. Brooks, B. R., Bruccoleri, R. E., Olafson, B. D., States, D. J., Swaminathan, S., and Karplus, M. (1983) CHARMM: a program for macromolecular energy minimization and dynamics calculations. *J. Comput. Chem.* **4**, 187-217
- 21. Bashford, D., and Karplus, M. (1990) p K_a 's of ionizable groups in proteins: atomic detail from a continuum electrostatic model. *Biochemistry* **29**, 10219-10225
- 22. Ishikita, H., and Knapp, E.-W. (2007) Protonation states of ammonia/ammonium in the hydrophobic pore of ammonia transporter protein AmtB. *J. Am. Chem. Soc.* **129**, 1210-1215
- 23. Ishikita, H. (2010) Origin of the p*K*_a shift of the catalytic lysine in acetoacetate decarboxylase. *FEBS Lett* **584**, 3464-3468
- 24. Nozaki, Y., and Tanford, C. (1967) Acid-base titrations in concentrated guanidine hydrochloride. Dissociation constants of the guamidinium ion and of some amino acids. *J Am Chem Soc* **89**, 736-742
- 25. Bassov, T., and Sheves, M. (1986) Alteration of pK_a of the bacteriorhodopsin protonated Schiff base. A study with model compounds. *Biochemistry* **25**, 5249-5258
- 26. Rousso, I., Friedman, N., Sheves, M., and Ottolenghi, M. (1995) pK_a of the protonated Schiff base and aspartic 85 in the bacteriorhodopsin binding site is controlled by a specific geometry between the two residues. *Biochemistry* **34**, 12059-12065
- 27. Tanokura, M. (1983) 1H-NMR study on the tautomerism of the imidazole ring of histidine residues. I. Microscopic pK values and molar ratios of tautomers in histidine-containing peptides. *Biochim Biophys Acta* **742**, 576-585
- 28. Tanokura, M. (1983) 1H-NMR study on the tautomerism of the imidazole ring of histidine residues. II. Microenvironments of histidine-12 and histidine-119 of bovine pancreatic ribonuclease A. *Biochim Biophys Acta* **742**, 586-596
- 29. Tanokura, M. (1983) 1H nuclear magnetic resonance titration curves and microenvironments of aromatic residues in bovine pancreatic ribonuclease A. *J Biochem* **94**, 51-62
- 30. Rabenstein, B., and Knapp, E. W. (2001) Calculated pH-dependent population and protonation of carbon-monoxy-myoglobin conformers. *Biophys J* **80**, 1141-1150
- 31. OSite, version 5.6, Schrödinger, LLC, New York, NY, 2010.
- 32. Saito, K., Ishida, T., Sugiura, M., Kawakami, K., Umena, Y., Kamiya, N., Shen, J.-R., and Ishikita, H. (2011) Distribution of the cationic state over the chlorophyll pair of photosystem II reaction center. *J. Am. Chem. Soc.* **133**, 14379-14388
- 33. Scott, A. P., and Radom, L. (1996) Harmonic vibrational frequencies: an evaluation of Hartree–Fock, Møller–Plesset, quadratic configuration interaction, density functional theory, and semiempirical scale factors. *J Phys Chem* **100**, 16502-16513
- 34. Mikenda, W. (1986) Stretching frequency versus bond distance correlation of O-D(H)...Y (Y = N, O, S, Se, Cl, Br, I) hydrogen bonds in solid hydrates. *J Mol Struct* **147**, 1-15
- 35. Richter, H. T., Brown, L. S., Needleman, R., and Lanyi, J. K. (1996) A linkage of the pK_a 's of asp-85 and glu-204 forms part of the reprotonation switch of bacteriorhodopsin. *Biochemistry* **35**, 4054-4062
- 36. Bergo, V. B., Ntefidou, M., Trivedi, V. D., Amsden, J. J., Kralj, J. M., Rothschild, K. J., and Spudich, J. L. (2006) Conformational changes in the photocycle of Anabaena sensory rhodopsin: absence of the Schiff base counterion protonation signal. *J Biol Chem* **281**, 15208-15214

- 37. Song, Y., Mao, J., and Gunner, M. R. (2003) Calculation of proton transfers in Bacteriorhodopsin bR and M intermediates. *Biochemistry* **42**, 9875-9888
- 38. Bombarda, E., Becker, T., and Ullmann, G. M. (2006) Influence of the membrane potential on the protonation of bacteriorhodopsin: insights from electrostatic calculations into the regulation of proton pumping. *J Am Chem Soc* **128**, 12129-12139
- 39. Metz, G., Siebert, F., and Engelhard, M. (1992) Asp85 is the only internal aspartic acid that gets protonated in the M intermediate and the purple-to-blue transition of bacteriorhodopsin. A solid-state 13C CP-MAS NMR investigation. *FEBS Lett* **303**, 237-241
- 40. Balashov, S. P., Govindjee, R., Imasheva, E. S., Misra, S., Ebrey, T. G., Feng, Y., Crouch, R. K., and Menick, D. R. (1995) The two pKa's of aspartate-85 and control of thermal isomerization and proton release in the arginine-82 to lysine mutant of bacteriorhodopsin. *Biochemistry* **34**, 8820-8834
- 41. Bashford, D., and Gerwert, K. (1992) Electrostatic calculations of the pKa values of ionizable groups in bacteriorhodopsin. *J Mol Biol* **224**, 473-486
- 42. Sampogna, R. V., and Honig, B. (1994) Environmental effects on the protonation states of active site residues in bacteriorhodopsin. *Biophys J* **66**, 1341-1352
- 43. Choi, A. R., Kim, S. Y., Yoon, S. R., Bae, K., and Jung, K. H. (2007) Substitution of Pro206 and Ser86 residues in the retinal binding pocket of Anabaena sensory rhodopsin is not sufficient for proton pumping function. *J Microbiol Biotechnol* 17, 138-145
- 44. Balashov, S. P., Imasheva, E. S., Govindjee, R., and Ebrey, T. G. (1996) Titration of aspartate-85 in bacteriorhodopsin: what it says about chromophore isomerization and proton release. *Biophys J* **70**, 473-481
- 45. Dioumaev, A. K., Richter, H. T., Brown, L. S., Tanio, M., Tuzi, S., Saito, H., Kimura, Y., Needleman, R., and Lanyi, J. K. (1998) Existence of a proton transfer chain in bacteriorhodopsin: participation of Glu-194 in the release of protons to the extracellular surface. *Biochemistry* 37, 2496-2506
- 46. Rammelsberg, R., Huhn, G., Lubben, M., and Gerwert, K. (1998) Bacteriorhodopsin's intramolecular proton-release pathway consists of a hydrogen-bonded network. *Biochemistry* **37**, 5001-5009
- 47. Goyal, P., Ghosh, N., Phatak, P., Clemens, M., Gaus, M., Elstner, M., and Cui, Q. (2011) Proton storage site in bacteriorhodopsin: new insights from quantum mechanics/molecular mechanics simulations of microscopic pK_a and infrared spectra. *J Am Chem Soc* **133**, 14981-14997
- 48. Alexov, E. G., and Gunner, M. R. (1999) Calculated protein and proton motions coupled to electron transfer: electron transfer from Q_A^- to Q_B in bacterial photosynthetic reaction centers. *Biochemistry* **38**, 8253-8270
- 49. Gerencser, L., and Maroti, P. (2001) Retardation of proton transfer caused by binding of the transition metal ion to the bacterial reaction center is due to pK_a shifts of key protonatable residues. *Biochemistry* **40**, 1850-1860
- 50. Ishikita, H., Morra, G., and Knapp, E.-W. (2003) Redox potential of quinones in photosynthetic reaction centers from *Rhodobacter sphaeroides*: dependence on protonation of Glu-L212 and Asp-L213. *Biochemistry* **42**, 3882-3892
- 51. Otto, H., Marti, T., Holz, M., Mogi, T., Stern, L. J., Engel, F., Khorana, H. G., and Heyn, M. P. (1990) Substitution of amino acids Asp-85, Asp-212, and Arg-82 in bacteriorhodopsin affects the proton release phase of the pump and the pK of the Schiff base. *Proc Natl Acad Sci U S A* **87**, 1018-1022
- 52. Brown, L. S., Bonet, L., Needleman, R., and Lanyi, J. K. (1993) Estimated acid dissociation constants of the Schiff base, Asp-85, and Arg-82 during the bacteriorhodopsin photocycle. *Biophys J* **65**, 124-130
- 53. Calimet, N., and Ullmann, G. M. (2004) The influence of a transmembrane pH gradient on protonation probabilities of bacteriorhodopsin: the structural basis of the back-pressure effect. *J Mol Biol* **339**, 571-589

FOOTNOTES

This research was supported by the JST PRESTO program (H.I.), Grant-in-Aid for Scientific Research from the Ministry of Education, Culture, Sports, Science and Technology (MEXT) of Japan (22740276 to K.S. and 20108014 to H.K.), Special Coordination Fund for Promoting Science and Technology of MEXT (H.I.), Takeda Science Foundation (H.I.), Kyoto University Step-up Grant-in-Aid for Young Scientists (H.I.), and Grant for Basic Science Research Projects from The Sumitomo Foundation (H.I.).

FIGURE LEGENDS

Figure 1. Overview of the BR (PDB code 1C3W) and ASR (PDB code 1XIO) structures. Red spheres indicate O atoms of water molecules. Yellow arrows indicate proton transfer pathways.

Figure 2. Residues and groups included in the QM regions for (a) BR and (b) ASR. Dotted lines indicate hydrogen bonds. (c) Geometry of moieties near Lys210 in ASR before (magenta) and after (yellow) the QM/MM optimization.

Figure 3. The geometric correlation of the O-D stretching frequencies with the $O_{acceptor} \cdots H-O_{donor}$ bond of H_2O (correlation with the $O_{donor}-O_{acceptor}$ distance). Each \Box represents the $O_{donor}-O_{acceptor}$ distance of the QM/MM optimized geometries (Tables 1 and 3) and the O-D stretching frequency calculated by a QM/MM numerical differentiation method (Tables 2 and 4). The solid curve was determined using an empirical equation (eq. 1) proposed by Mikenda (34).

Figure 4. H-bond geometries of water molecules near the Schiff base in BR and ASR. Pink arrows in model 1 indicate mode couplings of the two O-D bonds. Red dotted lines indicate "strong H bonds" reported in FTIR studies (7).

Figure 5. Potential-energy profiles along the proton transfer coordinates of O_{W402} – $H...O_{Asp85}$ in BR (black solid line) and O_{W402} – $H...O_{Asp75}$ in ASR (blue solid line). ΔE describes the difference in energy relative to the energy minimum. Boxed arrows indicate the shifts in the potential energy curve accompanied by the p K_a (Asp) change from ASR to BR. The marginal energy drop near 1.74 Å in O_{W402} –H of ASR was due to an alteration of the H-bond pattern; below 1.74 Å, Ser47 donates an H bond to the carboxyl O atom of Asp75, which is simultaneously H-bonded by W402. At 1.74 Å, the hydroxyl H atom is oriented toward another carboxyl O atom of Asp75 due to the unusual proximity of the H atom of W402 to Asp75.

Figure 6. Residues that contribute to differences between $pK_a(Asp)$ of (a) BR and (b) ASR.

Table 1. Comparison of the H-bond geometries near W402 in the BR crystal structures (PDB ID codes 1C3W at 1.55 Å resolution and 2NTU at 1.53 Å resolution), in the QM/MM geometry by Hayashi and Ohmine (8), and in the present study (models 1 and 2, Figure 4). Distances in Å, angles in °. n.d. = not determined. An underlined number indicates a significant deviation from the distance in the crystal structure. See Table S1 in the SI for the atomic coordinates of the QM/MM geometries.

	1C3W				2NTU		
Donoracceptor		QM/MM Hayashi & Ohmine ^a (model 1)		QM/MM (model 2)		QM/MM (model 1)	QM/MM (model 2)
W402Asp85	2.63	2.58	2.61	2.57	2.52	2.63	2.59
W402Asp212	2.85	<u>4.07</u>	2.92	2.99	3.01	2.90	2.95
W401Asp85	2.59	2.63	2.51	2.64	2.69	2.59	2.68
(OH without acceptor) b		(W401)	(W401)	(W406)		(W401)	(W406)
W406W401	2.75	2.77	2.76	3.06	2.69	2.78	3.07
W406Asp212	2.75	2.78	2.64	2.64	2.81	2.73	2.65
Lys216W402	2.87	2.63	2.78	2.76	2.94	2.80	2.77
Arg82W406	2.49	2.72	2.74	2.87	2.87	2.73	2.86
RMSD	n. d.	n. d.	0.19	0.18	n. d.	0.17	0.18
$angle(N_{Lys216}O_{W402}O_{Asp85})$	106	n. d.	108	108	111	111	111

^a See Ref. (8).

^b A water molecule in which one of the two O-H bonds does not possess the H-bond acceptor molecule; W401 in model 1 and W402 in model 2 (Figure 4).

Table 2. O–D stretching frequencies (in cm⁻¹) of the water molecules near W402 in BR. Connecting lines indicate the presence of coupling between the two modes. n.d. = not determined. R^2 = correlation coefficient.

(a) Model 1.

		1C3W	1C3W	1C3W	2NTU	2NTU
	FTIR-1 ^a	Hayashi & Ohmine ^b	QM/MM (model 1)	empirical eq. ^c (model 1)	QM/MM (model 1)	empirical eq. ^c (model 1)
W402Asp85	2171 (lowest)	2257	2221	2258	2247	2293
W402Asp212	2636	2618	2663	2564	2646	2548
W401Asp85	2323	2320	2133 J (lowest)	2173 (lowest)	2169 J (lowest)	2229 (lowest)
W401(n.d.)	2690	2721	2703	2727	2704	2727
W406W401	2599	2601	2515	2454	2514	2463
W406Asp212	2292	2468	2434	2375	2437	2418
R^2 to FTIR		0.89	0.75	0.75	0.76	0.71

^a See Ref. (7).

^b See Ref. (8).

^c See Ref. (34).

Table 2.(b) Model 2. The QM/MM results of model 2 on the basis of the 1C3W structure primarily discussed in the present study.

		1C3W	1C3W	2NTU	2NTU
	FTIR-2 a	QM/MM (model 2)	empirical eq. ^b (model 2)	QM/MM (model 2)	empirical eq. ^b (model 2)
W402Asp85	2171 (lowest)	2078 (lowest)	2201 (lowest)	2120 (lowest)	2240 (lowest)
W402Asp212	<u>2599</u>	2690	2592	2670	2572
W401Asp85	2323	2400	2311	2411	2359
W406(n.d.)	2690	2678	2727	2689	2727
W406W401	<u>2636</u>	2658	2620	2656	2622
W406Asp212	2292	2312	2303	2302	2328
p ² 4 POVDS		0.046	0.000	0.00	0.000
R ² to FTIR ^c		0.94 ^c	0.99°	0.96°	0.98°

^a See Ref. (7).

^b See Ref. (34).

^c Relative to "FTIR-2."

Table 3. H-bond geometries near W402 in the ASR crystal structure (PDB ID code 1XIO at 2.00 Å resolution) and the QM/MM geometry. n.d. = not determined. See Table S1 in the SI for the atomic coordinates of the QM/MM geometries. Distances in Å, angles in °.

	1XIO	QM/MM
Donoracceptor		Deprotonated Asp75
W402Asp75	2.74	2.71
W402Tyr51	2.92	2.97
Trp76W402	2.94	2.84
Lys210W402	3.02	3.00
$RMSD^a$	n. d.	0.17
$angle(N_{Lys210}O_{W402}O_{Asp75})$	83	92

^a Excluding Lys210 due to obvious displacement of the sidechain carbon atoms from the original crystal structure (Figure 2c).

Table 4. O–D stretching frequencies (in cm⁻¹) of W402 in ASR.

	deprotonated Asp75					
	FTIR ^a	QM/MM	Empirical eq.			
W402Asp75	>2500	2376	2392			
W402Tyr51	>2500	2573	2583			

^a See Ref. (11).

Table 5. Contribution of residues to the pK_a shifts of Asp85 in BR and Asp75 in ASR (relative to the bulk solvent) in the presence of the protonated Schiff base (in pK_a units). For clarity, residue pairs that are responsible for differences of <0.7 between pK_a (Asp85) and pK_a (Asp75) are not listed, except for the Glu204/Asp198 pair.

side = sidechain, b.b. = backbone.

BR	pK _a (A	sp85) :	= 1.5	ASR	pK _a (Asp75) = -5.1	difference
(1C3W)	side	b.b.	total	(1XIO)	side	b.b.	total	total
Ile52	-0.1	0.0	-0.1	Trp46	-0.6	-0.3	-0.9	0.8
Ala53	0.0	-0.5	-0.5	Ser47	-2.6	-0.6	-3.2	2.7
Tyr57	-0.5	-0.4	-0.9	Tyr51	0.2	-0.4	-0.2	-0.7
Arg82	-3.9	0.8	-3.1	Arg72	-2.1	0.9	-1.2	-1.9
Glu194	1.2	-0.1	1.1	Ser188	0.0	0.0	0.0	1.1
Glu204	0.0	0.1	0.1	Asp198	0.6	0.0	0.6	-0.5
Asp212	6.3	-0.1	6.2	Pro206			-0.1	6.3
Schiff			-9.3	Schiff			-10.3	1.0

Table 6. Key residues that alter the pK_a difference between Asp85 and Asp212 in the presence of the protonated Schiff base (in pK_a units).

side = side chain, b.b. = backbone, n.d. = not determined.

pK _a (Asp85) =	= 1.5	pK _a (Asp212	(2) = -2.0
side	b.b.	total	side	b.b.	total
-0.4	-0.8	-1.2	-0.1	-0.3	-0.3
-0.5	-0.3	-0.9	-3.2	-0.2	-3.4
0.0	0.7	0.8	0.0	0.1	0.1
-3.9	0.8	-3.1	-6.4	0.1	-6.3
n.d.	n.d.	n.d.	6.4	-0.4	6.0
-0.2	-0.6	-0.8	-0.8	-0.3	-1.0
-2.0	-0.9	-2.9	0.1	-0.2	-0.1
-0.3	-0.1	-0.4	-3.2	-0.1	-3.3
1.2	-0.1	1.2	1.8	-0.1	1.7
0.0	0.2	0.2	0.0	1.8	1.8
6.3	-0.1	6.2	n.d.	n.d.	n.d.
-9.5	0.2	-9.3	-9.3	0.6	-8.7
	side -0.4 -0.5 0.0 -3.9 n.d0.2 -2.0 -0.3 1.2 0.0 6.3	side b.b. -0.4 -0.8 -0.5 -0.3 0.0 0.7 -3.9 0.8 n.d. n.d. -0.2 -0.6 -2.0 -0.9 -0.3 -0.1 1.2 -0.1 0.0 0.2 6.3 -0.1	-0.4 -0.8 -1.2 -0.5 -0.3 -0.9 0.0 0.7 0.8 -3.9 0.8 -3.1 n.d. n.d. n.d0.2 -0.6 -0.8 -2.0 -0.9 -2.9 -0.3 -0.1 -0.4 1.2 -0.1 1.2 0.0 0.2 0.2 6.3 -0.1 6.2	side b.b. total side -0.4 -0.8 -1.2 -0.1 -0.5 -0.3 -0.9 -3.2 0.0 0.7 0.8 0.0 -3.9 0.8 -3.1 -6.4 n.d. n.d. 6.4 -0.2 -0.6 -0.8 -0.8 -2.0 -0.9 -2.9 0.1 -0.3 -0.1 -0.4 -3.2 1.2 -0.1 1.2 1.8 0.0 0.2 0.2 0.0 6.3 -0.1 6.2 n.d.	side b.b. total side b.b. -0.4 -0.8 -1.2 -0.1 -0.3 -0.5 -0.3 -0.9 -3.2 -0.2 0.0 0.7 0.8 0.0 0.1 -3.9 0.8 -3.1 -6.4 0.1 n.d. n.d. 6.4 -0.4 -0.2 -0.6 -0.8 -0.8 -0.3 -2.0 -0.9 -2.9 0.1 -0.2 -0.3 -0.1 -0.4 -3.2 -0.1 1.2 -0.1 1.2 1.8 -0.1 0.0 0.2 0.2 0.0 1.8 6.3 -0.1 6.2 n.d. n.d.

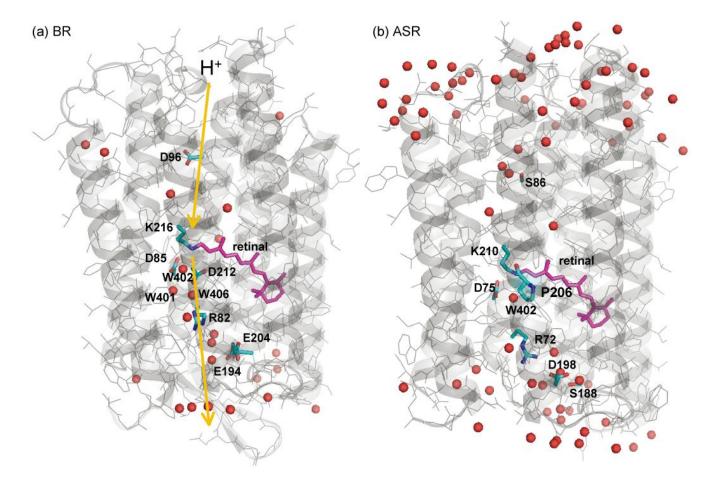


Figure 1.

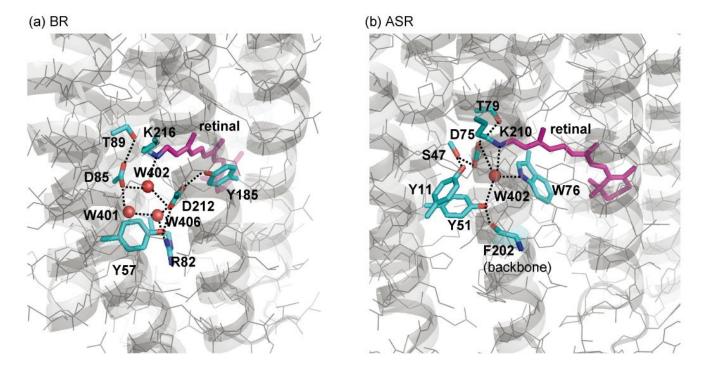


Figure 2 (a, b).

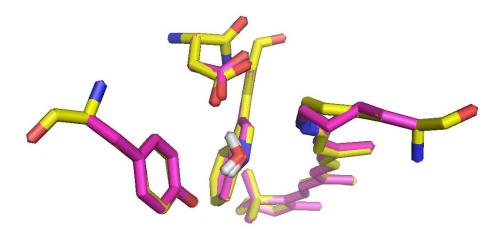


Figure 2 (c).

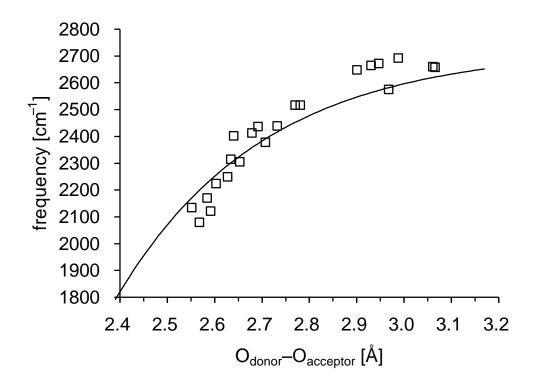


Figure 3.

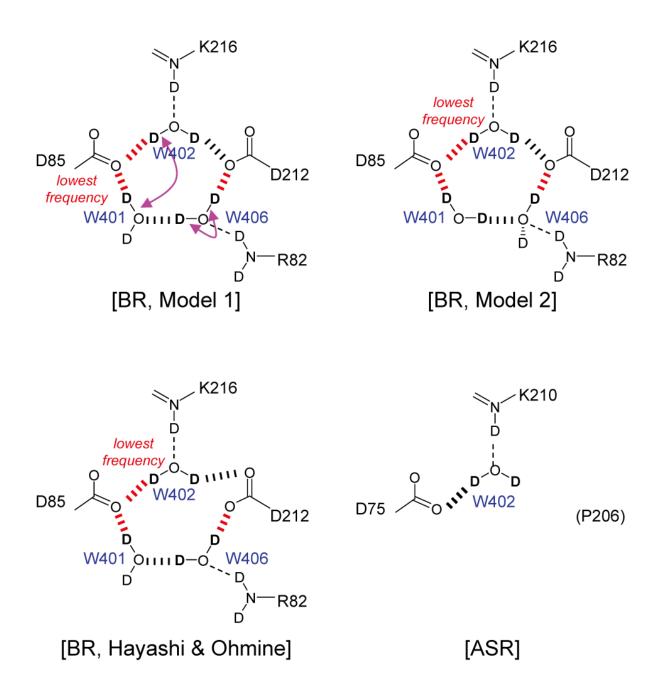


Figure 4.

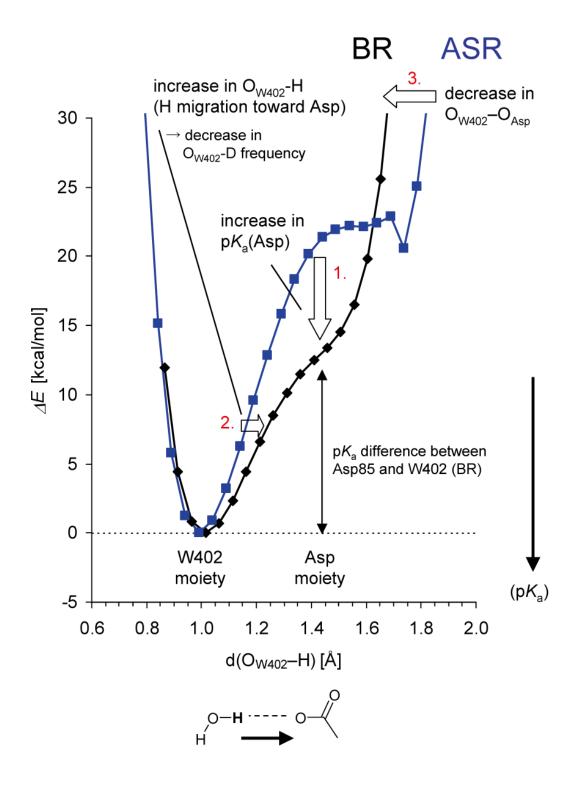


Figure 5.

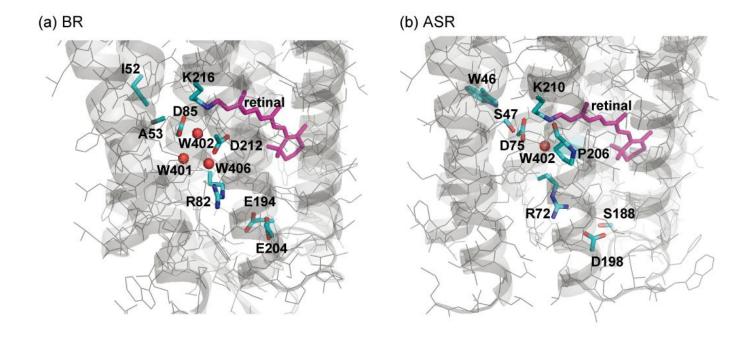


Figure 6.

Factors that differentiate the H-bond strengths of water near the Schiff bases

in bacteriorhodopsin and Anabaena sensory rhodopsin

Keisuke Saito, Hideki Kandori, and Hiroshi Ishikita*

Table S1. Atomic coordinates of the QM/MM optimized geometry for BR [W401, W402, W406, Tyr57, Arg82, Asp85, Thr89, Tyr185, Asp212, Lys216 (the Shiff base) and the retinal].

(a) 1C3W (model 1)

residue	atom	X	y	Z	
ARG A			14.733	36.929	-2.302
ARG A			15.162	37.353	-1.391
ARG A			14.176	37.735	-2.799
ARG A			15.861	36.483	-3.21
ARG A			15.528	36.352	-4.243
ARG A			16.291	35.541	-2.87
ARG A			16.925	37.572	-3.181
ARG A			17.371	37.628	-2.187
ARG A			16.494	38.545	-3.42
ARG A			17.958	37.321	-4 .161
ARG A			17.955	37.866	-5.04
ARG A			18.912	36.413	-4.065
ARG A			19.066	35.659	-2.979
ARG A			18.435	35.684	-2.155
ARG A			19.894	35.071	-2.935
ARG A			19.727	36.23	-5.134
ARG A			19.679	36.905	-5.893
ARG A			20.569	35.69	-5.009
ASP A			13.183	33.56	1.776
ASP A			13.478	34.483	1.278
ASP A			13.239	32.845	0.967
ASP A			14.378	33.28	2.695
ASP A			14.364	33.431	3.929
ASP A			15.428	32.917	2.042
ASP A			20.879	34.919	2.914
ASP A			20.234	34.146	3.344
ASP A			21.559	34.43	2.212
ASP A			19.886	35.803	2.158
ASP A			19.395	36.831	2.662
ASP A			19.491	35.319	1.033
TYR A			18.451	28.758	-1.114
TYR A			19.199	28.139	-1.589
TYR A	57 HB2		18.125	28.236	-0.217
TYR A	57 CG		19.121	30.08	-0.848
TYR A			18.793	30.923	0.216
TYR A	57 HD1		18.074	30.612	0.962
TYR A			19.357	32.189	0.332
TYR A	57 HE1		19.07	32.83	1.155
TYR A			20.257	32.65	-0.64
TYR A			20.72	33.915	-0.634
TYR A			20.274	34.489	0.098
TYR A			20.101	30.515	-1.752
TYR A			20.394	29.853	-2.564
TYR A			20.675	31.778	-1.657
TYR A			21.412	32.105	-2.382
TYR A			19.172	43.743	2.258
TYR A			18.377	43.664	1.513

TYR	A	185 HB2	18.767	44.326	3.088
TYR	Α	185 CG	19.52	42.353	2.737
TYR	Α	185 CD1	20.268	42.19	3.9
TYR	A	185 HD1	20.633	43.08	4.378
TYR	A	185 CE1	20.548	40.951	4.451
TYR	A	185 HE1	21.122	40.855	5.366
TYR	A	185 CZ	20.084	39.807	3.801
TYR	Α	185 OH	20.332	38.611	4.382
TYR	Α	185 HH	20.005	37.882	3.796
TYR	A	185 CD2	19.06	41.197	2.104
TYR	A	185 HD2	18.443	41.276	1.216
TYR	A	185 CE2	19.358	39.933	2.61
TYR	A	185 HE2	19.001	39.039	2.114
THR	Α	89 CB	13.525	34.153	7.186
THR	A	89 HB	14.346	34.097	7.913
THR	A	89 OG1	13.966	35.005	6.142
THR	A	89 HG1	13.87	34.535	5.292
THR	A	89 CG2	13.189	32.74	6.718
THR	A	89 HG21	12.422	32.755	5.943
THR	A	89 HG22	12.81	32.133	7.545
THR	Α	89 HG23	14.065	32.249	6.293
LYS	Α	216 CB	19.893	32.654	8.085
LYS	A	216 HB1	19.393	31.733	8.411
LYS	A	216 HB2	20.648	32.351	7.352
LYS	A	216 CG	18.877	33.605	7.446
LYS	A	216 HG1	19.386	34.456	6.987
LYS	A	216 HG2	18.239	34.032	8.227
LYS	Α	216 CD	17.978	32.944	6.407
LYS	Α	216 HD1	17.583	31.994	6.785
LYS	A	216 HD2	18.528	32.712	5.492
LYS	A	216 CE	16.805	33.859	6.066
LYS	A	216 HE1	16.292	34.19	6.969
LYS	A	216 HE2	16.06	33.387	5.427
LYS	A	216 NZ	17.246	35.053	5.368
LYS	Α	216 HZ1	17.404	34.926	4.357
HOH		402 O	17.687	33.915	2.868
HOH		402 H1	17.98	34.384	2.073
НОН		402 H2	16.81	33.489	2.627
НОН		401 O	16.112	33.207	-0.4
НОН		401 H1	15.746	33.082	0.528
НОН		401 H2	16.816	32.55	-0.483
HOH		406 O	17.486	35.589	-0.744
HOH		406 H1	16.939	34.786	-0.588
HOH		406 H2	18.131	35.647	-0.008
RTAL		301 C1	14.874	45.074	0.303
RTAL		301 C2	14.774	45.988	-0.938
RTAL		301 H21	14.308	45.416	-1.741
RTAL		301 H22	15.791	46.237	-1.265
RTAL		301 C3	14.02	47.282	-0.724
RTAL		301 H31	12.981	47.066	-0.471
RTAL		301 H32	13.995	47.853	-1.658
RTAL		301 C4	14.724	48.066	0.379
RTAL		301 H41	14.15	48.946	0.693
RTAL		301 H42	15.652	48.492	-0.025
RTAL		301 C5	15.062	47.233	1.599

RTAL	301 C6	15.157	45.873	1.58
RTAL	301 C7	15.498	45.176	2.812
RTAL	301 H71	15.837	45.798	3.63
RTAL	301 C8	15.497	43.842	3.061
RTAL	301 H81	15.129	43.141	2.326
RTAL	301 C9	16.055	43.227	4.233
RTAL	301 C10	16.193	41.863	4.193
RTAL	301 H101	15.834	41.354	3.301
RTAL	301 C11	16.824	41.027	5.149
RTAL	301 H111	17.126	41.424	6.114
RTAL	301 C12	17.031	39.71	4.84
RTAL	301 H121	16.723	39.382	3.853
RTAL	301 C13	17.49	38.689	5.711
RTAL	301 C14	17.499	37.406	5.192
RTAL	301 H141	17.461	37.302	4.12
RTAL	301 C15	17.349	36.236	5.932
RTAL	301 H15	17.194	36.268	7.005
RTAL	301 C16	13.524	44.339	0.492
RTAL	301 H161	13.238	43.888	-0.457
RTAL	301 H162	12.74	45.034	0.806
RTAL	301 H163	13.563	43.545	1.238
RTAL	301 C17	16.018	44.093	-0.04
RTAL	301 H171	16.959	44.637	-0.16
RTAL	301 H172	15.8	43.609	-0.995
RTAL	301 H173	16.179	43.311	0.697
RTAL	301 C18	15.215	48.039	2.857
RTAL	301 H181	14.243	48.181	3.345
RTAL	301 H182	15.611	49.025	2.625
RTAL	301 H183	15.866	47.594	3.603
RTAL	301 C19	16.581	44.061	5.361
RTAL	301 H191	15.867	44.825	5.666
RTAL	301 H192	17.494	44.579	5.059
RTAL	301 H193	16.808	43.445	6.225
RTAL	301 C20	17.929	38.991	7.105
RTAL	301 H201	17.237	39.646	7.628
RTAL	301 H202	18.89	39.516	7.053
RTAL	301 H203	18.086	38.088	7.695

(b) 1C3W (model 2)

residue	atom	X	У	z	
ARG A	82 CB		14.716	36.94	-2.32
ARG A	82 HB1		15.108	37.429	-1.422
ARG A	82 HB2		14.149	37.713	-2.855
ARG A	82 CG		15.871	36.492	-3.191
ARG A	82 HG1		15.553	36.313	-4.222
ARG A	82 HG2		16.312	35.568	-2.814
ARG A	82 CD		16.91	37.607	-3.198
ARG A	82 HD1		17.336	37.739	-2.201
ARG A	82 HD2		16.466	38.557	-3.494
ARG A	82 NE		17.976	37.324	-4.132
ARG A	82 HE		18.016	37.848	-5.026
ARG A	82 TIE 82 CZ		18.914	36.414	-3.944
ARG A	82 VH1		19.004	35.725	-2.804
ARG A	82 HH11		18.337	35.723	-2.036
	82 HH12				
ARG A			19.779	35.083	-2.677
ARG A	82 NH2		19.781	36.164	-4.949 5.764
ARG A	82 HH21		19.784	36.767	-5.764
ARG A	82 HH22		20.589	35.589	-4.743
ASP A	85 CB		13.185	33.544	1.792
ASP A	85 HB1		13.47	34.438	1.24
ASP A	85 HB2		13.249	32.788	1.024
ASP A	85 CG		14.391	33.314	2.718
ASP A	85 OD1		14.376	33.501	3.951
ASP A	85 OD2		15.441	32.952	2.073
ASP A	212 CB		20.87	34.929	2.914
ASP A	212 HB1		20.226	34.154	3.345
ASP A	212 HB2		21.545	34.442	2.208
ASP A	212 CG		19.871	35.814	2.165
ASP A	212 OD1		19.349	36.818	2.688
ASP A	212 OD2		19.496	35.354	1.023
TYR A	57 CB		18.454	28.766	-1.12
TYR A	57 HB1		19.207	28.151	-1.595
TYR A	57 HB2		18.132	28.244	-0.22
TYR A	57 CG		19.11	30.096	-0.859
TYR A	57 CD1		18.677	30.993	0.119
TYR A	57 HD1		17.859	30.744	0.78
TYR A	57 CE1		19.239	32.257	0.226
TYR A	57 HE1		18.867	32.943	0.973
TYR A	57 CZ		20.235	32.663	-0.668
TYR A	57 OH		20.71	33.934	-0.668
TYR A	57 HH		20.283	34.5	0.065
TYR A	57 CD2		20.161	30.49	-1.699
TYR A	57 HD2		20.519	29.792	-2.453
TYR A	57 CE2		20.732	31.756	-1.612
TYR A	57 HE2		21.529	32.055	-2.286
TYR A	185 CB		19.175	43.742	2.26
TYR A	185 HB1		18.377	43.663	1.516
TYR A	185 HB2		18.772	44.322	3.091
TYR A	185 CG		19.524	42.352	2.737

TVD		105 CD1	20.270	42 100	2 007
TYR	A	185 CD1	20.279	42.189	3.897
TYR	A	185 HD1	20.647	43.078	4.373
TYR	A	185 CE1	20.557	40.95	4.449
TYR	A	185 HE1	21.134	40.854	5.363
TYR	A	185 CZ	20.086	39.806	3.804
TYR	A	185 OH	20.332	38.61	4.388
TYR	A	185 HH	19.984	37.882	3.815
TYR	A	185 CD2	19.058	41.196	2.109
TYR	A	185 HD2	18.436	41.276	1.223
TYR	A	185 CE2	19.355	39.932	2.615
TYR	A	185 HE2	18.995	39.038	2.124
THR	A	89 CB	13.524	34.158	7.187
THR	A	89 HB	14.34	34.092	7.921
THR	A	89 OG1	13.971	35.019	6.155
THR	A	89 HG1	13.886	34.553	5.299
THR	A	89 CG2	13.187	32.749	6.707
THR	A	89 HG21	12.427	32.769	5.926
		89 HG21		32.709	
THR	A		12.801		7.529
THR	A	89 HG23	14.065	32.257	6.286
LYS	A	216 CB	19.897	32.655	8.084
LYS	A	216 HB1	19.398	31.733	8.408
LYS	A	216 HB2	20.653	32.354	7.352
LYS	A	216 CG	18.881	33.604	7.445
LYS	A	216 HG1	19.39	34.454	6.983
LYS	A	216 HG2	18.245	34.034	8.227
LYS	A	216 CD	17.977	32.942	6.411
LYS	A	216 HD1	17.587	31.99	6.789
LYS	A	216 HD2	18.52	32.713	5.49
LYS	A	216 CE	16.798	33.854	6.081
LYS	A	216 HE1	16.29	34.183	6.986
LYS	A	216 HE2	16.051	33.384	5.442
LYS	A	216 NZ	17.232	35.05	5.383
LYS	A	216 HZ1	17.388	34.918	4.369
НОН	•	402 O	17.669	33.922	2.906
НОН		402 H1	17.968	34.368	2.101
НОН		402 H2	16.789	33.486	2.653
HOH		401 O	15.754	33.480	-0.549
НОН		401 H1	15.588	32.944	
		401 H1 401 H2			0.422
HOH			16.345	33.777	-0.634
HOH		406 O	17.34	35.631	-0.467
НОН		406 H1	16.641	36.19	-0.093
HOH		406 H2	18.102	35.669	0.17
RTAL		301 C1	14.867	45.06	0.321
RTAL		301 C2	14.764	45.965	-0.927
RTAL		301 H21	14.293	45.386	-1.723
RTAL		301 H22	15.779	46.209	-1.26
RTAL		301 C3	14.014	47.264	-0.721
RTAL		301 H31	12.974	47.055	-0.467
RTAL		301 H32	13.992	47.829	-1.659
RTAL		301 C4	14.719	48.053	0.377
RTAL		301 H41	14.147	48.937	0.684
RTAL		301 H42	15.648	48.473	-0.03
RTAL		301 C5	15.054	47.227	1.603
RTAL		301 C6	15.151	45.867	1.592
RTAL		301 C7	15.499	45.176	2.826

RTAL	301 H71	15.836	45.803	3.642
RTAL	301 C8	15.51	43.843	3.078
RTAL	301 H81	15.142	43.138	2.346
RTAL	301 C9	16.079	43.231	4.248
RTAL	301 C10	16.214	41.868	4.212
RTAL	301 H101	15.845	41.358	3.324
RTAL	301 C11	16.844	41.028	5.167
RTAL	301 H111	17.152	41.421	6.131
RTAL	301 C12	17.039	39.709	4.859
RTAL	301 H121	16.722	39.383	3.874
RTAL	301 C13	17.491	38.683	5.729
RTAL	301 C14	17.487	37.402	5.212
RTAL	301 H141	17.442	37.298	4.14
RTAL	301 C15	17.337	36.229	5.949
RTAL	301 H15	17.183	36.259	7.022
RTAL	301 C16	13.52	44.326	0.519
RTAL	301 H161	13.243	43.838	-0.415
RTAL	301 H162	12.73	45.028	0.802
RTAL	301 H163	13.554	43.56	1.294
RTAL	301 C17	16.014	44.08	-0.015
RTAL	301 H171	16.956	44.626	-0.125
RTAL	301 H172	15.806	43.599	-0.974
RTAL	301 H173	16.169	43.295	0.722
RTAL	301 C18	15.209	48.042	2.857
RTAL	301 H181	14.24	48.173	3.353
RTAL	301 H182	15.589	49.031	2.617
RTAL	301 H183	15.872	47.608	3.599
RTAL	301 C19	16.609	44.069	5.37
RTAL	301 H191	15.845	44.736	5.772
RTAL	301 H192	17.434	44.699	5.028
RTAL	301 H193	16.971	43.443	6.181
RTAL	301 C20	17.934	38.98	7.123
RTAL	301 H201	17.233	39.618	7.657
RTAL	301 H202	18.885	39.523	7.072
RTAL	301 H203	18.109	38.074	7.702

(c) 2NTU (model 1)

residue	atom	X	y	Z	
ARG A	82 CB		14.748	37.211	-2.273
ARG A	82 HB1		15.184	37.638	-1.365
ARG A	82 HB2		14.193	38.019	-2.77
ARG A	82 CG		15.87	36.75	-3.19
ARG A	82 HG1		15.52	36.594	-4.217
ARG A	82 HG2		16.306	35.812	-2.842
ARG A	82 CD		16.932	37.845	-3.208
ARG A	82 HD1		17.39	37.935	-2.221
ARG A	82 HD2		16.491	38.808	-3.469
ARG A	82 NE		17.957	37.582	-4.198
ARG A	82 HE		17.942	38.111	-5.083
ARG A	82 CZ		18.929	36.69	-4.086
ARG A	82 NH1		19.096	35.953	-2.991
ARG A	82 HH11		18.491	35.985	-2.145
ARG A	82 HH12		19.937	35.38	-2.143
ARG A	82 NH2		19.749	36.504	-5.151
ARG A	82 HH21		19.749	37.164	-5.922
ARG A	82 HH22		20.597	35.974	-5.019
ASP A	85 CB		13.226	33.832	1.815
ASP A	85 HB1		13.226	34.753	1.309
ASP A	85 HB2		13.259	34.733	1.014
ASP A	85 CG		13.239	33.547	2.735
ASP A	85 OD1		14.429	33.679	3.974
ASP A	85 OD1 85 OD2		14.412	33.079	2.073
ASP A ASP A	212 CB 212 HB1		20.954 20.298	35.152 34.39	3.094
ASP A ASP A	212 HB2		20.298	34.39 34.642	3.527 2.413
ASP A	212 HB2 212 CG		19.991	34.042 36.029	2.413
ASP A	212 CG 212 OD1		19.561	37.115	2.283
ASP A	212 OD1 212 OD2		19.572	37.113	1.198
TYR A	57 CB		18.602	28.971	-1.201
TYR A	57 HB1		19.338	28.349	-1.706
TYR A	57 HB2		19.338	28.432	-0.306
TT 170 4	57 ПБ2 57 СG		19.272	30.289	-0.895
	57 CD1		18.921		0.187
TYR A TYR A	57 HD1		18.197	31.103 30.771	0.187
TYR A	57 CE1		19.47	30.771	0.348
TYR A	57 HE1		19.47	32.371	1.183
TYR A	57 CZ		20.388	32.863	-0.59
TYR A	57 CZ 57 OH		20.388	34.13	
TYR A	57 HH		20.347	34.13 34.671	-0.538 0.202
TYR A	57 CD2		20.397	30.756	
					-1.766
	57 HD2		20.581	30.12	-2.59
TYR A	57 CE2		20.831	32.019	-1.621
TYR A TYR A	57 HE2 185 CB		21.583 19.289	32.368	-2.322
				44.049	2.317
	185 HB1		18.483	43.962	1.584
TYR A	185 HB2		18.897	44.633	3.152
TYR A	185 CG		19.643	42.663	2.815
TYR A	185 CD1		20.383 20.742	42.505 43.393	3.987
TYR A TYR A	185 HD1				4.48 4.539
TYR A	185 CE1		20.658	41.263	4.339

TYR	Α	185 HE1	21.228	41.169	5.459
TYR	Α	185 CZ	20.197	40.115	3.892
TYR	A	185 OH	20.451	38.914	4.468
TYR	A	185 HH	20.14	38.186	3.873
TYR	A	185 CD2	19.176	41.503	2.192
TYR	A	185 HD2	18.552	41.577	1.308
TYR	A	185 TID2 185 CE2			
			19.467	40.239	2.703
TYR	A	185 HE2	19.106	39.345	2.21
THR	A	89 CB	13.567	34.365	7.361
THR	A	89 HB	14.402	34.288	8.071
THR	A	89 OG1	13.997	35.215	6.309
THR	A	89 HG1	13.927	34.738	5.46
THR	A	89 CG2	13.208	32.958	6.882
THR	A	89 HG21	12.446	32.984	6.101
THR	A	89 HG22	12.815	32.348	7.702
THR	A	89 HG23	14.083	32.459	6.463
LYS	A	216 CB	20.002	32.975	8.395
LYS	A	216 HB1	19.514	32.047	8.714
LYS	A	216 HB2	20.757	32.678	7.659
LYS	A	216 CG	18.983	33.908	7.732
LYS	A	216 HG1	19.487	34.774	7.732
LYS	A	216 HG2	18.312	34.312	8.496
LYS				33.228	
	A	216 CD	18.139		6.655
LYS	A	216 HD1	17.749	32.269	7.018
LYS	A	216 HD2	18.739	33.004	5.769
LYS	A	216 CE	16.955	34.104	6.244
LYS	A	216 HE1	16.39	34.434	7.117
LYS	A	216 HE2	16.252	33.59	5.589
LYS	A	216 NZ	17.378	35.296	5.529
LYS	A	216 HZ1	17.515	35.161	4.516
HOH		402 O	17.751	34.13	3.01
HOH		402 H1	18.114	34.562	2.222
НОН		402 H2	16.872	33.737	2.733
НОН		401 O	16.221	33.469	-0.389
НОН		401 H1	15.83	33.367	0.53
НОН		401 H2	16.916	32.8	-0.442
НОН		406 O	17.631	35.848	-0.691
НОН		406 H1	18.266	35.884	0.056
НОН		406 H2	17.078	35.046	-0.553
RTAL		301 C1		45.339	
			14.962		0.341
RTAL		301 C2	14.828	46.27	-0.888
RTAL		301 H21	14.365	45.702	-1.699
RTAL		301 H22	15.833	46.545	-1.23
RTAL		301 C3	14.054	47.547	-0.624
RTAL		301 H31	13.031	47.307	-0.328
RTAL		301 H32	13.982	48.138	-1.544
RTAL		301 C4	14.779	48.323	0.469
RTAL		301 H41	14.218	49.198	0.808
RTAL		301 H42	15.703	48.738	0.041
RTAL		301 C5	15.132	47.485	1.676
RTAL		301 C6	15.24	46.125	1.634
RTAL		301 C7	15.573	45.425	2.863
RTAL		301 H71	15.864	46.055	3.692
RTAL		301 C8	15.607	44.088	3.112
RTAL		301 H81	15.305	43.379	2.354
		201 1101	10.505		55 .

RTAL	301 C9	16.111	43.48	4.312
RTAL	301 C10	16.282	42.117	4.27
RTAL	301 H101	15.987	41.611	3.354
RTAL	301 C11	16.872	41.286	5.252
RTAL	301 H111	17.139	41.693	6.223
RTAL	301 C12	17.095	39.963	4.96
RTAL	301 H121	16.82	39.63	3.965
RTAL	301 C13	17.542	38.952	5.844
RTAL	301 C14	17.575	37.66	5.332
RTAL	301 H141	17.537	37.551	4.258
RTAL	301 C15	17.453	36.493	6.078
RTAL	301 H15	17.31	36.53	7.154
RTAL	301 C16	13.633	44.566	0.523
RTAL	301 H161	13.393	44.037	-0.401
RTAL	301 H162	12.818	45.256	0.759
RTAL	301 H163	13.668	43.831	1.328
RTAL	301 C17	16.125	44.388	-0.02
RTAL	301 H171	17.053	44.954	-0.14
RTAL	301 H172	15.913	43.905	-0.978
RTAL	301 H173	16.307	43.605	0.714
RTAL	301 C18	15.268	48.288	2.938
RTAL	301 H181	14.269	48.449	3.36
RTAL	301 H182	15.694	49.264	2.711
RTAL	301 H183	15.882	47.834	3.712
RTAL	301 C19	16.543	44.31	5.487
RTAL	301 H191	15.904	45.182	5.63
RTAL	301 H192	17.564	44.675	5.34
RTAL	301 H193	16.519	43.724	6.405
RTAL	301 C20	17.961	39.272	7.244
RTAL	301 H201	17.301	39.991	7.724
RTAL	301 H202	18.957	39.729	7.203
RTAL	301 H203	18.038	38.384	7.871

(d) 2NTU (model 2)

		atom	•			
residue ARG	A	82 CB	X	14.733	37.219	-2.288
	A	82 CB 82 HB1		15.136	37.219	-2.288
	A A	82 HB2		13.130	37.7 37.999	-2.815
	A	82 HB2 82 CG		15.88	36.76	-2.813
	A A	82 CG 82 HG1		15.88	36.76	-3.176 -4.197
				16.334	35.844	
_	A	82 HG2 82 CD			35.844 37.885	-2.792
	A	82 CD 82 HD1		16.912		-3.238
_	A	82 HD1 82 HD2		17.352	38.059	-2.251
_	A A	82 HD2 82 NE		16.451 17.97	38.819 37.594	-3.559 -4.185
	A A	82 NE 82 HE		17.97	37.394	-4.183 -5.086
	A A	82 HE 82 CZ		17.988	36.709	-3.086
_	A A	82 CZ 82 NH1				
_				19.056	36.026 36.134	-2.857
	A	82 HH11 82 HH12		18.425		-2.058
	A	-		19.863	35.418	-2.749
	A	82 NH2		19.802	36.472	-5.008
	A	82 HH21		19.782	37.064	-5.831
_	A	82 HH22		20.616	35.904	-4.814
	A	85 CB		13.228	33.821	1.827
	A	85 HB1		13.515	34.727	1.296
	A	85 HB2		13.265	33.073	1.043
	A	85 CG		14.433	33.557	2.758
	A	85 OD1		14.405	33.7	3.999
	A	85 OD2		15.486	33.219	2.107
	A	212 CB		20.947	35.16	3.094
	A	212 HB1		20.291	34.398	3.528
	A	212 HB2		21.63	34.651	2.409
	A	212 CG		19.979	36.039	2.292
	A	212 OD1		19.525	37.111	2.737
	A	212 OD2		19.575	35.507	1.189
	A	57 CB		18.607	28.985	-1.208
	A	57 HB1		19.348	28.367	-1.711
	A	57 HB2		18.295	28.45	-0.308
	A	57 CG		19.259	30.314	-0.915
	A	57 CD1		18.778	31.201	0.052
	A	57 HD1		17.928	30.947	0.667
	A	57 CE1		19.331	32.463	0.209
	A	57 HE1		18.925	33.138	0.948
	A	57 CZ		20.374	32.88	-0.623
	A	57 OH		20.853	34.15	-0.573
	A	57 HH		20.416	34.686	0.165
	A	57 CD2		20.35	30.722	-1.696
	A	57 HD2		20.748	30.036	-2.44
	A	57 CE2		20.915	31.989	-1.558
	A	57 HE2		21.745	32.298	-2.187
	A	185 CB		19.29	44.049	2.318
	A	185 HB1		18.482	43.962	1.586
	A	185 HB2		18.899	44.632	3.153
	A	185 CG		19.644	42.664	2.814
	A	185 CD1		20.391	42.508	3.981
	A	185 HD1		20.755	43.397	4.468
TYR A	A	185 CE1		20.665	41.267	4.537

TYR	Α	185 HE1	21.239	41.176	5.454
TYR	A	185 CZ	20.196	40.118	3.896
TYR	A	185 OH	20.451	38.918	4.475
TYR	A	185 HH	20.125	38.19	3.89
TYR	A	185 CD2	19.17	41.504	2.197
TYR	A	185 HD2	18.54	41.578	1.317
TYR	A	185 CE2	19.461	40.24	2.711
TYR	A	185 HE2	19.097	39.346	2.223
THR	A	89 CB	13.568	34.362	7.362
THR	A	89 HB	14.401	34.278	8.075
THR	Α	89 OG1	14.002	35.214	6.313
THR	A	89 HG1	13.942	34.736	5.462
THR	A	89 CG2	13.205	32.957	6.879
THR	A	89 HG21	12.446	32.987	6.094
THR	A	89 HG22	12.806	32.348	7.697
THR	A	89 HG23	14.079	32.453	6.462
LYS	A	216 CB	20	32.974	8.398
LYS		216 CB 216 HB1	19.506	32.974	8.721
	A				
LYS	A	216 HB2	20.754	32.669	7.663
LYS	A	216 CG	18.986	33.908	7.73
LYS	A	216 HG1	19.495	34.768	7.285
LYS	A	216 HG2	18.32	34.321	8.493
LYS	A	216 CD	18.134	33.224	6.661
LYS	Α	216 HD1	17.75	32.265	7.028
LYS	A	216 HD2	18.726	33	5.77
LYS	A	216 CE	16.943	34.096	6.26
LYS	A	216 HE1	16.383	34.423	7.138
LYS	A	216 HE2	16.238	33.582	5.607
LYS	A	216 NZ	17.357	35.292	5.546
LYS	A	216 HZ1	17.484	35.157	4.529
HOH	А	402 O	17.709	34.173	3.04
HOH		402 H1	18.096	34.173	2.24
HOH		402 H2	16.838	33.75	2.75
HOH		401 O	15.873	33.336	-0.542
НОН		401 H1	15.675	33.256	0.422
НОН		401 H2	16.494	34.08	-0.599
НОН		406 O	17.525	35.918	-0.446
HOH		406 H1	18.262	35.901	0.223
HOH		406 H2	16.826	36.47	-0.065
RTAL		301 C1	14.962	45.329	0.352
RTAL		301 C2	14.819	46.256	-0.88
RTAL		301 H21	14.352	45.683	-1.686
RTAL		301 H22	15.823	46.528	-1.23
RTAL		301 C3	14.048	47.534	-0.619
RTAL		301 H31	13.026	47.298	-0.32
RTAL		301 H32	13.976	48.124	-1.54
RTAL		301 C4	14.776	48.314	0.471
RTAL		301 H41	14.216	49.189	0.808
RTAL		301 H42	15.698	48.727	0.039
RTAL		301 C5	15.132	47.48	1.681
RTAL		301 C5 301 C6	15.132	46.119	1.642
RTAL		301 C6 301 C7	15.591	45.421	
					2.87
RTAL		301 H71	15.887	46.053	3.696
RTAL		301 C8	15.632	44.085	3.122
RTAL		301 H81	15.322	43.372	2.37

RTAL	301 C9	16.15	43.483	4.32
RTAL	301 C10	16.307	42.118	4.291
RTAL	301 H101	15.994	41.608	3.382
RTAL	301 C11	16.896	41.283	5.271
RTAL	301 H111	17.174	41.686	6.24
RTAL	301 C12	17.102	39.958	4.98
RTAL	301 H121	16.814	39.625	3.988
RTAL	301 C13	17.545	38.943	5.863
RTAL	301 C14	17.562	37.653	5.354
RTAL	301 H141	17.511	37.543	4.281
RTAL	301 C15	17.443	36.483	6.1
RTAL	301 H15	17.31	36.518	7.177
RTAL	301 C16	13.635	44.555	0.542
RTAL	301 H161	13.4	44.005	-0.371
RTAL	301 H162	12.815	45.246	0.761
RTAL	301 H163	13.669	43.837	1.363
RTAL	301 C17	16.124	44.379	-0.015
RTAL	301 H171	17.053	44.946	-0.13
RTAL	301 H172	15.915	43.905	-0.978
RTAL	301 H173	16.305	43.59	0.712
RTAL	301 C18	15.266	48.29	2.939
RTAL	301 H181	14.267	48.447	3.36
RTAL	301 H182	15.686	49.268	2.71
RTAL	301 H183	15.881	47.841	3.715
RTAL	301 C19	16.601	44.326	5.477
RTAL	301 H191	15.863	45.087	5.732
RTAL	301 H192	17.529	44.85	5.229
RTAL	301 H193	16.775	43.719	6.363
RTAL	301 C20	17.974	39.261	7.261
RTAL	301 H201	17.301	39.959	7.756
RTAL	301 H202	18.957	39.744	7.213
RTAL	301 H203	18.08	38.369	7.878

Table S2. Atomic coordinates of the QM/MM optimized geometry for ASR [W402, Tyr11, Tyr51, Asp75, Trp76, Thr79, Ser47, the backbone of Phe202, Lys210 (the Schiff base) and the retinal].

1XIO (deprotonated Asp75)

residue		atom	v	*7	7	
	A	11 CB	X	28.683	32.043	23.475
	A	11 CB 11 HB1		29.208	32.866	23.473
	A	11 HB2		29.208	32.457	23.932
		11 HB2 11 CG				
	A			29.664	30.969	23.082
	A	11 CD1		29.352	30.066	22.066
	A	11 HD1		28.405	30.148	21.539
	A	11 CE1		30.236	29.054	21.722
	A	11 HE1		29.956	28.336	20.969
	A	11 CZ		31.467	28.925	22.373
	A	11 OH		32.302	27.928	21.976
	A	11 HH		33.133	27.936	22.512
	A	11 CD2		30.878	30.802	23.759
	A	11 HD2		31.123	31.475	24.574
	A	11 CE2		31.782	29.801	23.416
	A	11 HE2		32.735	29.716	23.927
	A	51 CB		33.276	33.098	25.01
	A	51 HB1		32.369	33.707	25.086
TYR	A	51 HB2		33.13	32.21	25.638
	A	51 CG		33.493	32.727	23.547
	A	51 CD1		34.614	32.028	23.071
TYR	A	51 HD1		35.404	31.705	23.737
TYR	A	51 CE1		34.764	31.711	21.727
TYR	A	51 HE1		35.648	31.178	21.404
TYR	Α	51 CZ		33.767	32.05	20.812
TYR	A	51 OH		33.953	31.654	19.506
TYR	A	51 HH		33.22	31.982	18.944
TYR	A	51 CD2		32.518	33.062	22.602
TYR	Α	51 HD2		31.614	33.553	22.948
TYR	Α	51 CE2		32.632	32.735	21.25
	Α	51 HE2		31.849	32.996	20.548
	A	75 CB		38.833	29.43	22.443
	Α	75 HB1		38.814	30.216	21.689
	Α	75 HB2		38.631	29.939	23.383
	A	75 CG		37.618	28.525	22.171
	Α	75 OD1		37.662	27.273	22.244
	Α	75 OD2		36.54	29.182	21.953
	A	76 CB		40.705	27.733	17.857
	A	76 HB1		40.923	26.964	17.11
	A	76 HB2		41.52	28.465	17.807
	A	76 CG		39.4	28.405	17.529
	A	76 CD1		38.175	28.269	18.132
	A	76 HD1		37.904	27.718	19.022
	A	76 NE1		37.205	28.987	17.456
	A	76 HE1		36.34	29.236	17.436
	A	76 CE2		37.81	29.230	16.41
	A	76 CD2		39.193	29.037	16.423
	A	76 CE3		40.044	29.313	15.482
11/1	Δ.	10 CE3		40.044	47.74 l	13.404

TRP	Α	76 HE3	41.111	29.725	15.488
TRP	A	76 CZ3	39.514	30.836	14.569
TRP	Α	76 HZ3	40.169	31.345	13.87
TRP	Α	76 CZ2	37.267	30.535	15.463
TRP	Α	76 HZ2	36.204	30.759	15.452
TRP	A	76 CH2	38.136	31.134	14.551
TRP	A	76 HH2	37.755	31.856	13.836
THR	A	79 CB	38.473	23.841	21.882
THR	A	79 HB	37.622	23.177	
					21.659
THR	A	79 OG1	38.448	24.87	20.899
THR	A	79 HG1	38.395	25.736	21.353
THR	A	79 CG2	38.308	24.372	23.299
THR	A	79 HG21	39.101	25.078	23.545
THR	A	79 HG22	38.334	23.561	24.035
THR	Α	79 HG23	37.359	24.898	23.411
SER	Α	47 CB	35.096	27.085	24.338
SER	A	47 HB1	34.234	26.62	24.822
SER	Α	47 HB2	35.756	26.296	23.961
SER	Α	47 OG	34.614	27.899	23.283
SER	Α	47 HG1	35.344	28.389	22.814
LYS	A	210 CB	32.248	23.136	21.7
LYS	A	210 HB1	32.44	22.836	22.736
LYS	A	210 HB2	31.46	23.896	21.72
LYS	A	210 HB2 210 CG	33.52	23.732	21.72
LYS	A	210 CG 210 HG1	33.4	23.732	19.998
LYS	A	210 HG2	34.351	23.027	21.202
LYS	A	210 CD	33.912	25.075	21.673
LYS	A	210 HD1	34.034	24.981	22.759
LYS	A	210 HD2	33.124	25.814	21.506
LYS	A	210 CE	35.247	25.562	21.118
LYS	Α	210 HE1	35.994	24.769	21.174
LYS	Α	210 HE2	35.637	26.403	21.685
LYS	Α	210 NZ	35.227	26.014	19.715
LYS	Α	210 HZ1	35.132	27.03	19.564
PHE	Α	202 CA	31.509	32.626	15.652
PHE	Α	202 HA	31.679	31.61	15.31
PHE	A	202 C	31.066	32.505	17.082
PHE	Α	202 O	31.843	32.422	18.035
CYS	A	203 N	29.738	32.291	17.228
CYS	A	203 HN	29.099	32.666	16.538
НОН	11	402 O	35.297	29.007	19.553
НОН		402 H1	34.655	29.743	19.529
HOH		402 H2	35.765	29.143	20.421
		301 C1			
RTAL			41.013	29.34	10.326
RTAL		301 C2	41.593	30.413	9.367
RTAL		301 H21	41.876	31.293	9.956
RTAL		301 H22	40.798	30.755	8.69
RTAL		301 C3	42.752	29.901	8.522
RTAL		301 H31	43.559	29.515	9.158
RTAL		301 H32	43.182	30.707	7.919
RTAL		301 C4	42.232	28.783	7.63
RTAL	,	301 H41	43.051	28.282	7.101
RTAL	,	301 H42	41.609	29.205	6.826
RTAL	,	301 C5	41.418	27.751	8.376
RTAL	,	301 C6	40.871	27.97	9.613

RTAL	301 C7	40.162	26.866	10.261
RTAL	301 H71	40.012	25.985	9.646
RTAL	301 C8	39.687	26.785	11.534
RTAL	301 H81	39.82	27.621	12.206
RTAL	301 C9	38.957	25.685	12.123
RTAL	301 C10	38.455	25.913	13.382
RTAL	301 H101	38.671	26.889	13.812
RTAL	301 C11	37.625	25.092	14.2
RTAL	301 H111	37.396	24.071	13.904
RTAL	301 C12	37.11	25.618	15.358
RTAL	301 H121	37.359	26.653	15.577
RTAL	301 C13	36.321	24.963	16.353
RTAL	301 C14	35.941	25.734	17.44
RTAL	301 H141	36.093	26.804	17.366
RTAL	301 C15	35.553	25.244	18.7
RTAL	301 H15	35.623	24.183	18.924
RTAL	301 C16	41.961	29.212	11.546
RTAL	301 H161	42.022	30.174	12.068
RTAL	301 H162	42.967	28.917	11.236
RTAL	301 H163	41.622	28.473	12.272
RTAL	301 C17	39.642	29.893	10.784
RTAL	301 H171	38.928	29.904	9.956
RTAL	301 H172	39.771	30.924	11.123
RTAL	301 H173	39.191	29.346	11.611
RTAL	301 C18	41.306	26.466	7.592
RTAL	301 H181	42.24	25.899	7.673
RTAL	301 H182	41.185	26.701	6.53
RTAL	301 H183	40.485	25.807	7.875
RTAL	301 C19	38.729	24.409	11.358
RTAL	301 H191	38.228	24.596	10.403
RTAL	301 H192	38.113	23.708	11.921
RTAL	301 H193	39.684	23.917	11.135
RTAL	301 C20	35.918	23.524	16.19
RTAL	301 H201	36.748	22.875	15.905
RTAL	301 H202	35.183	23.45	15.381
RTAL	301 H203	35.45	23.125	17.09