TITLE:
Diet-induced changes in Ucp1 expression in bovine adipose tissues.

AUTHOR(S):
Asano, Hiroki; Yamada, Tomoya; Hashimoto, Osamu; Umemoto, Takenao; Sato, Ryo; Ohwatari, Shiori; Kanamori, Yohei; Terachi, Tomohiro; Funaba, Masayuki; Matsui, Tohru

CITATION:
Asano, Hiroki ...[et al]. Diet-induced changes in Ucp1 expression in bovine adipose tissues.. General and comparative endocrinology 2013, 184: 87-92

ISSUE DATE:
2013-04

URL:
http://hdl.handle.net/2433/171241

RIGHT:
© 2013 Elsevier Inc.; この論文は出版社版ではありません。引用の際には出版社版をご確認ご利用ください。; This is not the published version. Please cite only the published version.
Diet-induced changes in Ucp1 expression in bovine adipose tissues

Hiroki Asano¹, Tomoya Yamada²*, Osamu Hashimoto³, Takenao Umemoto¹, Ryo Sato³, Shiori Ohwatari¹, Yohei Kanamori¹, Tomohiro Terachi¹, Masayuki Funaba† and Tohru Matsui¹

¹Division of Applied Biosciences, Kyoto University Graduate School of Agriculture, Kyoto 606-8502, Japan.
²National Institute of Livestock and Grassland Science, Nasushiobara 329-2793, Japan.
³Laboratory of Experimental Animal Science, Kitasato University, School of Veterinary Medicine, Towada 034-8628, Japan.

*These authors equally contributed to this study.
†To whom correspondence should be addressed:
Masayuki Funaba, Ph.D.
Division of Applied Biosciences
Kyoto University Graduate School of Agriculture
Kitashirakawa Oiwakecho, Kyoto 606-8502, Japan
Tel: +81-75-753-6055
Fax: +81-75-753-6344
E-mail: mfunaba@kais.kyoto-u.ac.jp
Abstract

Brown adipocytes, which regulate non-shivering thermogenesis, have been believed to exist in a limited number of mammalian species, and only under limited physiological conditions. Recent discoveries indicate that adult humans possess a significant number of functional brown adipocytes. This study explores the regulatory emergence of brown adipocytes in white adipose tissue (WAT) depots of fattening cattle. RT-PCR analyses indicated significant expression of *Ucp1*, a brown adipocyte-specific gene, in the WAT of 31-month-old Japanese Black steers. Immunohistochemical analysis revealed that *Ucp1*-positive small adipocytes were dispersed in the subcutaneous WAT. Next, we examined expression level of *Ucp1* and other brown adipocyte-selective genes such as *Pgc1α, Cidea, Dio2, Cox1, Cox7a1* and *Cox8b* in WAT of 30-month-old steers fed either diet with low protein/energy content (roughage diet) or that with high protein/energy content (concentrate diet) for 20 months. *Ucp1* expression in the subcutaneous WAT was significantly higher in the concentrate diet group than in the roughage diet group. Furthermore, the higher *Ucp1* expression levels were limited to the subcutaneous WAT, and no differences between groups were detected in the mesenteric, perirenal, intermuscular or intramuscular WAT. Expression of *Dio2, Cox1* and *Cox8b* was higher in the subcutaneous WAT but not in the mesenteric WAT of the concentrate diet group. Furthermore, expression of *Prdm16*, a positive regulator of differentiation toward brown adipocyte-lineage cells, and expression of *leptin*, a molecule that enhances activity of brown adipocytes, were significantly higher in the subcutaneous WAT of the concentrate diet group. This study demonstrates the presence of brown adipocytes in WAT depots of fattening cattle, and suggests the diet-related modulation of expression of genes predominantly expressed in brown adipocytes.
1. Introduction

There are two major types of adipose tissues in mammals: white and brown. White adipose tissue (WAT), which is dispersed throughout the body in mammals [13], is specialized for the storage of excess energy. WAT contains all of the enzymatic machinery necessary to produce triacylglycerols from fatty acids, either those synthesized de novo or imported from circulating lipoproteins. In addition, WAT plays a central role in the regulation of energy balance by acting as the site of the synthesis and secretion of molecules called adipokines, including leptin [11, 30]. In contrast, brown adipose tissue (BAT) is specialized to dissipate chemical energy in the form of heat in response to cold or excess feeding [1, 14, 22, 23, 38]. The thermogenic function of BAT results from the expression of a series of genes related to high mitochondrial content, as well as elevated cellular respiration largely uncoupled from ATP synthesis. The uncoupling occurs through expression of uncoupling protein 1 (Ucp1), a brown adipocyte-specific mitochondrial protein that promotes proton leak across the inner mitochondrial membrane [4, 13].

Thermogenesis in brown adipocytes has hitherto been considered to occur in a limited number of animal species, including small rodents, and under the limited (patho)physiological status such as newborn humans and patients with phenochromocytoma [4, 13]. Recently, a significant amount of a functional brown adipocyte depot was identified in adult humans via integrated positron emission tomography-computed tomography (PET-CT) studies using an $^{18}$F-labeled glucose analogue, fluorodeoxyglucose, as a tracer, and by the immunohistochemical analyses to detect Ucp1 [9, 31, 45, 47]. The finding that the sizes of the BAT depots are inversely correlated with body mass index [9, 31, 45, 47] has fuelled considerable interest in the therapeutic potential of brown adipocytes in obesity and obesity-related diseases [44].
Ucp1-positive brown adipocytes are also found interspersed in the WAT of rodents and adult humans, which are therefore called “brown in white” (brite) adipocytes, inducible brown fat cells, or beige cells [5, 18, 32]. Furthermore, Ucp1 mRNA was detected in the subcutaneous WAT of adult humans [29, 42, 49]. In view of the large amount of WAT depots, the brite adipocytes may be the major brown adipocyte throughout the human body [42].

Beef cattle are raised as industrial animals, and fattening efficiency is one of the determining factors of the economy of beef production. Unlike the case of adult humans, the emergence of brite adipocytes, which dissipate energy as heat, is not preferable in beef cattle. Thus, determining the factors affecting the induction and activity of brite adipocytes is important in beef cattle and humans alike, although the presence of brite adipocytes has not yet been established in fattening cattle. While Ucp1 expression was detected in the subcutaneous WAT of fetal calves [20, 37], the expression level was decreased to the detection limit at birth. Expression of Ucp1 could not be detected in the subcutaneous, perirenal and intermuscular WAT of mature cattle [20, 25, 26, 39, 40]. However, in previous studies, Ucp1 expression was examined by Northern blot or slot blot analyses, techniques that are generally insensitive methods compared with quantitative real-time RT-PCR analyses. Therefore, it is possible that the Ucp1 expression in WAT of fattening cattle was previously overlooked.

The objectives of the present study are to clarify the expression of Ucp1 in WAT depots of fattening cattle, and effects of diets on Ucp1 expression. Our findings demonstrate that Ucp1 is expressed in the WAT of fattening cattle and that the expression level in the subcutaneous WAT is increased by consuming a diet with higher protein and energy density. In view of the substantial parallel increase in expression of Ucp1 and that of the
other brown adipocyte-selective genes, we propose the diet-related emergence of brite adipocytes in fattening cattle.

2. Materials and methods

2.1. Animals and feeds

In Experiment 1, four Japanese Black steers aged 21 months were fed roughage (rice straw) with 38% total digestible nutrients (TDN) and 5% crude protein (CP), and a concentrate mixture consisting of barley, corn, wheat bran, rice bran and soybean meal with 74% TDN and 12% CP on an ad libitum basis for 10 months. The steers were raised in a stall covered with sawdust. They were allowed free access to drinking water and a mineral block (Cowstone, Nihon Zenyaku Kogyo, Koriyama, Japan). The perirenal and subcutaneous WAT depots were collected from the steers aged 31 months in a commercial slaughterhouse, where steers were exsanguinated after stunning. Tissue samples were promptly collected. The perirenal WAT depot samples were frozen in dry ice and stored at -80°C until samples underwent total RNA extraction. The subcutaneous WAT depot was washed with phosphate-buffered saline (PBS) and fixed for immunohistochemical analysis.

In Experiment 2, 8 Japanese Black steers were used. Experiment 2 was the same experiment reported by Yamada and Nakanishi [48]; data on daily feed intake and body composition were previously shown [48]. The steers were raised in a stall covered with sawdust. They were allowed free access to drinking water and a mineral block (Cowstone, Nihon Zenyaku Kogyo, Koriyama, Japan). Feeds were individually provided by door feeding system (Orion Machinery, Kyoto, Japan). The steers were fed roughage (orchardgrass hay) with 56% TDN and 8% CP and a concentrate mixture
consisting of corn, barley, wheat bran, rice bran and soybean meal (Nasuno for Wagyu Fattening; JA Higashi-nihon Kumiai Shiryou, Ota, Gunma, Japan) with 88% TDN and 15% CP as described below. At 10 months of age, the steers were allotted by body weight to one of two groups: the roughage diet group (n = 4) or the concentrate diet group (n = 4). The roughage diet consisted of 35% roughage and 65% concentrate mixture, whereas the concentrate diet contained 10% roughage and 90% concentrate mixture on a TDN basis. The roughage was given on an ad libitum basis to steers in the roughage diet group. The amount of concentrate in the roughage diet group was determined as follows: the amount of TDN required for achievement of body weight gain of 0.7 kg/day was obtained from the Japanese Feeding Standard for Beef Cattle [27], and the concentrate that corresponded to 65% of the required TDN was provided to the steers. To eliminate the influence of the total TDN intake between groups, the steers were pair-fed for 20 months; weekly TDN intake was measured in the roughage diet group, and amount of feed with average TDN content in the former week in the roughage diet group was provided to steers in the concentrate diet group. At 30 months of age, steers were slaughtered in a commercial slaughterhouse, and adipose samples from five types of WAT depots (subcutaneous, mesenteric, perirenal, intermuscular, and intramuscular) were collected [48]. All animals received humane care as outlined in the Guide for the Care and Use of Experimental Animals (National Institute of Livestock and Grassland Science).

2.2. Immunohistochemistry

Rabbit polyclonal antibody to human Ucp1 (ab10983) was obtained from Abcam (Cambridge, MA) to examine immunolocalization in the subcutaneous WAT. According to the manufacturer, this antibody is predicted to recognize bovine Ucp1. After fixation with Bouin’s solution for 24 h at room temperature, the samples were
dehydrated, embedded in paraffin, and sectioned to 4 µm thickness. Endogenous peroxidase was blocked using 0.3% H₂O₂ in methanol for 20 min at room temperature. The sections were washed with PBS, and treated using blocking solution (Histofine SAB-PO(R); Nichirei Biosciences, Tokyo, Japan) for 10 min at room temperature. After washing with PBS, the sections were incubated with the anti-Ucp1 antibody (diluted 1:400) overnight at 4°C. The sections were then washed with PBS and incubated with a biotinylated goat anti-rabbit secondary antibody (Histofine SAB-PO(R)) for 10 min at room temperature. After washing with PBS, the sections were incubated with peroxidase-conjugated streptavidin for 5 min at room temperature. After washing with PBS, the DAB substrate kit (Nichirei Biosciences) was applied to the sections for 2 min at room temperature, followed by counterstaining with hematoxylin. The sections were then dehydrated and mounted. The experiments were repeated at least three times, and the positive staining was reproducibly detected.

2.3. RNA isolation, conventional RT-PCR and quantitative RT-PCR

Total RNA isolation and cDNA synthesis were conducted as described by Yamada and Nakanishi [48]. The cDNA, reverse-transcribed from 10 ng of total RNA, was used as a template for conventional RT-PCR or quantitative RT-PCR (qRT-PCR). The oligonucleotide primers for conventional RT-PCR and qRT-PCR are presented in Table 1. Conventional PCR was performed in a total volume of 10 µl containing 1× Ex-Taq buffer with 2.0 mM MgCl₂, 0.2 mM of each dNTP, 0.2 µM of each primer, and 1.25 U of an Ex-Taq HS DNA polymerase (TaKaRa, Kyoto, Japan). The PCR profile of conventional RT-PCR is as follows: after denature for 10 sec at 95°C, 35 cycles consisting of 5 sec at 95°C and 20 sec at 60°C. The PCR products were separated in a 2% agarose gel in 1× TAE and visualized with ethidium bromide. In Experiment 2, the qRT-PCR was performed as described previously [12]. The Cₜ value was determined,
and the abundance of gene transcripts was calculated from the C\textsubscript{t} value by normalizing against $Hprt1$; $Hprt1$ expression is frequently used to evaluate expression level of gene of interest as a reference [28]. Expression of $Ucp1$, $Prdm16$ and leptin was examined in 5 WAT depots, whereas that of $Pgc1\alpha$, $Cidea$, $Dio2$, $Cox1$, $Cox7al$ and $Cox8b$ was done in the subcutaneous WAT and the mesenteric WAT. The expression in the roughage diet group in each WAT depot was set to 1, and the expression in the concentrate diet group was expressed as the value relative to that in the roughage diet group.

2.4. \textit{Statistical analyses}

Data are expressed as the mean ± SEM. Data were log-transformed to provide an approximation of a normal distribution before analysis. Differences between the dietary groups were examined using unpaired $t$-test. Differences of $P < 0.05$ were considered significant. A tendency to a difference was considered to be present when $0.05 \leq P < 0.10$.

3. Results

3.1. \textit{Expression of Ucp1 in WAT of fattening cattle}

Experiment 1 was performed as the first step to explore the presence of brite adipocytes in fattening cattle. Conventional RT-PCR was conducted to examine $Ucp1$ expression using total RNA prepared from the perirenal WAT depot of 31-month-old cattle (Fig. 1). A band with the expected size (235 bp) was reproducibly detected only in the sample treated with reverse transcriptase.

A representative result on immunohistochemical analyses to detect Ucp1 expressing cells is shown in Fig. 2. Ucp1-positive small adipocytes were scatteredly located
between white adipocytes in the subcutaneous WAT of 31-month-old cattle (Fig. 2).

3.2. Regulatory expression of Ucp1 in subcutaneous WAT related to diet

In Experiment 2, because the steers were pair-fed, intake of TDN and CP was comparable between the dietary groups [48]. In addition, body weight and daily weight gain were not different between the groups [48]. Furthermore, there were no differences in the weights of carcass bone, lean, and adipose tissue between the groups [48].

We evaluated diet-related changes in expression of Ucp1 in WAT of fattening cattle. Expression of Ucp1 in the subcutaneous WAT was significantly higher in the concentrate diet group than in the roughage diet group ($P = 0.01$, Fig. 3). In contrast, the expression in the other regions of WAT depots was not significantly different between the groups, although it tended to be higher in the concentrate diet group in the intramuscular WAT ($P = 0.09$).

Furthermore, gene expression levels of Pgc1α, Cidea, Dio2, Cox1, Cox7a1 and Cox8b were examined; these genes are involved in up-regulating Ucp1 expression and in activating brown adipocytes [35]. Consistent with increased expression of Ucp1, expression levels of the brown adipocyte-selective genes in the subcutaneous WAT were generally higher in the concentrate diet group than in the roughage diet group. Specifically, expression levels of Dio2 ($P = 0.03$), Cox1 ($P = 0.007$) and Cox8b ($P = 0.009$) were significantly higher in the concentrate diet group (Fig. 4). However, expression levels of the brown adipocyte-selective genes were comparable between the groups in the mesenteric WAT.

Prdm16 and leptin are molecules involved in the differentiation and activation of brown
adipocytes, respectively [7, 33]. Dietary effects on the expression of Prdm16 and leptin were similar to those on Ucp1 expression, as higher expression in the concentrate diet group was detected in the subcutaneous WAT depots ($P = 0.006$ and $P = 0.003$, respectively, Fig. 5). In addition, leptin expression in the intramuscular WAT depot was higher in the concentrate diet group than in the roughage diet group ($P = 0.02$, Fig. 5B).

4. Discussion

This study clarifies that 1) brite adipocytes are present in WAT depots of fattening cattle, 2) expression levels of Ucp1 is increased in response to feeding the concentrate diet, which is limited in the subcutaneous WAT, and 3) expression levels of brown adipocyte-selective genes are also increased in the subcutaneous WAT but not in the mesenteric WAT of steers fed the concentrate diet. In adult humans, the expression level of Ucp1 in WAT depots is affected by body mass index and insulin sensitivity, implicating that brite cell activity is modulated by metabolism [42]. The emergence of brite adipocytes is also affected by environmental and pharmacological factors [2, 8, 10], as cold acclimation or CL316,243, a β3-adrenergic receptor agonist [50], induced brown adipocytes in WAT [2, 17, 46]. Furthermore, brown adipocytes in WAT depots are also induced in genetically engineered mice, e.g., mice forced expression of Prdm16 in adipocytes [34]. This study indicates that diet can modulate expression of Ucp1 and brown adipocyte-selective genes, suggesting further physiological control over the emergence or activation of brite adipocytes.

The diet-related changes in expression levels of Pgc1α, Cidea, Dio2, Cox1, Cox7a1 and Cox8b in the subcutaneous WAT were basically similar to those of subcutaneous Ucp1 (Fig. 4); the expression levels in the concentrate diet group were generally higher than
those in the roughage diet group. Especially, the higher expression of \( \text{Dio2}, \text{Cox1} \) and \( \text{Cox8b} \) was statistically significant. All these gene products are involved in function of brown adipocytes as heat producing cells; in murine BAT, expression of Dio2, an enzyme catalyzing the reaction of thyroxine to bioactive triiodothyronine [3], is increased during cold exposure [36], and triiodothyronine stimulates thermogenesis in BAT through activation of sympathetic nervous system [21]. In addition, expression of Cox1, a cyclooxygenase, is up-regulated in the subcutaneous WAT depot in mice during cold exposure, and the increased expression was required for up-regulation of \( \text{Ucp1} \) expression and heat production [24]. Furthermore, consistent with the fact that brown adipocytes are rich in mitochondria [4], expression of \( \text{Cox8b} \), a component of mitochondrial cytochrome c oxidase, is higher in BAT depot than in WAT depot [35]. Taken higher \( \text{Ucp1} \) expression in the subcutaneous WAT of steers fed the concentrate diet with the results together, number of functional brown adipocytes in the subcutaneous WAT possibly increased in response to feeding the concentrate diet.

The present study revealed that the diet-related modulation of \( \text{Ucp1}, \text{Dio2}, \text{Cox1} \) and \( \text{Cox8b} \) expression was detected in the subcutaneous WAT but not in the mesenteric WAT. The cold-induced emergence of \( \text{Ucp1} \)-positive adipocytes was detected in the inguinal WAT (a representative subcutaneous WAT), but not in the parametrial WAT (a representative visceral WAT) in mice [2]. Treatment with Fgf21 induced \( \text{Ucp1} \) expression in the inguinal WAT but not in the epididymal WAT, another representative visceral WAT [10]. Thus, the more prominent regulation of brite adipocyte induction in the subcutaneous WAT than in the visceral WAT may be a common feature in mammals.

\( \text{Prdm16} \) and leptin are possibly involved in regulating the emergence of brite adipocytes.
Expression of Prdm16 in the subcutaneous WAT was higher in the concentrate diet group than in the roughage diet group, which coincided with the dietary effect on expression of Ucp1 and brown adipocyte-selective genes. Prdm16 is a transcriptional co-regulator that stimulates the development of brown adipocytes [19, 33, 34]. Treatment with leptin stimulated energy expenditure [16] and expression of Ucp1 in BAT as well as WAT depots [7] in leptin-deficient ob/ob mice. Leptin-induced up-regulation of the Ucp1 expression was reduced in melanocortin receptor 4-deficient mice [51]. Furthermore, leptin increased the production of α-melanocyte-stimulating hormone in the hypothalamus, indicating the involvement of the leptin-melanocortin pathway in activating brown adipocytes [15]. Thus, at least two reasons are possible for the concentrate diet-induced Ucp1 expression in the subcutaneous WAT: 1) brown adipocyte differentiation is locally stimulated through up-regulation of Prdm16 expression, and 2) enhanced expression of leptin, which is produced in and secreted from white adipocytes [43], systemically acts as a stimulator of brown adipocyte activation.

Dispersed brite adipocytes in large subcutaneous WAT masses are suggested to cumulatively represent substantial brown adipocyte activity in adult humans, although they are not detected by the integrated PET-CT method [42]. Expression levels of Ucp1 in the subcutaneous WAT were significantly affected by diet, but body weight gain and body composition in cattle used in this study were comparable between the groups [48]. We postulate that the concentrate diet-induced activation of brite adipocytes is masked by the enhanced anabolic activity in cattle. In support of this, plasma concentrations of insulin and Igf-1, both with potent anabolic effects [6], linearly increased with concentrate proportions in cattle feed under the condition of equal energy ingestion between groups, suggesting an intrinsic anabolic effect of the concentrate diet [41].
Thus, suppressing the induction of brite adipocytes in fattening cattle maintained on the concentrate diet may improve the fattening efficiency significantly.

Here we show the presence of brown adipocytes in WAT depots of fattening cattle based upon the expression of brown adipocyte-selective genes, and the regulatory expression of the genes in response to diet. The present results point out a novel possibility on the concentrate diet-specific emergence or activation of the subcutaneous brite adipocytes. To uncover the precise role of brown adipocytes in WAT depots in relation to the diet, further studies are needed to clarify 1) the molecular bases of the concentrate diet-induced up-regulation of Prdm16 and leptin expression, 2) a detailed explanation as to why the diet-induced modulation of Ucp1 and Prdm16 expression occurs in the subcutaneous WAT but not in the visceral WAT, and 3) the physiological significance of the induced brown adipocytes.

Acknowledgements
This work was supported by a Grant-in-Aid for Scientific Research (23580368) from The Japan Society for the Promotion of Science.

References


Metabolism and morphology of brown adipose tissue from Brahman and Angus newborn calves.


[26] G.S. Martin, D.K. Lunt, K.G. Britain, S.B. Smith, Postnatal development of


[48] T. Yamada, N. Nakanishi, Effects of the roughage/concentrate ratio on the
expression of angiogenic growth factors in adipose tissue of fattening Wagyu steers.

[49] X. Yang, S. Enerbäck, U. Smith, Reduced expression of FOXC2 and brown
1182-1191.

[50] T. Yoshida, N. Sakane, Y. Wakabayashi, T. Umekawa, M. Kondo, Anti-obesity and
anti-diabetic effects of CL 316,243, a highly specific β3-adrenocceptor agonist, in

Bannon, R.L. Mynatt, T.W. Gettys, Targeted deletion of melanocortin receptor
subtypes 3 and 4, but not CART, alters nutrient partitioning and compromises
Figure legends

Fig. 1. Expression of Ucp1 in perirenal WAT depot of fattening cattle
Total RNA was isolated from the perirenal WAT depot of cattle aged 31 months, and
cDNA was prepared by treatment with (RT+) or without (RT-) reverse transcriptase.
PCR was performed to detect Ucp1. The PCR products were electrophoresed in agarose
gels, followed by staining with ethidium bromide. A representative result is shown.

Fig. 2. Immunolocalization of Ucp1 in subcutaneous WAT depot of fattening cattle
The subcutaneous WAT from cattle 31 months of age was fixed and sectioned, followed
by immunochemical analyses to determine localization of Ucp1. Bar: 100 μm.

Fig. 3. Diet-related changes in Ucp1 expression in WAT depots of fattening cattle
Fattening cattle were fed either the roughage diet or the concentrate diet for 20 months.
At 30 months of age, subcutaneous (sc), mesenteric (mesen), perirenal (pr),
intermuscular (inter) and intramuscular (intra) WAT depots were collected, and Ucp1
expression was examined by qRT-PCR. Expression of Ucp1 was normalized to that of
Hprt1, and the expression in the roughage diet group in each WAT depot was set to 1.
Data are shown as the mean ± SE (n = 4). *: P < 0.05.

Fig. 4. Diet-related changes in expression of brown adipocyte-selective genes in WAT depots of fattening cattle
Fattening cattle were fed either the roughage diet or the concentrate diet for 20 months.
Expression levels of brown adipocyte-selective genes in the subcutaneous WAT and the
mesenteric WAT were examined by qRT-PCR. The expression was normalized to
*Hprt1* expression, and the expression in the roughage diet group in each WAT depot was set to 1. Data are shown as the mean ± SE (n = 4). * and **: *P* < 0.05 and *P* < 0.01, respectively.

Fig. 5. Diet-related expression of Prdm16 and leptin in WAT depots of fattening cattle

Fattening cattle were fed either the roughage diet or the concentrate diet for 20 months. At 30 months of age, subcutaneous (sc), mesenteric (mesen), perirenal (pr), intermuscular (inter) and intramuscular (intra) WAT depots were collected, and expression of *Prdm16* (A) and *leptin* (B) was examined by qRT-PCR. The expression was normalized to *Hprt1* expression, and the expression in the roughage diet group in each WAT depot was set to 1. Data are shown as the mean ± SE (n = 4). * and **: *P* < 0.05 and *P* < 0.01, respectively.
Table 1. Oligonucleotide PCR primers for conventional RT-PCR and quantitative RT-PCR

<table>
<thead>
<tr>
<th>Oligonucleotide</th>
<th>Conventional RT-PCR</th>
<th>Quantitative RT-PCR</th>
<th>GenBank accession number</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>5'-primer</td>
<td>3'-primer</td>
<td></td>
</tr>
<tr>
<td>Conventional RT-PCR</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ucp1</td>
<td>5'-agggactactcccaatcgaca-3’</td>
<td>5'-gttgggcacacctgtgtgtact-3’</td>
<td>XM_003587124</td>
</tr>
<tr>
<td>Cidea</td>
<td>5'-agcaagacctttggtccact-3’</td>
<td>5'-gaactctctctggtctccaccac-3’</td>
<td>NM_001083449</td>
</tr>
<tr>
<td>Coxl</td>
<td>5'-gtttctgctgctgetcc-3’</td>
<td>5'-gggcaagaaggcaacact-3’</td>
<td>NM_001105323</td>
</tr>
<tr>
<td>Cox7a1</td>
<td>5'-cggagaaccgagtagctgaa-3’</td>
<td>5'-atagcaggttctgctgac-3’</td>
<td>NM_176674</td>
</tr>
<tr>
<td>Coxb8</td>
<td>5'-ccgactccacacatgcc-3’</td>
<td>5'-aagctcacagagcccaat-3’</td>
<td>NM_001114517</td>
</tr>
<tr>
<td>Dio2</td>
<td>5'-atgcacacctctgcttc-3’</td>
<td>5'-ggcagctgttagctgaagg-3’</td>
<td>NM_001010992</td>
</tr>
<tr>
<td>Hprt1</td>
<td>5'-gtgattagctgtggaacc-3’</td>
<td>5'-ccatgaggaataaacctcttc-3’</td>
<td>NM_001034035</td>
</tr>
<tr>
<td>Leptin</td>
<td>5'-acatctcacaagcagac-3’</td>
<td>5'-ggcatgaaatccacccagta-3’</td>
<td>NM_173928</td>
</tr>
<tr>
<td>Pgc1α</td>
<td>5'-acctccattttggactccag-3’</td>
<td>5'-agcgcggctatatctcga-3’</td>
<td>NM_177945</td>
</tr>
<tr>
<td>Prdm16</td>
<td>5'-gagggctgctcatccaaag-3’</td>
<td>5'-agcatcaccacagcttc-3’</td>
<td>XM_001788152</td>
</tr>
<tr>
<td>Ucp1</td>
<td>5'-ctggtggtgctgatataca-3’</td>
<td>5'-tggtggtgatgctcaat-3’</td>
<td>XM_003587124</td>
</tr>
</tbody>
</table>
Cattle no: 1394 1398
RT: + - + -
A Self-archived copy in Kyoto University Research Information Repository
https://repository.kulib.kyoto-u.ac.jp
**Figure 3**

A Self-archived copy in Kyoto University Research Information Repository
https://repository.kulib.kyoto-u.ac.jp

**Ucp1**

![Bar chart showing relative mRNA levels of Ucp1 in different tissues: sc, mesen, pr, inter, intra. The chart compares Roughage and Concentrate conditions with error bars.]
Figure 4

A Self-archived copy in
Kyoto University Research Information Repository
https://repository.kulib.kyoto-u.ac.jp
Figure 5

A) **Prdm16**

B) **Leptin**

A Self-archived copy in Kyoto University Research Information Repository
https://repository.kulib.kyoto-u.ac.jp