

TNF α , PDGF and TGF β synergistically induce synovial lining hyperplasia via inducible PI3K δ

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ABSTRACT

Objectives To determine the mechanism underlying hypertrophic synovium in rheumatoid arthritis (RA).

Methods We examined micromass cultures of fibroblast-like synoviocytes (FLSs) stimulated with tumor necrosis factor α (TNF α), platelet-derived growth factor (PDGF), and/or transforming growth factor β (TGF β). The hypertrophic architecture of the micromasses, expression of phosphoinositide 3 kinase (PI3K) isoforms, and persistent activation of PI3K-Akt pathways were investigated. FLSs transfected with siRNA were also examined in the micromass cultures.

Results The combination of TNF α , PDGF and TGF β (TPT condition) induced obvious hypertrophic architecture of the intimal lining layer in FLSs in micromass cultures, and was accompanied by upregulated expression of matrix metalloproteinase-3 (MMP3), Cadherin-11, and PI3K δ . In monolayer FLSs, the TPT condition enhanced the expression of PI3K δ and persistent activation of the PI3K-Akt pathway. Knockdown of PI3K δ significantly inhibited the formation of the hypertrophic synovial lining in the TPT condition.

Conclusions These results collectively indicate that inducible PI3K δ plays a crucial role in persistent activation of PI3K-Akt in FLSs, and in the formation of a hypertrophic synovial lining. PI3K δ may be an alternative treatment target for the regulation of proliferative synovium in RA.

Keywords:

Fibroblast-like synoviocytes

PI3K/Akt pathway

PI3K δ

PDGF

TGF β

Introduction

Synovial hyperplasia refers to rheumatoid arthritis (RA) lesions responsible for cartilage degeneration, osteoclast induction, and subsequent joint deformities [1].

Clinically, the swollen joint count of patients with RA is associated with the progression of joint damage in RA more than systematic inflammation is [2]. Although there are various types of cells in RA synovium and joints, including fibroblast-like synoviocytes (FLSs), macrophages, T cells, B cells, dendritic cells, neutrophils, and osteoclasts, and while each is considered crucial for RA pathogenesis, FLSs play a key role in synovial hyperplasia by constructing the main framework of RA synovium [1,3,4,5].

Platelet-derived growth factor (PDGF) and transforming growth factor β (TGF β) are abundant in the synovium and synovial fluid in RA [6,7,8]. PDGF is considered to be involved in the pathogenesis of RA as a growth factor of FLSs [7,8]. On the other hand, TGF β is more complicated; it has an inhibitory effect in lymphocytes and macrophages [9], but induces proliferation and aggrecanase production in FLSs [10,11]. The combination of PDGF and TGF β synergistically activates extracellular signal-regulated kinase and phosphoinositide 3 kinase (PI3K) pathways of FLSs and

enhances the production of inflammatory mediators in monolayer FLSs [12].

Monolayer culture or invasion assays of FLSs are frequently used but have difficulty in the analysis of factors relating to hypertrophic architecture formed by FLSs. Kiener et al. showed that a micromass culture of human FLSs formed lining/sublining architecture resembling synovial tissues, and found that TNF α and PDGF contributed to the hypertrophic architecture of intimal lining layer to some extent [13]. However, further mechanisms underlying the hypertrophy of intimal lining driven by FLSs remain to be determined.

In this study, we show that the combination of TNF α , PDGF, and TGF β synergistically induces obvious hypertrophic architecture in FLS micromass culture, and that inducible PI3K δ plays a key role in the persistent activation of the PI3K-Akt pathway, and in the formation of hypertrophic FLS architecture.

Materials and Methods

Isolation of fibroblast-like synoviocytes (FLSs)

Ethical approval for using human samples for this study was granted by the Ethics Committee of Kyoto University Graduate School and Faculty of Medicine. Written informed consent was obtained from all study participants. RA was diagnosed in accordance with the criteria of the American College of Rheumatology [14]. FLSs from patients with RA were prepared and cultured in DMEM (Sigma-Aldrich, St. Louis, MO) supplemented with penicillin (Sigma), streptomycin and 10% FBS (Sigma), as previously described [15].

Micromass culture of FLSs

Micromass cultures of FLSs were prepared, using a process previously reported [13]. Briefly, FLSs were suspended in ice-cold Matrigel Matrix (BD Biosciences, Bedford, MA) at a density of 5×10^6 cells/ml. Droplets (40 μ l) of the suspension were placed onto poly-2-hydroxyethylmethacrylate (poly-HEMA; Sigma)-coated culture dishes. Gelation was allowed to occur for 45 min at 37°C. The gel was then overlaid with culture medium. The floating 3-D culture was maintained for 2–3 weeks with a medium change twice a week.

For stimulation experiments, FLS micromasses were cultured in basal medium containing 10 ng/ml of tumor necrosis factor α (TNF α), 10 ng/ml of platelet-derived growth factor (PDGF), and/or 10 ng/ml of transforming growth factor β (TGF β) (each from Pepro Tech; Rocky Hill, NJ).

Histological analysis

Micromasses were processed as 20- μ m-thick frozen sections and stained with hematoxylin and eosin. Light microscopic images were taken using ECLIPSE 80i (Nikon, Japan) with a 10 \times dry lens. The area of the intimal lining layer, including villous structures intruding toward the center of the micromass, was measured using ImageJ (NIH).

Cell proliferation assay

FLSs at 50% confluent were cultured for 5 days with or without indicated cytokines. Cell proliferation was measured with CellTiter 96® AQueous One Solution Cell Proliferation Assay (Promega) according to manufacturer's protocol.

Quantitative real-time PCR (qRT-PCR)

Reverse transcription was performed using a Transcriptor Universal cDNA Master (Roche Applied Science, Indianapolis, IN). qRT-PCR was performed using FastStart Universal SYBR Green Master (Roche Applied Science) on an Applied Biosystems 7500 Thermocycler according to the manufacturer's protocol. The following primer sequences were used: *GAPDH*: forward, 5'-TCTCGCTCCTGGAAGATGGT-3'; reverse, 5'-GGAAGGTGAAGGTCGGAGTC-3'; *MMP3*: forward, 5'-GATACCCAAGAGGCATCCAC-3'; reverse, 5'-CTGGCTCCATGGAATTTCTC-3'; *Cadherin11*: forward, 5'-AGAGAGCCCAGTACACGTTGA-3'; reverse, 5'-TTGGCATGATAGGTCTCGTGC-3'; *PI3K α* : forward, 5'-CCACGACCATCATCAGGTGAA-3'; reverse, 5'-CCTCACGGAGGCATTCTAAAGT-3'; *PI3K β* : forward, 5'-AGAGCACTTGGTAATCGGAGG-3'; reverse, 5'-CTTCCCCGGCAGTATGCTTC-3'; *PI3K γ* : forward, 5'-GGCGAAACGCCCATCAAAA-3'; reverse, 5'-GACTCCCGTGCAGTCATCC-3'; and *PI3K δ* : forward, 5'-AAGGAGGAGAATCAGAGCGTT-3'; and reverse, 5'-GAAGAGCGGCTCATACTGGG-3'. The expression of each mRNA was normalized by the expression of GAPDH.

Western blot analysis

Western blot analysis was performed as previously described [1]. Briefly, 10–20 ng of cell lysate was subjected to 10% SDS-PAGE and transferred onto a nitrocellulose membrane (Schleicher & Schuell, Dassel, Germany). After blocking with 1% skim milk or 2% BSA, the membrane was probed with rabbit anti- β -tubulin (clone 11H10), rabbit anti-Akt (clone 11E7), or rabbit anti-phospho-Akt (Ser473: clone D9E) (all purchased from Cell Signaling Technology; Beverly, MA), incubated with horseradish protein-conjugated goat anti-rabbit IgG (Cayman Chemical, Ann Arbor, MI), and visualized using Pierce ECL Western Blotting Substrate (Thermo Fisher Scientific, Rockford, IL), ECL Plus Western Blotting Detection Reagent (GE Healthcare UK, Buckinghamshire, UK), or ECL Prime Western Blotting Detection Reagent (GE Healthcare UK), as appropriate.

Cell-based enzyme-linked immunosorbent assay (ELISA)

Cell ELISA was performed as previously described [3]. Briefly, FLSs were cultured for 24 h, stimulated with cytokines, fixed with 4% paraformaldehyde, permeabilized with TBS containing 0.001% Triton X, quenched with PBS containing 0.1 M glycine and 1% H₂O₂, and blocked with TBS with 2% BSA solution, incubated with primary antibody as for Western blottings, incubated with horseradish protein-conjugated goat anti-rabbit IgG, and reacted with a substrate solution containing 3,3',5,5'-tetramethylbenzidine (Sigma). The reaction was stopped with 2 N sulfuric acid. The optical density was measured at 450–630 nm with a microplate reader. Linearity of ELISA was confirmed using serially diluted samples.

siRNA

FLSs were transfected with double-stranded siRNA (Stealth RNAi; Life Technologies) for 48 h using a TransIT-TKO transfection reagent (Mirus) as per the manufacturer's protocol. The siRNA sequences are as follows; siCTL: sense, 5'-AAGUGAAUGUGCGUUUGGAACGUGC-3'; antisense, 5'-GCACGUUCCAAACGCACAUUCACUU-3'; siPI3K δ , 5'-GGGCCAACCTCATGCTGTTTGACTA-3'; and antisense, 5'-TAGTVAAAVAGCATGAGGTTGGCCC-3'.

Results

The combination of TNF α , PDGF, and TGF β induces hypertrophy of the intimal lining layer in micromass cultures of FLSs.

Hypertrophy of synovial tissue, especially in the lining layer, is characteristic of lesions observed in RA patients. To determine whether the combination of PDGF and TGF β has a synergistic effect in the formation of hypertrophic synovium, we conducted micromass culture of human FLSs under stimulation with TNF α , PDGF, and/or TGF β . Micromass cultures in a normal medium showed lining/sublining architecture, as previously reported (Fig. 1A). The individual use of TNF α , PDGF and TGF β in the culture medium at a concentration of 10 ng/ml slightly thickened the intimal lining layer (Fig. 1B, C and D), and higher concentrations (100 ng/ml) of each cytokine had a similar effect as when stimulating at a concentration of 10 ng/ml (data not shown). Interestingly, PDGF or TGF β frequently showed villous architecture of the intimal lining intruding toward the center of FLS micromass cultures (Fig. 1C, D and G). The addition of TNF α to PDGF or TGF β (Fig. 1E and F) did not enhance the hypertrophy

effect of PDGF or TGF β , whereas the combination of TNF α , PDGF, and TGF β (TPT condition) significantly increased the thickness of the intimal lining and villus formation (Fig. 1H and I). Thus, the combination of TNF α , PDGF, and TGF β , each present abundantly in the synovial fluid, induced the hyperplastic architecture of FLSs.

When monolayer FLSs were cultured with these cytokine conditions, each cytokine stimulation significantly enhanced cell proliferation. Consistent with the result of micromass culture, TPT condition enhanced the proliferation of monolayer FLSs most (Fig. 1J).

PI3K δ was upregulated in FLS micromasses cultured in the TPT condition.

In the hyperplastic synovium of RA, various molecules are elevated and contribute to the pathogenesis of RA. Metalloproteinases, including MMP3, contribute to cartilage erosion and tendon rupture by degenerating the collagen matrix [17-19]. Cadherin-11 plays a crucial role in synovial structure by supporting cell-to-cell junctions. In the micromass organ culture, the expression of MMP3 was apparently correlated with the addition of TNF α (Fig. 2A), whereas Cadherin-11 was expressed most in the TPT condition (Fig. 2B). The elevation of PI3K δ is also reported in RA synovium [19]. Among the isoforms of PI3K, the expression of PI3K δ significantly increased in micromass culture under the TPT condition (Fig. 2C-E), whereas PI3K γ was undetectable in almost all samples (data not shown). Thus, micromass culture of FLSs accompanying hypertrophic architecture showed the upregulation of molecules characteristic of RA hyperplastic synovial tissues.

TPT condition enhanced the expression of PI3K δ and persistently activated the PI3K-Akt pathway in monolayer FLSs.

To further determine the signal pathway relating to the TPT condition using monolayer culture of FLSs, we investigated the similarity between monolayer and micromass cultures of FLSs at first. Stimulation of monolayer FLSs showed an expression pattern of PI3K isoforms and MMP3 similar to that of micromass cultures of FLSs; the expression of PI3K δ and MMP3 was significantly increased by the stimulation of the TPT condition (Fig. 3A and Supplementary Figure 1), whereas stimulation of monolayer FLSs with the TPT condition failed to enhance the expression of Cadherin-11 (Supplementary Figure 2). Taken together with the result of proliferation assay (Fig. 1J), monolayer cultures of FLSs apparently reflect the phenomena of micromass cultures to some extent. At least, considering similar expression pattern of PI3K δ between monolayer and micromass cultures, it is reasonable to investigate the PI3K-Akt pathway using monolayer FLSs .

We stimulated monolayer FLSs with the TPT condition and investigated the activation of the PI3K-Akt pathway. As previously reported, TNF α did not enhance the phosphorylation of Akt, whereas the other conditions, including PDGF alone, the combination of TNF α and PDGF, and the TPT condition, each similarly increased phosphorylated Akt (P-Akt) 6 h after the stimulation (Fig. 3B). The TPT condition also increased the amount of P-Akt 24 h after the stimulation compared with other conditions (Fig. 3B). The time course of P-Akt, measured by cell-ELISA, showed that its increase under stimulation with PDGF was similar to that under stimulation with TNF α and PDGF together, and with the TPT condition for the first 6 h. In the subsequent 18 h, the amount of P-Akt under the condition of PDGF, and with the

combination of PDGF and TNF α , gradually decreased, but continued to increase in the TPT condition (Fig. 3C). The amount of P-Akt in the TPT condition was significantly higher than that of other conditions at both 24 h and 48 h after stimulation (Fig. 3D and E). These data indicate that stimulation of monolayer FLSs with the TPT condition enhances PI3K δ and persistently activates the PI3K-Akt pathway.

Knockdown of PI3K δ in monolayer FLSs attenuated the activation of the PI3K-Akt pathway induced by the TPT condition.

To determine the involvement of PI3K δ in the activation of the PI3K-Akt pathway, we knocked down the expression of PI3K δ . First, we confirmed that transfection of siPI3K to monolayer FLSs specifically downregulated the mRNA of PI3K δ but not that of PI3K α , nor PI3K β (Fig. 4A). PI3K δ siRNA significantly attenuated the proliferation of monolayer FLSs (Fig. 4B) and the phosphorylation of Akt induced by the TPT condition at each time point during 48 h (Fig. 4C). These results indicate that inducible PI3K δ of FLSs plays a crucial role in FLS proliferation and in the persistent activation of the PI3K-Akt pathway induced by the TPT condition.

PI3K δ is involved in hyperplastic synovial lining.

We have shown that the TPT condition induces hypertrophy of the intimal lining layer and also enhances the expression of PI3K δ in Figures 1 and 2. To determine whether PI3K δ is functionally involved in the formation of hypertrophic synovial lining, PI3K δ -knocked down FLSs were conducted 3-D culture under the TPT condition. We confirmed that the knockdown of PI3K δ persisted for at least 10 days (data not shown).

Knockdown of PI3K δ significantly inhibited the thickening of the intimal lining layer induced by the TPT condition (Fig. 4D and E). Thus, these results collectively suggest that the TPT condition induces hypertrophic architecture in synovium by elevating the expression of PI3K δ , and then activating the PI3K-Akt pathways.

Discussion

Clinical reports showed that synovial hyperplasia of RA contributed to the deformation of joints [2]. However, there is no clear understanding of how FLSs, which consist of a main framework of synovium, are regulated to lead to hyperplastic synovium in RA. In this study, we have shown that the combination of TNF α , PDGF, and TGF β synergistically induced obvious hypertrophic architecture in FLS micromass cultures accompanying upregulation of PI3K δ expression, and that inducible PI3K δ plays a crucial role in the activation of the PI3K-Akt pathway and in the hypertrophy of the FLS intimal lining.

A normal synovium is an acellular structure with a thin intimal lining. In RA, a massive increase of macrophage-like synoviocytes and FLSs accompanies the infiltration of T cells and B cells to lead to hyperplastic RA synovium. Interestingly, micromass cultures of FLSs form lining/sublining architecture similar to normal synovial architecture, whereas skin fibroblasts form a solid structure. This indicates that FLSs, by their nature, play a crucial role in forming architectural structures of synovial tissue [13]. Furthermore, the addition of cytokines to micromass cultures induced hypertrophic architecture. Thus, micromass cultures of FLSs are suitable experimental systems for determining the factors affecting FLSs to induce hyperplastic architecture.

TGF β , TNF α , and PDGF are abundantly present in the RA synovial environment [20-22]. TGF β is a kind of growth factor, but unlike PDGF, it has inhibitory effects and proliferative effects depending on the target cells [9-11,23]. In monolayer FLSs, the combination of PDGF and TGF β synergistically enhances the production of MMP3, IL-8, and MIP1a induced by TNF α [12], and activates the PI3K-Akt pathway [24]. In addition, we have shown that this combination also has a synergistic effect in the thickening of the synovial lining. Long-lasting activation of the PI3K-Akt pathway (Fig. 3C) might contribute to the architectural change in micromasses of FLSs.

PI3K has constitutive isoforms of PI3K α and PI3K β , and inducible isoforms of PI3K γ and PI3K δ . A deficiency of the constitutive isoforms, PI3K α or PI3K β , is embryonic lethal [25,26], implying the difficulty of treating RA by targeting these molecules. On the other hand, PI3K δ is preferentially expressed by lymphoid cells [27] and RA hypertrophic synovium [19], and a deficiency of PI3K δ does not affect the lifespan, and ameliorates arthritis in mice [28,29]. Although B cells were considered to be the most responsible cells for this amelioration, our study showed that inducible PI3K δ of FLSs is also involved in the activation of the PI3K-Akt pathway and synovial lining hyperplasia (Fig. 4B and 4C).

In this study, we addressed the signaling pathway of TNF α , PDGF and TGF β using monolayer FLSs. MMP3 and PI3K δ showed similar expression in both monolayer and micromass cultures, suggesting that the analysis of the PI3K-Akt pathway using monolayer FLSs is reasonable. On the other hand, Cadherin-11 failed to increase in monolayer FLSs under TPT condition. The discrepancy in Cadherin-11 might be partly

attributed to a correlation between Cadherin-11 and adherens junction at the site of cell-cell contact [30]. The TPT condition enhanced the proliferation of monolayer FLSs most (Fig. 1J), whereas the difference of proliferation between cytokine conditions was not so obvious as expected from the results of micromass cultures (Fig. 1I). Furthermore, TNF α apparently contributes most to the proliferation of FLS compared to PDGF and TGF β (Fig. 1J), whereas least to synovial hyperplasia (Fig. 1I). This indicates that micromass cultures have different aspect from monolayer culture and may have an advantage in the analysis of synovial hyperplasia compared to monolayer culture or invasion assays, and that factors other than cell proliferation may be involved in the synovial hyperplasia.

This study has several limitations. The factors connecting PI3K-Akt pathway and the synovial hyperplasia remain to be determined. As reported in tumor cells, the enhancement of adhesion via PI3K-Akt pathway might contribute to the synovial hyperplasia in TPT condition [31,32]. The influence of other cytokines, such as IL-1 β or IL-6, on synovial lining hyperplasia was not investigated. The mechanism underlying the upregulation of PI3K δ in the TPT condition also remains to be determined. However, we have shown that the TPT condition clearly contributes to an architectural change in the synovial lining layer via inducible PI3K δ , possibly by persistent activation of the PI3K-Akt pathway using FLS micromass culture. Further clarification of the mechanism by which FLSs regulate the 3-D architecture should lead to an alternative treatment for RA.

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Conflict of interest None.

Figure legends

Fig. 1. Micromass cultures of RA FLSs with cytokine stimulation. **A–H**, The frozen sections of micromass were stained with hematoxylin and eosin. Human FLS micromasses were cultured for 21 days in the presence of basal medium (A), with TNF α (B), TGF β (C), PDGF (D), PDGF and TNF α (E), TNF α and TGF β (F), PDGF and TGF β (G), and a combination of TNF α , PDGF and TGF β (TPT condition) (H). The concentration of each cytokine was 10 ng/ml. Insets indicate scaled-up images of intimal linings. Open and closed arrowheads indicate villous architecture and the border between the thickening lining layer and sublining layer, respectively. **I**, Percentages of area for intimal linings in total area of a micromass. **J**, Cell proliferation of monolayer FLSs for 5 days under the conditions as in A–H. Data are shown as mean \pm SEM of sextuplet. * = $P < 0.05$ in one-way ANOVA.

Fig. 2. Expression of markers relating to RA synovium in FLS micromasses under the TPT condition. **A–E**, Expression of MMP3 (A), Cadherin-11 (B), and PI3K isoforms (C–E), in micromass culture of RA FLSs in response to TNF α , TGF β , and PDGF. Quantitative PCR was performed to quantify mRNA levels in micromass cultures. Data are normalized to GAPDH. Expression of PI3K γ was not detectable (not shown).

MMP3, Caherin-11, and PI3K δ were expressed most in the TPT condition. PI3K α and β were not affected by stimulation. Data are shown as mean \pm SEM of sextuplet. * = $P < 0.05$ in one-way ANOVA.

Fig. 3. Analysis of PI3K-Akt pathway in monolayer FLSs under the TPT condition. **A**, Expression of PI3K δ mRNA in FLSs under the indicated condition for 24 h. Data are shown as mean \pm SEM of quadrants. * = $P < 0.05$ in one-way ANOVA. **B**, Expression of phospho-Akt (P-Akt), total-Akt (T-Akt), and β -tubulin protein in FLSs cultured under indicated conditions for 6 h or 24 h was analyzed by Western blot assay. **C–E**, Expression of P-Akt and T-Akt was measured by ELISA. Relative P-Akt expression during 48 h after cytokine stimulation (C), and at 24 h (D), and 48 h (E), are shown. Data are shown as mean \pm SEM of triplicate. * = $P < 0.05$ in two-way ANOVA.

Fig. 4. Analysis of PI3K-Akt pathway and formation of hyperplastic architecture in PI3K δ -knocked down FLSs. **A**, Expression of PI3K isoforms in monolayer FLSs after transfection with PI3K δ siRNA or control siRNA. RA FLSs were incubated for 24 h, then transfected for 48 h. Cells were cultured in basal medium for 4 days, then mRNA was isolated. Data are shown as mean \pm SEM of sextuplet. * = $P < 0.05$ in Student's t-test. **B**, Proliferation of PI3K δ -knocked down or control FLSs cultured in monolayer for 5 days. Data are shown as mean \pm SEM of quintuplets. * = $P < 0.05$ in Student's t-test. **C**, The relative expression of phospho-Akt in monolayer transfected FLSs under the TPT condition was measured by cell ELISA. Data are shown as mean \pm SEM of sextuplet. * = $P < 0.05$ in two-way ANOVA. **D and E**, Micromasses of FLSs transfected

with siPI3K δ or siCTL were cultured in the TPT condition for 14 days. Hematoxylin and eosin stain of micromasses (D) and the percentages of lining layer area (E) are shown. Data are shown as mean \pm SEM of sextuplet. * = $P < 0.05$ in Student's t-test.

References

1. G.S. Firestein, Evolving concepts of rheumatoid arthritis. *Nature*. 2003;423:356-61.
2. D. Aletaha, F. Alasti, J.S. Smolen, Rheumatoid arthritis near remission: clinical rather than laboratory inflammation is associated with radiographic progression. *Ann. Rheum. Dis.* 2011;70:1975–80.
3. K. Murata, H. Yoshitomi, M. Furu, M. Ishikawa, H. Shibuya, H. Ito, S. Matsuda, miR-451 downregulates neutrophil chemotaxis via p38 mitogen-activated protein kinase. *Arthritis Rheum.* DOI: 10.1002/art.38269
4. S. Kobayashi, K. Murata, H. Shibuya, M. Morita, M. Ishikawa, M. Furu, et al., A Distinct Human CD4+ T Cell Subset That Secretes CXCL13 in Rheumatoid Synovium. *Arthritis Rheum.* 2013;65:3063–72.
5. S. Kubo, K. Yamaoka, M. Kondo, K. Yamagata, J. Zhao, S. Iwata, Y. Tanaka, The JAK inhibitor, tofacitinib, reduces the T cell stimulatory capacity of human monocyte-derived dendritic cells. *Ann. Rheum. Dis.* DOI: 10.1136/annrheumdis-2013-203756.
6. R. Lafyatis, N.L. Thompson, E.F. Remmers, K.C. Flanders, N.S. Roche, S.J. Kim, Transforming growth factor-beta production by synovial tissues from rheumatoid patients and streptococcal cell. *J. Immunol.* 1989;143:1142-8.
7. E.F. Remmers, H. Sano, R.L. Wilder, Platelet-derived growth factors and heparin-binding (fibroblast) growth factors in the synovial tissue pathology of rheumatoid arthritis. *Arthritis Rheum.* 1991;21:191–9.
8. S.C. Thornton, S.B. Por, R. Penny, M. Richter, L. Shelley, S.N. Breit, Identification of the major

- fibroblast growth factors released spontaneously in inflammatory arthritis as platelet derived growth factor and tumour necrosis factor-alpha. *Clin. Exp. Immunol.* 1991; 86:79–86.
9. M.O. Li, Y.Y. Wan, S. Sanjabi, A.K.L. Robertson, R.A. Flavell, Transforming growth factor-beta regulation of immune responses. *Ann. Rev. Immunol.* 2006;24: 99–146.
 10. S.M. Wahl, J.B. Allen, H.L. Wong, S.F. Dougherty, L.R. Ellingsworth, Antagonistic and agonistic effects of transforming growth factor-beta and IL-1 in rheumatoid synovium. *J. Immunol.* 1990;145:2514–9.
 11. Y. Yamanishi, D.L. Boyle, M. Clark, R.A. Maki, M.D. Tortorella, E.C. Arner, et al., Expression and regulation of aggrecanase in arthritis: the role of TGF-beta. *J. Immunol.* 2002;168:1405–12.
 12. S. Rosengren, M. Corr, D.L. Boyle, Platelet-derived growth factor and transforming growth factor beta synergistically potentiate inflammatory mediator synthesis by fibroblast-like synoviocytes. *Arthritis Res. Ther.* 2010;12:R65.
 13. H.P. Kiener, G.F.M. Watts, Y. Cui, J. Wright, T.S. Thornhill, M. Sköld, et al., Synovial fibroblasts self-direct multicellular lining architecture and synthetic function in three-dimensional organ culture. *Arthritis Rheum.* 2010;62:742–52.
 14. Arnett FC, Edworthy SM, Bloch DA, McShane DJ, Fries JF, Cooper NS, et al., The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum.* 1988;31:315-24.
 15. S. Tanida, H. Yoshitomi, M. Ishikawa, T. Kasahara, K. Murata, H. Shibuya, et al., IL-27-producing CD14(+) cells infiltrate inflamed joints of rheumatoid arthritis and regulate inflammation and chemotactic migration. *Cytokine.* 2011;55: 237–44.
 16. K. Nishitani, H. Ito, T. Hiramitsu, R. Tsutsumi, S. Tanida, T. Kitaori, et al., PGE2 inhibits MMP expression by suppressing MKK4-JNK MAP kinase-c-JUN pathway via EP4 in human articular chondrocytes. *J. Cell. Biochem.* 2010;109:425–33.
 17. P.P. Minghetti, W.D. Blackburn, Effects of sulfasalazine and its metabolites on steady state messenger RNA concentrations for inflammatory cytokines, matrix metalloproteinases, and tissue inhibitors of metalloproteinase in rheumatoid synovial fibroblasts. *J. Rheumatol.*

- 2000;27:653-60.
18. D.M. Lee, H.P. Kiener, S.K. Agarwal, E.H. Noss, G.F.M. Watts, O. Chisaka, et al., Cadherin-11 in synovial lining formation and pathology in arthritis. *Science* 2007;315:1006–10.
 19. B. Bartok, D.L. Boyle, Y. Liu, P. Ren, S.T. Ball, W.D. Bugbee, et al., PI3 kinase δ is a key regulator of synoviocyte function in rheumatoid arthritis. *Am. J. Pathol.* 2012;180: 1906–16.
 20. H.L. Wright, R.C. Bucknall, R.J. Moots, S.W. Edwards, Analysis of SF and plasma cytokines provides insights into the mechanisms of inflammatory arthritis and may predict response to therapy. *Rheumatology (Oxford, England)*. 2012;51: 451–9.
 21. M. Lotz, D.A. Carson, J. Kekow, Transforming growth factor-beta and cellular immune responses in synovial fluids. *J. Immunol.* 1990;144:4189–4194.
 22. S. Monier, T. Reme, C. Cognot, Q.L. Gao, A.T. Encinoza, M. Cuchacovich, et al., Growth factor activity of IL-6 in the synovial fluid of patients with rheumatoid arthritis. *Clin. Exp. Rheumatol.* 1994;12:595-602.
 23. H.J. Zhang, H.Y. Wang, H.T. Zhang, J.M. Su, J. Zhu, H.B. Wang, et al., Transforming growth factor- β 1 promotes lung adenocarcinoma invasion and metastasis by epithelial-to-mesenchymal transition. *Mol. Cell. Biochem.* 2011;355:309–14.
 24. G. Kim, J.-B. Jun, K.B. Elkon, Necessary role of phosphatidylinositol 3-kinase in transforming growth factor beta-mediated activation of Akt in normal and rheumatoid arthritis synovial fibroblasts. *Arthritis Rheum.* 2002;46:1504–11.
 25. L. Bi, I. Okabe, D.J. Bernard, A. Wynshaw-Boris, R.L. Nussbaum, Proliferative defect and embryonic lethality in mice homozygous for a deletion in the p110alpha subunit of phosphoinositide 3-kinase. *J. Biol. Chem.* 1999;274:10963–8.
 26. L. Bi, I. Okabe, D.J. Bernard, R.L. Nussbaum, Early embryonic lethality in mice deficient in the p110beta catalytic subunit of PI 3-kinase. *Mamm. Genome.* 2002;13:169–72.
 27. K. Okkenhaug, A. Bilancio, G. Farjot, H. Priddle, S. Sancho, E. Peskett, et al., Impaired B and T cell antigen receptor signaling in p110delta PI 3-kinase mutant mice. *Science.* 2002;297:1031–4.

28. E. Clayton, G. Bardi, S.E. Bell, D. Chantry, C.P. Downes, A. Gray, et al., A crucial role for the p110delta subunit of phosphatidylinositol 3-kinase in B cell development and activation. *J. Exp. Med.* 2002;196:753–63.
29. T.M. Randis, K.D. Puri, H. Zhou, T.G. Diacovo, Role of PI3Kdelta and PI3Kgamma in inflammatory arthritis and tissue localization of neutrophils. *Eur. J. Immunol.* 2008;38:1215–24.
30. Kiener HP, Stipp CS, Allen PG, Higgins JM, Brenner MB. The cadherin-11 cytoplasmic juxtamembrane domain promotes alpha-catenin turnover at adherens junctions and intercellular motility. *Mol Biol Cell.* 2006;17(5):2366–76.
31. Su Y, Zheng L, Wang Q, Bao J, Cai Z, Liu A. The PI3K/Akt pathway upregulates Id1 and integrin $\alpha 4$ to enhance recruitment of human ovarian cancer endothelial progenitor cells. *BMC cancer.* 2010;10:459.
32. Cavallaro U, Christofori G. Cell adhesion and signalling by cadherins and Ig-CAMs in cancer. *Nat Rev Cancer.* 2004;4(2):118–32.

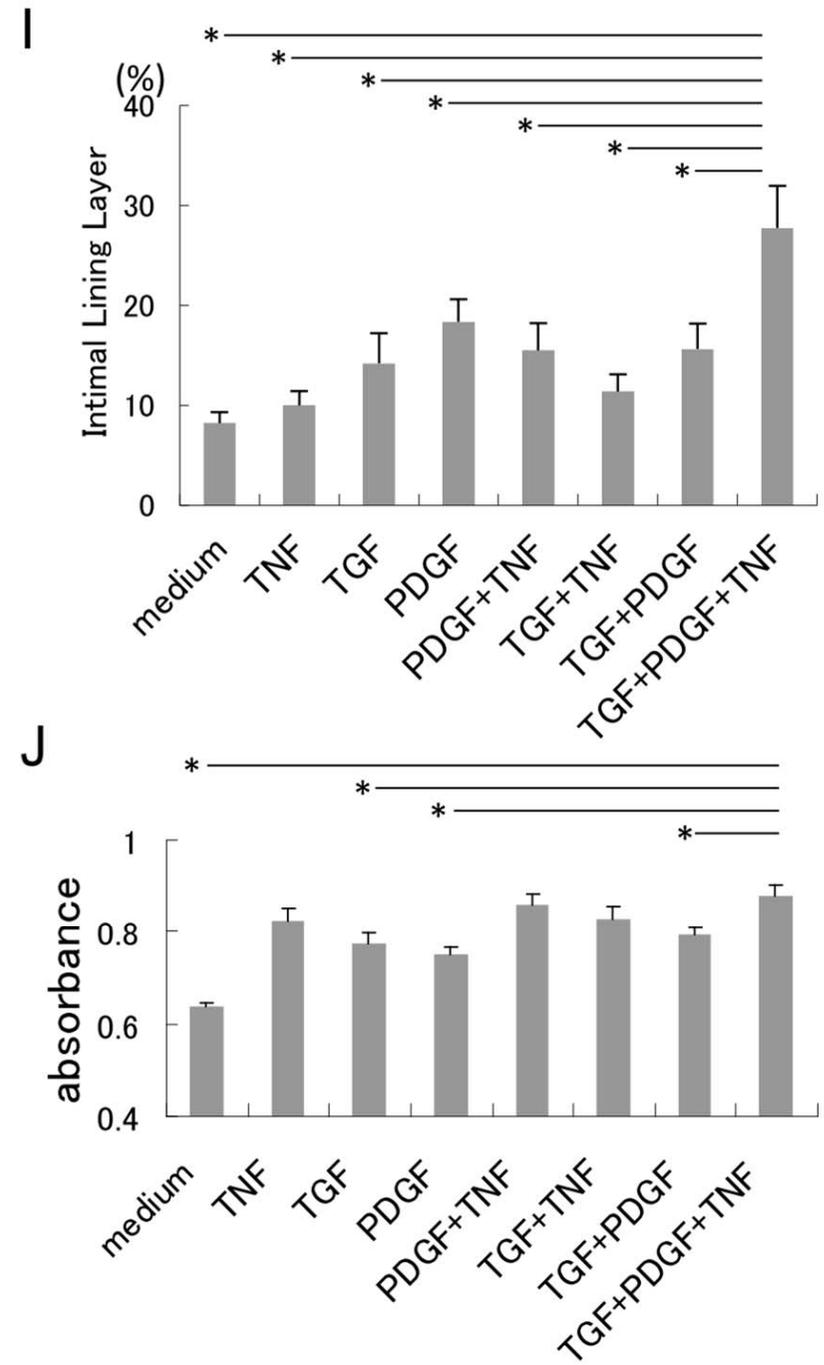
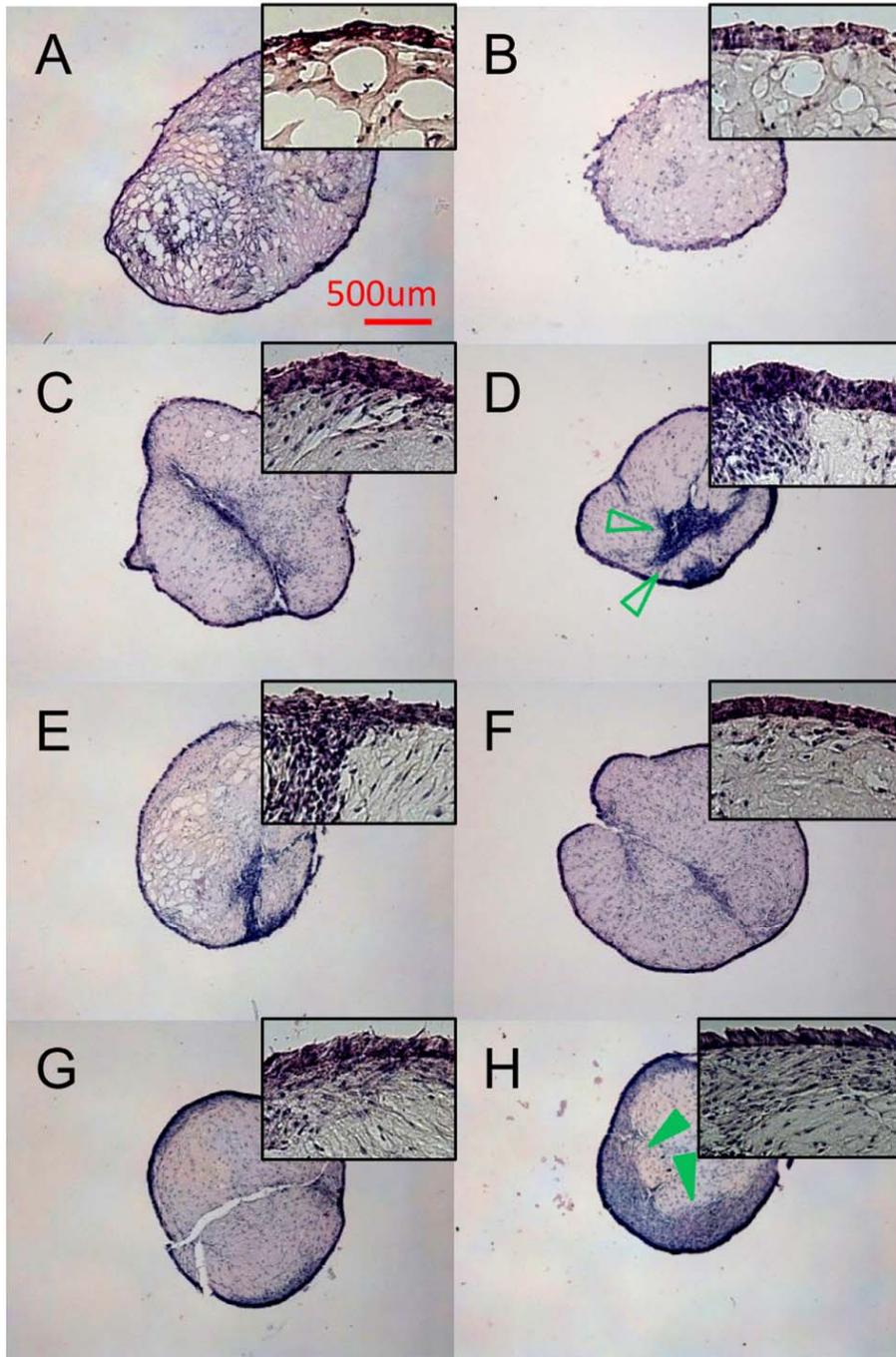


Fig. 1

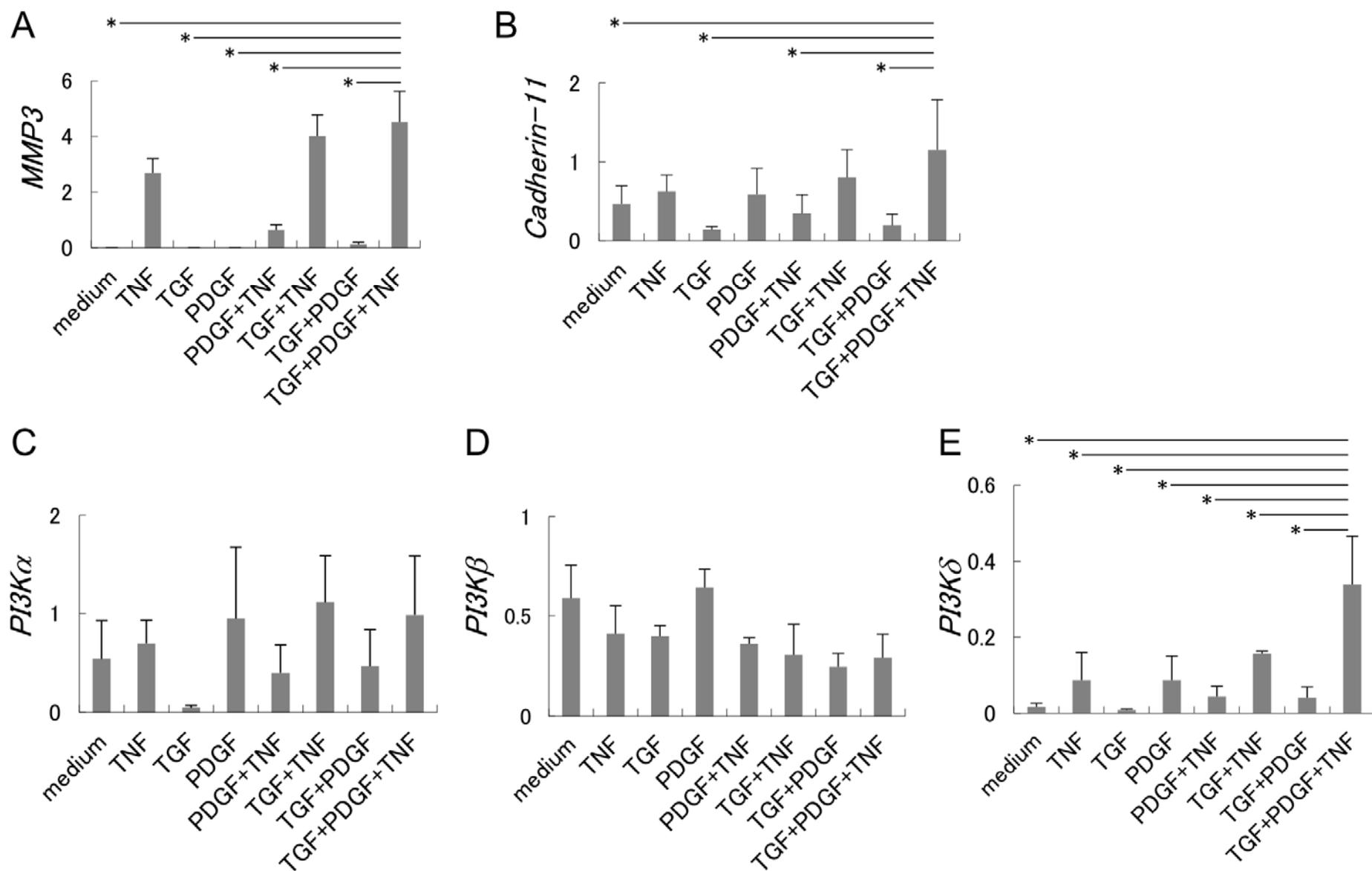


Fig. 2

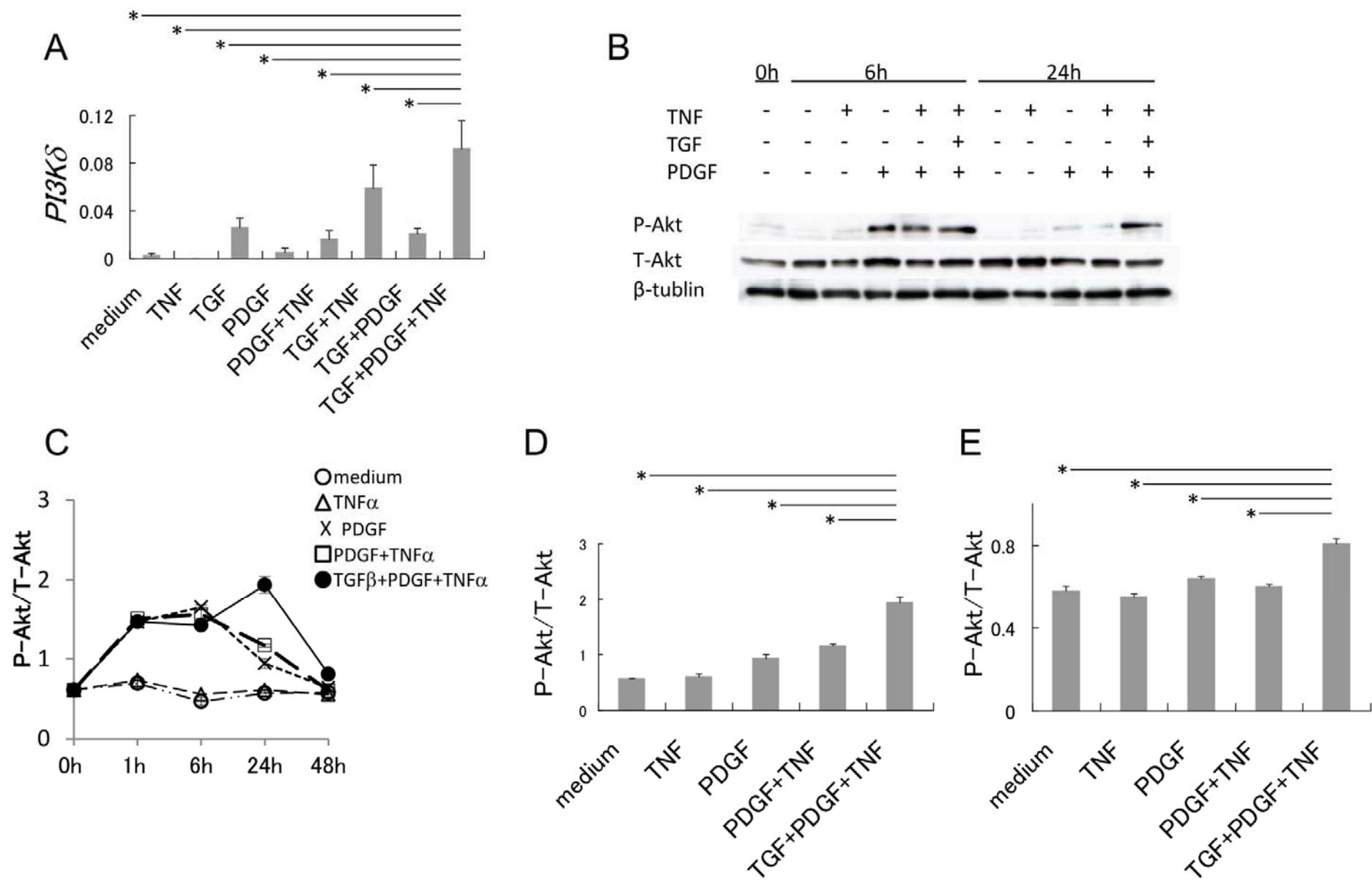


Fig. 3

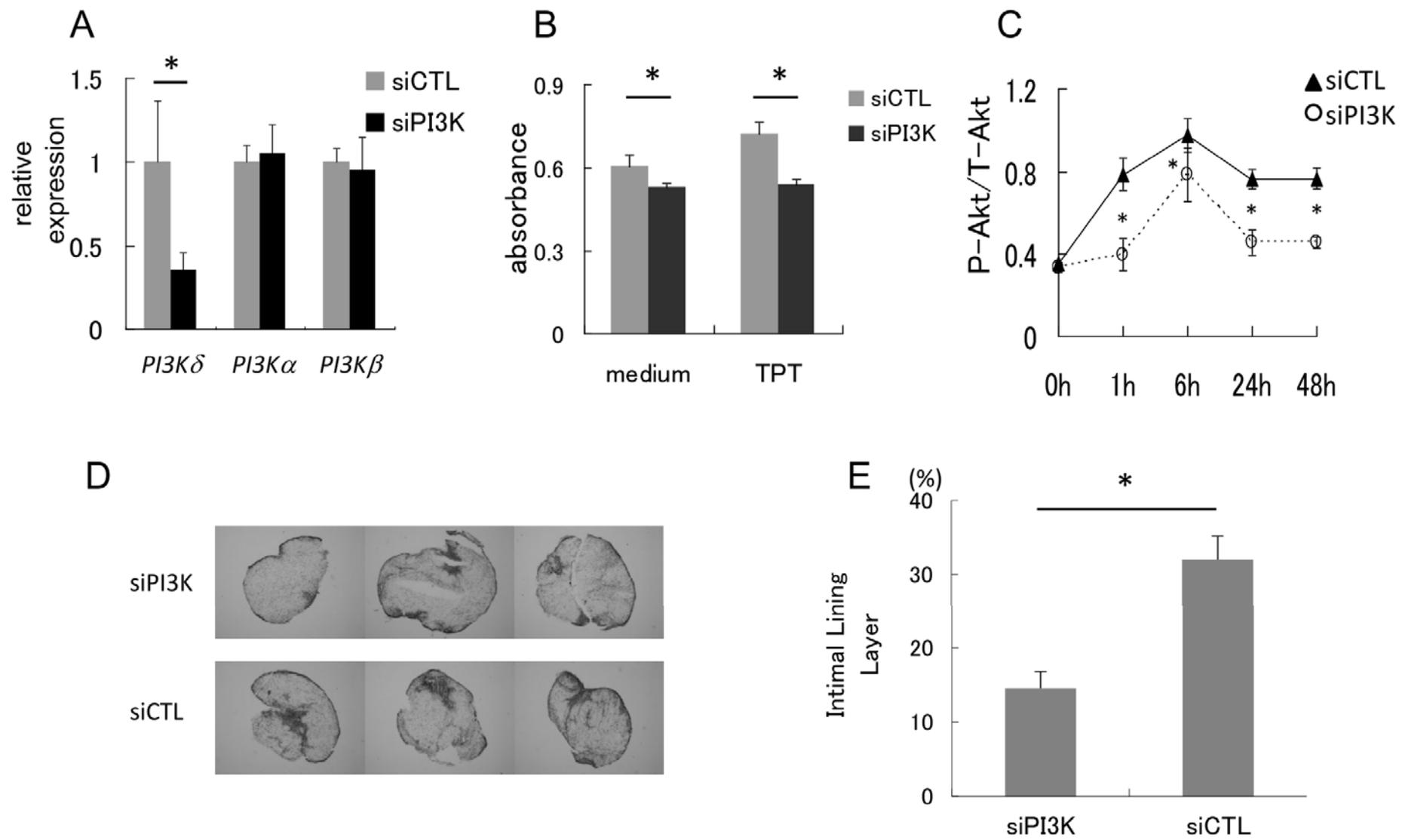
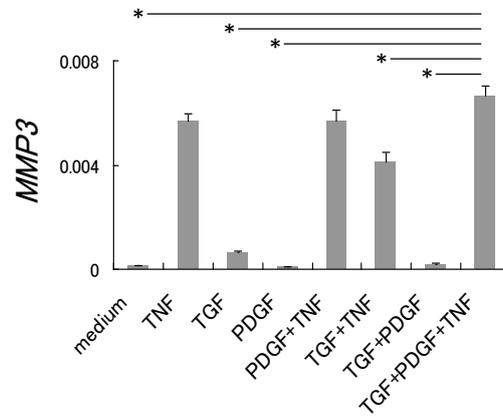
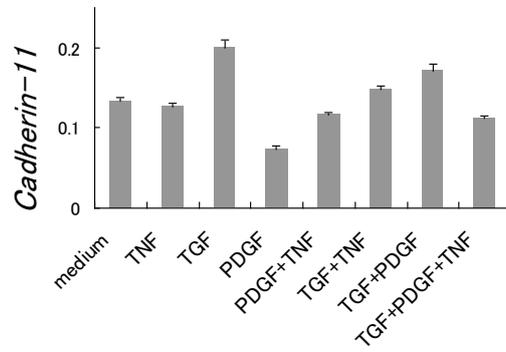


Fig. 4



Supplementary Figure 1. Expression of MMP3 mRNA in monolayer RA FLSs cultured with $\text{TNF}\alpha$, $\text{TGF}\beta$, and/or PDGF for 48 h. Data are shown as mean \pm SEM of sextuplet. * = $P < 0.05$ in Student's t-test.



Supplementary Figure 2. Expression of Cadherin-11 mRNA in monolayer RA FLSs cultured with TNF α , TGF β , and/or PDGF for 48 h. Data are shown as mean \pm SEM of sextuplet.