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Palmitate induces reactive oxygen species production and β-cell dysfunction by activating nicotinamide adenine dinucleotide phosphate oxidase through Src signaling

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Keywords
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INTRODUCTION
In pancreatic β-cells, glucose metabolism regulates exocytosis of insulin granules through metabolism-secretion coupling1. Reactive oxygen species (ROS) is one of the most important factors that impair glucose-induced insulin secretion (GIIS) in β-cells.

ABSTRACT
Aims/Introduction: Chronic hyperlipidemia impairs pancreatic β-cell function, referred to as lipotoxicity. We have reported an important role of endogenous reactive oxygen species (ROS) overproduction by activation of Src, a non-receptor tyrosine kinase, in impaired glucose-induced insulin secretion (GIIS) from diabetic rat islets. In the present study, we investigated the role of ROS production by Src signaling in palmitate-induced dysfunction of β-cells.

Materials and Methods: After rat insulinoma INS-1D cells were exposed to 0.6 mmol/L palmitate for 24 h (palmitate exposure); GIIS, ROS production and nicotinamide adenine dinucleotide phosphate oxidase (NOX) activity were examined with or without exposure to 10 μmol/L 4-amino-5-(4-chlorophenyl)-7-(t-butyl)pyrazolo[3,4-d]pyrimidine (PP2), a Src inhibitor, for 30 or 60 min.

Results: Exposure to PP2 recovered impaired GIIS and decreased ROS overproduction as a result of palmitate exposure. Palmitate exposure increased activity of NOX and protein levels of NOX2, a pathological ROS source in β-cells. Palmitate exposure increased the protein level of p47phox, a regulatory protein of NOX2, in membrane fraction compared with control, which was reduced by PP2. Transfection of small interfering ribonucleic acid of p47phox suppressed the augmented p47phox protein level in membrane fraction, decreased augmented ROS production and increased impaired GIIS by palmitate exposure. In addition, exposure to PP2 ameliorated impaired GIIS and decreased ROS production in isolated islets of KK-Ay mice, an obese diabetic model with hyperlipidemia.

Conclusions: Activation of NOX through Src signaling plays an important role in ROS overproduction and impaired GIIS caused by chronic exposure to palmitate, suggesting a lipotoxic mechanism of β-cell dysfunction of obese mice.
production through non-mitochondrial and mitochondrial pathways has been proposed. In the mitochondrial pathway, ROS is generated in the electron transport chain associated with the mitochondrial membrane potential. However, in pathophysiological conditions, nicotinamide adenine dinucleotide phosphate (NADPH) oxidase (NOX), an important non-mitochondrial ROS source, could play an important role in ROS generation in β-cells.

Deleterious effects of chronic hyperlipidemia on β-cell function are referred to as lipotoxicity. Chronic exposure to palmitate, a long-chain saturated fatty acid, impairs GIIS with an increase in production of ROS in β-cells. Recently, an important role of NOX in ROS production in β-cells by palmitate exposure has been proposed. In addition, both oxidative stress markers and NOX expression are increased in islets of obese diabetic rodents with hyperlipidemia. We have proposed that endogenous overproduction of ROS involving activation of Src, a non-receptor tyrosine kinase, plays an important role in impaired metabolism-secretion coupling in islets of diabetic Goto–Kakizaki (GK) rats. An important role of Src in activation of NOX has been reported in various cells. In the present study, to elucidate the mechanism of lipotoxicity in β-cells more precisely, we investigated involvement of Src in ROS production derived from NOX and impaired GIIS caused by chronic exposure to palmitate.

MATERIALS AND METHODS

Materials
Palmitate obtained from Nacalai (Kyoto, Japan) was dissolved in 95% ethanol at stock concentration of 100 mmol/L. The specific Src inhibitor, 4-amino-5-(4-chlorophenyl)-7-(1-butyl)pyrazolo[3,4-d]pyrimidine (PP2), was purchased from Tocris (Ellisville, MO, USA).

Cell Culture and Mouse Islet Isolation
Rat insulinoma cell line INS-1D cells were cultured as previously described with or without palmitate in the presence of 0.5% bovine serum albumin (BSA) for 24 h.

Male KK-A1 mice and control C57/BL6 mice (Clea Japan, Tokyo, Japan) were maintained and used in accordance with the guidelines of the animal care committee of Kyoto University. All experiments were carried out with mice aged 8–10 weeks. Pancreatic islets were isolated as previously described.

Insulin Secretion
Insulin secretion from INS-1D cells was determined as previously described. INS-1D cells cultured on 24-well plates coated with 0.001% poly-L-ornithine were washed with Krebs–Ringer bicarbonate HEPES (KRBH) buffer composed of (in mmol/L) 140 NaCl, 3.6 KCl, 0.5 MgSO\(_4\), 0.5 Na\(_2\)HPO\(_4\), 1.5 CaCl\(_2\), 2 NaHCO\(_3\) and 10 HEPES (pH 7.4) with 0.1% BSA and 2 mmol/L glucose, pre-incubated at 37°C for 30 min in KRBH with 2 mmol/L glucose and then incubated at 37°C for 30 min in KRBH with 2 mmol/L glucose and 10 mmol/L glucose. Insulin release from intact islets was measured using batch incubation using KRBH supplemented with 0.2% BSA as previously described.

ROS Measurements
ROS was measured according to the method previously described. INS-1D cells and isolated islets were incubated in KRBH medium containing 2 mmol/L glucose and 10 mmol/L 5-(and 6-) chloromethyl-2',7'-dichlorodihydrofluorescein diacetate (CM-H\(_2\)DCFDA; Invitrogen, Carisbad, CA, USA) for 1 h at 37°C, and then were rapidly frozen, stored at −80°C and thawed. Fluorescence of the supernatant was quantified using a reader (Powerscan HT; DS Pharma Biomedical, Suita, Japan) with excitation wavelength at 485 nm and emission at 530 nm, which was corrected by subtracting parallel blanks.

Isolation of Total Ribonucleic Acid and Quantitative Reverse Transcription Polymerase Chain Reaction
Total ribonucleic acid (RNA) was isolated from INS-1D cells using RNeasy mini kit (Qiagen, Hilden, Germany). cDNA was prepared by reverse transcriptase (Superscript II; Invitrogen) with an oligo (dT) primer. The rat sequences of forward and reverse primers to NOX2/gp91\(^{\text{phox}}\) and β-actin (as an inner control) were as follows: (NOX2/gp91\(^{\text{phox}}\): 5'-TGA CTC GGT TGG CTG GCA TC-3'; 5'-CGC AAA GGT ACA GGA ACA TGG G-3'; β-actin: 5'-CAA TGA GCG GTT CCG ATG CC -3'; 5'-AAT GCC TGG GAT CAT GGT GG -3'). AmpliTaq Gold (Applied Biosystems, Foster, CA, USA) was used as a DNA polymerase for reverse transcription polymerase chain reaction (RT–PCR). SYBR Green PCR Master Mix (Applied Biosystems) was prepared for quantitative RT–PCR run. The thermal cycling conditions were denaturation at 95°C for 10 min followed by 40 cycles at 95°C for 30 s and 60°C for 30 s.

Immunoblot Analysis
For immunoblotting, cells were washed with phosphate-buffered saline (PBS) containing protease inhibitor cocktail (Complete; Roche, Mannheim, Germany) and phosphatase inhibitor cocktail (Calbiochem, Darmstadt, Germany), suspended in 1 mL of PBS containing protease inhibitor and phosphatase inhibitor, and homogenized as previously described. Membrane fraction was prepared as described previously. INS-1D cells were washed three times with PBS, suspended in buffer A consisting of 50 mmol/L Tris (pH 7.5), 1 mmol/L ethylenediaminetetraacetic acid, and containing protease inhibitor cocktail, phosphatase inhibitor cocktail and 5 mol/L sodium pyrophosphate, homogenized and then centrifuged at 10,000 g at 4°C for 1 h. The pellets were resuspended in 500 μL of buffer A and stored at −80°C until immunoblot analysis or NOX activity assay. Protein (20 μg per sample) was separated on a 15% polyacrylamide gel and transferred to a polyvinylidene difluoride membrane. After blocking with Tris-buffered saline.
(10 mmol/L Tris/HCl and 100 mmol/L NaCl, pH 7.5) containing 0.1% Tween 20 and 5% BSA (blocking buffer) at room temperature (25°C) for 2 h, blotted membranes were incubated overnight at 4°C with anti-p418 Src antibody (Biosource, Camarillo, CA, USA) at 1:1000 dilution, anti-NCFL (p47^phox^) antibody (Abcam, Cambridge, UK) at 1:1000 dilution, anti-NOX2/gp91^phox^ antibody (Abcam) at 1:1000 dilution, anti-flotillin-1 antibody (Cell Signaling Technology, Danvers, MA, USA) at 1:1000 dilution in blocking buffer and subsequently with anti-rabbit or anti-mouse immunoglobulin G horse-radish peroxidase-conjugated secondary antibody (GE Healthcare, Little Chalfont, UK) diluted at 1:5000 at room temperature for 2 h before detection using ECL prime™ (GE Healthcare). Band intensities were quantified with Multi Gauge software (Fujifilm, Tokyo, Japan).

**NOX Activity Assay**

NOX activity was measured by a luminescence assay in a cuvette containing 50 mmol/L phosphate buffer (pH 7.0), 1 mmol/L ethylene glycol tetraacetic acid, 150 mmol/L sucrose, 500 μmol/L lucigenin as the electron acceptor and 100 μmol/L NADPH as the substrate (total volume 900 μL) as previously described. No activity was measured in the absence of NADPH. In some experiments, PP2 (final concentration 10 μmol/L) was added to the cuvette 10 min before readings. The reaction was started by the addition of 100 μL of membrane fraction (50–300 μg protein). Photon emission was measured every 15 s for 15 min in a luminometer (GloMax 20/20

**Figure 1** | Effect of palmitate exposure on glucose-induced insulin secretion (GIIS) and reactive oxygen species (ROS) production in INS-1D cells. Values are mean ± standard error of the mean (n = 4 in each bar). After INS-1D cells were cultured with or without various concentrations of palmitate (Palm) for 24 h, GIIS and ROS production were measured. (a) Effect of palmitate exposure on GIIS. GIIS was examined in the presence of 2 mmol/L (white bar) and 10 mmol/L glucose (black bar) for 30 min (n = 4 in each bar). *P < 0.01 vs 10 mmol/L glucose, culture without palmitate; †P < 0.01 vs 2 mmol/L glucose, culture without palmitate. (b) Effect of palmitate exposure on ROS production. After INS-1D cells were incubated in medium containing 2 mmol/L glucose and 10 μmol/L CM-H_2DCFDA for 60 min, ROS production was measured. ROS production was also measured using INS-1D cells cultured with 0.6 mmol/L palmitate plus ROS scavengers (0.1 mmol/L vitamin E + 0.2 mmol/L vitamin C [Vit]) or anti-oxidant mimics (10 mmol/L tempol [superoxide dismutase mimic] + 10 μmol/L ebselen [glutathione peroxidase mimic] [T + E]) for 24 h. Culturing with these agents suppressed enhanced ROS production by exposure to 0.6 mmol/L palmitate (n = 5 in each bar). *P < 0.01 vs culture without palmitate; †P < 0.01 vs culture with 0.6 mmol/L palmitate. (c) Effect of Src inhibitor on impaired GIIS by 0.6 mmol/L palmitate exposure. GIIS was measured in the presence of 2 mmol/L (white bar) and 10 mmol/L glucose (black bar) with or without 10 μmol/L 4-amino-5-[4-chlorophenyl]-7-[t-buty]pyrazolo[3,4-d]pyrimidine (PP2) for 30 min (n = 4 in each bar). *P < 0.01 vs 10 mmol/L glucose without PP2, culture without palmitate; †P < 0.01 vs 10 mmol/L glucose without PP2, culture with 0.6 mmol/L palmitate. (d) Effect of Src inhibitor on augmented ROS production by 0.6 mmol/L palmitate exposure. After INS-1D cells were incubated in medium containing 2 mmol/L glucose and 10 μmol/L CM-H_2DCFDA with or without 10 μmol/L PP2 for 60 min, ROS production was measured (n = 5 in each bar). *P < 0.01 vs without PP2, culture with 0.6 mmol/L palmitate. (e) Effect of Src small interfering ribonucleic acid (siRNA) on protein expression and GIIS in INS-1D cells cultured with palmitate. After INS-1D cells transfected with control and Src siRNA were cultured with 0.6 mmol/L palmitate for 24 h, protein levels and GIIS were measured. Representative immunoblots were presented.
Luminometer; Promega, Fitchburg, WI, USA), which was corrected by a subtracting blank.

**Small Interfering RNA Transfection**

Stealth™ small interfering (si)RNAs were synthesized by Invitrogen. The sequences of siRNAs specific for rat NCF1 (p47phox) were as follows: 5′-GGU GAA GCC AUC GAG GUC AUU CAU A-3′, 5′-UAU GAA UGA CCU CGA UGG CUU CAC C-3′. The sequences of siRNAs specific for rat Src were as follows: 5′-GGG AGC GGC UGC AGA UUG UCA AUA A-3′, 5′-UUA UUG ACA AUC UGC AGC CGC UCC C-3′. The sequences of control siRNAs were as follows: 5′-ACC AAC AAC AGU UUG GGA AUA GGG A-3′, 5′-U CCC UAU UCC CAA ACU GUU GUU GGU -3′. Cultured INS-1D cells were trypsinized, suspended with RPMI 1640 medium without antibiotics, mixed with Opti-MEM (Invitrogen) containing siRNA and Lipofectamine 2000 (Invitrogen), plated on dishes or wells and then incubated at 37°C. The final contents of INS-1D cell, RPMI 1640, Opti-MEM, siRNA and Lipofectamine 2000 were 1.9106 cells/mL, 75% v/v, 25% v/v, 80 nmol/L and 0.3% v/v, respectively. The medium was changed to RPMI 1640 3–4 h after transfection. All experiments using siRNA-transfected INS-1D cells were carried out 72 h after transfection.

**Statistical Analysis**

The data are expressed as mean ± standard error of the mean. Statistical significance was calculated by the unpaired Student’s t-test. P < 0.05 was considered significant.

**RESULTS**

**Src Inhibition Recovers Impaired Glucose-Induced Insulin Secretion and Decreases Augmented ROS Production as a Result of Exposure to Palmitate**

Exposure to palmitate (C16:0) concentration dependently decreased GIIS and increased ROS production (Figure 1a,b). Oleic acid (C18:1) slightly increased ROS production, but arachidonic acid (C20:4 n-6) did not increase ROS production.
Palmitate Exposure Causes NOX2 Activation by Src Activation

Palmitate exposure caused Src activation, shown by an increased protein level of Tyr418-phosphorylated Src (Figure 2a). NOX activity was prominently increased by palmitate exposure (Figure 2b). Palmitate exposure also increased the messenger RNA level and protein level of NOX2 (Figure 2c,d). Exposure to 10 μmol/L PP2 for 30 min reduced augmented ROS production (Figure 2b), but did not affect the protein level of NOX2 (Figure 2d). Palmitate exposure caused an increase in the protein level of p47phox in membrane fraction that was reduced by exposure to PP2 for 30 min (Figure 2e). Palmitate exposure caused a decrease in the protein level of p47phox in cytosol fraction that was increased by exposure to PP2 for 30 min (Figure S2).

p47phox Knockdown Ameliorates Impaired Insulin Secretion and ROS Overproduction by Palmitate Exposure

The increased protein level of p47phox in membrane fraction by palmitate exposure was reduced by transfection with p47phox siRNA (Figure 3a). Downregulation of the p47phox level was also observed without palmitate exposure by p47phox knockdown (Figure S3). p47phox Knockdown ameliorated impaired GIIS, and decreased augmented ROS production by palmitate exposure (Figure 3b).

DISCUSSION

Exposure to palmitate, a saturated non-esterified fatty acid decreased GIIS and increased ROS production (Figure 1a,b), as

![Figure 3](http://onlinelibrary.wiley.com/journal/jdi)
previously described. We have previously described an important role of endogenous ROS production that involves Src activation in impaired GIIS in diabetic islets. In the present study, we investigated the effects of PP2, a specific Src inhibitor, on impaired GIIS and augmented ROS production by lipotoxicity. Exposure to PP2 ameliorated impaired GIIS and decreased augmented ROS production by palmitate exposure (Figure 1c,d). In addition, Src downregulation impaired impairment of GIIS by palmitate exposure (Figure 1e). These results suggest that Src activation might be involved in impaired GIIS and augmented ROS production as a result of palmitate exposure.

Src is a non-receptor tyrosine kinase that is associated with the cell membrane and plays important roles in various signal transductions. Its activity is regulated by intramolecular interactions that depend on tyrosine phosphorylation and phosphorylation of Tyr by the kinase domain that results in Src activation. Palmitate exposure caused Src activation indicated by an increased protein level of Tyr-phosphorylated Src (Figure 2a). In a recent study, it was proposed that palmitate alters membrane distribution of Src, causing it to partition into intracellular membrane subdomains, where it likely becomes autophosphorylated and activated, NOX2 (gp91phox and p22phox), an isoform of NOX, and its related subunits (p47phox and p67phox) are expressed in β-cells. Interestingly, exposure to palmitate affects expression level of p47phox in islets. In addition, impaired GIIS and an increase in apoptosis by chronic exposure to palmitate are restored by suppression of NOX2 with a decrease in ROS level in the β-cell line. The effects of palmitate exposure on NOX2 and involvement of Src were therefore examined. NOX activity was prominently increased by palmitate exposure (Figure 2b). PP2 reduced augmented ROS production (Figure 2b), but did not affect protein level of NOX2 (Figure 2d). These results suggest that Src activation might affect regulatory factor(s) of NOX. An important role of Src in activation of NOX2 through translocation of p47phox to plasma membrane has been reported in various cells. In addition, cigarette smoke particle-phase extract induces Src activation, which causes NOX2, Src and p47phox complex formation that increases NOX activity in smooth muscle cells. Palmitate exposure was found to cause an increase in protein level of p47phox in membrane fraction that was reduced by PP2 (Figure 2e). Taken together, these findings show that palmitate exposure increases NOX activity mainly by increasing translocation of p47phox to plasma membrane through Src signaling.

To elucidate the role of p47phox in impaired insulin secretion and ROS overproduction by palmitate exposure, the effect of p47phox knockdown on GIIS and ROS production was examined. Knockdown ameliorated impaired GIIS and decreased augmented ROS production by palmitate exposure (Figure 3B). These results show that p47phox is involved in impaired GIIS and ROS overproduction by palmitate exposure.

Src inhibition by PP2 treatment completely recovered palmitate-induced GIIS impairment despite partial reduction of NOX activation and p47phox expression in the membrane in INS-1D cells. Explanation of these findings is difficult using the data in the present study, but one possibility is that palmitate exposure
also decreases vulnerability to ROS in β-cell function. In addition, it is proposed that Src is constantly activated by forming an intracellular disulphide bond derived from ROS exposure, which could affect the concentration-dependent effect of Src inhibitors. Suppression of ROS content was partial despite complete suppression of the p47phox level by knockdown. These phenomena might be explained by the fact that palmitate exposure increases NOX2 expression in addition to the p47phox level (Figure 2c,d).

NOX2 expression is upregulated in islets from obese diabetic mice1,12 and rats11. The pathophysiological significance of the results in the present study was examined using isolated islets of KK-Ay mice, an obese diabetic model with hyperlipidemia. Impairment of GIIS from islets of KK-Ay mice was ameliorated by PP2 (Figure 4a). In addition, ROS production and the level of p47phox protein in membrane fraction in the islets of KK-Ay mice was reduced by exposure to PP2 (Figure 4b,c). These results suggest that activation of NOX through Src signaling might be involved in impairment of GIIS from islets of KK-Ay mice.

The ameliorating effect of Src inhibition on GIIS is less in islets of obese KK-Ay mice than it is in islets of non-obese GK rats13. As shown in Figure S4, Src activation is similar in KK-Ay mice islets to that in GK rat islets.14 This indicates that the differing effect of Src inhibition on GIIS is not derived from the different level of Src activation. Impairment of GIIS is more prominent in KK-Ay mice islets than in GK rat islets. Another mechanism of impaired GIIS independent of ROS overproduction might exist in obese mice islets. Severe impairment of GIIS is also observed in islets of ob/ob mice, another obese diabetic model, in which upregulation of UCP-2 derived from upregulation of TBP-2 plays an important role in impaired GIIS.15 In contrast, we observed neither UCP-2 upregulation nor TBP-2 upregulation in GK rat islets (unpublished observation).

In conclusion, activation of NOX through Src signaling plays an important role in ROS overproduction and impaired GIIS caused by chronic exposure to palmitate, suggesting a lipotoxic mechanism of β-cell dysfunction in obese mice.

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REFERENCES


SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article:

Figure S1 | Effect of oleic acid (Ole) and arachidonic acid (Ara) exposure on reactive oxygen species (ROS). After INS-1D cells were cultured with 0.1 mmol/L palmitate, 0.6 mmol/L palmitate, 0.1 mmol/L Ole and 0.1 mmol/L Ara for 24 h, ROS production was measured. Values are mean ± standard error of the mean; n = 4 in each bar. *P < 0.01 vs control without free fatty acid.

Figure S2 | Representative immunoblots of p47phox in the cytosol fraction from two independent experiments. After INS-1D cells were cultured with or without palmitate (Palm) for 24 h and incubated with Krebs–Ringer bicarbonate HEPES with or without 10 μmol/L 4-amino-5-(4-chlorophenyl)-7- (t-butyl)pyrazolo[3,4-d]pyrimidine (PP2) for 30 min, immunoblots of the cytosol fraction were carried out.

Figure S3 | Effect of p47phox small interfering ribonucleic acid (siRNA) on basal p47phox level in membrane fraction without palmitate exposure. After INS-1D cells transfected with control, and p47phox siRNA were cultured without palmitate for 24 h, protein levels were measured. Immunoblot was carried out using membrane fraction. Data are expressed relative to control values transfected with control siRNA; n = 3 in each bar.

Figure S4 | Representative immunoblots of Src in islets of KK-Ay mice and control C57BL6 mice from two independent experiments. Src activation was detected by Tyr418-phosphorylated Src. The numbers of the side of each panel express the ratio to value of control islets.