1 Original Paper

- 2 Title: Patterns of morphological variation in enamel-dentin junction and outer-enamel surface of human
 3 molars
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24 Abstract

25	Tooth crown patterning is governed by the growth and folding of the inner enamel
26	epithelium (IEE) and the following enamel deposition forms outer enamel surface (OES). We
27	hypothesized that overall dental crown shape and covariation structure is determined by processes that
28	configurate shape at enamel-dentin junction (EDJ), the developmental vestige of IEE, and tested this
29	hypothesis by comparing patterns of morphological variation between EDJ and OES in human
30	maxillary permanent first molar (UM1) and second deciduous molar (um2). Using geometric
31	morphometric methods, we described morphological variation and covariation between EDJ and OES,
32	and evaluated the strength of two components of phenotypic variability: canalization and morphological
33	integration, in addition to the relevant evolutionary flexibility, i.e., the ability to respond to selective
34	pressure. The strength of covariation between EDJ and OES was greater in um2 than UM1, and the way
35	that multiple traits covary between EDJ and OES were different between these teeth. The variability
36	analyses showed that EDJ had less shape variation and a higher level of morphological integration than
37	OES, which indicated that canalization and morphological integration acted as developmental
38	constraints. These tendencies were greater in UM1 than um2. On the other hand, EDJ and OES had the
39	comparable level of evolvability in these teeth. Amelogenesis could play a significant role in tooth shape
40	and covariation structure, and its influence was not constant among teeth, which may be responsible for
41	the differences in the rate and/or period of enamel formation.
42	

43 Key Words: developmental constraints, geometric morphometrics, morphological variability,

44 evolvability, odontogenesis

45 Introduction

46	Dental morphological characteristics such as cusps, accessory cusps, and ridges on the
47	occlusal surface have been used extensively in the studies of hominoid evolution and phylogenetic
48	relationships (Miller, 1918; Simons and Pilbeam, 1972; Dean, 2000; Pilbrow, 2006; Matsumura et al.,
49	2011). Tooth crown morphology is determined through two developmental processes (Avishai et al.,
50	2004; Skinner and Gunz, 2010; Smith et al., 2011). The first process is the growth and folding of the
51	inner enamel epithelium (IEE) during the bell stage. This morphogenesis (= tooth crown patterning) is
52	governed by interactions between the IEE and underlying mesenchymal tissues. The final configuration
53	of the IEE is preserved as the enamel-dentin junction (EDJ). The second process is biomineralization by
54	the enamel-forming ameloblasts and dentin-forming odontoblasts. Ameloblatsts are derived from the
55	IEE cells and odontoblasts from the dental papilla cells. Enamel formation starts at the cusp tips, and
56	proceeds apically to complete the outer-enamel surface (OES).
57	Recent micro-CT dental analyses have revealed that crown morphological traits of the
58	completed EDJ are modified or masked through the process of enamel deposition (Skinner et al., 2009,
59	2010; Ortiz et al., 2012), and that the extent of modification varies depending, in part, if not totally, on
60	the enamel thickness (Ortiz et al., 2012). This raises a concern about whether or not shared derived
61	features and homoplastic features of similarity at the OES can be properly discriminated (Hunter and
62	Jernvall, 1995; Collard and Wood, 2000; Finarelli and Clyde, 2004). Additionally, by examining
63	enamel thickness variation and its heritability in pedigreed baboon molars, Hlusko et al. (2004) showed
64	that enamel thickness could change rapidly under moderate or low selective pressure over evolutionarily
(5	short periods increasing the potential for homonlasy. Although the OES morphology is directly related

66	to dental functions such as occlusion and feeding and thus is a direct target of natural selection, the
67	morphology of EDJ has been considered to be more conservative evolutionally and a more reliable
68	representation of the phenotype for estimating phylogenetic relationships (Kraus, 1952; Korenhof,
69	1960; Smith et al., 1997; Sasaki and Kanazawa, 1999; Smith et al., 2000; Olejniczak et al., 2007).
70	So far researchers have explored to which extent enamel formation influences on the crown
71	morphology by comparing EDJ with OES (Kraus, 1952; Nager, 1960; Korenhof, 1960, 1961; Sakai
72	and Hanamura, 1971; Skinner et al., 2008, 2009; Ortiz et al., 2012). However, these studies mainly have
73	focused on discrete dental traits. Although a few studies tried to evaluate general morphological
74	difference between EDJ and OES quantitatively by using intercusp distance (Smith et al., 1997) or
75	surface complexity (Skinner et al., 2010), complex dental crown topography of EDJ and OES has not
76	been clarified in detail. Examining morphological variation and covariation between EDJ and OES
77	enables us to understand the effects of morphological change caused by enamel formation.
78	Additionally, given the different developmental backgrounds between the EDJ and OES, it is
79	likely that the patterns of phenotypic variability differ between these structures. Phenotypic variability is
80	defined as the tendency or potential of an organism to vary (Wagner and Altenberg, 1996; Wagner et al.,
81	1997; Willmore et al., 2007). Therefore, it determines the potential range or distribution of
82	morphological variation, and ultimately affects the tempo and mode of evolutionary change. The recent
83	literature about phenotypic variability has paid the greatest attention to canalization and morphological
84	integration (Wagner and Altenberg, 1996; Hallgimmson et al., 2002; Willmore et al., 2007;
85	Hallgrímsson et al., 2009). Canalization is generally considered a property of an organism that limits
86	phenotypic variation by buffering developmental processes against both environmental and genetic

87	perturbations (Wagner et al., 1997; Willmore et al., 2007). Morphological integration refers to the
88	tendency for different characters to covary as a result of common underlying developmental factors
89	(Hallgrímsson et al., 2002), which constrains the production of phenotypic variation (Wagner and
90	Altenberg, 1996; Chernoff and Magwene, 1999). Canalization and morphological integration are
91	interrelated and can act as developmental constraints (Alberch, 1982; Maynard Smith et al., 1985;
92	Hallgrímsson et al., 2002). Since the morphological integration framework is directly connected to the
93	rate and direction of evolutionary change (Cheverud, 1996; Wagner and Altenberg, 1996), some studies
94	have focused on quantification of the intervening effect of morphological integration on evolutionary
95	trajectory (Lande, 1979; Lande and Arnold, 1983). The resultant data have led to recent studies that
96	evaluated evolvability (the ability of a population or species to respond to selection: Hansen, 2003)
97	using the simulation of evolutionary responses to selection (Marroig et al., 2009; Villmoare et al., 2011;
98	Lewton, 2012; Grabowski, 2013). The relationships and interactions among developmental processes,
99	variability and variation, mediated by the feedback loop of natural selection, are critically involved in
100	evolutionary change (Willmore et al., 2007). Comparison of the pattern of variability between EDJ and
101	OES helps to infer how the production of morphological variation is regulated in each of these
102	components.
103	In this study, we explore the relationship between the crown morphology and odontogenesis
104	through quantitative analyses of the EDJ and OES morphology. We hypothesized that overall dental
105	crown shape and covariation structure are determined by processes that configurate shape at the EDJ. If
106	this hypothesis is rejected, a significant role of enamel formation for patterning of crown morphological
107	variation must be presumed. To test this hypothesis, we described morphological variation and

108	covariation between EDJ and OES and revealed how much variation in the OES shape is explained by
109	the EDJ shape variation. Consequently, we evaluated the strength of two components of phenotypic
110	variability: canalization and morphological integration, in addition to the relevant evolutionary
111	flexibility.
112	Canalization: if EDJ shows larger variation, it means that more variable morphology is created during
113	the early phase of the tooth development, and subsequent enamel formation acts as stabilizing
114	developmental process that buffers the deviation from mean shape. On the other hand, if OES shows
115	larger variation, it indicates that enamel formation brings about not only homogeneous enamel
116	distribution above the EDJ after the morphogenesis, but also some modification of the OES associated
117	with the increased variation.
118	Morphological integration: if either during morphogenesis or the enamel formation process, some
119	developmental factors produce higher morphological integration of one of these structures (whether
120	EDJ or OES). Combined with the result regarding canalization, this analysis will help to determine what
121	factors play important roles in generating or reducing morphological variance.
122	Evolutionary flexibility: in relation to the above two components of phenotypic variability, we
123	specifically compared how the developmental constraints exert influence on the ability of the response
124	to selection in EDJ versus in OES.
125	This study focused on EDJ and OES shape variation of maxillary permanent first molar
126	(UM1) and second deciduous molar (um2). Although UM1 and um2 share similar patterns of occlusal
127	morphology that are elaborated through the same developmental processes, their developmental timing,
128	speed and period are different (Nanci, 2013). The differences between UM1 and um2 will provide a

better understanding of the relationship between odontogenesis and crown morphological variability.

131 Materials and Methods

132	The samples used in this study comprised fully formed but unworn UM1 and um2 crowns
133	obtained from archaeological sites in Japan. The total sample (57 UM1 and 48 um2) consisted of
134	samples from the Jomon (14500-300 BC; n=8 and 5), Medieval (13-15C AD; n=13 and 8), and Edo
135	(17-19C AD; n=36 and 35) periods. Although the total sample was from a mixture of populations from
136	different periods and regions, the aim of this study was to investigate differences and patterns of
137	variability produced by a common tooth formation process of the Holocene human, and mixing these
138	samples does not violate the objective of this study. In order to maximize sample size, no discrimination
139	between right and left teeth was made, but only a single tooth was used from each individual. All
140	specimens were regarded as left side. Right molar images were transformed into the mirror image using
141	ImageJ software (NIH, USA). Sex was unknown for most of the samples, since they were taken from
142	juvenile individuals.
143	Each specimen was μ CT scanned (ScanXmateA080S, Comscantecno, Japan) with a pixel
144	size and slice interval of 31–32 μ m (80 kV, 125 μ A). To facilitate tissue segmentation, the image stack
145	for each tooth was filtered using a median filter followed by a kuwahara filter, and enamel and dentin
146	tissues were segmented by the seed region growing method in ImageJ. Triangular mesh models of the
147	3D EDJ and OES of each specimen were reconstructed using Analyze 6.0 (Mayo Clinic, USA) with the
148	marching cube method. Subsequent procedures were done using the software Rapidform 2004 (INUS
149	Technology, Korea).

150	We treated the EDJ and the OES as biologically corresponding structures in order to
151	compare variability between them directly, and digitized (semi)landmarks on both of them in the same
152	way as follows. We digitized main cusp tips (paracone, protocone, metacone, and hypocone) at the OES
153	and the dentin horn tips at the EDJ, and the lowest points on the ridges at both the OES and the EDJ,
154	connecting the two cusps as landmarks. Each ridge on both the OES and the EDJ was divided into eight
155	sections by the cusp tips and the lowest points, respectively. For each section, a given number of
156	semi-landmarks was digitized equi-distantly, as illustrated in Figure 1. The number of semi-landmarks
157	on the EDJ and the OES were determined to satisfy two criteria, namely, that each corresponding
158	section in the EDJ and the OES had the same number of (semi)landmarks, and that the contributions of
159	the section between the (semi)landmarks to the curve were approximately equal to each other (Skinner
160	et al., 2009; Skinner and Gunz, 2010). The dataset consisted of four configurations (UM1EDJ,
161	UM1OES, um2EDJ and um2OES), and each of them had a total of 8 landmarks and 48
162	semi-landmarks.
163	Semi-landmarks are not considered to be homologous landmarks unless they are slid
164	(Bookstein, 1997). The minimum bending energy algorithm (Bookstein, 1997; Gunz et al., 2005) was
165	adopted. This data processing was performed by W. Y. using MATHEMATICA 8 (www.
166	wolfram.com). Each homologous set of landmarks was converted to shape coordinates by Generalized
167	Procrustes Analysis (GPA; Rohlf and Slice, 1990), which was performed using MorphoJ version 1.05d
168	(Klingenberg, 2011).
169	

170 Morphometric analysis

171	Covariation between EDJ and OES was analyzed using 2B-PLS. This method compares two
172	morphological data sets by using a singular value decomposition of the cross-covariation matrix, finds
173	new pairs of axes that account for the maximum amount of covariance between both data sets and
174	visualizes the main associated morphological changes. The RV coefficient was used to evaluate the
175	strength of multivariate correlations between data sets. This coefficient is a multivariate analogue of the
176	squared correlation coefficient (Escoufier, 1973; Klingenberg, 2008). The significances of both the
177	correlation between the scores for each pair of PLS axes and RV coefficient were evaluated by means of
178	resampling tests with 1000 random permutations. These procedures were carried out with MorphoJ
179	software (Klingenberg, 2011).
180	A principal component analysis of Procrustes shape coordinates was used to extract main
181	patterns of morphological variation across EDJ and OES in both UM1 and um2. Using first few PC
182	scores of EDJ and OES, we performed a regression analysis between these two structures to test
183	whether shape variation of OES can be predicted by that of EDJ.
184	The difference in multivariate morphological change vector from EDJ to OES between UM1
185	and um2 was assessed by calculating the length and direction of shape change using a residual
186	randomization procedure outlined in Collyer and Adams (2007). The length of a vector describes the
187	overall amount of morphological change and the direction of a vector describes the way in which
188	multiple traits covary. Observed vector lengths and directions were compared with 999 random
189	permutations plus the observed value to assess significance.
190	

191 Variability analysis

192	Among-individual phenotypic variation is the most common measurement of canalization.
193	Canalization is generally inferred from a reduction of the observed phenotypic variance. Here we
194	quantified both size and shape variance within each of the four configurations. For size, Centroid size
195	(CS) of each configuration was calculated. Coefficient of variation (CV) of the LogCS was used to
196	compare size variation, and tested as suggested by Sokal and Braumann (1980). For comparison of
197	shape variability among configurations, the square root of the sum of the squared distances between
198	Procrustes transformed coordinates of each cusp and its landmark mean configuration was used as the
199	measure of shape variation. To test whether there was a significant difference of variability between the
200	EDJ and the OES within the same tooth class, a nonparametric Kruskall-Wallis test and
201	multiple-comparison test were performed.
202	To compare the overall strength of morphological integration, we followed Wagner (1984)
203	in using the variance of the eigenvalues for the variance-covariance matrix as the measure of integration.
204	This measure of integration captures whether shape variance can be explained by a small number of
205	principal components, or whether variance is more evenly distributed across principal components. The
206	former case would be considered more integrated and the latter less integrated. Variance of eigenvalues
207	(VE) was compared between the EDJ and the OES within the same tooth using bootstrap resampling
208	methods (Manly, 1997). For each of the EDJ and the OES, the original data matrix was bootstrapped
209	1000 times, a variance-covariance matrix was derived from each bootstrap sample, and VE was
210	calculated from each of the 1000 variance-covariance matrices. For each of the 1000 VE replicates, the
211	difference between the EDJ and the OES was calculated. This created a distribution of differences in
212	VE replicates that was then zero-centered. Each of the zero-centered differences was then compared to

- the observed difference in VE between the EDJ and theOES. The two-tailed P value was calculated as the number of times the difference from the zero-centered distribution was equal to or greater than the observed difference, divided by the number of bootstrap replicates (Manly, 1997).
- The ability of EDJ and OES morphology to respond to selection was evaluated using mean flexibility (f) (Marroig et al. 2009), which is derived from Lande's (1979) multivariate selection equation:

219 $\Delta z = G \beta$

220 where G is the genetic covariance matrix, β is a selection vector, and Δz is the response vector. Here 221 the phenotypic covariance matrix P is substituted for G because previous studies established structural 222 similarity between them (e.g., Cheverud, 1996; Porto et al., 2009). The covariance matrix for each of 223 EDJ and OES was subjected to 1,000 randomly generated selection vectors and the angle between the 224 selection and response vectors was calculated for each time. The mean cosine of angles in 1000 repeats 225 is called the mean flexibility (Marroig et al., 2009), which describes the degree to which the response 226 and selection vectors are aligned in multivariate space. Response and selection vectors that are parallel 227 (i.e., when the cosine of the angle between them is 1.0) indicate a structure that is more responsive to 228 selection, i.e., more evolvable. A larger angle between the response and selection vectors is indicative of 229 less evolvability. In general, high levels of evolvability measures, such as evolutionary flexibility, tend 230 to be associated with low levels of integration measures (e.g., VE). Pairwise comparisons of 231 evolutionary flexibility between EDJ and OES within the same tooth class were performed as described 232 for VE; the distribution of vector correlations obtained from the covariance matrix and 1,000 random 233 selection vectors for EDJ and OES were compared using the difference of means test and accompanied

by a two-tailed P value. All statistical analyses were performed using R version 2.13.1 (R Development
Core Team, 2011).

236

237 Results

238 Morphometric analysis

239 Covariation between EDJ and OES is higher in um2 (RV=0.914; P<0.001) than in UM1 240 (RV=0.794; P<0.001). 2B-PLS analysis in UM1 revealed that the first axis explained 49.43% of total 241 shape covariance and that corresponding shape change mainly involves the contraction of buccal side 242 and expansion of distolingual cusp (hypocone) for both EDJ and OES (Table 1; Fig. 2a). The second 243 axis also revealed that EDJ and OES showed similar shape change that contraction of mesiobuccal cusp 244 (paracone) and contraction of distal side (Fig. 2b). In um2, the first singular axis of correspondence to 245 the comparison of EDJ and OES revealed a correlated reduction of mesiolingual-distobuccally and expansion of mesiobuccal-distolingually (Fig. 2c). The second axis also revealed significant shape 246 247 change of reduction of mesial cusps and reduction of distal cusps for both EDJ and OES (Fig. 2d). 248 In the PCA, the first two principal components account for 34.85% of the total variation 249 (Figure 3a; Table 2). Positive scores of PC1 are associated with relatively high and sharp cusp tips and 250 lingually located hypocone. Its negative values correspond to relatively-gentle and inner located cusp 251 tips with deep intercuspal grooves. Positive PC2 scores are associated with mesial expansion and 252 contraction of protocone and negative ones with mesial contraction with lingually expanded protocone. 253 PC1 corresponds to the distinction between EDJ and OES, whereas PC2 separates between UM1 and 254 um2. Figure 3b and 3c illustrates the regressions of first two PCs for EDJ and OES in both teeth. The

255	adjusted R- squared value is lower in UM1 than that in um2 for both PC1 (0.249 vs.0.700) and PC2
256	(0.842 vs. 0.907), which indicated that the OES shape variation is better predicted by EDJ shape
257	variation in um2 than in UM1.
258	The tooth specific morphological change vectors between EDJ and OES were not
259	statistically different in length (ΔD =0.004; P=0.27). However, the angle between these vectors was
260	significantly greater than expected by chance ($\theta = 27.62^\circ$; P<0.001: Fig. 3a).
261	
262	Variability analysis
263	Canalization
264	The coefficients of variation of the LogCS for each configuration (UM1EDJ, UM1OES,
265	um2EDJ and um2OES) was not significantly different from each other, although OES tended to be
266	more variable than EDJ in both the UM1 and um2 tooth classes (Figure 4a). On the other hand, shape
267	variability was significantly different among these configurations, and pair-wise tests showed that only
268	in UM1 was there a significant difference in shape variability between EDJ and OES (Figure 4b).
269	
270	Morphological integration
271	The variance of the eigenvalue (VE) was significantly greater for EDJ than for OES in UM1,
272	but not in um2 (Figure 4c). The greater VEs for EDJ were seen in both UM1 and um2, indicating that
273	EDJ was more integrated than OES.
274	
275	Evolutionary flexibility

The mean cosine between the selection vector and the response vector for OES tended to be greater than that for EDJ, but a significant difference was not detected between them in either tooth class (Figure 4d). This meant that there was no difference in the extent to which EDJ and OES would be influenced by the selection vector.

280

281 Discussion

282 Both UM1 and um2 showed significantly correlated shape changes between EDJ and OES 283 corresponding to singular axes. Enamel formation does not alter the basic morphology of the dentine 284 horn and EDJ ridges and corresponding features (cusp tips and ridges) on OES. Our results accord with 285 previous studies that dental traits seen in EDJ can be observed at OES (Korenhof, 1961, 1982; Nager, 286 1960; Sakai and Hanamura, 1973; Corruccini, 1998; Sasaki and Kanazawa, 1999; Skinner et al., 2008; 287 2009), which supports the major role of the EDJ in their origin and degree of dental crown traits. 288 However, this does not necessarily mean that tooth shape and covariation structure are predetermined 289 by processes that configurate tooth shape at EDJ. Comparisons between um2 and UM1 revealed 290 different influences of enamel formation on the OES morphology. In um2, OES shape variation is better 291 predicted from EDJ shape variation. Thus, multivariate covariation between EDJ and OES is higher 292 compared to UM1. This result suggests that morphological change caused by enamel formation is more 293 stable and less vulnerable to random perturbations in um2. This could be attributed to the difference in 294 the enamel thickness (Grine, 2005), the rate of enamel formation (Shellis, 1984) and/or period of 295 enamel formation (Liversidge and Molleson, 2004). While the amount of overall morphological change 296 induced by enamel formation does not differ between UM1 and um2, the direction of change described

297	by traits covariation marks a significant difference. Given the different period of formation between
298	UM1 and um2 (Nanci, 2013), it may be expected that they show resembling directions of
299	morphological change with different amounts of morphological change. However, the result is converse,
300	suggesting a complex nature of crown enamel formation. For example, Grine (2005) noted that the
301	difference in enamel thickness between the paracone tip and the protocone tip was greater in um2 rather
302	than in UM1. The difference in patterns of enamel distribution between UM1 and um2 might affect the
303	way of covariation between EDJ and OES. Thus, enamel formation has a significant effect on patterns
304	of morphological change, probably according to tooth-specific developmental parameter, though it does
305	not cause a drastic change in morphology during odontogenesis.
306	The lack of significant difference in size variation between EDJ and OES in both tooth
307	classes examined here suggests that the strength of canalization on size is almost constant throughout
308	the processes of morphogenesis and the subsequent enamel formation period. A recent developmental
309	study revealed that molar crown sizes were regulated by intrinsic factors from mesenchymal tissues (Cai
310	et al., 2007) and adjacent molars during development (Kavanagh et al., 2007). Several dental metrics
311	studies confirmed that tooth crown size was less variable than intercusp distance and/or cusp size owing
312	to stronger genetic control (Townsend et al., 2003; Harris and Dihn, 2006), which would be also
313	supported by experimental evidence that cusp density (intercusp distances) was likely to be polygenic
314	(Harjunmaa et al., 2012). The present analysis of EDJ and OES at the dentin horns/cusp tips and ridges
315	provided the insight about intercusp distances that their size variation might not be altered largely by
316	enamel formation. Additionally, the spatial relationship with the surrounding tissues, including the
317	maxillary bone and/or other tooth germs, and the available space for tooth growth (Boughner, 2011)

318	may be involved in the canalization of crown size during odontogenesis. The extent of the deviation
319	from mean size in EDJ and OES were not significantly different, and therefore both EDJ size
320	differences and OES size differences among groups being compared can be used as a reliable measure
321	of phylogenetic relatedness.
322	In the case of UM1, shape variation of OES was greater than that of EDJ. This result
323	suggests that canalization of crown shape may be weakened during the process of enamel formation.
324	Kraus and Jordan (1965) argued that early stages of tooth development were mediated by genes that are
325	more evolutionarily stable than those associated with calcification. Hlusko's (2004) simulation model
326	indicated that enamel thickness could change rapidly under appropriate selective pressure. The present
327	result obtained at the cusp tips and ridges is in accord with these studies and implies that shape (e.g.,
328	intercusp topological relationship) variation is more susceptible to modifications resulting from enamel
329	formation than size variation, which might be likely to cause homoplasy that would confuse
330	phylogenetic reconstructions (note here "size" refers to the centroid size of the cuspal tips and ridges and
331	not commonly used crown size proxies like maximum mesiodistal x buccolingual dimensions).
332	The result of VE analysis showed that EDJ was more integrated than OES in UM1, although
333	the same was not supported statistically in um2. Molar crown morphogenesis is a morphodynamic
334	process in which inductive events and morphogenetic processes act at the same time, and is regulated by
335	interactions between the epithelial and underlying mesenchymal tissues. Cusp initiation and patterning
336	in tooth germ is an iterative process that repeatedly utilizes the same set of genes and signaling pathways,
337	which would lead to higher morphological integration in EDJ. On the other hand, the pattern of enamel
338	formation is the end product of a sequence proceeding from ameloblast differentiation from the IEE

cells, to secretion of enamel proteins including amelogenins and enamelins, and finally organization of
the enamel crystallites into enamel rods or prisms (Boyde, 1964, 1989). Topological developmental
parameters, such as the rate and the duration of enamel apposition and/or ameloblast extension and
termination (Simmer et al., 2010), might impact the OES formation, which could lead to weaker
morphological integration in OES.

344 It is predicted that stronger integration between traits acts as a limitation on producing 345 phenotypic variation (Wagner and Altenberg, 1996). The results of the canalization and morphological 346 integration analyses presented here are consistent with this prediction, i.e., the more strongly integrated 347 EDJ shows smaller variability. The set of genes expressed during morphogenesis of the tooth are also 348 used in different organs, including hair, pancreas, mammary gland, salivary gland, thymus, vibrissae, 349 and others (Fincham et al., 2000; Jernvall and Jung, 2000). Mutations in coding region that alter the 350 function or activity of proteins are likely to have widespread and potentially many negative effects on 351 development and fitness, and may thus be under considerable constraint (Carroll, 2008). Size and shape 352 of EDJ are thus more likely to be stabilized in order to reduce the risks of negative pleiotropic side 353 effects. The high level of integration in EDJ can be regarded as a relatively rigorous developmental 354 constraint during odontogenesis. Meanwhile, the set of genes that contribute to enamel formation, such 355 as amelogenin, enamelin, ameloblastin, and enamelysin genes, is highly specialized, and can easily 356 modify the OES morphology during the enamel formation process. Morphological change of the OES, 357 which has less developmental constraint, can easily be brought about by neutral evolution by 358 non-natural selective genetic factors such as random genetic drift.

359

The observed pattern of morphological integration and the results of evolutionary flexibility

360	analyses presented here are not consistent with those of previous studies, in which low levels of
361	integration accompanied high levels of evolvability (Marroig et al., 2009; Porto et al., 2009; Lewton,
362	2012). The developmental constraints due to canalization and morphological integration act more
363	strongly on the shape of EDJ than on that of OES in UM1, while there is no significant difference in the
364	evolutionary flexibility between EDJ and OES. This may result from the relatively integrated
365	covariance structure of each cusp (for both EDJ and OES). Since the secondary enamel knot that
366	functions as a signaling center and regulates cusp formation at the future cusp tip acts as a
367	"developmental module" (Jernvall and Jung, 2000), it can directly affect the covariance structure of EDJ,
368	and indirectly affect that of the overlying OES. In the case of the human tooth, if the crown covariance
369	structure is divided into individual cusp units, this patterning cascade mode of cusp development
370	facilitates the ability to respond to selective challenges (Jernvall and Jung, 2000), and enables the
371	maintenance of a certain level of evolvability at EDJ despite existence of developmental constraints.
372	The comparable level of evolutionary flexibility between EDJ and OES suggests that both of them can
373	be utilized as an equally effective proxy for inferring phylogenetic relationships that would result from
374	selective pressure.
375	Overall, the difference of each measurement (canalization, morphological integration and
376	evolutionary flexibility) between the EDJ and OES in the present study was greater in UM1 than in um2.
377	The process of enamel formation is more likely to influence crown morphological variability and
378	evolvability in UM1 than in um2, which can be explained by the duration and/or thickness of enamel
379	formation. Compared to UM1, the enamel deposition period of um2 is shorter and the enamel is thinner
380	(Nanci, 2013). Therefore enamel formation may exert less influence on shape change in um2, which

381 may be related to the conservation of primitive morphology, as discussed in previous studies (Dahlberg, 382 1945; Butler, 1956, 1971; Suzuki and Sakai, 1973; Saunders and Mayhall, 1982). Since not only 383 morphology but also variability would be likely to differ between EDJ and OES, a tooth crown that has 384 a longer period of enamel formation and/or thicker enamel would require careful evaluation for 385 phylogenetic studies. This study compared patterns of canalization, morphological integration, and evolutionary 386 387 flexibility between the EDJ and the OES in UM1 and um2 in order to explore their possible effects on 388 phylogenetic reconstructions. Our results suggest that a tooth crown that has thicker enamel and/or a 389 longer period of enamel formation can be more variable in shape at the OES, where similarity can be 390 due to homoplasy. Recent advances in imaging techniques have made it possible to approach the details

391 of developmental trajectories reflected in the teeth of fossil species (Avishai et al., 2004; Smith et al.,

392 2011). Understanding the morphological variability and evolvability produced by the developmental

393 process is an important step in validating phylogenetic hypotheses based on the OES morphology alone.

394

395 Concluding Remarks

Both morphometric and variability analyses indicate that tooth shape and covariation
structure is not only determined by processes that contribute to tooth shape at the EDJ, but also
amelogenesis can play a significant role in them. The influence of enamel formation on morphological
variation and patterns of variability is not constant among teeth, which may be responsible for the
differences in the rate and/or period of enamel formation.

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411	
412	Author contributions
413	W.M. and W.Y. designed the research and performed the analysis. W.M., T.N., and M.A.

414 collected the data. W.M., H.O., and M.N. wrote the manuscript.

415 **References**

- 416 Alberch P (1982) Developmental constraints in evolutionary processes. In: Development in evolution
- 417 (ed. Bonner JT), pp. 313–332. Berlin and New York: Springer-Verlag.
- 418 Avishai G, Müller R, Gabet Y, Bab I, Zilberman U, Smith P (2004) New approach to quantifying
- 419 developmental variation in the dentition using serial microtomographic imaging. *Microsc Res Tech* 65,
- 420 263–299.
- 421 Bookstein FL (1997) Landmark methods for forms without landmarks: morphometrics of group
- 422 differences in outline shape. *Med Image Anal* 1, 225–243.
- 423 Boughner JC (2011) Making space for permanent molars in growing baboon (*Papio anubis*) and great
- 424 ape (Pan paniscus and P. troglodytes) mandibles: Possible ontogenetic strategies and solutions. Anat
- 425 *Res* Int doi.org/10.1155/2011/484607.
- 426 Boyde A (1964) The structure and development of mammalian enamel. PhD dissertation. University of
- 427 London.
- 428 Boyde A (1989) Enamel. In: Handbook of Microscopic Anatomy, Volume 6: Teeth (eds. Oksche A
- 429 and Vollrath L), pp. 309–473. Berlin and New York: Springer-Verlag.
- 430 Butler PM (1956) The ontogeny of molar pattern. *Biol Rev* 31, 30–70.
- 431 Butler PM (1971) Growth of human tooth germs. In: *Dental Morphology and Evolution* (ed. Dahlberg
- 432 AA), pp. 3–14. Chicago: University of Chicago Press.
- 433 Cai J, Cho SW, Kim JY, Lee MJ, Cha YG, Jung HS (2007) Patterning the size and number of tooth and
- 434 its cusps. *Dev Biol* 304, 499–507.
- 435 Carroll SB (2008) Evo-devo and expanding evolutionary synthesis: A genetic theory of morphological

- 436 evolution. *Cell* 134, 25–36.
- 437 Chernoff B, Magwene PM (1999) Morphological Integration: Forty Years Later. In: Morphological
- 438 Integration (eds. Olsen EC, Miller RL), pp. 319–353. Chicago: University of Chicago Press.
- 439 Cheverud JM (1996) Developmental integration and the evolution of pleiotropy. Am Zool 36, 44–50.
- 440 Collard M, Wood B (2000) How reliable are human phylogentic hypotheses? Proc Natl Acad Sci USA
- 441 97, 5003–5006.
- 442 Collyer ML, Adams DC (2007) Analysis of two-state multivariate phenotypic change in ecological
- 443 studies. *Ecology* 88, 683–692.
- 444 Corruccini RS (1998) The dentino-enamel junction in primate mandibular molars. In: Human Dental
- 445 Development, Morphology, and Pathology: A Tribute to Albert A. Dahlberg (ed. Lukacs JR), pp. 1–16.
- 446 Portland: University of Oregon Anthropological Papers.
- 447 Dahlberg AA (1945) The changing dentition of man. JAm Dent Assoc 32, 676–690.
- 448 Dean MC (2000) Progress in understanding hominoid dental development. J Anat 197, 77–101.
- 449 Escoufier Y (1973) Le traitement des variables vectorielles. *Biometrics* 29, 751–760.
- 450 Finarelli JA, Clyde WC (2004) Reassessing hominoid phylogeny: evaluating congruence in the
- 451 morphological and temporal data. *Paleobiol* 30, 614–651.
- 452 Fincham AG, Luo W, Moradian-Oldak J, Paine ML, Snead ML, Zeichner-David M (2000) Enamel
- 453 biomineralization: the assembly and dissassembly of the protein extracellular organic matrix. In:
- 454 Development, Function and Evolution of Teeth (eds. Teaford MF, Meredith-Smith M, Ferguson MWJ),
- 455 pp. 37–61. Cambridge: Cambridge University Press.
- 456 Grabowski MW (2013) Hominin obstetrics and the evolution of canstraints. *Evol Biol* 40, 57–75.

- 457 Grine FE (2005) Enamel thickness of deciduous and permanent molars in modern Homo sapiens. Am J
- 458 *Phys Anthropol* 126, 14–31.
- 459 Gunz P, Mitteroecker P, Bookstein FL (2005) Semilandmarks in three dimensions. In: Modern
- 460 Morphometrics in Physical Anthropology (ed. Slice DE), pp. 73–98. New York: Kluwer
- 461 Academic/Plenum Publishers.
- 462 Hallgrímsson B, Willmore K, Hall BK (2002) Canalization, developmental stability, and morphological
- 463 integration in primate limbs. *Am J Phys Anthropol* 35, 131–158.
- 464 Hallgrímsson B, Jamniczky H, Young NM, et al. (2009) Deciphering the palimpsest: Studying the
- 465 relationship between morphological integration and phenotypic covariation. *Evol Biol* 36, 355–376.
- 466 Hansen TF (2003) Is modularity necessary for evolvability? Remarks on the relationship between
- 467 pleiotropy and evolvability. *Biosystems* 69, 83–94.
- 468 Harjunmaa E, Kallonen A, Voutilainen M, Hämäläinen K, Mikkola ML, Jernvall J (2012) On the
- 469 difficulty of increasing dental complexity. *Nature* 483, 324–327.
- 470 Harris EF, Dinh DP (2006) Intercusp relationships of the permanent maxillary first and second molars
- 471 in American whites. *Am J Phys Anthropol* 130, 514–28.
- 472 Hlusko LJ (2004) Integrating the genotype and phenotype in hominid paleontology. Proc Natl Acad Sci
- 473 *USA* 101, 2653–2657.
- 474 Hlusko LJ, Suwa G, Kono R, Mahaney MC (2004) Genetics and the evolution of primate enamel
- 475 thickness: A baboon model. Am J Phys Anthropol 124, 223–233.
- 476 Hunter JP, Jernvall J (1995) The hypocone as a key innovation in mammalian evolution. Proc Natl
- 477 *Acad Sci U S A* 92, 10718–10722.

- 478 Jernvall J, Jung HS (2000) Genotype, phenotype, and developmental biology of molar tooth characters.
- 479 *Year Phys Anthropol* 43, 171–190.
- 480 Kavanagh KD, Evans AR, Jernvall J (2007) Predicting evolutionary patterns of mammalian teeth from
- 481 development. *Nature* 449, 427–432.
- 482 Klingenberg CP (2008) Morphological integration and developmental modularity. Annu Rev Ecol Evol
- 483 Syst 39, 115–132.
- 484 Klingenberg CP (2011) MorphoJ: an integrated software package for geometric morphometrics. *Molec*
- 485 *Ecol Res* 11, 353–357.
- 486 Korenhof CAW (1960) Morphogenetical Aspects of the Human Upper Molar. Utrecht:
- 487 Uitgeversmaatschappij Neerlandia.
- 488 Korenhof CAW (1961) The enamel-dentine border: a new morphological factor in the study of the
- 489 (human) molar pattern. Proc Koninkl Nederl Acad Wetensch 64B, 639–664.
- 490 Korenhof CAW (1982) Evolutionary trends of the inner enamel anatomy of deciduous molars from
- 491 Sangiran (Java, Indonesia). In: Teeth: Form, Function and Evolution (ed. Kurtén B), pp. 350–365. New
- 492 York: Columbia University Press.
- 493 Kraus BS (1952) Morphologic relationships between enamel and dentin surfaces of lower first molar
- 494 teeth. J Dent Res 31, 248–256.
- 495 Kraus BS, Jordan RE (1965) The Human Dentition Before Birth. Philadelphia: Lea and Febiger.
- 496 Lande R (1979) Quantitative genetic analysis of multivariate evolution, applied to brain: Body size
- 497 allometry. *Evolution* 33, 402–416.
- 498 Lande R, Arnold SJ (1983) The measurement of selection on correlated characters. Evolution 37, 1210–

499 1226.

- 500 Lewton KL (2012) Evolvability of the primate pelvic girdle. *Evol Biol* 39, 126–139.
- 501 Liversidge HM, Molleson T (2004) Variation in crown and root formation and eruption of human
- 502 deciduous teeth. Am J Phys Anthropol 123, 172–180.
- 503 Manly BFJ (1997) Randomization, bootstrap and Monte Carlo methods in biology. London: Chapman
- 504 & Hall.
- 505 Matsumura H, Domett KM, O'reilly DJW (2011) On the origin of pre-Angkorian peoples: perspectives
- 506 from cranial and dental affinity of the human remains from Iron Age Phum Snay, Cambodia. *Anthropol*
- 507 *Sci* 119, 67–79.
- Maynard Smith J, Burian R, Kauffman S, et al. (1985) Developmental constraints and evolution. *Q Rev Biol* 60, 265–287.
- 510 Marroig G, Shirai L, Porto A, de Oliveira F, De Conto V (2009) The evolution of modularity in the
- 511 mammalian skull II: Evolutionary consequences. *Evol Biol* 36, 136–148.
- 512 Miller GS (1918) The Piltdown jaw. *Am J Phys Anthropol* 1, 25–52.
- 513 Nager G, (1960) Der vergleich zwischen dem räumlichen verhalten des dentinkronenreliefs und dem
- 514 schmelzrelief der zahnkrone. Acta Anat 42, 226–250.
- 515 Nanci A (2013) Ten Cate's Oral Histology: Development, Structure and Function, 8th Edition. St.
- 516 Louis: Mosby Elsevier.
- 517 Olejniczak AJ, Gilbert CG, Martin LB, Smith TM, Ulhaas L, Grine FE (2007) Morphology of the
- 518 enamel-dentine junction in sections of anthropoid primate maxillary molars. *J Hum Evol* 53, 292–301.
- 519 Ortiz A, Skinner MM, Bailey SE, Hublin JJ (2012) Carabelli's trait revisted: an examination of

- 520 mesiolingual features at the enamel-dentine junction and enamel surface of Pan and Homo sapiens
- 521 upper molars. *J Hum Evol* 63, 586–596.
- 522 Pilbrow V (2006) Population systematics of chimpanzees using molar morphometrics. *J Hum Evol* 51,
 523 646–662.
- 524 Porto A, de Oliveira FB, Shirai LT, de Conto V, Marroig G (2009) The evolution of modularity in the
- 525 mammalian skull I: Morphological integration patterns and magnitudes. *Evol Biol* 36, 118–135.
- 526 R Development Core Team (2011) R: A Language and Environment for Statistical Computing. Vienna:
- 527 R Foundation for Statistical Computing. http://cran.R-project.org.
- 528 Rohlf FJ, Slice D (1990) Extensions of the Procrustes method for the optimal superimposition of
- 529 landmarks. *Syst Zool* 39, 40–59.
- 530 Sakai T, Hanamura H (1971) A morphology study of enamel-dentin border on the Japanese dentition.
- 531 Part V. Maxillary molar. J Anthropol Soc Nippon 79, 297–322.
- 532 Sakai T, Hanamura H (1973) A morphology study of enamel-dentin border on the Japanese dentition.
- 533 Part VII. General conclusion. J Anthropol Soc Nippon 81, 87–102.
- 534 Sasaki K, Kanazawa E (1999) Morphological traits on the dentino-enamel junction of lower deciduous
- 535 molar series. In: Dental morphology 1998: Proceedings of the 11th international symposium on dental
- 536 *morphology* (eds. Mayhall J, Heikkinen T), pp. 167–178. Oulu: Oulu University Press.
- 537 Saunders SR, Mayhall JT (1982) Developmental patterns of human morphological traits. Archs Oral
- 538 Biol 27, 45–49.
- 539 Shellis RP (1984) variations in growth of the enamel crown in human teeth and a possible relationship
- 540 between growth and enamel structure. *Archs Oral Biol* 29, 697–705.

- 541 Simmer JP, Papagerakis P, Smith CE, et al. (2010) Regulation of dental enamel shape and hardness. J
 542 Dent Res 89, 1024–1038.
- 543 Simons EL, Pilbeam D (1972) Hominoid paleoprimatology. In: The Functional and Evolutionary
- 544 *Biology of Primates* (ed. Tuttle R), pp. 36–62. Chicago: Aldine-Atherton.
- 545 Skinner MM, Wood BA, Boesch C, et al. (2008) Dental trait expression at the enamel-dentine junction
- of lower molars in extant and fossil hominoids. *J Hum Evol* 54, 173–186.
- 547 Skinner MM, Wood BA, Hublin JJ (2009) Protostylid expression at the enameledentine junction and
- 548 enamel surface of mandibular molars of Paranthropus robustus and Australopithecus africanus. J Hum
- 549 Evol 56, 76–85.
- 550 Skinner MM, Evans A, Smith T, et al. (2010) Brief Communication: Contributions of enamel-dentine
- 551 junction shape and enamel deposition to primate molar crown complexity. Am J Phys Anthropol 142,
- 552 157–163.
- 553 Skinner MM, Gunz P (2010) The presence of accessory cusps in chimpanzee lower molars is consistent
- 554 with a patterning cascade model of development. J Anat 217, 245–253.
- 555 Smith P, Gomorri JM, Spitz S, Becker J (1997) Model for the examination of evolutionary trends in
- tooth development. Am J Phys Anthropol 102, 283–294.
- 557 Smith P, Gomori JM, Shaked R, Haydenblit R, Joskowicz L (2000) A computerized approach to
- 558 reconstruction of growth patterns in hominid molar teeth. In: Proceedings of the 11th International
- 559 Symposium on Dental Morphology (eds. Mayhall J, Heikkinen T), pp. 388–397. Oulu: Oulu University
- 560 Press.
- 561 Smith P, Avishai G, Muller R, Gabet Y (2011) Computerized reconstruction of prenatal growth

- 562 trajectories in the dentition: Imprications for the taxonomic status of Neandertals. In: Continuity and
- 563 discontinuity in the peopling of Europe: One hundred fifty years of Neanderthal study (eds. Condemi S,
- 564 Weniger G-C), pp. 165–173. New York: Springer Science+Business Media B.V.
- 565 Sokal RR, Braumann CA (1980) Significance tests for coefficients of variation and variability profiles.
- 566 Syst Zool 29, 50–66.
- 567 Suzuki M, Sakai T (1973) Occlusal surface pattern of the lower molars and the second deciduous molar
- among living Polynesians. Am J Phys Anthropol 39, 305–315.
- 569 Townsend G, Richards L, Hughes T (2003) Molar intercuspal dimensions: genetic input to phenotypic
- 570 variation. J Dent Res 82, 350–355.
- 571 Villmoare B, Fish J, Jungers W (2011) Selection, morphological integration, and strepsirrhine
- 572 locomotor adaptations. *Evol Biol* 38, 88–99.
- 573 Wagner GP (1984) On the eigenvalue distribution of genetic and phenotypic dispersion matrices:
- 574 Evidence for a nonrandom organization of quantitative character variation. J Math Biol 21, 77–95.
- 575 Wagner GP, Altenberg L (1996) Complex adaptations and the evolution of evolvability. *Evolution* 50,
- 576 967–976.
- 577 Wagner GP, Booth G, Bagheri-Chaichian H (1997) A population genetic theory of canalization.
- 578 *Evolution* 51, 329–347.
- 579 Willmore KE, Young N, Richtsmeier JT (2007) Phenotypic variability: its components, measurement
- 580 and underlying developmental processes. *Evol Biol* 34, 99–120.

581 Tables

Table 1 Results of PLS analyses between EDJ and OES corresponding to UM1 and um2

		UM1			um2	
	% Total	Correlation	P-value ¹	% Total	Correlation	P-value ¹
	Cov.	coefficient		Cov.	coefficient	
1	49.43	0.951	< 0.001	43.14	0.974	<0.001
2	17.39	0.933	< 0.001	25.11	0.970	< 0.001
3	14.65	0.908	< 0.001	17.76	0.954	< 0.001
4	10.22	0.879	< 0.001	6.52	0.948	< 0.001

¹Randomiztion rounds: 1000

582

Table 2 Results of principal component analysis with the total sample

	Eigenvalue	% Explained variance	% Cumulative variance
1	0.0016	19.99	19.99
2	0.0012	14.86	34.85
3	0.0009	11.80	46.64
4	0.0007	9.08	55.73
5	0.0005	6.86	62.58
6	0.0005	6.68	69.26
7	0.0004	5.31	74.58
8	0.0002	3.14	77.71
9	0.0002	2.90	80.61
10	0.0002	2.35	82.96

584 **Figure legends**

585 Figure 1. Digital image of permanent maxillary first molar crown (lingual view). (a) EDJ ridge curve

586 digitized on the EDJ surface. (b) OES ridge curve digitized on the OES. Red circles are landmarks, and

587 yellow circles are semi-landmarks. Numbers appended to each section of the ridge curve refer to the

588 equally-spaced interpolated semi-landmarks.

589

Figure 2. Scatter plots representing the first and second pairs of PLS axes between EDJ and OES within the same tooth class. (a) PLS1 UM1, (b) PLS2 UM1, (c) PLS1 um2, (d) PLS2 um2. Shape deformation corresponding to each axis is provided to the left of x-axes or above y-axes. Each shape deformation is represented in colored line whose scale factor used for is 0.1 and mean shape is represented in gray line.

595	Figure 3. Pri	ncipal com	ponent plot	s for shape	variation bet	ween EDJ a	nd OES of	f both UM1	and um2.
	<i>L</i>)								

(a) Plots of PC1 versus PC2 scores. Variance explained by PC1 and PC2 is 34.85% of total variance.

597 Shape deformation corresponding to the positive or negative loadings of each axis is provided to the left

598 and right for x-axes or the above and bottom for y-axes. Each shape deformation is represented in

599 colored line whose scale factor used for is 0.1 and mean shape is represented in gray line. Arrows show

600 morphological change vectors from mean shape represented in large symbols of EDJ to that of OES for

- each tooth class. (b) Relationship between EDJ and OES for PC1 in both UM1 and um2. The slope and
- 602 intercept of the regression line for UM1 are 0.804 and -0.070, respectively (r=0.51, P<0.001). The slope
- and intercept of the regression line for um2 are 0.876 and -0.068, respectively (r=0.84, P<0.001). (c)
- Relationship between EDJ and OES for PC2 in both UM1 and um2. The slope and intercept of the

605	regression line for UM1 are 0.863 and -0.002, respectively (r=0.92, P<0.001). The slope and intercept of
606	the regression line for um2 are 0.918 and 0.007, respectively (r=0.95, P<0.001).
607	

608	Figure 4. (a) Bar graph showing the size variation for four configurations (UM1EDJ, UM1OES,
609	um2EDJ and um2OES). Significance test for coefficient of variation for LogCS among them reveals
610	that there is no significant difference (P>0.05). (b) Bar graph showing mean of properustes distance
611	from each mean shape for shape variance of four configurations (UM1EDJ, UM1OES, um2EDJ and
612	um2OES), and the error bars show standard deviations. The Kruskall-Wallis test reveals a significant
613	difference among them (P<0.001). A nonparametric multiple-comparison test between EDJ and OES
614	within the same tooth class reveals that the difference is highly significant in UM1 (P<0.001). (c) Bar
615	graph showing the scaled variances of eigenvalue for morphological integration for four configurations
616	(UM1EDJ, UM1OES, um2EDJ and um2OES). The error bars shown are standard deviations obtained
617	by resampling the original datasets with replacement 1000 times. Bootstrap tests between EDJ and OES
618	within the same tooth class reveal that the difference is highly significant only in UM1 (P=0.009). (d)
619	Bar graph showing the evolutionary flexibility for four configurations (UM1EDJ, UM1OES, um2EDJ
620	and um2OES). The error bars shown are standard deviations obtained by resampling the original
621	datasets with replacement 1000 times. Bootstrap tests between EDJ and OES within the same tooth
622	class reveal that there is no significant difference (P>0.05).



















