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Dynamics of receptor-operated Ca\textsuperscript{2+} currents controlled via the PI(4,5)P\textsubscript{2}-PLC signaling pathway

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Transient receptor potential canonical (TRPC) channels are Ca\textsuperscript{2+}-permeable, nonselective cation channels that carry receptor-operated Ca\textsuperscript{2+} currents (ROCs) triggered by receptor-induced, phospholipase C (PLC)-catalyzed hydrolysis of phosphatidylinositol 4,5-bisphosphate [PI(4,5)P\textsubscript{2}]. Within the vasculature, TRPC channel ROCs contribute to smooth muscle cell depolarization, vasoconstriction, and vascular remodeling. However, TRPC channel ROCs exhibit a variable response to receptor-stimulation, and the regulatory mechanisms governing TRPC channel activity remain obscure. The variability of ROCs may be explained by their complex regulation by PI(4,5)P\textsubscript{2} and its metabolites, which differentially affect TRPC channel activity. To resolve the complex regulation of ROCs, the use of voltage-sensing phosphoinositide phosphatases and model simulation have helped to reveal the time-dependent contribution of PI(4,5)P\textsubscript{2} and the possible role of PI(4,5)P\textsubscript{2} in the regulation of ROCs. These approaches may provide unprecedented insight into the dynamics of PI(4,5)P\textsubscript{2} regulation of TRPC channels and the fundamental mechanisms underlying transmembrane ion flow. Within that context, we summarize the regulation of TRPC channels and their coupling to receptor-mediated signaling, as well as the application of voltage-sensing phosphoinositide phosphatases to this research. We also discuss the controversial bidirectional effects of PI(4,5)P\textsubscript{2} using a model simulation that could explain the complicated effects of PI(4,5)P\textsubscript{2} on different ROCs.

Keywords: receptor-operated calcium current, TRPC channels, PIP2, voltage-sensing phosphatase, Ca\textsuperscript{2+} signaling, smooth muscle

INTRODUCTION ~RECEPTOR-OPERATED Ca\textsuperscript{2+} CURRENTS~

Calcium is a ubiquitous and fundamental messenger that triggers numerous downstream cellular events, including hormone secretion, vasoconstriction, and activity-dependent gene expression, to name a few (Berridge, 2012). There are several mechanisms by which Ca\textsuperscript{2+} signals are generated (Parekh and Putney, 2005). These include voltage-dependent Ca\textsuperscript{2+} influx, Ca\textsuperscript{2+} release from intracellular stores, and store-operated Ca\textsuperscript{2+} influx in response to depletion of the Ca\textsuperscript{2+} stores. In this paper, we will focus on phospholipase C (PLC)-driven Ca\textsuperscript{2+} influx, often described as receptor-operated Ca\textsuperscript{2+} currents (ROCs). The Ca\textsuperscript{2+} signal mediated by ROCs differs from that provided by voltage-dependent Ca\textsuperscript{2+} influx (Somlyo and Somlyo, 1994). However, comparatively little is known about the properties of ROCs and their underlying mechanisms. It is known that in various vertebrate cell types ROCs are generated through the breakdown of phosphatidylinositol 4,5-bisphosphate [PI(4,5)P\textsubscript{2}] by PLC upon receptor stimulation and are marked by Ca\textsuperscript{2+}/Na\textsuperscript{+} influx (Bolton, 1979; Putney and Tomita, 2012). The role of ROCs has also been studied in the Drosophila phototransduction pathway, wherein TRP channels mediate cation currents in response to photoreceptor activation (Hardie, 2011; Montell, 2011).

Several TRP channel homologs, known as transient receptor potential canonical (TRPC; canonical), have been cloned from mammalian (Ramsey et al., 2006). Among of these, TRPC2, 3, 6, 7 channels can be activated by diacylglycerol (DAG), a potent lipid messenger produced from PI(4,5)P\textsubscript{2} by PLC activation (Hofmann et al., 1999). It has been suggested that activation of TRPC4, 5 is PLC-dependent, but with no detectable contribution of DAG (Schaefer et al., 2000). Nevertheless, the linkage between PLC-coupled receptors and TRPC channels is almost universally accepted, and the resulting Ca\textsuperscript{2+} influx is considered to be a ROC (Figure 1A).

PHYSIOLOGICAL PATHWAY FOR ROCs OF TRPC CURRENTS

Transient receptor potential canonical channels are also widely distributed in various other tissues (Beech, 2013). Thus, the upstream of TRPC channels can be diverse, reflecting the physiological context. Within the autonomic nervous system, receptor ligands (e.g., noradrenaline and acetylcholine) are released as transmitters from efferent and afferent sympathetic and parasympathetic nerve fibers at target organs. The effects of noradrenaline are mediated via activation of adrenergic receptors, including the α1A receptor, which is abundantly expressed on venous smooth muscle cells, and has been shown to induce ROCs through...
TRPC6 channels (Inoue et al., 2001). In cerebellar granule cells, brain-derived neurotrophic factor (BDNF)-induced Ca^{2+} elevation by TRPC channels has been shown to play an essential role in nerve growth cones guidance (Li et al., 2005). Furthermore, pathological contributions of ROCs of TRPC channels have been shown in development of hypertrophy (Onohara et al., 2006; Wu et al., 2010; Seo et al., 2014) and genetic kidney disease ‘focal segmental glomerulosclerosis (FSGS)’ (Mukerji et al., 2007). To emphasize physiological contribution of ROCs, a list of the agonists, receptors, PLC subtypes, and TRPC channels, and their confirmed linkage is presented in the supplementary material (Table S1).

**DYNAMICS OF ROCs**

Transient receptor potential canonical channel ROCs can exhibit slow \( \alpha \)-shaped time dependence or an initial rapid spike followed by a sustained response, depending on the strength of the receptor stimulation (Figure 1B). The activation phase of ROCs shows a facilitative or growing curve, irrespective of the agonist concentration applied (Figure 1B,a). It has been demonstrated that cytosolic Ca^{2+} potentiates activity of TRPC5 channels (Blair et al., 2009), however, the facilitative responses induced by receptor agonists are less clear to PI(4,5)P2. For example, PI(4,5)P2 exerts an inhibitory effect on the Drosophila TRPC channel, which prompted an intriguing proposal that reductions in the PI(4,5)P2 concentration due to PLC hydrolysis may be sufficient to evoke ROCs. In a recent study, however, reducing PI(4,5)P2 levels in the absence of PLC activity through rapamycin-induced yeast PI(4,5)P2 phosphatase had no effect on TRPL channel activation (Lev et al., 2012).

The decay phase following the peak exhibits an even more curious. Under a low-dose of an agonist application, ROCs gradually disappear without a plateau phase (Figure 1B,b). On the other hand, at higher agonist doses ROCs often demonstrate fast inactivation followed by a plateau phase (Figure 1B,c). The plateau phase of ROCs also appears in Drosophila photoreceptors, where it is known to be dependent on the intensity of the light stimulation. When the light stimulus is very dim, the photoreceptor-operated currents decay to baseline without a clear plateau phase. Brighter stimuli elicit a plateau phase and shortened the decay time from the plateau to baseline (Minke, 1982).

**CONTROVERSIAL PI(4,5)P2 EFFECTS IN TRP CHANNELS**

Phosphatidylinositol 4,5-bisphosphate is located in the inner leaflet of the plasma membrane. In addition to being a substrate for hydrolysis by PLC, PI(4,5)P2 plays key roles in the regulation of cytoskeletal organization, cell motility, and a number of ion conducting proteins (Ball, 2013). In mammals, 20 of the 28 known TRP channel subtypes are regulated by PI(4,5)P2. However, studies of PI(4,5)P2 regulation have often reached differing conclusions. For example, in TRPV1, one of the best characterized members of the TRP superfamily, PI(4,5)P2 may positively or negatively regulate channel activity (for review, see Rohacs, 2013). In addition, TRPV4 channels, recently further added to the controversy surrounding PI(4,5)P2 function, PI(4,5)P2 appears to suppress TRPV4 channel activity by binding to an ankyrin domain, while on the other hand PI(4,5)P2 can also facilitate TRPV4 channel activity through binding to a N-terminal region separate from the ankyrin domain (Garcia-Elias et al., 2013; Taka-hashi et al., 2014), which implies domain-specific regulation of TRPV4 activation. Furthermore, the effect of PI(4,5)P2 on TRPV4 channel activation can depend on the stimulus in the heat or osmo vs chemical (4-α-PPD) stimulations. Under physiological conditions, the breakdown of PI(4,5)P2 by PLC is required for activation of TRPC and TRPL channels. Nonetheless, as with the aforementioned TRPV channels, the effect of PI(4,5)P2 is controversial. It is noteworthy that the controversial reports, which are well-reviewed (Rohacs, 2013), utilized different cell settings and different approaches to manipulating PI(4,5)P2 levels, which raises questions as to whether the PI(4,5)P2 manipulations in these studies are comparable (Mori and Inoue, 2014).
VSP AS A MODERN TOOL FOR STUDYING PI(4,5)P₂ REGULATION

Voltage-sensing phosphatase is a newly standardized tool for studying ion channel regulation by phosphoinositides that provide high temporal resolution and is controllable through the membrane potential. So far, two VSPs from aquatic species have been being applied to ion channel studies. *Ciona intestinalis* (Ci) VSP was the first to be identified as a voltage-sensing phosphoinositide phosphatase (Murata et al., 2005). Identified later was the VSP from *Danio rerio* (Dr), which exhibits only small differences in the substrate specificity from Ci-VSP, mainly in its voltage-sensitivity (Hossain et al., 2008).

Membrane depolarization under voltage-clamp activates VSPs within a few milliseconds, enabling detection of transient effects on ion channels (Figure 2A, onset). Hille’s group utilized Dr-VSP, and by measuring onset time course of the current reduction, they estimated the residence time of PI(4,5)P₂ on the KCNQ2/3 channels to be less than 10 ms, which is more than 400 times faster than the previous estimation (Falkenburger et al., 2010). Because VSPs are only activated during membrane depolarization, upon repolarization of the membrane, PI(4,5)P₂ levels, and thus channel currents, are able to recover due the presence of endogenous phosphatidylinositol 4-phosphate 5-kinase (PIP5K), which catalyzes position 5 of the inositol ring (Di Paolo and De Camilli, 2006). The time course of the channel current recovery (Figure 2A, recovery) is illuminating and reflects the time constant for the functional re-association of PI(4,5)P₂ with the ion channel. Intriguingly, the recovery constant for the KCNQ2/3 channel and N-type Ca²⁺ channel inhibition is more than 10 s, which is much slower than in TRPC3/6/7 channels (1–4 s) or TRPM8 channels (4 s; Falkenburger et al., 2010; Suh et al., 2010; Yudin et al., 2011; Imai et al., 2012). According to the time constants of onset and recovery, higher affinity of PI(4,5)P₂ for TRPC3 and TRPC6 channels has suggested than that for the KCNQ2/3 or N-type Ca²⁺ channels.

To determine the affinity of PI(4,5)P₂ for ion channels in another voltage clamp study, we controlled the magnitude of PI(4,5)P₂ reductions by Dr-VSP activation using a step-pulse protocol, which enabled us to determine the affinities of PI(4,5)P₂ for TRPC3/6/7 channels. The Kₐ values obtained ranged from 1 to 10 μM (Itsuki et al., 2014), indicating higher affinity than that of PI(4,5)P₂ for KCNQ2/3 channels (EC₅₀ ≈ 87 μM; Zhang et al., 2003). This raises the possibility that the affinity of PI(4,5)P₂ for TRPC channels could be too strong to exert regulatory effects, given the physiological PI(4,5)P₂ concentration. However, the PI(4,5)P₂ concentration in cells has been estimated to range from 5 to 10 μM and may vary by around 50% upon activation of signaling (Xu et al., 2003; McLaughlin and Murray, 2005). This concentration range clearly overlaps the dissociation constants for TRPC channels and is consistent with the physiological relevance of PI(4,5)P₂ regulation. Furthermore, evidence of physical coupling between PLC and TRPC channels has been provided (van Rossum et al., 2005), local PI(4,5)P₂ depletion may possible to happen nearby TRPC channels.

COMPARISON OF TRPC6 CHANNELS INHIBITION BETWEEN CI-, DR-, AND GG-VSPs

Voltage-sensing phosphatase In addition to sea ascidian (Ci) and zebrafish (Dr), VSPs have also been identified in chicken (*Gallus gallus*, Gg; Yamaguchi et al., 2014). Among these three VSPs,
When we then fit simulations generated with the new model to the data from TRPC6 currents, we found that bi-directional PI(4,5)P2 regulation was possible if the negative effect site for PI(4,5)P2 was different from the positive effect site (Table S2). Phosphatidylinositol 4,5-bisphosphate association sites as well as its physiological importance remains to be addressed in future studies. TRPC channel ROCs show great variation, and the mechanisms and physiological consequences of that variation are not yet fully understood. Precise detection of signals elicited by receptor stimulation and clear cut evidence of the regulatory factors, including PI(4,5)P2, Ca2+, will aid our understanding of ROCs within pathophysiological responses.

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**SUPPLEMENTARY MATERIAL**

The Supplementary Material for this article can be found online at: http://www.frontiersin.org/journal/10.3389/fphar.2015.00022/abstract

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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