

1 **Ecology has contrasting effects on genetic variation within species versus rates of**
2 **molecular evolution across species in water beetles**

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16

17 **Abstract**

18 Comparative analysis is a potentially powerful approach to study effects of ecological traits on
19 genetic variation and rate of evolution across species. However, the lack of suitable datasets means
20 that comparative studies of correlates of genetic traits across an entire clade have been rare. Here,
21 we use a large DNA-barcode data set (5062 sequences) of water beetles to test the effects of species
22 ecology and geographical distribution on genetic variation within species and rates of molecular
23 evolution across species. We investigated species traits predicted to influence the genetic
24 characteristics, such as surrogate measures of species population size, latitudinal distribution and
25 habitat types, taking phylogeny into account. Genetic variation of cytochrome oxidase I of water
26 beetles was positively correlated with occupancy (counts of sites of species presence) and
27 negatively with latitude, whereas substitution rates across species depended mainly on habitat types,
28 and running water specialists had the highest rate. These results are consistent with theoretical
29 predictions from nearly neutral theories of evolution, and suggest that the comparative analysis
30 using large databases can give insights into correlates of genetic variation and molecular evolution.

31

32 *Key words:* CO1; genetic variation; rate of evolution; PGLS; water beetles

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34

35 **Introduction**

36 Genetic variation is a key parameter determining how populations evolve. Consequently, a central
37 goal of population genetic studies is to understand the factors that control genetic variation in
38 populations (Leffler et al. 2012). The neutral theory of molecular evolution predicts genetic
39 variation based on mutation and drift alone. Under the assumption of selective neutrality of alleles,
40 genetic variation of a population is proportional to the product of effective population size and
41 mutation rate (Kimura 1984). Theories have been developed to incorporate the other main factors
42 that can affect neutral genetic variation, such as population structure (Charlesworth et al. 2003) and
43 demographic history (Knowles 2006). These parameters are widely used to interpret population
44 genetic data. However, there have been relatively few comparative tests of which factors correlate
45 with genetic variation across multiple species (Bazin et al. 2006). A comparative study can reveal
46 which parameters identified by theory are most important in explaining levels of genetic variation
47 across species. However, this requires detailed data across a large enough genetic sample for
48 numerous closely related species.

49 Neutral theories of molecular variation have also been applied to the study of DNA
50 substitution rates across species. Phylogenetic dating methods using molecular clocks have found
51 considerable rate variation among taxa, which led to interest in the causes of that variation
52 (Bromham & Penny 2003; Lanfear et al. 2010). Many studies have investigated the importance of
53 generation times, metabolic rates, environmental energy inputs and other potential correlates of
54 neutral or nearly neutral substitution rates (Davies et al. 2004; Welch et al. 2008; Santos 2012).
55 However, population genetic parameters such as population size have been hard to incorporate
56 because of the general lack of sequence data across a sufficiently broad sample of species.
57 Estimates of effective population sizes from relatively few well-known model organisms have been
58 used for comparison with molecular variation (Gossmann et al. 2012), but these have limited
59 potential to tease apart the effects of multiple factors. Matched pairs of island and mainland
60 populations have demonstrated an elevated rate of nonsynonymous substitution (interpreted as

61 fixation of mildly deleterious mutations) in island populations with small effective population size
62 (Woolfit & Bromham 2005). The matched species pair design is elegant for investigating a single
63 factor, but cannot easily allow the investigation of multiple factors simultaneously if each pair is
64 distantly related from each other pair.

65 The increasing availability of within-species DNA sequence of putatively neutral markers
66 for large clades, associated with the rise of DNA barcoding, offers the potential for comparative
67 analysis of population genetic variation and substitution rates in concert. For example, if mutation
68 rates alone vary systematically among different species, then genetic variation within species should
69 correlate with substitution rates over longer evolutionary timescales. However, if population size
70 variation is more important, then we predict different associations: genetic variation should
71 correlate positively with measures of population size (Manier & Arnold 2006), whereas substitution
72 rates should be independent of population size under strict neutrality or increase with decreasing
73 population size if variants are nearly neutral (i.e. mildly deleterious, Ohta 1992). Alternatively, we
74 might find no evidence of predicted correlations if the parameters affecting present-day genetic
75 variation have been inconsistent over the longer evolutionary timescales relevant for substitution
76 rate differences.

77 Dispersal is another important factor to determine species' genetics by controlling
78 connectivity between local populations and the chance of colonization into newly opened habitats.
79 The effects of species' dispersal on genetic variation have been widely studied, and higher dispersal
80 ability is commonly associated with less structured populations (Burney & Brumfield 2009,
81 Papadopoulou et al. 2011; Riginos et al. 2014). For example, Riginos et al. (2014) reported that
82 more dispersive "pelagic spawner" fish have less structured populations than benthic guarders. The
83 effect of dispersal on the rate of evolution is less widely studied. Faster evolution in less dispersive
84 species is observed in darkling beetles (Papadopoulou et al. 2009; Papadopoulou et al. 2011), but
85 this has not been formally tested in a comparative analysis. The broad database of barcoding
86 sequences enables us to test the relative importance of these multiple factors.

87 Here, we use a large sequence dataset of mitochondrial DNA of water beetles (Baselga et al.
88 2013) to test the effect of species ecology on their genetic traits taking other confounding factors
89 into account. Mitochondrial genes have been most widely sampled for comparative studies of
90 molecular evolution because of their favorable characteristics: supposed neutrality, simple maternal
91 inheritance without recombination and easy amplification. Although the use of mtDNA for a
92 population genetic marker has been questioned (Ballard & Whitlock 2004; Meiklejohn et al. 2007;
93 Galtier et al. 2009), because of introgression and the potential for selective sweeps that violate
94 assumption of neutrality, we know of no nuclear DNA dataset of equivalent breadth and depth
95 within a single clade.

96 The data set includes not only genetic information of species but also geographic
97 distribution and habitat descriptions of samples, which can be used to measure other factors
98 influencing their genetic traits. For example, as a surrogate estimate of the population size of each
99 species, which is a fundamental parameter affecting genetic variation but hard to measure in natural
100 populations, we use species occupancy and distribution estimated from geographical collection
101 records since species occupancy typically correlates well with abundance (Gaston et al. 2000;
102 Blackburn et al. 2006; Gaston & He 2011). Latitudinal distribution of species, which is reported to
103 affect the rate of evolution, is obtained from sampling records. The genetic information also allows
104 DNA-based species delimitation and circumvents the lack of reliable taxonomic identification of
105 evolutionary units, which often hinders a large-scale sequence survey. Moreover, most of the
106 sampled species in the data set belong to the same family of water beetles (87% of samples belong
107 to Dytiscidae) and it is possible to reconstruct their species phylogeny without major gaps in species
108 sampling for the study region. Phylogeny can therefore be taken into account when judging the
109 correlations among traits (Freckleton et al. 2002).

110 Habitat type is an ecological trait commonly used as a surrogate of dispersal ability in
111 aquatic insects (Marten et al. 2006). Water beetles provide a useful system for investigating the
112 effects of habitat types on genetic traits. Water beetles inhabit two distinctive types of water bodies,

113 standing and running water, which are expected to have contrasting effects on species traits (Ribera
114 2008). Lentic (standing water) habitat is more ephemeral in evolutionary time scale than lotic
115 (running water) habitat, hence lentic species are expected to have greater ability to colonize new
116 habitat. This difference in dispersal propensity associated with habitat type has various effects,
117 ranging from species range size to degree of gene flow and evolutionary species turnover (Ribera
118 2008). Lentic species have greater colonization ability and larger ranges as a result of their higher
119 flight ability relative to lotic species (Abellan and Ribera 2011; Arribas et al. 2012). For instance, it
120 was reported that lentic species are on average 3.4 times larger range size than lotic species in
121 *Hydroporus* diving beetles (Abellan & Ribera 2011). We predict that lentic species should therefore
122 hold greater genetic variation than less dispersive lotic species. Small population sizes of lotic
123 species might lead to a relative excess of non-neutral changes, however, because of the lower
124 efficacy of purifying selection in smaller populations (Subramanian 2013). Alternatively, the
125 greater variation in ephemeral habitats may be counteracted by frequent extinctions and loss of
126 abundance on a local level. We will test this hypothesis using the water beetle data taking other
127 potential factors into account.

128

129 **Material and methods**

130 *Species delimitation and phylogenetic inference*

131 The water beetle dataset consists of 5062 CO1 sequences of a maximum of 700bp (Genbank
132 accession numbers JN840019-JN845080; see Baselga et al. 2013 for detailed descriptions of sample
133 collection and sequencing). Quality filtering by checking in-frame stop codons and removal of
134 identical haplotypes resulted in 2106 unique haplotypes. These sequences were used for DNA based
135 species delimitation. The gene tree of CO1 was reconstructed using RAxML7.0.3 (Stamatakis
136 2006) and made ultrametric with Pathd8 (Britton et al. 2007). Putative species groups were
137 delineated with the generalized mixed Yule-coalescent method (GMYC, Pons et al. 2006)

138 implemented in the ‘splits’ package (Ezard et al. 2009). The congruence between field identification
139 of diving beetle taxonomic species and GMYC delimited species was measured by counting the
140 exact matches of memberships between the two groupings. For convenience we refer to GMYC-
141 delimited groups as ‘species’ throughout.

142 Following species delimitation, a species tree of GMYC units was reconstructed to obtain a
143 working phylogeny for comparative analyses. One CO1 sequence was randomly taken from
144 samples of each species, and additional loci were retrieved from Genbank to improve the reliability
145 of phylogenetic reconstruction. Sequences of mitochondrial 16S and nuclear ribosomal RNA 18S,
146 28S and protein-coding *wingless* genes, were searched using the taxonomic names of the CO1
147 samples. The downloaded sequences were separately aligned using Muscle 3.8 (Edgar 2004) and
148 the resulting alignments were concatenated into a single data matrix. When 2 or more GMYC-
149 delimited species had the same taxonomic names, only one GMYC species was randomly chosen
150 and the downloaded sequences were attached to it. Incorrect concatenations due to mis-
151 identification were checked by comparing a resulting phylogeny and taxonomic literature (listed in
152 Table S1). The tree search described in the following paragraph was run twice and a final species
153 tree was built from the corrected matrix.

154 Markov chain Monte Carlo (MCMC) sampling of tree topology and substitution parameters
155 were conducted with MrBayes 3.2.2 (Ronquist et al. 2012). Four independent MCMC chains of 8
156 million generations were run with the GTR+I+G model, selected by jModelTest2 (Darriba et al.
157 2012), with 3 partitions and the first 4 million generations were discarded as burn-in. The effective
158 sample sizes (ESS) of the parameters were examined with Tracer 1.5 (Rambaut & Drummond
159 2007). The ESSs of all parameters reached more than 100 within the 8 million generations allowed
160 for the searches. A maximum clade credibility (MCC) tree was taken from the MCMC samples by
161 TreeAnnotator 1.61. Because no time calibration points were available for this diving beetle group,
162 relative divergence time was estimated using BEAST (Drummond et al. 2006) with the uncorrelated
163 log-normal relaxed clock. The GTR+I+G model with 3 partitions was again used as the model of

164 sequence evolution, and the mean substitution rate for the CO1 partition was set to 1.0. An MCMC
165 of fifty million generations was run and convergence of parameters was checked by Tracer.

166 To assess incongruence between the genealogy of mitochondrial and nuclear loci, a test of
167 tree topology was conducted. Three maximum likelihood topologies were separately estimated for
168 mtDNA, 18S and whole concatenated alignments using RAxML 7.0.3 with GTR+I+G model. Then,
169 likelihood values of the three topologies were calculated with re-optimization of parameters for
170 each alignment. The significance between likelihoods of the 3 topologies was tested with the
171 Shimodaira-Hasegawa test (SH test) with 10000 bootstrap replicates (Shimodaira & Hasegawa
172 1999). Significant reduction of likelihood between topologies indicates incongruence of the loci.
173 Taxa used for the tree search were reduced so that all taxa have complete alignments without
174 missing characters.

175

176 ***Data extraction***

177 *Habitat types, occupancy and geographic distribution*

178 Habitat types of the species were assigned by mining sampling records. First, the habitat types of
179 samples were determined according to descriptions of sampling sites in Ribera & Vogler (2000): for
180 example, lotic habitat for “river” or “creek” and lentic habitat for “pond” or “lake”. Then,
181 frequencies of samples from lotic and lentic habitats were counted, and species with more than 90%
182 of either lotic or lentic samples were marked as “Running” or “Standing” species respectively.
183 Species composed of mixed samples were assigned to the “Both” category.

184 The sequences were sampled at about 190 unique sites across Europe, and 4999 of 5062
185 sequences have GPS records of sampled localities. The sampling sites were clustered into 23 broad
186 regions of about 50 km in diameter according to their geographic positions in Baselga et al. (2013).
187 Geographic positions of regions and proportion of lotic or lentic samples collected from the regions

188 are summarized in Figure S1 and Table S2. Species occupancy was measured by counting the
189 number of the geographic regions where the specimens from the current study were sampled. The
190 latitudinal ranges of species, which we call range size, were obtained by subtracting the minimum
191 from the maximum latitude of species sampling records. The median and minimum latitude of
192 species samples were used as measures of their latitudinal distributions, which has been used as a
193 variable in studies of the effects of environmental energy on molecular rates (Davies et al. 2004).
194 Geographic locations of samples lacking GPS information were approximated with the mean values
195 of the regions within which the samples were collected. The number of samples was recorded for
196 each species to incorporate the effect of sampling, since species with large sample size may have
197 larger genetic variation than rarely sampled species. Habitat type and geographic records of samples
198 are summarized in supplementary data A (available at Dryad: doi:10.5061/dryad.926pq)

199

200 Genetic variation and rate of evolution of mtDNA

201 Genetic variation measured as nucleotide diversity (π) of CO1 for each species was estimated from
202 intra-specific genetic distances. Identical haplotypes that were removed for the construction of
203 GMYC groups were assigned back to each group. Pairwise genetic distances under the HKY model
204 were then calculated for the CO1 sequences within each species represented by 3 or more sequences,
205 and their mean values used as a measure of nucleotide diversity within the species. Genetic
206 distances were calculated for the whole CO1 alignment including all codon positions (π_{overall}) and
207 third codon positions ($\pi_{3\text{rd}}$). The mean genetic distances were arcsine square-root transformed
208 because their distribution was positively skewed. Species with less than 3 sequences were excluded,
209 and the remaining 191 species were used for downstream analyses.

210 The branch lengths (d) of a CO1 gene tree were used as a measure of rate of molecular
211 evolution. Branch lengths of the gene tree were recalculated using RAxML 7.0.3 with the GMYC
212 species tree as a binary constraint. The branch lengths on the gene tree were summed up through

213 the path from tip to root for each sample, and their median for each species was taken as
214 representative measures of substitution rate of the group. The branch lengths were measured on the
215 trees from the entire alignment (d_{overall}) and the third codon positions ($d_{3\text{rd}}$). To account for the effect
216 of node density on branch length estimation (node density effect, Hugall & Lee 2007), the number
217 of nodes on the path from tip to root was recorded for each species and included in regression
218 analyses described below.

219

220 ***Phylogenetic regression and model averaging***

221 To test the predicted correlation of the habitat types with species ecology and distribution,
222 univariate regressions with each parameter as a response variable in turn and habitat types as an
223 explanatory variable were conducted first. Then, the multivariate regression models with genetic
224 measures (genetic variation and substitution rate) as response variables and all species traits
225 including habitat types as explanatory variables were constructed to analyse the collective effects of
226 the variables on the genetic properties. Species measures are summarized in supplementary data B
227 (Dryad: doi:10.5061/dryad.926pq).

228 Phylogenetic generalized least squares (PGLS) described in Freckleton et al. (2002) were
229 used in order to incorporate the effect of phylogenetic dependence on the correlations of the
230 variables. Pagel's λ (Pagel 1999) was estimated for all explanatory and response variables and the
231 degree of phylogenetic dependency was examined before the correlation between each variable and
232 habitat was tested. The *ppls* function in the 'caper' package (Orme 2012) was used for the PGLS
233 analysis. For the multivariate phylogenetic regressions including all variables, model averaging
234 following Burnham & Anderson (2002) was used to assess the effects of multiple variables. The
235 95% confidence sets were constructed from all possible models without interaction terms, and the
236 sum of Akaike weights, weighted average of estimates and standard errors were obtained for each
237 parameter from the models within the confidence set. The explanatory variables in the maximal

238 model included terms for habitat type, species occupancy, latitude, sample size, the number of
239 nodes and genetic variation or rate of evolution. All statistical analyses were conducted by using R
240 (R Core Team 2013) with the aid of packages ‘ape’ (Paradis et al. 2004) and ‘phangorn’ (Schliep
241 2011) for phylogenetic analyses and ‘pegas’ (Paradis 2010) for population genetic analyses.

242

243 **Results**

244 *Delimitation and species phylogeny*

245 The GMYC analysis delimited 274 putative species groups, of which 45% matched exactly with
246 taxonomic identifications, i.e. all members of a taxonomic species were included in a single GMYC
247 group that did not include any other taxonomic species. Taxonomic species were over-split in 28%
248 of cases and lumped with other taxonomic species in 32% of cases. The numbers of sequences
249 retrieved from Genbank that matched with taxonomic names of the studied species were 100 and 51
250 for 16S and 18S respectively (Accession numbers of downloaded sequences are in supplementary
251 data C available at Dryad: doi:10.5061/dryad.926pq). These sequences were concatenated into a
252 single alignment with 1854 characters, in which 650 sites were parsimony informative (344/700,
253 233/546 and 73/608 for CO1, 16S and 18S) and 47% of nucleotides were missing. Other nuclear
254 markers, 28S and *wingless* had only 2 and 11 matched sequences, and were excluded from the
255 phylogenetic analysis.

256 The Bayesian tree search resulted in an MCC tree in which 113 out of 272 internal nodes
257 had more than 95% of support (Fig. S1; a tree file is available at Dryad: doi:10.5061/dryad.926pq).
258 Basal relationships among water beetle genera were less resolved than intra-generic relationships
259 (median posterior probability 0.77 and 0.88 respectively). Three of the main families of water
260 beetles (Noteridae, Haliplidae and Gyrinidae) formed monophyletic clades with more than 95%
261 support and the largest family, Dytiscidae, was monophyletic with 66% of posterior probability.
262 The SH test with the reduced data showed significant differences on the likelihoods between the

263 mitochondrial and nuclear species trees (Table S3), indicating possible incongruence between
264 mitochondrial and nuclear genealogies. Because the ML tree of mtDNA and the concatenated data
265 were not significantly different, however, the effect of incongruence on the concatenation is small
266 and the concatenated tree was used for further analyses.

267

268 *Species ecology, distribution and genetic variables*

269 Mean pairwise genetic distances and median branch lengths on the gene tree were obtained for 191
270 out of the 274 GMYC species that contained more than 3 samples (sample sizes per GMYC species
271 ranged from 3 to 356, median 13; 9 species had 3 samples only, 6 species had >100 samples). The
272 mean genetic distances ranged from 0.0 to 0.019 (median 0.004) for the entire alignment (π_{overall}),
273 and from 0.0 to 0.055 (median 0.0085) for third codon positions ($\pi_{3\text{rd}}$). Branch lengths ranged from
274 0.31 to 0.73 (median 0.45) and from 0.62 to 1.90 (median 1.01) respectively (d_{overall} and $d_{3\text{rd}}$). The
275 number of nodes from tip to root ranged between 4 and 20 (median 11). Among the 191 GMYC
276 species, there were 38 species categorized as "Running" water species and 86 as "Standing". The
277 remaining 63 groups were found in both lotic and lentic habitats and marked as "Both", and 4
278 groups did not have any available habitat descriptions. The most widespread species, mainly
279 consisting of samples identified as *Agabus bipustulatus*, was sampled from 20 out of 23 geographic
280 regions, whereas 38 groups were found in only one geographic region.

281 The estimates of Pagel's λ for habitat type and median latitude were significantly different
282 from both 0 and 1 ($\lambda = 0.67$ and 0.45 respectively), indicating partial phylogenetic dependence of
283 these variables (Figure 1). Estimated λ of branch length and node numbers was 1.0 and 0.99
284 respectively as they were measured on paths from root to tip along the phylogeny. There was no
285 phylogenetic dependence of the other variables including occupancy, latitudinal range size,
286 minimum latitude, sample size and genetic variations.

287

288 *Phylogenetic regressions*

289 *Univariate phylogenetic regression*

290 There were significant differences in species latitudinal extent, and median and minimum
291 latitude between lotic and lentic groups (Table 1). Lotic species had significantly smaller range
292 sizes than the other groups including the "both". Lentic species were distributed further north than
293 the other groups regarding their southern limits (min. latitude) and distributional centres (median
294 latitude). The median latitude of lotic species was significantly lower than in the other two groups.
295 Neither genetic variation nor branch length had significant correlation with habitat types, except
296 that median branch length of the tree from the full dataset (d_{overall}) showed a significant difference
297 between lotic and other groups. The sample sizes were not correlated with habitat type, which
298 indicates that there was no sampling bias from different habitat types.

299 *Multivariate phylogenetic regression with model averaging*

300 When genetic variation was a response variable, species occupancy had the largest effect
301 with sum of Akaike weight 1.0 (Table 2), which means it was included in all models within the
302 95% confidence set. Out of 63 candidate models, 23 were included within the confidence set. Model
303 averaged Pagel's λ was zero, indicating no phylogenetic dependency on genetic variation. Among
304 the explanatory variables, the model averaged estimate of species occupancy was significantly
305 greater than zero, indicating positive effects of species occupancy on genetic variation. The second
306 largest effect was median latitude with Akaike weight 0.93. The alternate use of minimum or
307 median of latitude and occupancy or latitudinal range did not affect the trend of model averaging
308 (Table S4). The measures of latitudinal distribution were the second largest factors in both cases,
309 and both occupancy and latitudinal range had significant positive effects on the genetic variation.
310 Sample size and habitat had the third and fourth largest effects, but their estimates were not
311 significantly different from zero. The relative importance of the explanatory variables using the
312 third codon position resulted in patterns similar to the full dataset. Species occupancy and minimum

313 latitude exhibited the highest, significant correlations, and the relative importance of latitude was
314 0.99, which is higher than in the analysis with the total dataset (Table 2). Model averaged Pagel's λ
315 was 0.0004.

316 For the model with branch lengths as a response, the number of nodes had the largest weight,
317 1.0, which suggests a strong correlation between branch lengths and the number of nodes from tip
318 to root (Table 3). Out of 63 candidate models, 21 were included in the 95% confidence set. Model
319 averaged Pagel's λ was 0.99. Apart from the effect of node density, habitat type was the second
320 largest weight, 0.84, and the rate of evolution for lotic species was significantly higher than lentic
321 and "both". Latitude was the third largest weight, but its effect was not significant. With the third
322 codon positions, no variables except for the node number had significant effect on the branch
323 lengths. The number of models included in the 95% confidence set was 42, showing low
324 explanatory power of the candidate models. Habitat type dropped the strong effects observed in the
325 entire alignment.

326

327 **Discussion**

328 Habitat type had significant effects on both species range size and latitudinal distribution. Lotic
329 species had significantly smaller range size (measured by latitudinal extent) and more southern
330 distribution than lentic species. These patterns were consistent with well-supported effects of
331 habitat type on dispersal propensity and distribution (Ribera & Vogler 2000; Hof et al. 2006).

332 Lentic habitats are expected to be more ephemeral than lotic habitats, hence populations of lentic
333 species only persist if they disperse more frequently to reach new suitable habitats, which predicts
334 broader ranges and faster colonization of northern latitudes after the last ice age.

335 Occupancy and latitudinal range, in turn, displayed the strongest positive effect on genetic variation
336 within species (Table 2, Figure 2). Species with larger ranges and higher occupancy, which are
337 associated with lentic habitats, contained more genetic variation than species with smaller ranges.

338 There are well-established reports that the species occupancy and range size are tightly correlated
339 with its abundance (Gaston et al. 2000; Blackburn et al. 2006; Gaston & He 2011). Therefore, the
340 effect of occupancy on the genetic variation of CO1 is interpreted under the theory of neutral
341 molecular evolution that predicts neutral genetic variation to be proportional to effective population
342 size. The association of population size to mitochondrial genetic variation has been questioned
343 (Bazin et al. 2006) based on a comparison of levels of genetic variation in vertebrates and
344 invertebrates (assumed to have low and high effective population size, respectively). Rate
345 heterogeneity or frequent selective sweeps are proposed as possible alternative determinants of
346 genetic variation of mtDNA (Nabholz et al. 2008; Nabholz et al. 2009). Frequent fixation of
347 adaptive mutation can reduce genetic variation and decouple it from population size (Gillespie
348 2001). However, the strong effect of abundance together with the weak effect of substitution rate
349 (the product of mutation rate and fixation rate) on π observed in this study supported the traditional
350 view of positive association between population size and genetic variation. An alternative
351 explanation might be that species with larger ranges are older and have had time to accumulate
352 more genetic variation, if speciation tended to result in new species with small ranges and low
353 variation. However, there was no significant correlation between genetic variation and age of
354 species measured on species tree (Figure S2).

355 The second predictor of genetic variation was latitude, yet contrary to the expected pattern if
356 high environmental energy promoted higher mutation rates (Davies et al. 2004), we find that
357 minimal (low) latitude was correlated with lower π . It might also be expected for high latitude
358 populations to show low genetic variation due to recent colonization and expansion after the ice
359 ages (Hewitt 2000), but we found no such relationship with latitude either. One possible explanation
360 is that the residual variation explained by latitude could be a surrogate of additional range size
361 extending outside the sample area. It has been reported that high latitude species of water beetles
362 have far wider range size than southern species, often encompassing the Holarctic (e.g. Abellan &
363 Ribera 2011), and the genetic variation in the northern species could be higher than expected by

364 observed range size alone. Another possibility is that species at higher latitudes might be starting to
365 diversify between local populations, yet too recently to have diverged into reciprocally
366 monophyletic genetic clusters detectable by GMYC, while southern populations have diversified,
367 and therefore are categorized as different species with small ranges and consequently small π . Weir
368 and Schluter (2007) found evidence of faster speciation rates at higher latitudes in birds, perhaps
369 explained by new opportunities for speciation in empty areas recently re-colonised since glacial
370 retreat.

371 Next, we considered the factors correlated with substitution rates across species. Habitat
372 type had the strongest effect on the rate of evolution, but unlike intra-specific genetic variation that
373 was highest in lentic species with larger population size, the highest substitution rate was found in
374 lotic species. The higher rate of evolution of lotic species could be explained by more frequent
375 fixation of non-neutral mutations in small populations. If lotic species have smaller and more
376 structured populations because of their low dispersal ability, fixation of deleterious alleles within
377 local demes can increase because of stronger genetic drift (Woolfit & Bromham 2005). This
378 scenario is supported by the fact that the effect of habitat type was not observed for the rate of third
379 codon positions, which represent more neutral substitutions. Therefore, the effect of habitat types on
380 whole alignment should be attributable to the first and second codon positions, and a separate
381 analysis confirmed it (Table S5). Unexpectedly under this scenario, we found no direct negative
382 correlation of substitution rate with occupancy. This may be explained by the fact that population
383 size (measured by occupancy) is an evolutionarily labile trait ($\lambda=0.10$) not consistent across a long
384 time scale, while habitat type as a more phylogenetically conserved trait ($\lambda=0.67$), is a better
385 predictor of long-term rate variation.

386 The significant positive correlation between the rate of evolution and the number of nodes
387 on the path from tip to root indicated the node density effect. The model averaging analysis without
388 controlling for node density resulted in different patterns of relative importance of explanatory
389 variables (Table S6). Relative importance of habitat type was smaller than the analysis with node

390 number included. In extreme cases, the effect of habitat type could possibly be overshadowed by
391 biases introduced by node density effects due to uneven sampling of species. This result is
392 consistent with studies showing that using total path lengths as a measure of rate of evolution can be
393 compromised by the node density artifact (Hugall & Lee 2007). Methods to filter out the node
394 density effect have been debated and remain controversial (Webster et al. 2003; Hugall & Lee 2007;
395 Lanfear et al. 2010). In this study, a clear linear relationship between branch lengths and the number
396 of nodes was detected (Figure 3). Thus, the inclusion of the node number as an explanatory variable
397 into the regression modeling should moderate the node density effect.

398 A latitudinal gradient of the rate of molecular evolution is a commonly observed pattern of
399 molecular evolution (Davies et al. 2004; Allen et al. 2006; Gillman et al. 2009). However, although
400 latitude still had a negative effect on the rate of evolution, it was not significant in this study. The
401 cause of the latitudinal gradient of molecular evolution has been widely debated, often associated
402 with the cause of the latitudinal gradient of species diversity (reviewed in Mittelbach et al. 2007).
403 Hypotheses include the high energy input that results in higher metabolic rates and more mutagenic
404 events at lower latitudes or more frequent fixation of deleterious alleles in small tropical
405 populations. The lotic species in this study were distributed at lower latitudes on average and had
406 higher rate of evolution. The gradient of rates might be a consequence of different life history traits
407 distributed along latitude.

408 Lastly, we do not observe direct correlations between genetic variation and rate of evolution,
409 which would be expected if mutation rate were the only dominant factor controlling both genetic
410 variation and substitution. Rather, the patterns are explained by occupancy and habitat types,
411 parameters associated to effective population size. Also, the rate at the third codon positions, which
412 is a closer approximation of mutation rate, was not correlated with any variables considered in this
413 study. This evidence supports the scenario that mutation rates are relatively constant or haphazard
414 across species and both present-day genetic variation and long term substitution rates are controlled
415 by demographic factors, which are, in turn, controlled by species ecology.

416 The working phylogeny used for the comparative analysis has nodes with low support
417 values (posterior probabilities < 0.9), and this may have introduced the errors in the regression
418 analysis. Nevertheless, because genetic variation did not show strong phylogenetic dependency, the
419 effect of a suboptimal tree is probably minimal for the regression model of genetic variation. For
420 the rate of evolution, the inaccuracy may have introduced biases as the branch lengths were
421 estimated along the species phylogeny. However, the different rates between lotic and lentic groups
422 observed in this study are unlikely to be an artifact. There were no differences in the levels of node
423 support on the trees between lotic and lentic groups (Fig S3), therefore there should not be
424 systematic biases affecting branch lengths of each type.

425 Our analyses were conducted with species solely delimited with DNA sequences, which
426 allowed us to compare patterns of genetic variation between comparable units with positive
427 evidence of independent evolution. Complete identification of specimens to taxonomic species
428 would have been useful for comparison but was not feasible here with available resources. DNA-
429 based species delimitation is particularly useful when taxonomic information is not easily accessible,
430 for example in a rapid survey of unstudied biota (Monaghan et al. 2009) or in taxonomically
431 difficult groups such as fungi or meiofauna (Tang et al. 2012; Millanes et al. 2014). Because of the
432 recent advances of the sequencing technology, such as "metabarcoding" (Ji et al. 2013), data
433 without complete taxonomic identification is becoming more common in biodiversity surveys. The
434 correlations between species' ecological and genetic parameters detected in this study were largely
435 consistent with predictions from conventional studies, which supports the use of DNA-based
436 delimitation as a tool to study data without clear taxonomic information.

437 We conclude that different correlates explain variation within species versus rates of
438 molecular evolution between species. Neutral theory predicts higher genetic variation in larger
439 populations but that substitution rates of neutral mutations are independent of population size and
440 substitution rates of nearly-neutral mutations are faster in smaller populations. Our results broadly
441 support these alternative predictions: genetic variation was greatest in larger populations, whereas

442 substitution rates were fastest in lotic species (which have smaller populations on average than
443 lentic species). While we cannot identify mechanisms from correlations, and it is possible that other
444 factors correlated with these variables influenced the patterns, our results show how comparative
445 studies of DNA barcoding-type data can provide insights into correlates of molecular evolution.

446

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452

453 **Data accessibility**

454 The CO1 sequences are available at Genbank under accession numbers JN840019 – JN845080.
455 Supplementary data were deposited in Dryad (doi: doi:10.5061/dryad.926pq).

456

457 **References**

- 458 1. Abellán, P. & Ribera, I. 2011 Geographic location and phylogeny are the main determinants
459 of the size of the geographical range in aquatic beetles. *BMC Evol. Biol.* **11**, 344.
460 (doi:10.1186/1471-2148-11-344)
- 461 2. Allen, A. P., Gillooly, J. F., Savage, V. M. & Brown, J. H. 2006 Kinetic effects of
462 temperature on rates of genetic divergence and speciation. *Proc. Natl. Acad. Sci.* **103**, 9130–
463 9135. (doi:10.1073/pnas.0603587103)
- 464 3. Arribas, P., Velasco, J., Abellán, P., Sánchez-Fernández, D., Andújar, C., Calosi, P., Millán,
465 A., Ribera, I. & Bilton, D. T. 2012 Dispersal ability rather than ecological tolerance drives
466 differences in range size between lentic and lotic water beetles (Coleoptera: Hydrophilidae).
467 *J. Biogeogr.* **39**, 984–994. (doi:10.1111/j.1365-2699.2011.02641.x)
- 468 4. Ballard, J. W. O. & Whitlock, M. C. 2004 The incomplete natural history of mitochondria.
469 *Mol. Ecol.* **13**, 729–744. (doi:10.1046/j.1365-294X.2003.02063.x)
- 470 5. Baselga, A., Fujisawa, T., Crampton-Platt, A., Bergsten, J., Foster, P. G., Monaghan, M. T.
471 & Vogler, A. P. 2013 Whole-community DNA barcoding reveals a spatio-temporal
472 continuum of biodiversity at species and genetic levels. *Nat. Commun.* **4**, 1892.
473 (doi:10.1038/ncomms2881)
- 474 6. Bazin, E., Glémin, S. & Galtier, N. 2006 Population size does not influence mitochondrial
475 genetic diversity in animals. *Science* **312**, 570–2. (doi:10.1126/science.1122033)
- 476 7. Blackburn, T. M., Cassey, P. & Gaston, K. J. 2006 Variations on a theme: sources of
477 heterogeneity in the form of the interspecific relationship between abundance and
478 distribution. *J. Anim. Ecol.* **75**, 1426–39. (doi:10.1111/j.1365-2656.2006.01167.x)
- 479 8. Britton, T., Anderson, C. L., Jacquet, D., Lundqvist, S. & Bremer, K. 2007 Estimating
480 Divergence Times in Large Phylogenetic Trees. *Syst. Biol.* **56**, 741–752.
481 (doi:10.1080/10635150701613783)
- 482 9. Bromham, L. & Penny, D. 2003 The modern molecular clock. *Nat. Rev. Genet.* **4**, 216–224.
483 (doi:10.1038/nrg1020)

- 484 10. Burney, C. W. & Brumfield, R. T. 2009 Ecology predicts levels of genetic differentiation in
485 neotropical birds. *Am. Nat.* **174**, 358–68. (doi:10.1086/603613)
- 486 11. Burnham, K. P. & Anderson, D. R. 2002 *Model Selection and Multimodel Inference: A*
487 *Practical Information-Theoretic Approach*. 2nd edn. New York, USA: Springer
488 Science+Business Media, Inc.
- 489 12. Charlesworth, B., Charlesworth, D. & Barton, N. H. 2003 The effects of genetic and
490 geographic structure on neutral variation. *Annu. Rev. Ecol. Evol. Syst.* **34**, 99–125.
491 (doi:10.1146/annurev.ecolsys.34.011802.132359)
- 492 13. Darriba, D., Taboada, G. L., Doallo, R. & Posada, D. 2012 jModelTest 2: more models, new
493 heuristics and parallel computing. *Nat. Methods* **9**, 772. (doi:10.1038/nmeth.2109)
- 494 14. Davies, T. J., Savolainen, V., Chase, M. W., Moat, J. & Barraclough, T. G. 2004
495 Environmental energy and evolutionary rates in flowering plants. *Proc. R. Soc. B Biol. Sci.*
496 **271**, 2195–2200. (doi:10.1098/rspb.2004.2849)
- 497 15. Drummond, A. J., Ho, S. Y. W., Phillips, M. J. & Rambaut, A. 2006 Relaxed phylogenetics
498 and dating with confidence. *PLoS Biol.* **4**, e88. (doi:10.1371/journal.pbio.0040088)
- 499 16. Edgar, R. C. 2004 MUSCLE: a multiple sequence alignment method with reduced time and
500 space complexity. *BMC Bioinformatics* **5**, 113. (doi:10.1186/1471-2105-5-113)
- 501 17. Ezard, T., Fujisawa, T. & Barraclough, T. G. 2009 SPLITS: SPecies' Limits by Threshold
502 Statistics.
- 503 18. Freckleton, R. P., Harvey, P. H. & Pagel, M. 2002 Phylogenetic analysis and comparative
504 data: a test and review of evidence. *Am. Nat.* **160**, 712–26. (doi:10.1086/343873)
- 505 19. Galtier, N., Nabholz, B., Glémin, S. & Hurst, G. D. D. 2009 Mitochondrial DNA as a marker
506 of molecular diversity: a reappraisal. *Mol. Ecol.* **18**, 4541–50. (doi:10.1111/j.1365-
507 294X.2009.04380.x)
- 508 20. Gaston, K. J., Blackburn, T. M., Greenwood, J. J. D., Gregory, R. D., Quinn, R. M. &
509 Lawton, J. H. 2000 Abundance-occupancy relationships. *J. Appl. Ecol.* **37**, 39–59.
510 (doi:10.1046/j.1365-2664.2000.00485.x)

- 511 21. Gaston, K. J. & He, F. 2011 Species occurrence and occupancy. In *Biological Diversity:*
512 *frontiers in measurement and assessment* (eds A. E. Magurran & B. J. McGill), pp. 141–151.
513 Oxford: Oxford University Press.
- 514 22. Gillespie, J. H. 2000 Genetic drift in an infinite population. The pseudohitchhiking model.
515 *Genetics* **155**, 909–19.
- 516 23. Gillman, L. N., Keeling, D. J., Ross, H. A. & Wright, S. D. 2009 Latitude, elevation and the
517 tempo of molecular evolution in mammals. *Proc. R. Soc. B Biol. Sci.* **276**, 3353–9.
518 (doi:10.1098/rspb.2009.0674)
- 519 24. Gossmann, T. I., Keightley, P. D. & Eyre-Walker, A. 2012 The effect of variation in the
520 effective population size on the rate of adaptive molecular evolution in eukaryotes. *Genome*
521 *Biol. Evol.* **4**, 658–67. (doi:10.1093/gbe/evs027)
- 522 25. Hewitt, G. 2000 The genetic legacy of the Quaternary ice ages. *Nature* **405**, 907–13.
523 (doi:10.1038/35016000)
- 524 26. Hof, C., Brändle, M. & Brandl, R. 2006 Lentic odonates have larger and more northern
525 ranges than lotic species. *J. Biogeogr.* **33**, 63–70. (doi:10.1111/j.1365-2699.2005.01358.x)
- 526 27. Hugall, A. F. & Lee, M. S. Y. 2007 The likelihood node density effect and consequences for
527 evolutionary studies of molecular rates. *Evolution* **61**, 2293–307. (doi:10.1111/j.1558-
528 5646.2007.00188.x)
- 529 28. Ji, Y. et al. 2013 Reliable, verifiable and efficient monitoring of biodiversity via
530 metabarcoding. *Ecol. Lett.* , 1245–1257. (doi:10.1111/ele.12162)
- 531 29. Kimura, M. 1984 *The neutral theory of molecular evolution*. Cambridge: Cambridge
532 University Press.
- 533 30. Knowles, L. L. 2006 Statistical Phylogeography. *Annu. Rev. Ecol. Evol. Syst.* **40**, 593–612.
534 (doi:10.1146/annurev.ecolsys.38.091206.095702)
- 535 31. Lanfear, R., Welch, J. J. & Bromham, L. 2010 Watching the clock: studying variation in
536 rates of molecular evolution between species. *Trends Ecol. Evol.* **25**, 495–503.
537 (doi:10.1016/j.tree.2010.06.007)

- 538 32. Leffler, E. M., Bullaughey, K., Matute, D. R., Meyer, W. K., Ségurel, L., Venkat, A.,
539 Andolfatto, P. & Przeworski, M. 2012 Revisiting an old riddle: what determines genetic
540 diversity levels within species? *PLoS Biol.* **10**, e1001388.
541 (doi:10.1371/journal.pbio.1001388)
- 542 33. Manier, M. K. & Arnold, S. J. 2006 Ecological correlates of population genetic structure: a
543 comparative approach using a vertebrate metacommunity. *Proc. R. Soc. B Biol. Sci.* **273**,
544 3001–9. (doi:10.1098/rspb.2006.3678)
- 545 34. Marten, A., Brändle, M. & Brandl, R. 2006 Habitat type predicts genetic population
546 differentiation in freshwater invertebrates. *Mol. Ecol.* **15**, 2643–51. (doi:10.1111/j.1365-
547 294X.2006.02940.x)
- 548 35. Meiklejohn, C. D., Montooth, K. L. & Rand, D. M. 2007 Positive and negative selection on
549 the mitochondrial genome. *Trends Genet.* **23**, 259–63. (doi:10.1016/j.tig.2007.03.008)
- 550 36. Millanes, A. M., Truong, C., Westberg, M., Diederich, P. & Wedin, M. 2014 Host Switching
551 Promotes Diversity in Host-Specialized Mycoparasitic Fungi: Uncoupled Evolution in the
552 Biatropsis-Usnea System. *Evolution (N. Y.)*, 1–18. (doi:10.1111/evo.12374)
- 553 37. Mittelbach, G. G. et al. 2007 Evolution and the latitudinal diversity gradient: speciation,
554 extinction and biogeography. *Ecol. Lett.* **10**, 315–331. (doi:10.1111/j.1461-
555 0248.2007.01020.x)
- 556 38. Nabholz, B., Glémin, S. & Galtier, N. 2009 The erratic mitochondrial clock: variations of
557 mutation rate, not population size, affect mtDNA diversity across birds and mammals. *BMC*
558 *Evol. Biol.* **9**, 54. (doi:10.1186/1471-2148-9-54)
- 559 39. Nabholz, B., Mauffrey, J.-F., Bazin, E., Galtier, N. & Glemin, S. 2008 Determination of
560 mitochondrial genetic diversity in mammals. *Genetics* **178**, 351–61.
561 (doi:10.1534/genetics.107.073346)
- 562 40. Ohta, T. 1992 The Nearly Neutral Theory of Molecular Evolution. *Annu. Rev. Ecol. Syst.* **23**,
563 263–286. (doi:10.2307/2097289)
- 564 41. Orme, C. D. L. 2012 Caper: comparative analyses of phylogenetics and evolution in R.

- 565 42. Pagel, M. 1999 Inferring the historical patterns of biological evolution. *Nature* **401**, 877–84.
566 (doi:10.1038/44766)
- 567 43. Papadopoulou, A., Anastasiou, I., Keskin, B. & Vogler, A. P. 2009 Comparative
568 phylogeography of tenebrionid beetles in the Aegean archipelago: the effect of dispersal
569 ability and habitat preference. *Mol. Ecol.* **18**, 2503–2517. (doi:10.1111/j.1365-
570 294X.2009.04207.x)
- 571 44. Papadopoulou, A., Anastasiou, I., Spagopoulou, F., Stalimerou, M., Terzopoulou, S., Legakis,
572 A. & Vogler, A. P. 2011 Testing the species--genetic diversity correlation in the Aegean
573 archipelago: toward a haplotype-based macroecology? *Am. Nat.* **178**, 241–55.
574 (doi:10.1086/660828)
- 575 45. Paradis, E. 2010 pegas: an R package for population genetics with an integrated–modular
576 approach. *Bioinformatics* **26**, 419–420. (doi:10.1093/bioinformatics/btp696)
- 577 46. Paradis, E., Claude, J. & Strimmer, K. 2004 APE: Analyses of Phylogenetics and Evolution
578 in R language. *Bioinformatics* **20**, 289–290. (doi:10.1093/bioinformatics/btg412)
- 579 47. Pons, J., Barraclough, T. G., Gomez-Zurita, J., Cardoso, A., Duran, D. P., Hazell, S.,
580 Kamoun, S., Sumlin, W. D. & Vogler, A. P. 2006 Sequence-based species delimitation for
581 the dna taxonomy of undescribed insects. *Syst. Biol.* **55**, 595–609.
582 (doi:10.1080/10635150600852011)
- 583 48. Rambaut, A. & Drummond, A. J. 2007 Tracer v1.4.
- 584 49. Ribera, I. 2008 Habitat constraints and the generation of diversity in freshwater
585 macroinvertebrates. In *AQUATIC INSECT: Challenges to Populations* (eds J. Lancaster & R.
586 A. Briers), pp. 289–311. Wallingford: CAB International.
- 587 50. Ribera, I. & Vogler, A. P. 2000 Habitat type as a determinant of species range sizes: the
588 example of lotic–lentic differences in aquatic Coleoptera. *Biol. J. Linn. Soc.* **71**, 33–52.
589 (doi:10.1111/j.1095-8312.2000.tb01240.x)
- 590 51. Riginos, C., Buckley, Y. M., Blomberg, S. P. & Treml, E. a 2014 Dispersal capacity predicts
591 both population genetic structure and species richness in reef fishes. *Am. Nat.* **184**, 52–64.
592 (doi:10.1086/676505)

- 593 52. Ronquist, F. et al. 2012 MrBayes 3.2: efficient Bayesian phylogenetic inference and model
594 choice across a large model space. *Syst. Biol.* **61**, 539–42. (doi:10.1093/sysbio/sys029)
- 595 53. Santos, J. C. 2012 Fast molecular evolution associated with high active metabolic rates in
596 poison frogs. *Mol. Biol. Evol.* **29**, 2001–18. (doi:10.1093/molbev/mss069)
- 597 54. Schliep, K. P. 2011 phangorn: phylogenetic analysis in R. *Bioinformatics* **27**, 592–3.
598 (doi:10.1093/bioinformatics/btq706)
- 599 55. Shimodaira, H. & Hasegawa, M. 1999 Multiple Comparisons of Log-Likelihoods with
600 Applications to Phylogenetic Inference. *Mol. Biol. Evol.* **16**, 1114–1116.
- 601 56. Stamatakis, A. 2006 RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses
602 with thousands of taxa and mixed models. *Bioinformatics* **22**, 2688–2690.
603 (doi:10.1093/bioinformatics/btl446)
- 604 57. Subramanian, S. 2013 Significance of population size on the fixation of nonsynonymous
605 mutations in genes under varying levels of selection pressure. *Genetics* **193**, 995–1002.
606 (doi:10.1534/genetics.112.147900)
- 607 58. Tang, C. Q., Leasi, F., Obertegger, U., Kieneker, A., Barraclough, T. G. & Fontaneto, D. 2012
608 The widely used small subunit 18S rDNA molecule greatly underestimates true diversity in
609 biodiversity surveys of the meiofauna. *Proc. Natl. Acad. Sci. U. S. A.* **109**, 16208–12.
610 (doi:10.1073/pnas.1209160109)
- 611 59. Team, R. C. 2013 R: A language and environment for statistical computing.
- 612 60. Wakeley, J. 2004 Metapopulation models for historical inference. *Mol. Ecol.* **13**, 865–875.
613 (doi:10.1111/j.1365-294X.2004.02086.x)
- 614 61. Wakeley, J. & Aliacar, N. 2001 Gene genealogies in a metapopulation. *Genetics* **159**, 893–
615 905.
- 616 62. Webster, A. J., Payne, R. J. H. & Pagel, M. 2003 Molecular phylogenies link rates of
617 evolution and speciation. *Science* **301**, 478. (doi:10.1126/science.1083202)
- 618 63. Weir, J. T. & Schluter, D. 2007 The latitudinal gradient in recent speciation and extinction
619 rates of birds and mammals. *Science* **315**, 1574–6. (doi:10.1126/science.1135590)

- 620 64. Welch, J. J., Bininda-Emonds, O. R. P. & Bromham, L. 2008 Correlates of substitution rate
621 variation in mammalian protein-coding sequences. *BMC Evol. Biol.* **8**, 53.
622 (doi:10.1186/1471-2148-8-53)
- 623 65. Woolfit, M. & Bromham, L. 2005 Population size and molecular evolution on islands. *Proc.*
624 *R. Soc. B Biol. Sci.* **272**, 2277–82. (doi:10.1098/rspb.2005.3217)
- 625

626 **Captions of figures**

627 Figure 1.

628 Phylogenetic dependency of traits measured as maximum likelihood estimates of Pagel's λ . The
629 95% confidence intervals of the estimates are shown by error bars.

630

631 Figure 2.

632 The effect of occupancy on genetic variation within species. The solid line is a regression line
633 estimated by PGLS ($y = 0.009*x+0.052$, $p=0.0013$, $R^2=0.05$, Pagel's $\lambda =0.0$).

634

635 Figure 3.

636 The effect of number of nodes on branch lengths from tip to root. Habitat type of species is
637 indicated by 3 types of symbols (Black dot: Lotic, Open square: Lentic and Grey cross: "Both").

638 The solid lines are regression lines estimated by PGLS for lotic and lentic species ($y =$
639 $0.013*x+(lotic: 0.34, lentic: 0.33)$, $p<<0.001$, $R^2=0.24$, $\lambda=1.0$). The interaction term between habitat
640 type and node number was not significant.

641

642

643 **Tables**

644 Table 1. Univariate PGLS regression analysis of effect of habitat types on species traits (only
 645 estimates of lotic and lentic species are shown).

		Lotic	Lentic	P-value		R ²	Pagel's λ
Occupancy(log)		1.00	1.07	0.65		0.045	0.11
Latitude	Extent	5.97	10.24	0.01	*	0.060	0.01
	Median	42.71	52.36	0.00	***	0.195	0.23
	Minimum	40.38	47.33	0.00	***	0.169	0.00
Sample size(log)		2.55	2.65	0.61		0.008	0.00
π_{overall}		0.059	0.066	0.17		0.006	0.00
π_{3rd}		0.089	0.101	0.25		0.001	0.06
d_{overall}		0.46	0.43	0.01	*	0.026	0.99
d_{3rd}		1.15	1.10	0.17		0.00	1.00

646

647

648 Table 2. Relative importance and model averaged estimators of species traits from PGLS models of
 649 genetic variation (π_{overall} and π_{3rd}). The parameter estimates which are significantly different from
 650 zero are indicated by bold numbers, and the highest relative importance was underlined.

		Overall			Third		
		Σw_i	$\beta \pm \text{SE}$		Σw_i	$\beta \pm \text{SE}$	
Occupancy		<u>1.00</u>	0.013	0.0054	<u>1.00</u>	0.023	0.0087
Med.latitude		0.93	0.0007	0.0003	0.99	0.0014	0.0005
Med. <i>d</i>		0.26	0.0028	0.031	0.60	-0.023	0.014
Habitat	Lotic	0.50	0.0098	0.0065	0.55	0.019	0.012
	Lentic	0.50	0.0081	0.0050	0.55	0.013	0.0081
Sample size		0.58	-0.0059	0.0037	0.55	-0.0093	0.0061
Node number		0.30	0.0003	0.0006	0.27	-0.0001	0.0009

651 Σw_i : Sum of Akaike weights of variables

652 $\beta \pm \text{SE}$: Model averaged estimates and standard errors

653

654 Table 3. Relative importance and model averaged estimators of species traits from PGLS models of
 655 branch length (d_{overall} and d_{3rd}).

		Overall			Third		
		Σw_i	$\beta \pm \text{SE}$		Σw_i	$\beta \pm \text{SE}$	
Occupancy		0.38	-0.0061	0.0067	0.24	-0.0009	0.012
Med.latitude		0.58	-0.0007	0.0005	0.27	-0.0007	0.0014
Mean π		0.28	0.055	0.094	0.29	-0.084	0.17
Habitat	Lotic	0.84	0.025	0.010	0.24	0.045	0.032
	Lentic	0.84	-0.0011	0.0062	0.24	0.0013	0.018
Sample size		0.45	0.0048	0.0043	0.25	0.0014	0.0080
Node number		<u>1.00</u>	0.013	0.0018	<u>1.00</u>	0.025	0.0059

656





