

Title	In Vivo Linking of Membrane Lipids and the Anion Transporter Band 3 with Thiourea-modified Amphiphilic Lipid Probes.
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Citation	Scientific reports (2015), 5
Issue Date	2015
URL	http://hdl.handle.net/2433/207514
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Type	Journal Article
Textversion	publisher

SCIENTIFIC REPORTS

OPEN

In Vivo Linking of Membrane Lipids and the Anion Transporter Band 3 with Thiourea-modified Amphiphilic Lipid Probes

Received: 12 June 2015
Accepted: 29 October 2015
Published: 30 November 2015

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Membrane proteins interact with membrane lipids for their structural stability and proper function. However, lipid–protein interactions are poorly understood at a molecular level especially in the live cell membrane, due to current limitations in methodology. Here, we report that amphiphilic lipid probes can be used to link membrane lipids and membrane proteins *in vivo*. Cholesterol and a phospholipid were both conjugated to a fluorescent tag through a linker containing thiourea. In the erythrocyte, the cholesterol probe fluorescently tagged the anion transporter band 3 via thiourea. Tagging by the cholesterol probe, but not by the phospholipid probe, was competitive with an anion transporter inhibitor, implying the presence of a specific binding pocket for cholesterol in this ~100kDa protein. This method could prove an effective strategy for analyzing lipid–protein interactions *in vivo* in the live cell membrane.

How membrane lipids and membrane proteins interact with each other is a fundamental question in biology, as lipid–protein (LP) interactions are expected to play important roles in the cell membrane^{1–3}. Biochemical analyses have unveiled specific functions of membrane lipids; for example, conferring thermal stability to proteins and modulating the affinity between proteins and ligands. However, membrane lipids can regulate protein properties either directly by binding to a specific pocket, or indirectly by changing the physical properties of the membrane surrounding the protein, such as the curvature, lateral pressure, and bilayer thickness. So far, several dozen membrane proteins have been found to interact with membrane lipids through the analysis of X-ray crystal structures⁴. Recently, ion-mobility mass spectrometry was used to screen for lipids that specifically bind to membrane proteins and thereby increase structural stability⁵. However, we have to carefully interpret data regarding the physiological role of the LP interactions detected *in vitro*, whereas confirming these membrane lipid interactions with proteins in the live cell membrane remains technically challenging.

For linking small molecule ligands and their cellular receptors, chemical strategies utilizing various reactive groups have been employed^{6–10}. Active site–directed irreversible inhibitors have been used to label target proteins⁷, and are sometimes used to tag shared mechanistic and structural features of large enzyme classes; for example, fluorophosphonates can tag serine hydrolase family proteins^{8,9}. On the other hand, the decoration of high-affinity ligands with reactive functions enables labeling of the receptor molecule in the proximity of the active site. The quantity, activity, and subcellular localization of the corresponding protein can therefore be detected using these approaches. Theoretically, such irreversible–modification approaches may be applicable to membrane lipids for detecting LP interactions. So far, a couple of photoreactive lipid probes have been utilized for global analysis of *in vivo* LP interactions^{11–13}.

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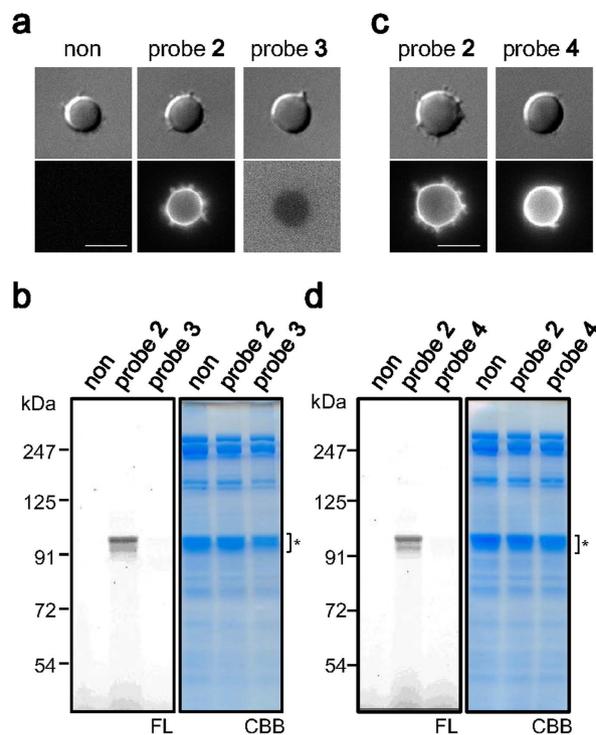


Figure 2. Tagging of the anion transporter band 3 by the cholesterol probe 2. (a,b) Binding and tagging with the cholesterol probe 2. Erythrocytes were treated with probe 2 or 3 (400 nM) at 37 °C, and observed under fluorescence microscopy (a). Bright field (DIC) and fluorescent images are shown. Scale bar, 5 μm. After 8 h incubation, cells were lysed and analyzed by SDS-PAGE (b). The fluorescence and CBB images of the gel are shown. Asterisk indicates band 3. (c,d) Erythrocytes were treated with the cholesterol probe 2 or 4, and analyzed as in (a,b).

junctional complex, and as free dimers^{22,23}, and within each class are subpopulations with further structural heterogeneity²⁴.

Reactivity of thiourea with amine. To our knowledge, thiourea has never previously been utilized for cross-linking biological molecules. The involvement of thiourea was investigated by synthesizing another cholesterol probe 4 in which an amide group was used to conjugate fluorescein to the linker. Probe 4 bound to the erythrocytes to a level comparable to that of probe 2 (Fig. 2c). However, no tagging of band 3 was detected, thus confirming the reactivity of thiourea in the cell membrane (Fig. 2d). We next synthesized a model compound and analyzed the reactivity of thiourea under aqueous conditions. When we mixed the model molecule 5 with an excess amount of *n*-propylamine, the amount of 5 gradually decreased to give rise to a new peak (6) observed by LC-MS analysis (Fig. 3). We purified the newly-emerged compound 6 and analyzed its structure by MS and NMR spectra. The ion peak in ESI MS revealed the molecular formula of C₄₁H₄₆N₄O₉, which lacked the S atom and had additional C₃H₇N compared with 5 (m/z 739.3334 [M + H]⁺, Δmmu = 0.4, calcd for C₄₁H₄₇N₄O₉). In addition, the ¹³C NMR spectrum of compound 6 did not show a quaternary carbon signal (δ_C 182.5 ppm) corresponding to the thiourea carbon in compound 5. Instead, a new carbon signal (δ_C 156.4 ppm) and a set of signals corresponding to the propyl group were detected. These data indicated that thiourea and *n*-propylamine reacted to yield guanidine (Fig. 3), which was supported by the fully assigned ¹H NMR data. Because the reactivity of thiourea was low, the specific tagging by probe 2 led us to hypothesize that cholesterol binds with high affinity to band 3.

Biophysical analyses had predicted the strong binding of cholesterol to band 3, an interaction that may regulate the structure and transporter activity of this protein^{25–27}. So far however, nothing has been reported regarding the location of the binding pocket. To investigate the tagging site of the cholesterol probe 2, a competitive tagging assay using 4,4′-diisothiocyanostilbene-2,2′-disulfonic acid (DIDS), an anion transporter inhibitor, was examined. When erythrocytes were treated with DIDS, we found that tagging by cholesterol probe 2 was significantly suppressed (85% inhibition after 24 h incubation) (Fig. 4a). DIDS is known to form covalent bonds with two lysine residues, e.g. Lys 539 and Lys851 in human band 3, although Lys851 is only reactive at high pH (>8)²⁸. In sheep erythrocytes, Lys539 is conserved (Lys557 in sheep), whereas Lys851 is replaced by Arg869, suggesting that Lys557 reacted with thiourea in the cholesterol probe 2 (Fig. 5, S1). Alternatively, tagging by cholesterol probe 2 may be

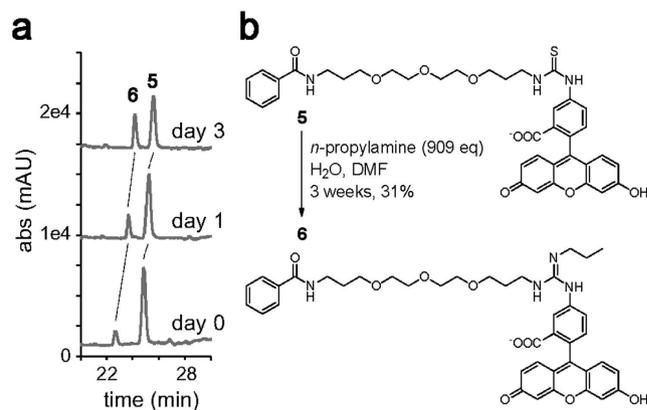


Figure 3. Reactivity of thiourea with amine. (a) Time course analysis of the reaction converting compound 5 to 6. Compound 5 was incubated with *n*-propylamine (3473 eq) and analyzed by LC-MS. UV chromatograms at 488 nm are shown. (b) Chemical structures of model compounds 5 and 6. The thiourea compound 5 was converted to the guanidine compound 6 in the presence of excess *n*-propylamine.

inhibited allosterically by DIDS. In fact, band 3 is expected to be fixed in an outward conformation by stilbene compounds such as DIDS²⁹, which likely inhibit anion transport activity in an allosteric manner³⁰. It is noted that cysteine, another nucleophilic amino acid residue, was not necessary for tagging band 3 since pretreatment of erythrocytes with excess amount of *N*-ethylmaleimide did not inhibit it (Figure S3).

Ligand-dependent tagging of band 3. We finally expanded our approach to another membrane lipid, 1,2-dipalmitoyl-*sn*-glycero-3-phosphoethanolamine (DPPE) (7). We designed probe 8 with the same structural features as that of the cholesterol probe 2. The DPPE probe 8 bound to erythrocytes to a similar level as cholesterol probes 2 and 4 (Figure S4). Interestingly, probe 8 also specifically tagged band 3 (Fig. 4b, S4). The kinetics of the tagging was similar to that of the cholesterol probe 2 (Figure S4), while the tagging efficiency was slightly higher than that of probe 2 (0.21% vs 0.073%) (Figure S2). However, in this case, the competitive effect of DIDS was minor (11.3% inhibition at 24 h; Fig. 4b). This suggested that the binding site of the DPPE probe 8 is not overlapped with the DIDS binding site, or not affected by the conformational change of band 3 by DIDS (Fig. 5). Otherwise, there might be multiple hydrophobic pockets for probe 8, and the overall tagging level was not decreased upon binding of DIDS. Although we can only speculate the binding mode of the DPPE probe 8, it is clear that band 3 has multiple lipid binding sites that were occupied by lipid probes 2 and 8. The atomic basis of these interactions would be more precisely analyzed by X-ray crystal structure analysis.

Discussion

The approach presented here to link lipids and proteins in the cell membrane has several unique features. First, the lipid probes we used are amphiphilic. Amphiphilicity can suppress membrane permeability, which is a key point when considering the specific tagging of membrane molecules. In fact, when the lipid probes were treated with the erythrocyte lysate, proteins other than band 3 were also tagged (Figure S5). This indicates that the reactivity of thiourea was regulated by the amphiphilicity of the probe, which would enable the introduction of other reactive groups to optimize the tagging rate and the modes of tagging. Next, we applied a novel chemical tagging strategy to the endogenous membrane molecules. Pioneering studies have shown that membrane proteins can be efficiently tagged by chemical probes, which are designed based on soluble inhibitors^{31,32}. In contrast, our results demonstrated that we could analyze the interactions between endogenous lipids and membrane proteins, in spite of the highly competitive environment *in vivo*. The interactions between cholesterol and membrane proteins are expected to be stabilized by three interactions: hydrophobic interactions, hydrogen bonding interactions, and CH- π interactions^{33,34}. Although the cholesterol probe 2 cannot form hydrogen bonding due to the absence of the free hydroxyl group at C-3, the hydrophobic and CH- π interactions by cholest-5-en structure in probe 2 could mimic the interaction between cholesterol and proteins. Finally, the thiourea-containing lipid probes revealed a tentative binding site for cholesterol in band 3 (Fig. 5). Although we can speculate that there are multiple interactions between cholesterol and band 3, our cholesterol probes tagged a specific binding site. This might be due to the limited reactivity of thiourea; tagging by thiourea requires the presence of specific amino acids, i.e., lysine, located in the close proximity to the lipid probe. In contrast, cholesterol probes possessing photoreactive diazirine allowed researchers to identify several hundred cholesterol-binding proteins using proteomic analyses¹². The combination of

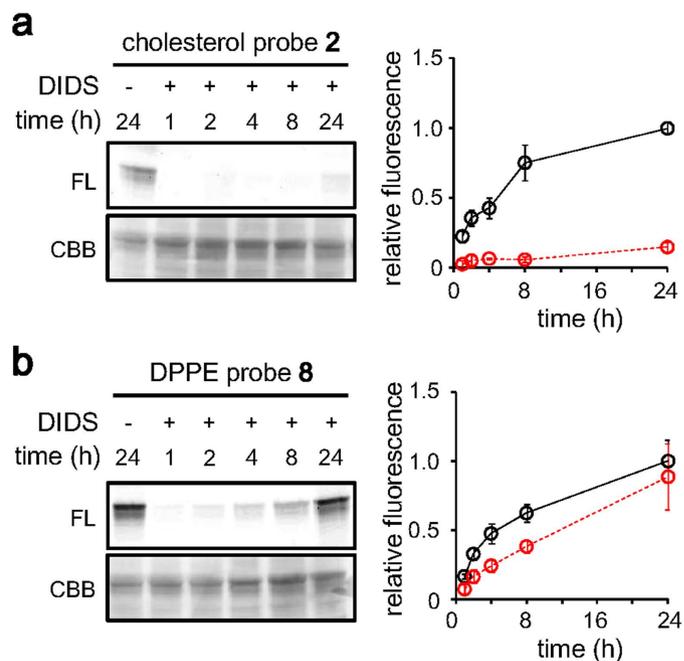


Figure 4. Competitive tagging of band 3 by lipid probes. Erythrocytes were pre-treated with a transporter inhibitor DIDS (100 μ M) for 30 min, then time-dependent tagging was examined using the cholesterol probe 2 (a) or the DPPE probe 8 (b). The concentration of the probes was 400 nM. The fluorescence intensities relative to that obtained after 24 h incubation in the absence of DIDS are shown. The tagging level in the absence (black solid line, Figures S2 and S4) or in the presence (red dashed line) of DIDS is shown. Means \pm SD of 3–6 experiments are shown.

narrowly-reactive probes for analyzing specific interactions with highly-reactive probes for comprehensive analysis would have significant advantages for LP interactome analyses³⁵.

In summary, we have demonstrated that the chemical tagging strategy is feasible for analyzing LP interactions in the cell membrane. Amphiphilic lipid probes can be used to analyze a specific LP interaction, and this may prove applicable to comprehensive proteomic analyses by tuning the probe reactivities. Expanding this approach to exogenous molecules targeting membrane lipids, such as antibiotics, is also an attractive challenge.

Methods

General considerations for organic synthesis. All reagents and solvent were used as received from commercial suppliers and were used without further purification. IR spectra were measured using an FTIR spectrometer equipped with a ZnSe ATR plate. Optical rotations were determined using the sodium D line (589 nm). NMR spectra were measured on a 500 MHz instrument. ¹H and ¹³C chemical shifts (δ) are shown in parts per million (ppm), and coupling constants (J) are in hertz (Hz). The following abbreviations are used to describe multiplicities: s, singlet; d, doublet; t, triplet; dd, doublet of doublets; dt, doublet of triplets; tt, triplet of triplets; m, multiplet; br, broad; ovl, overlapped. Mass spectral data were collected with ESI IT-TOF MS. Flash column chromatography was performed over Silica Flash F60 (SiliCycle) using an elution system as described for each experiment. Synthesis procedures and physico-chemical properties are described in the Supplementary Information file.

General materials and methods for Biochemical/Biological Experiments. Sheep whole blood was purchased from Kohjin Bio Co., Ltd. and used within two weeks after blood collection. Fluorescent images of SDS-PAGE gels were acquired with Typhoon 9400 (GE Healthcare). Fluorescent images of cells were acquired using an Olympus IX81 microscope equipped with UPLSAPO 100 \times objective (Olympus, Tokyo, Japan).

Tagging of anion transporter band 3 with probes. Sheep whole blood (0.5 mL) was centrifuged (300 g \times 5 min, unless stated) and supernatant was discarded. The precipitated red blood cells (RBC) were washed with 0.7 mL of ice-cold phosphate buffer (10 mM phosphate, 150 mM NaCl, pH 7.4) for 5 times, and re-suspended in 0.5 mL of phosphate buffer. RBC suspensions (1 mL, 10⁸ cells/mL) were mixed with lipid probes (0.4 μ M, 0.4% DMF), and incubated at 37 $^{\circ}$ C for 8 h with continuous mixing. Microscope analysis was performed within 30–60 min after addition of probes. After incubation, RBC suspensions were centrifuged, washed with 1 mL of ice-cold phosphate buffer twice, and mixed with

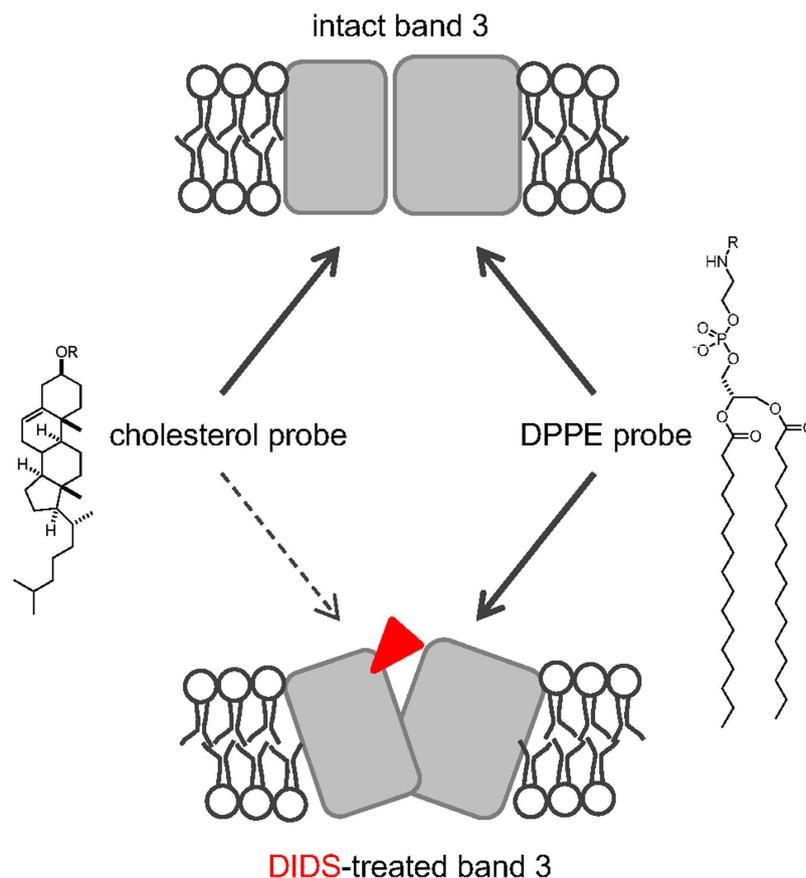


Figure 5. Schematics for the lipid-band 3 interactions. Both cholesterol and DPPE probes can tag band 3. After DIDS treatment (red triangle), only tagging by cholesterol probe was abolished. Covalent modification of Lys557 by DIDS might directly inhibit tagging of the lysine by the cholesterol probe. Otherwise, overall conformational change by DIDS might inhibit interaction between cholesterol probe and band 3.

80 μ L of SDS-PAGE sample buffer. After boiling at 100 $^{\circ}$ C for 5 min, 20 μ L of the sample was subjected to SDS-PAGE (8%) and the fluorescent images were acquired (Ex: 488 nm; Em 532 nm.). All proteins were stained by CBB and the signal intensity was calculated using MetaMorph software (Molecular Devices).

Competition assay with DIDS. Washed RBC solutions ($\times 10^8$ cells/mL) were prepared as described. RBCs were treated with DIDS (final concentration: 100 μ M in 0.4% DMF) on ice for 30 min in the dark. After incubation, RBC solution was centrifuged, and washed with phosphate buffer (1 mL) twice followed by washing with a BSA solution (0.5 w/v% BSA in PBS). After extensive PBS wash of RBCs (1 mL each, three times), RBCs were suspended in 1 mL of phosphate buffer and treated with lipid probes as described above.

Reactivity test of the thiourea-containing model compound 5. Compound 5 (2 mg/mL, 2.8 mM) was mixed with *n*-propylamine (400 μ L, 3473 eq), 4,4'-bis(2-methyl-2-propanyl)biphenyl (0.4 mg/mL), DMF (50 μ L), and water (50 μ L), which was stirred at 37 $^{\circ}$ C. After incubation, the reaction mixture (10 μ L) was mixed with 200 μ L of water, lyophilized, and dissolved in 500 μ L of DMF. A portion (1 μ L) of the prepared sample was subjected to LC-MS analysis. The analytes were separated by an ODS column (Senshu-PAK PEGASIL ODS SP100, $\phi 3 \times 100$ mm, 30 $^{\circ}$ C) using a gradient elution system (50–100% aq MeOH, 200 μ L/min).

References

- Phillips, R., Ursell, T., Wiggins, P. & Sens, P. Emerging roles for lipids in shaping membrane-protein function. *Nature* **459**, 379–385 (2009).
- Lee, A. G. Lipid-protein interactions in biological membranes: a structural perspective. *Biochim. Biophys. Acta* **1612**, 1–40 (2003).
- Oates, J. & Watts, A. Uncovering the intimate relationship between lipids, cholesterol and GPCR activation. *Curr. Opin. Struct. Biol.* **21**, 802–807 (2011).

4. Yeagle, P. L. Non-covalent binding of membrane lipids to membrane proteins. *Biochim. Biophys. Acta* **1838**, 1548–1559 (2014).
5. Laganowsky, A. *et al.* Membrane proteins bind lipids selectively to modulate their structure and function. *Nature* **510**, 172–175 (2014).
6. Takaoka, Y., Ojida, A. & Hamachi, I. Protein organic chemistry and applications for labeling and engineering in live-cell systems. *Angew. Chem. Int. Ed. Engl.* **52**, 4088–4106 (2013).
7. Gersch, M., Kreuzer, J. & Sieber, S. A. Electrophilic natural products and their biological targets. *Nat. Prod. Rep.* **29**, 659–682 (2012).
8. Cravatt, B. F., Wright, A. T. & Kozarich, J. W. Activity-based protein profiling: from enzyme chemistry to proteomic chemistry. *Annu. Rev. Biochem.* **77**, 383–414 (2008).
9. Moellering, R. E. & Cravatt, B. F. How chemoproteomics can enable drug discovery and development. *Chem. Biol.* **19**, 11–22 (2012).
10. Otsuki, S. *et al.* Chemical tagging of a drug target using 5-sulfonyl tetrazole. *Bioorg. Med. Chem. Lett.* **23**, 1608–1611 (2013).
11. Gubbens, J. *et al.* Photocrosslinking and click chemistry enable the specific detection of proteins interacting with phospholipids at the membrane interface. *Chem. Biol.* **16**, 3–14 (2009).
12. Hulce, J. J., Cognetta, A. B., Niphakis, M. J., Tully, S. E. & Cravatt, B. F. Proteome-wide mapping of cholesterol-interacting proteins in mammalian cells. *Nat. Methods* **10**, 259–264 (2013).
13. Haberkant, P. *et al.* *In vivo* profiling and visualization of cellular protein-lipid interactions using bifunctional fatty acids. *Angew. Chem. Int. Ed. Engl.* **52**, 4033–4038 (2013).
14. Simons, K. & Vaz, W. L. Model systems, lipid rafts, and cell membranes. *Annu. Rev. Biophys. Biomol. Struct.* **33**, 269–295 (2004).
15. Paila, Y. D. & Chattopadhyay, A. Membrane cholesterol in the function and organization of G-protein coupled receptors. *Subcell. Biochem.* **51**, 439–466 (2010).
16. Patel, K. R., Li, M. P., Schuh, J. R. & Baldeschwieler, J. D. The Pharmacological Efficacy of a Rigid Non-Phospholipid Liposome Drug Delivery System. *Biochim. Biophys. Acta* **797**, 20–26 (1984).
17. Miyajima, K., Baba, T. & Nakagaki, M. Hemolytic-Activity of Polyoxyethylene Cholesteryl Ethers. *Colloid and Polymer Science* **265**, 943–949 (1987).
18. Ishiwata, H., Sato, S. B., Vertut-Doi, A., Hamashima, Y. & Miyajima, K. Cholesterol derivative of poly(ethylene glycol) inhibits clathrin-independent, but not clathrin-dependent endocytosis. *Biochim. Biophys. Acta* **1359**, 123–135 (1997).
19. Sato, S. B. *et al.* Distribution and transport of cholesterol-rich membrane domains monitored by a membrane-impermeant fluorescent polyethylene glycol-derivatized cholesterol. *J. Biol. Chem.* **279**, 23790–23796 (2004).
20. Fairbanks, G., Steck, T. L. & Wallach, D. F. Electrophoretic analysis of the major polypeptides of the human erythrocyte membrane. *Biochemistry* **10**, 2606–2617 (1971).
21. Cooper, R. A. Influence of increased membrane cholesterol on membrane fluidity and cell function in human red blood cells. *J. Supramol. Struct.* **8**, 413–430 (1978).
22. Mankelaw, T. J., Satchwell, T. J. & Burton, N. M. Refined views of multi-protein complexes in the erythrocyte membrane. *Blood Cells Mol. Dis.* **49**, 1–10 (2012).
23. Burton, N. M. & Bruce, L. J. Modelling the structure of the red cell membrane. *Biochem. Cell Biol.* **89**, 200–215 (2011).
24. Kodippili, G. C. *et al.* Analysis of the mobilities of band 3 populations associated with ankyrin protein and junctional complexes in intact murine erythrocytes. *J. Biol. Chem.* **287**, 4129–4138 (2012).
25. Klappauf, E. & Schubert, D. Band 3-protein from human erythrocyte membranes strongly interacts with cholesterol. *FEBS Lett.* **80**, 423–425 (1977).
26. Schubert, D. & Boss, K. Band 3 protein-cholesterol interactions in erythrocyte membranes. Possible role in anion transport and dependency on membrane phospholipid. *FEBS Lett.* **150**, 4–8 (1982).
27. Muhlebach, T. & Cherry, R. J. Influence of cholesterol on the rotation and self-association of band 3 in the human erythrocyte membrane. *Biochemistry* **21**, 4225–4228 (1982).
28. Okubo, K., Kang, D., Hamasaki, N. & Jennings, M. L. Red blood cell band 3. Lysine 539 and lysine 851 react with the same H2DIDS (4,4'-diisothiocyanodihydrostilbene-2,2'-disulfonic acid) molecule. *J. Biol. Chem.* **269**, 1918–1926 (1994).
29. Takazaki, S. *et al.* The functional role of arginine 901 at the C-terminus of the human anion transporter band 3 protein. *J. Biochem.* **139**, 903–912 (2006).
30. Salhany, J. M. Mechanistic basis for site-site interactions in inhibitor and substrate binding to band 3 (AE1): evidence distinguishing allosteric from electrostatic effects. *Blood Cells Mol. Dis.* **27**, 901–912 (2001).
31. Chang, J. W., Moellering, R. E. & Cravatt, B. F. An activity-based imaging probe for the integral membrane hydrolase KIAA1363. *Angew. Chem. Int. Ed. Engl.* **51**, 966–970 (2012).
32. Miki, T. *et al.* LDAl-based chemical labeling of intact membrane proteins and its pulse-chase analysis under live cell conditions. *Chem. Biol.* **21**, 1013–1022 (2014).
33. Epand, R. M., Thomas, A., Brasseur, R. & Epand, R. F. Cholesterol interaction with proteins that partition into membrane domains: an overview. *Subcell. Biochem.* **51**, 253–278 (2010).
34. Cang, X. *et al.* Mapping the functional binding sites of cholesterol in beta2-adrenergic receptor by long-time molecular dynamics simulations. *J. Phys. Chem. B* **117**, 1085–1094 (2013).
35. Haberkant, P. & Holthuis, J. C. Fat & fabulous: bifunctional lipids in the spotlight. *Biochim. Biophys. Acta* **1841**, 1022–1030 (2014).

Acknowledgements

This work was supported in part by a Grant-in-Aid from the Japan Society for the Promotion of Science (JSPS) and the Ministry of Education, Culture, Sports, Science, and Technology of Japan (MEXT), and the Ministry of Health, Labour and Welfare of Japan (MHLW).

Author Contributions

S.N. and H.K. designed the experiments and wrote the manuscript; A.M. performed most experiments; N.K. and N.T. supported the organic synthesis and the tagging experiments.

Additional Information

Supplementary information accompanies this paper at <http://www.nature.com/srep>

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Moriyama, A. *et al.* *In Vivo* Linking of Membrane Lipids and the Anion Transporter Band 3 with Thiourea-modified Amphiphilic Lipid Probes. *Sci. Rep.* **5**, 17427; doi: 10.1038/srep17427 (2015).



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