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Fructose induces glucose-dependent insulinotropic polypeptide, glucagon-like peptide-1 and insulin secretion: Role of adenosine triphosphate-sensitive K⁺ channels

Yusuke Seino¹*, Hidetada Ogata², Ryuya Maekawa², Takako Izumoto², Atsushi Iida², Norio Harada³, Takashi Miki⁴, Susumu Seino⁵, Nobuya Inagaki³, Shin Tsunekawa², Yutaka Oiso², Yoji Hamada¹

Departments of ¹Metabolic Medicine, and ²Endocrinology and Diabetes, Nagoya University Graduate School of Medicine, Nagoya, ³Department of Diabetes, Endocrinology and Nutrition, Graduate School of Medicine, Kyoto University, Kyoto, ⁴Department of Medical Physiology, Graduate School of Medicine, Chiba University, Chiba, and ⁵Division of Molecular and Metabolic Medicine, Kobe University Graduate School of Medicine, Kobe, Japan

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*Correspondence
Yusuke Seino
Tel.: +81-52-744-2191
Fax: +81-52-744-2191
E-mail address: yusuke@med.nagoya-u.ac.jp

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INTRODUCTION
Glucose-dependent insulinotropic polypeptide (GIP) and glucagon-like peptide-1 (GLP-1) are incretin hormones secreted from enteroendocrine K-cells and L-cells by nutrients such as carbohydrate¹,².

Adenosine triphosphate-sensitive K⁺ (KATP) channels play an important role in glucose-induced insulin secretion from pancreatic β-cells³. It has been reported that K-cells and L-cells express glucokinase and KATP channels identical to those expressed in pancreatic β-cells⁴–⁵. In addition, facilitative glucose transporter 5 (GLUT5), which absorbs fructose from intestinal lumen to cytosol⁶, is abundantly expressed in K-cells, L-cells and β-cells. However, the role of fructose and the involvement of the KATP channel in the secretion of GIP, GLP-1 and insulin in vivo are poorly understood.

In the present study, we investigated the contributions of fructose and the KATP channel in the secretion of these hormones utilizing KATP channel-deficient mice.

MATERIALS AND METHODS
Mice
C57BL/6J mice (Kir6.2⁺/⁺ mice) and mice lacking the KATP channel (Kir6.2⁻/⁻ mice) were used. We carried out all animal experiments according to the protocol approved by the Nagoya University Institutional Animal Care and Use Committee.

Plasma Biochemical Analyses
Blood glucose levels were measured with ANTSENSE II (Bayer Medical, Leverkusen, Germany). Plasma total GIP and GLP-1 levels were measured using the GIP (TOTAL) ELISA kit (Merck...
Millipore, Billerica, MA, USA) and an electrochemiluminescent sandwich immunoassay (Meso Scale Discovery, Gaithersburg, MD, USA) as previously described7,8. Plasma insulin levels were determined by an ELISA kit (Morinaga, Tokyo, Japan).

**Induction of Diabetes**

As described previously7, streptozotocin (STZ; 150 mg/kg body-weight) was given intraperitoneally to Kir6.2+/+ mice after a 16-h fast.

**Diazoxide and Fructose Administration**

After 16 h of food deprivation, 240 mg/kg bodyweight of diazoxide (Wako, Osaka, Japan) was given orally7.9. 90 min after diazoxide administration, 6 g/kg bodyweight of fructose was given orally.

**MIN6 Experiment**

MIN6-K8 β-cells were cultured and stimulated for 30 min by various materials after pre-incubation for 30 min in HEPES-Krebs buffer with 2.8 mmol/L glucose, and released insulin was evaluated by insulin assay kit as previously reported9.

**Statistical Analysis**

Statistical analysis was carried out by unpaired, two-tailed Student’s t-test or two-way ANOVA.

**RESULTS**

**Fructose Induces GIP Secretion in the Diabetic State**

We first examined whether fructose stimulates GIP secretion. In Kir6.2+/+ mice, fructose tended to, but not significantly, stimulate GIP secretion in a normal state, but significantly enhanced the GIP secretion in the STZ-induced diabetic state (Figure 1a). To investigate the involvement of the KATP channel in fructose-induced GIP secretion in the diabetic state, we examined the effect of the KATP channel activator, diazoxide, on fructose-induced GIP secretion. Pretreatment of diazoxide did not affect fructose-induced GIP secretion in the diabetic state (Figure 1b). Fructose-induced GLP-1 levels at 15 min were not different under the normoglycemic condition and hyperglycemic condition (Figure 1c).

**KATP Channels Are Not Involved in Fructose-Induced GLP-1 Secretion In Vivo**

We next investigated whether the KATP channel participates in fructose-induced GLP-1 secretion in vivo, by utilizing Kir6.2−/− mice. Both in Kir6.2+/+ and Kir6.2−/− mice, fructose significantly stimulated GLP-1 secretion more than twofold at 15 min of fructose administration (Figure 2b). In contrast, fructose did not stimulate GIP secretion in Kir6.2−/− mice at all (Figure 2a).

**KATP Channels Are Involved in Fructose-Induced Insulin Secretion In Vivo and In Vitro**

To assess whether fructose-induced insulin secretion requires the KATP channel pathway, we investigated blood glucose levels and serum insulin levels during oral fructose tolerance test in both Kir6.2+/+ and Kir6.2−/− mice. The blood glucose levels were significantly higher in Kir6.2−/− mice than in Kir6.2+/+ mice (Figure 2c). Fructose significantly stimulated insulin secretion in Kir6.2+/+ mice at 15 min, but not in Kir6.2−/− mice at
tended to decrease insulin secretion at 8.3 mmol/L glucose ($P = 0.05$). The addition of 20 mmol/L fructose significantly potentiated insulin secretion at 8.3 mmol/L glucose, and diazoxide completely blocked the insulin response (Figure 3c).

Pretreatment of diazoxide did not affect fructose-induced GLP-1 secretion at 15 min in either Kir6.2$^{+/+}$ mice or Kir6.2$^{-/-}$ mice (Figure 3d).

**DISCUSSION**

The mechanism by which fructose stimulates gut hormone secretion is not well known. In the present study, we investigated the role of the $K_{ATP}$ channels in fructose-induced GIP, GLP-1 and insulin secretion in vivo.

We previously reported that the $K_{ATP}$ channels in K-cells are in a closed state under the normoglycemic condition in vivo, and are in an open state under the hyperglycemic condition$^7$. The increase of ATP produced by metabolism of glucose closes the $K_{ATP}$ channels in the K-cells under the hyperglycemic condition and enhances glucose-induced GIP secretion, suggesting that $K_{ATP}$ channels in K-cells contribute to glucose-induced GIP secretion under the hyperglycemic condition. However, the present results show that this mechanism is not involved in fructose-induced GIP secretion in the diabetic state and that the $K_{ATP}$ channels in K-cells do not contribute to fructose-induced GIP secretion under the hyperglycemic condition. In previous reports, 3 g/kg fructose did not stimulate GIP secretion in C57BL/6j mice, but did stimulate GIP secretion in obese type 2 diabetic model $ob/ob$ mice$^{10,11}$. The mechanism of such fructose-induced GIP secretion in various diabetic models remains to be elucidated.

In the present study, fructose was found to significantly induce GLP-1 secretion in Kir6.2$^{-/-}$ mice, and pretreatment of diazoxide did not block fructose-induced GLP-1 secretion at 15 min and fructose-induced GLP-1 secretion was not enhanced under the hyperglycemic condition. These results show that the $K_{ATP}$ channel is not required for fructose-induced GLP-1 secretion in vivo. However, a previous in vitro study using GLUTag cells found that fructose-induced GLP-1 secretion was entirely $K_{ATP}$ channel-dependent$^{12}$. This discrepancy could be due to the nature of the GLUTag cell line and/
or the fact that GLP-1 secretion is regulated by various factors, such as nutrients, intestinal hormones, neuropeptides and neuronal signal in vivo\textsuperscript{13-16}.

It is reported that activation of sweet taste receptors in pancreatic \(\beta\)-cells stimulates insulin secretion through the phospholipase C pathway\textsuperscript{17,18}. Kyriacis \textit{et al.} also reported that insulin secretion was not induced by glucose catalyzed from fructose, but by activation of the sweet taste receptor in a glucose-dependent manner through transient receptor potential cation channel, subfamily M, member 5\textsuperscript{17}. In the present study, the fructose-induced insulin secretion seen in Kir6.2\textsuperscript{+/+} mice was not observed at all in Kir6.2\textsuperscript{−/−} mice, and diazoxide completely blocked fructose-induced insulin secretion \textit{in vivo} and \textit{in vitro}. These results show that the \(K_{\text{ATP}}\) channel in \(\beta\)-cells plays an essential role in the fructose-induced insulin secretion. In contrast, we previously showed that insulin secretion mediated by the vagal nerve \textit{in vivo} was \(K_{\text{ATP}}\) channel-independent\textsuperscript{19}, and it was reported previously that insulin secretion through activation of the phospholipase C pathway differed from that induced by carbachol, the activator of the muscarinic receptor\textsuperscript{18}. These findings suggest that the \(K_{\text{ATP}}\) channel-dependent phospholipase C-transient receptor potential cation channel, subfamily M, member 5 pathway is involved in fructose-induced insulin secretion \textit{in vivo}.

In conclusion, fructose stimulates GLP-1 secretion under normoglycemia, but enhances GIP secretion under the hyperglycemic condition, both of which modifications are in a \(K_{\text{ATP}}\) channel-independent manner. \(K_{\text{ATP}}\) channels play an essential role in the insulin secretion induced by fructose \textit{in vivo}.

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**DISCLOSURE**

The authors declare no conflict of interest.

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