

## Basic Study

## Female spontaneously diabetic Torii fatty rats develop nonalcoholic steatohepatitis-like hepatic lesions

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### Abstract

**AIM:** To investigate the histological features of the liver in spontaneously diabetic Torii (SDT) fatty rats compared with age-matched Sprague-Dawley (SD) rats.

**METHODS:** Female SDT *Lep<sup>fa</sup>* (SDT fatty) rats and age-matched SD rats were fed *ad libitum*. Body weight and biochemical parameters, such as serum glucose, triglyceride (TG), total cholesterol (TC), alanine aminotransferase (ALT) and aspartate aminotransferase (AST) levels as well as fatty acid and TG accumulation in the liver were evaluated at 8 wk of age in the non-fasting state and at 8-wk intervals from 8 to 40 wk of age. Histopathological examinations of the liver were performed using hematoxylin and eosin and Sirius Red staining as well as double staining for ED-1 and toluidine blue. The expression of genes involved in TG synthesis, inflammation, and fibrosis was examined in the liver.

**RESULTS:** SDT fatty rats showed significantly increased body weight compared with SD rats. Serum glucose, TG, and TC levels were significantly higher in SDT fatty rats compared with SD rats. The serum AST and ALT levels in SDT fatty rats were significantly elevated at 8 wk of age compared with the levels in SD rats. Hepatic TG content was marked in SDT fatty rats from 8 to 32 wk of age. Histopathologically, severe hepatosteatosis accompanied by inflammation was observed at 8 wk of age, and fibrosis started to occur at 32 wk of age. Furthermore, Sirius Red and ED-1 staining were increased in the liver at 32 wk of age. Hepatic gene expression related to TG synthesis, inflammation and fibrosis tended to increase in SDT fatty rats compared with SD rats, and the gene expression related to TG secretion was decreased in SDT fatty rats compared with SD rats.

**CONCLUSION:** Female SDT fatty rats have the potential to become an important animal model of nonalcoholic steatohepatitis with type 2 diabetes and obesity.

**Key words:** Nonalcoholic steatohepatitis; Spontaneously diabetic Torii fatty rat; Fibrosis; Fatty liver; Metabolic syndrome

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**Core tip:** Nonalcoholic steatohepatitis (NASH) is recognized as a major risk for progression to cirrhosis and liver failure or to hepatocellular carcinoma. In this study, we investigated the histological features of the liver in female spontaneously diabetic Torii (SDT) fatty rats. The SDT fatty rats exhibited pathophysiological features of NASH, and fibrosis appeared in the liver without dietary manipulation. Female SDT fatty rats have the potential to become an important animal model of NASH with type 2 diabetes and obesity, a condition for which few models currently exist.

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## INTRODUCTION

Nonalcoholic fatty liver disease (NAFLD) is presently well recognized as the most common chronic liver disease in the Western world<sup>[1]</sup>. NAFLD is strongly associated with central obesity, type 2 diabetes, dyslipidemia, hypertension, and insulin resistance<sup>[1]</sup>. NAFLD represents the pathology of fatty liver, including simple steatosis, nonalcoholic steatohepatitis (NASH),

and cirrhosis<sup>[2]</sup>. In Western countries, 4%-22% of NAFLD cases lead to hepatocellular carcinoma<sup>[3-6]</sup>. The pathogenesis of NASH remains poorly defined, and effective pharmacological therapies for NASH have not been approved.

To understand the complicated features of NAFLD/NASH, animal models offer a source of important information. It has been reported that *ob/ob* mice, *db/db* mice, and KK-*A<sup>y</sup>* mice, as NAFLD animal models, exhibit spontaneous hepatic steatosis based on insulin resistance and obesity, but these mice do not progress to liver fibrosis when fed a normal diet<sup>[7-9]</sup>. In contrast, methionine and choline-deficient (MCD) diet-induced NASH is accompanied by hepatic inflammation and fibrosis in *ob/ob* mice and *db/db* mice<sup>[10,11]</sup>.

Spontaneously diabetic Torii *Lepr<sup>fa</sup>* (SDT fatty) rats, which are generated by introducing the *fa* allele of the Zucker fatty rat into the SDT rat genome, represent a new model of obese type 2 diabetes. SDT fatty rats exhibit hyperphagia that leads to obesity associated with hyperglycemia, hyperinsulinemia, and hyperlipidemia at a young age compared with SDT rats<sup>[12,13]</sup>. Compared with SDT rats, this early onset of diabetes in SDT fatty rats is considered to be caused by obesity resulting from hyperphagia. Furthermore, female SDT rats also exhibit hyperglycemia at the same young age as male SDT fatty rats<sup>[14,15]</sup>.

SDT fatty rats have been used to investigate the effects of a high-fat diet in a previous study. In this prior study, the rats fed a high-fat diet had improved hyperglycemia and increased serum glucagon-like peptide-1 (GLP-1) levels after refeeding<sup>[16]</sup>. Moreover, histopathological observations showed improvement in the fatty liver and pancreatic abnormalities. This previous report showed beneficial effects on diabetes in SDT fatty rats fed a high-fat diet.

With regard to the metabolic responses to a high-fat diet in Sprague-Dawley (SD) rats, steatohepatitis associated with hepatic fatty changes, inflammation, and mitochondrial lesions has been confirmed<sup>[17]</sup>. C57BL6 mice fed a high-fat diet also develop steatohepatitis<sup>[18]</sup>. However, the degree of hepatic fibrosis is less severe in animals fed a high-fat diet compared with choline-deficient diet models.

In the present study, we investigated the histological features of the liver in SDT fatty rats without manipulating the diet in comparison with age-matched SD rats. In addition to the pathophysiology, the biochemical parameters, hepatic lipid content, and expression of genes were examined.

## MATERIALS AND METHODS

### Animals

This experiment was conducted in compliance with the Guidelines for Animal Experimentation of Japan Tobacco Biological/Pharmacological Research Laboratories. The animal protocol was designed to

minimize pain or discomfort to the animals. Female SDT fatty rats from our colony were used in the study. At 8 wk of age, SDT fatty rats were divided into 5 groups for necropsy at 8, 16, 24, 32, and 40 wk of age. SD rats [Crj: CD (SD); Charles River Japan, Yokohama, Japan], as control animals, at 8 wk of age were also divided into the same 5 groups as the SDT fatty rats. In the experimental period, one SDT fatty rat in the 24-wk necropsy group died due to unidentified causes, leaving 4 animals in this group. The rats were housed individually in suspended bracket cages in a climate-controlled room with a temperature of 23 °C ± 3 °C, humidity 55% ± 15%, and a 12-h dark-light cycle, and the rats had free access to a commercial diet (CRF-1, Charles River Japan, Yokohama, Japan) and water. The rats were anesthetized with isoflurane inhalation before the procedures.

### **Biological parameters**

Body weight and biochemical parameters, such as serum glucose, triglyceride (TG), total cholesterol (TC), alanine aminotransferase (ALT), and aspartate aminotransferase (AST) levels, were measured at 8 wk of age and thereafter at 8-wk intervals until 40 wk of age. Blood samples were collected from the tail vein of the rats. The samples were stored in ice water until measurement. Serum glucose, TG, TC, ALT, and AST levels were measured as biochemical parameters using commercial kits (Roche Diagnostics, Basel, Switzerland) in an automatic analyzer (Hitachi 7170S; Hitachi, Tokyo, Japan).

### **Tissue sampling and histopathology**

Necropsy was performed at 8, 16, 24, 32, and 40 wk of age ( $n = 5$  for each age, except 24 wk of age with  $n = 4$ ). All animals were sacrificed by exsanguination under isoflurane anesthesia. The livers were sampled for measurement of gene expression, hepatic lipid content, and histopathology. Samples for determination of gene expression and hepatic lipid content were stored at -80 °C until analysis. For histopathology, the livers were immediately fixed in 10% neutral-buffered formalin. After resection, the tissue was paraffin-embedded by standard techniques and sectioned (3 to 5 μm). The sections were stained with hematoxylin and eosin (HE). The samples were all examined histopathologically in a blind manner, and the findings were graded from normal (-) to severe (3+).

### **Immunostaining**

The paraffin-embedded tissue sections were also stained with Sirius Red and toluidine blue (TB), and immunohistochemistry was performed for ED-1 (CD68: 1:100, BMA BIOMEDICALS, Switzerland) and alpha-smooth muscle actin ( $\alpha$ -SMA; 1A4, 1:50, Dako, Denmark).

### **Hepatic TG and FFA content**

An approximate 100 mg portion of the liver, 0.5 mL of methanol, and zirconia beads were added to tubes. The liver portion was homogenized using a mixer mill (MM300 Retch) (25 Hz for 10 min). To the homogenized solution, 1 mL of chloroform was added and mixed thoroughly. The mixture was then centrifuged (10000  $g$  for 5 min at 4 °C), and the resultant supernatant was collected. Solvents contained in 0.5 mL of the supernatant were dried under a stream of nitrogen gas. To the residue, 0.5 mL of 2-propanol was added, and the residue was then dissolved again. The TG concentration of the 2-propanol solution was determined using the biochemistry automatic analyzer (Hitachi 7170S; Hitachi, Tokyo, Japan). The free fatty acid (FFA) concentration was analyzed using a NEFA-C kit (Wako Pure Chemical Industries, Ltd., Osaka, Japan).

### **mRNA quantification with real-time quantitative PCR**

Total RNA was extracted from the liver with the GenElute Mammalian Total RNA Miniprep Kit (Sigma-Aldrich, St. Louis, MO, United States) at 8, 16, 24, 32, and 40 wk of age. RNA was transcribed into cDNA using M-MLV reverse transcriptase and random primers (Invitrogen, Carlsbad, CA, United States). The reaction mixture was incubated for 10 min at 25 °C, 1 h at 37 °C, and 5 min at 95 °C. Real-time PCR quantification was performed in a 50 μL reaction mixture with an automated sequence detector combined with ABI Prism 7700 Sequence Detection System software (Applied Biosystems, Foster City, CA, United States). The reaction mixture contained 50 ng of synthesized cDNA, 3.5 mmol/L MgCl<sub>2</sub>, 0.3 μmol/L primers, 0.1 μmol/L probes, and 1.25 units of Ampli Taq Gold<sup>®</sup>. The cycle parameters included 10 min at 95 °C followed by 40 cycles of 15 s at 95 °C and 60 s at 60 °C. The following primers and FAM-conjugated probes were designed using Primer Express software (Applied Biosystems): SREBP-1c (forward, CGACTACATCCGCTTCTTACAGC; reverse, TTTTGTGAGCACTTCGCAGG; probe, CAGCAACCAGAACTCAAGCAGGAGAACC), DGAT2 (forward, GCTGATAGCTGCTCTCTACTTCACC; reverse, TGTGATCTCCTGCCACCTTTCT; probe, TGGCATTGACTGGAACACGCCCA), FAS (forward, ACTGAACGGCATTACTCGGTCC; reverse, GTGTCCCATGTTGGATTTGGTG; probe, TTCCGCCAGAGCCCTTTGTTAATTGG), MTP (forward, GGACGTTGTGTTACTGTGGAGG; reverse, ACATTGACAGCCGTTATCGTGA; probe, GATCCCAGCGGTGACGATGATCAACTG), and  $\beta$ -actin (purchased from Applied Biosystems). The expression of the following genes were confirmed using Taqman Gene Expression Assays on Demand and Universal Maser Mix: ACC (Rn00573474\_m1),  $\alpha$ -SMA (Rn01759928\_g1), MCP-1 (Rn00580555\_m1), TGF- $\beta$  (Rn99999016\_m1), TNF- $\alpha$  (Rn99999017\_m1), and

collagen type 1 (Rn01463848\_m1).

### Statistical analysis

The results of the biological parameters are expressed as the mean  $\pm$  SD. Statistical analysis of differences between mean values was performed using an *F*-test followed by the Student *t*-test or the Aspin-Welch *t*-test. Differences were considered significant at  $P < 0.05$ . The statistical methods of this study were reviewed by Dr. Hisayo Morinaga from Biological/Pharmacological research laboratories, Central Pharmaceutical Research Institute, Japan Tobacco Inc.

## RESULTS

### Body weight and biochemical parameters

SDT fatty rats showed obesity at 8 wk of age. The mean bodyweight at 8 wk was significantly higher in SDT fatty rats compared with SD rats ( $328 \pm 40$  g vs  $216 \pm 8$  g;  $P < 0.01$ ). Obesity was sustained in SDT fatty rats until 24 wk of age (Figure 1A). The serum glucose levels in SDT fatty rats were higher than those in SD rats at 8 wk of age, and the increase was sustained until 40 wk of age (Figure 1B). The serum TG levels in SDT fatty rats increased until 24 wk of age, and they slightly decreased after 32 wk (Figure 1C). The serum TC levels in SDT fatty rats increased throughout the observation period (Figure 1D). The serum TG and TC levels in SDT fatty rats were higher than those in SD rats until 40 wk of age. Furthermore, the serum AST and ALT levels were significantly elevated at 8 wk, and these high levels were sustained until 40 wk of age (Figure 1E and F).

### Liver weight and hepatic TG and FFA content

The liver weights were increased in SDT fatty rats until 24 wk of age and slightly decreased thereafter, but they were greater than those in SD rats throughout the experiment, except at 24 wk of age (Figure 2A). The hepatic TG content in SDT fatty rats was higher than that in SD rats at 8 wk of age ( $129 \pm 77$  mg/g tissue vs  $10 \pm 2$  mg/g tissue;  $P < 0.01$ ). The hepatic TG content was markedly elevated until 16 wk of age in SDT fatty rats, and gradually decreased to the level in SD rats (Figure 2B). The hepatic FFA content showed a similar change to hepatic TG content in SDT fatty rats because it increased until 16 wk and gradually decreased after 32 wk of age (Figure 2C).

### Histopathological analyses

The liver histopathology was examined by HE staining. Histopathologically, severe changes (3+) of the liver, including fatty and vacuolar changes, were observed in SDT fatty rats at 8 wk of age. Moderate changes (1+–2+) of hypertrophy and very slight or slight changes in inflammation were also observed in SDT fatty rats at 8 wk of age (Table 1). Moreover, moderate changes (2+) indicating fibrosis were observed in SDT

rats at 32 wk of age.

Fibrosis was also confirmed by Sirius Red staining (Figure 3A–D). Consistent with the HE staining, similar findings of fibrosis were observed in SDT fatty rats. However, these changes were not observed in SD rats (Table 1). Activated hepatic stellate cells (HSCs) in the liver were investigated by  $\alpha$ -SMA immunostaining. The SD rats showed few HSCs at 32 wk of age. Consistent with the presence of hepatic fibrosis, there were increased numbers of  $\alpha$ -SMA-positive, activated HSCs in SDT fatty rats (Figure 3E and F). To examine the macrophages and mast cells in the liver, liver sections were immunostained with ED-1 and TB. Increased macrophage and mast cell counts, as indicated by ED-1- and TB-positive stained cells, respectively, were found in SDT fatty rats at 32 wk of age, and these cells were scarcely observed in SD rats (Figure 3G and H).

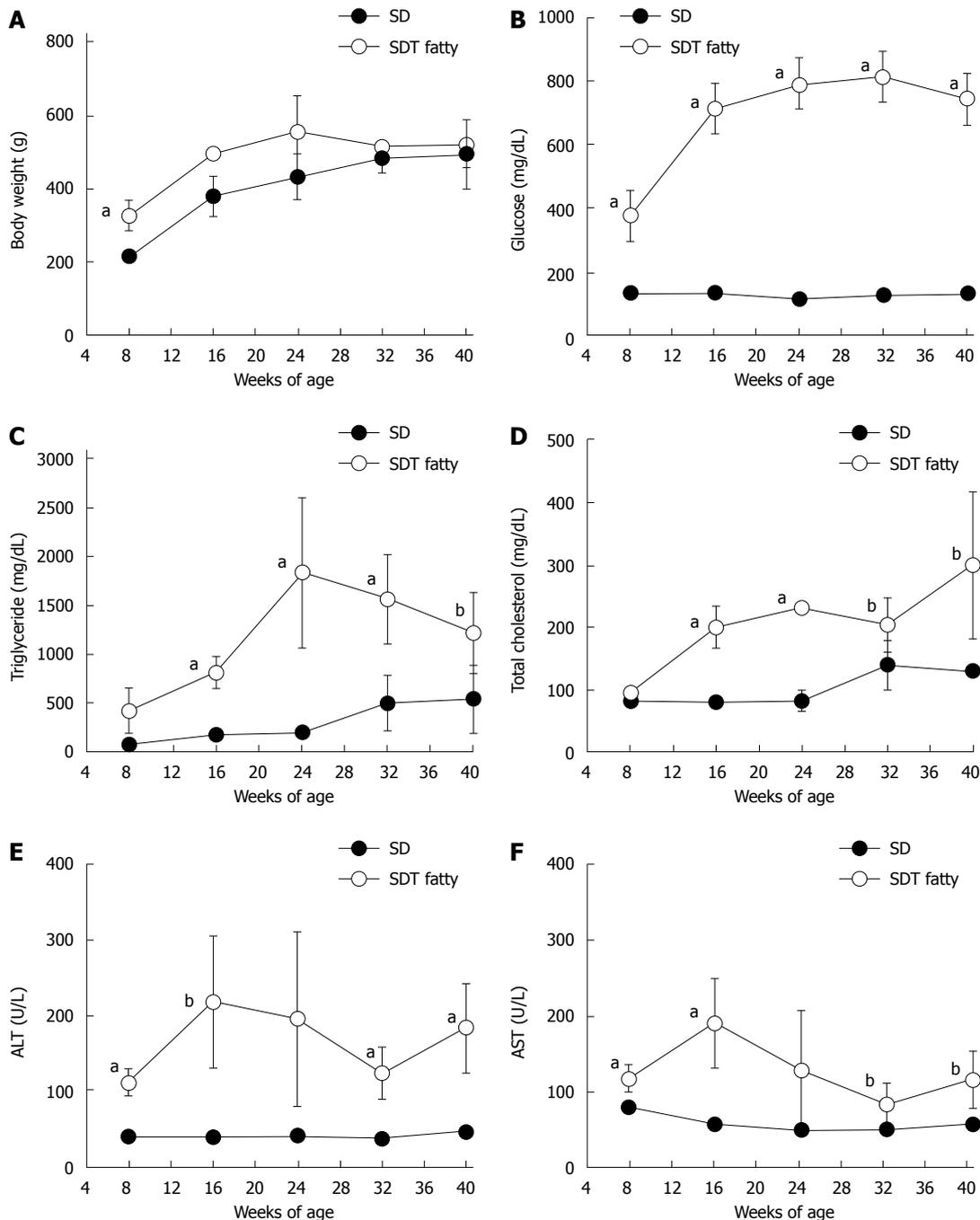
### Liver mRNA expression

The mRNA levels of lipogenic genes (*SREBP-1c*, *FAS*, *ACC*, *DGAT2*, and *MTP*), inflammatory genes (*MCP-1* and *TNF- $\alpha$* ), and fibrotic genes (*collagen Type 1*, *TGF- $\beta$* , and  *$\alpha$ -SMA*) in the liver were measured by real-time PCR at 8-wk intervals. At 8 wk of age, the expression of lipogenic genes (such as *SREBP-1c*, *FAS*, and *ACC*) increased in SDT fatty rats compared with SD rats (Figure 4A–C). The *DGAT2* mRNA level was not increased in SDT fatty rats (Figure 4D), and the *MTP* mRNA level, which is related to secretion of very low-density lipoprotein (VLDL), was decreased by 64% in SDT fatty rats compared with SD rats (Figure 4E). After 16 wk of age, SDT fatty rats showed decreased expression of lipogenic genes (such as *SREBP-1c*, *FAS*, and *ACC*). The expression of lipogenic genes was comparable between SDT fatty rats and SD rats, but the *DGAT2* and *MTP* mRNA levels in SDT fatty rats were lower than those in SD rats.

The *TNF- $\alpha$*  and *MCP-1* inflammatory genes were increased by 16.1% and 38.9%, respectively, in SDT fatty rats at 16 wk of age (Figure 4F and G). The mRNA levels of fibrotic genes, namely collagen type 1 and  $\alpha$ -SMA, tended to increase in SDT fatty rats compared with SD rats at 16 and 24 wk of age. The *TGF- $\beta$*  mRNA levels in SDT fatty rats also tended to increase at 32 wk of age.

## DISCUSSION

In a previous study, we investigated diabetes mellitus in female SDT fatty rats by comparing these rats with age-matched female SDT rats. Female SDT fatty rats showed metabolic abnormalities such as hyperglycemia, hyperinsulinemia, and dyslipidemia at a young age, which were similar to the abnormalities observed in male SDT fatty rats<sup>[14]</sup>. Furthermore, diabetic complications such as renal lesion and cataracts were observed at 16 wk of age in female SDT fatty rats, and were comparable to the complications



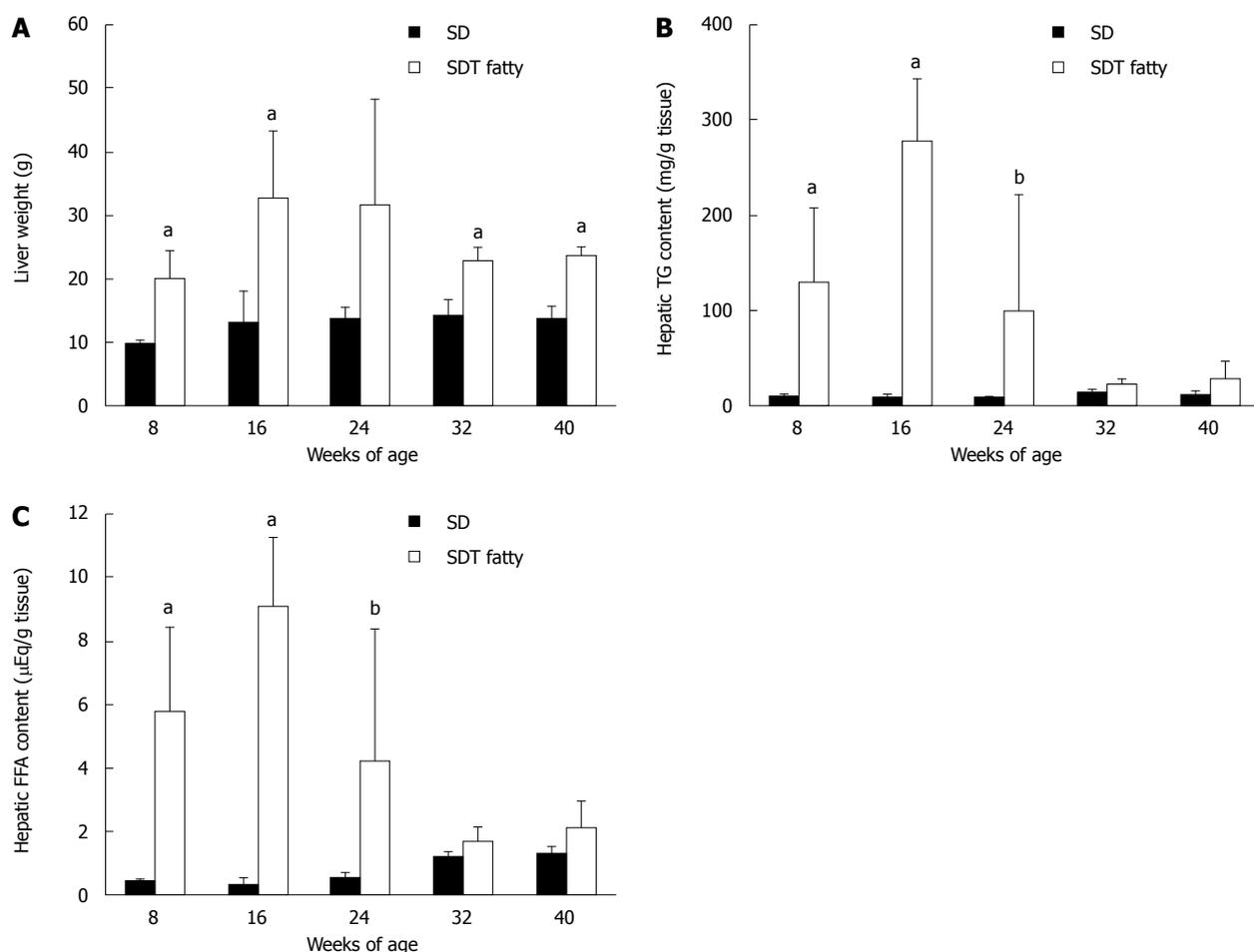
**Figure 1** Changes in body weight and biological parameters in spontaneously diabetic Torii fatty rats and Sprague-Dawley rats. A: Body weight; B: Glucose; C: Triglyceride; D: Total cholesterol; E: Alanine aminotransferase; F: Aspartate aminotransferase. Data are shown as mean  $\pm$  SD ( $n = 4-5$ ). <sup>a</sup> $P < 0.05$ , <sup>b</sup> $P < 0.01$ , vs Sprague-Dawley rats. SDT: spontaneously diabetic Torii; SD: Sprague-Dawley.

observed in male SDT fatty rats<sup>[15]</sup>. Sexual differences in SDT fatty rats did not exist in the progression of diabetes. Thus, female SDT fatty rats are an important animal model of metabolic syndrome with diabetes

In Zucker diabetic fatty rats, hepatic steatosis is observed without dietary manipulation, but hepatic fibrosis is not observed<sup>[19]</sup>. Although *ob/ob* and *db/db* mice develop hepatic steatosis associated with obesity and insulin resistance, these mice do not spontaneously progress to significant steatohepatitis<sup>[8,9]</sup>. *Ob/ob* and *db/db* mice show significant

steatohepatitis development when fed a MCD diet, but only *db/db* mice fed a MCD diet have increased fibrosis in the liver<sup>[10,11]</sup>. *Db/db* mice require other stimulation in addition to the background condition, such as obesity, diabetes, and dyslipidemia, to trigger progression to fibrosis<sup>[10]</sup>.

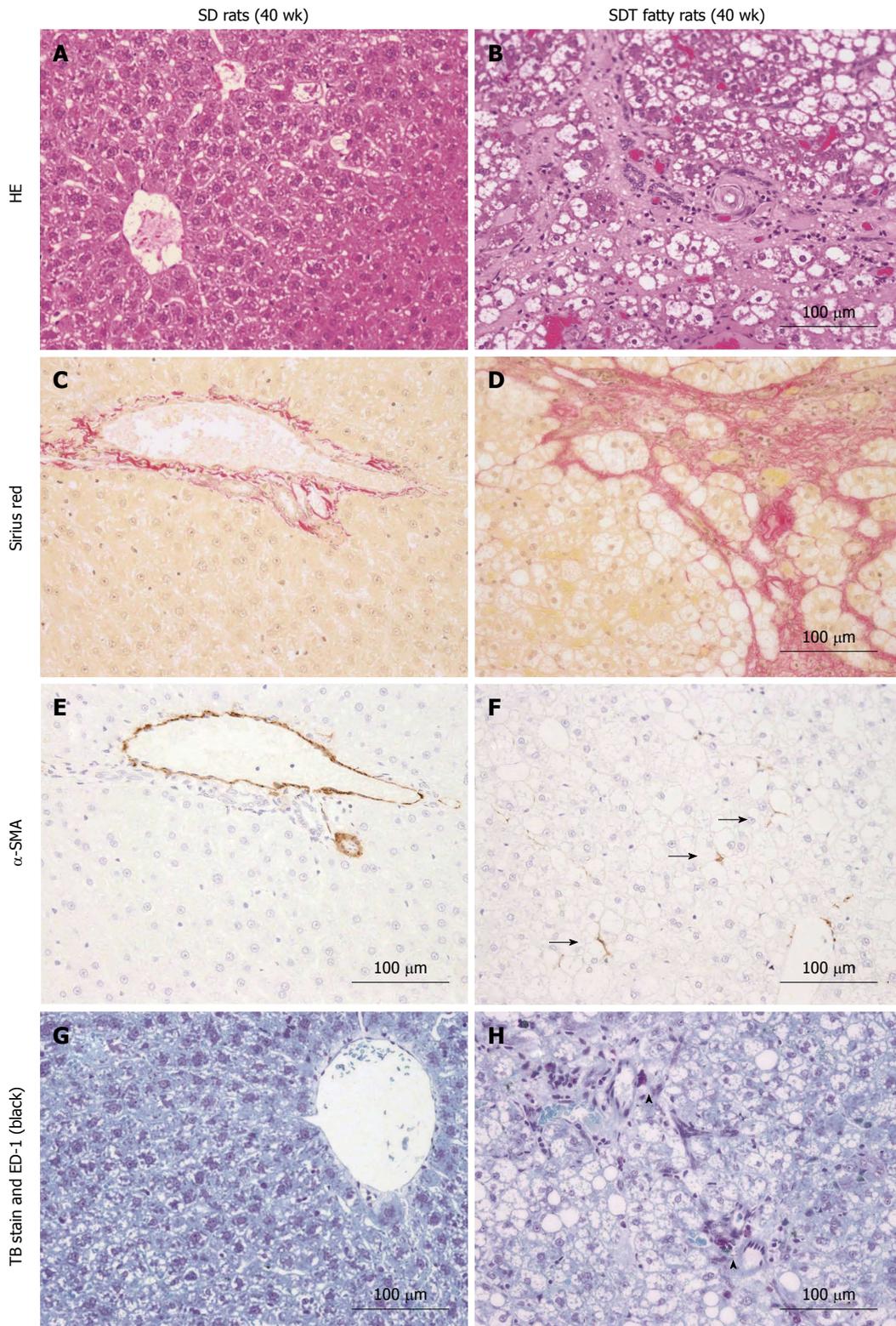
In the present study, we investigated the pathophysiological changes of the liver in female SDT fatty rats compared with SD rats. We used SD rats as the control, instead of SDT rats, because we wanted to focus on the investigation of pathophysiological



**Figure 2** Changes in liver weight, hepatic triglyceride, and fatty acid contents in Spontaneously Diabetic Torii fatty rats and Sprague-Dawley rats. A: Liver weight; B: Hepatic triglyceride content; C: Hepatic fatty acid content. Data are shown as the mean ± SD (n = 4-5). Data are shown as the mean ± SD (n = 4-5). <sup>a</sup>P < 0.05, <sup>b</sup>P < 0.01, vs SD rats. FFA: Free fatty acid; TG: Triglyceride.

**Table 1** Microscopy findings of the liver in female spontaneously diabetic Torii fatty rats and Sprague-Dawley rats

	8 wk		16 wk		24 wk		32 wk		40 wk	
	SD	SDT fatty	SD	SDT fatty	SD	SDT fatty	SD	SDT fatty	SD	SDT fatty
Hepatosteatosis (Vacuolar change/fatty change)										
-	5	0	5	0	5	0	5	0	5	0
±	0	0	0	0	0	0	0	0	0	0
+	0	0	0	0	0	0	0	0	0	0
2+	0	4	0	2	0	2	0	3	0	3
3+	0	1	0	3	0	2	0	2	0	2
Hypertrophy of hepatocytes (with vacuolation/fatty change)										
-	5	1	5	0	5	0	5	0	5	1
±	0	0	0	0	0	0	0	0	0	0
+	0	2	0	2	0	2	0	1	0	2
2+	0	2	0	3	0	2	0	3	0	1
3+	0	0	0	0	0	0	0	1	0	1
Fibrosis, focal										
-	5	4	5	5	5	4	5	1	5	1
±	0	0	0	0	0	0	0	0	0	0
+	0	1	0	0	0	0	0	1	0	2
2+	0	0	0	0	0	0	0	3	0	2
3+	0	0	0	0	0	0	0	0	0	0
Infiltration, inflammatory cell, periportal										
-	5	2	5	4	5	3	5	3	5	3
±	0	3	0	1	0	1	0	0	0	1
+	0	0	0	0	0	0	0	2	0	1
2+	0	0	0	0	0	0	0	0	0	0
3+	0	0	0	0	0	0	0	0	0	0

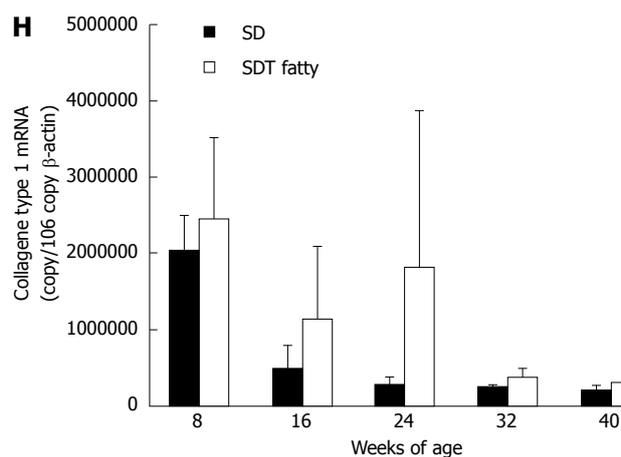
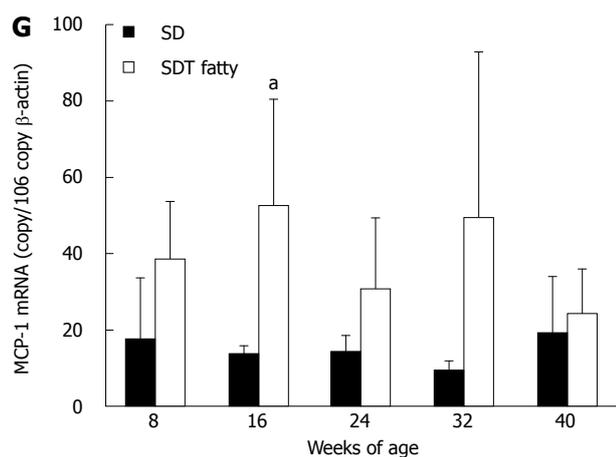
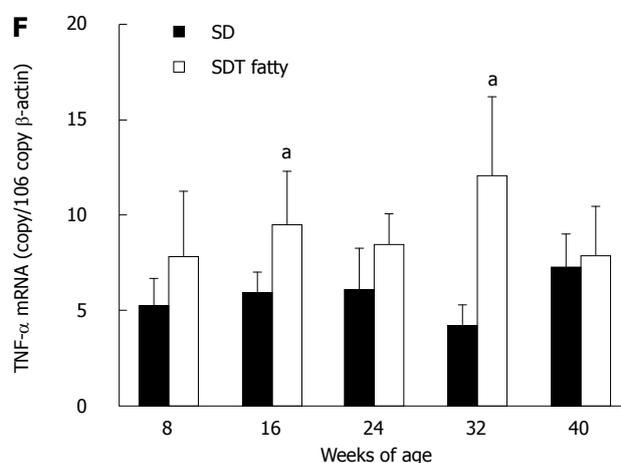
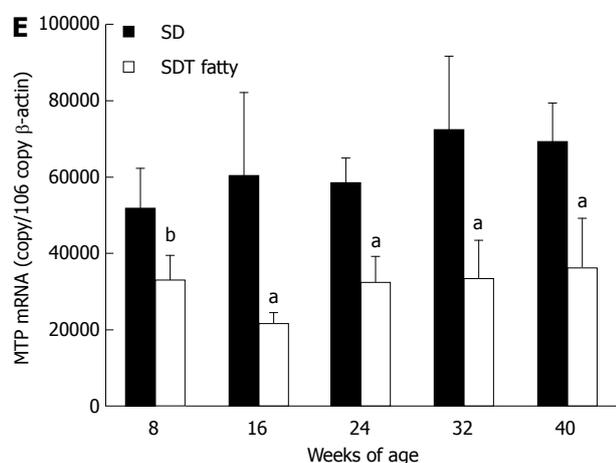
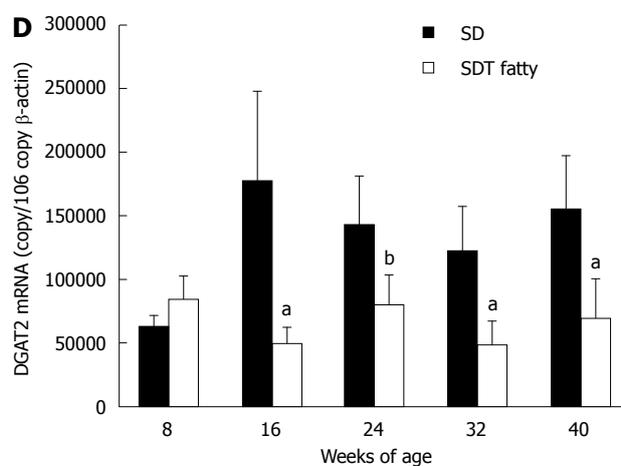
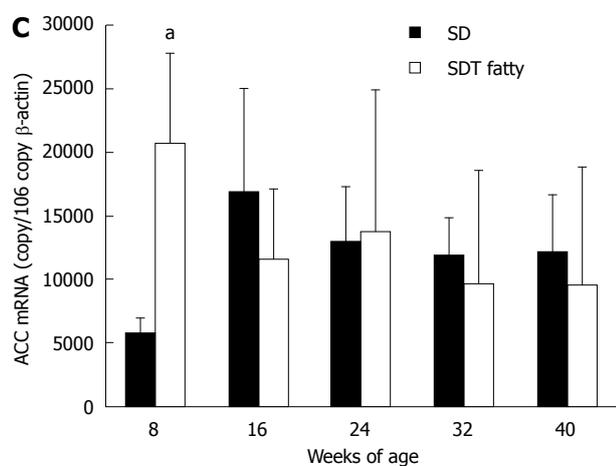
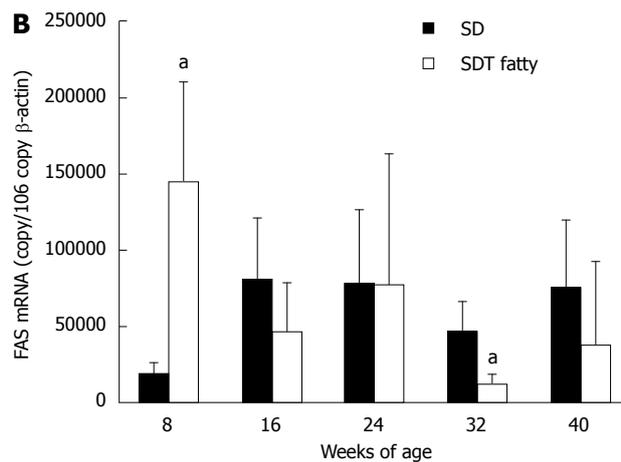
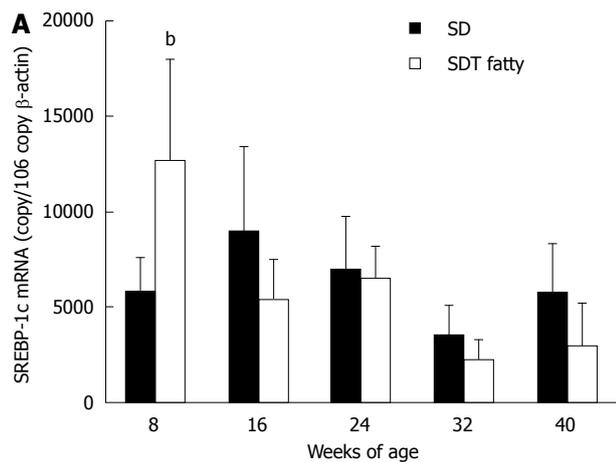


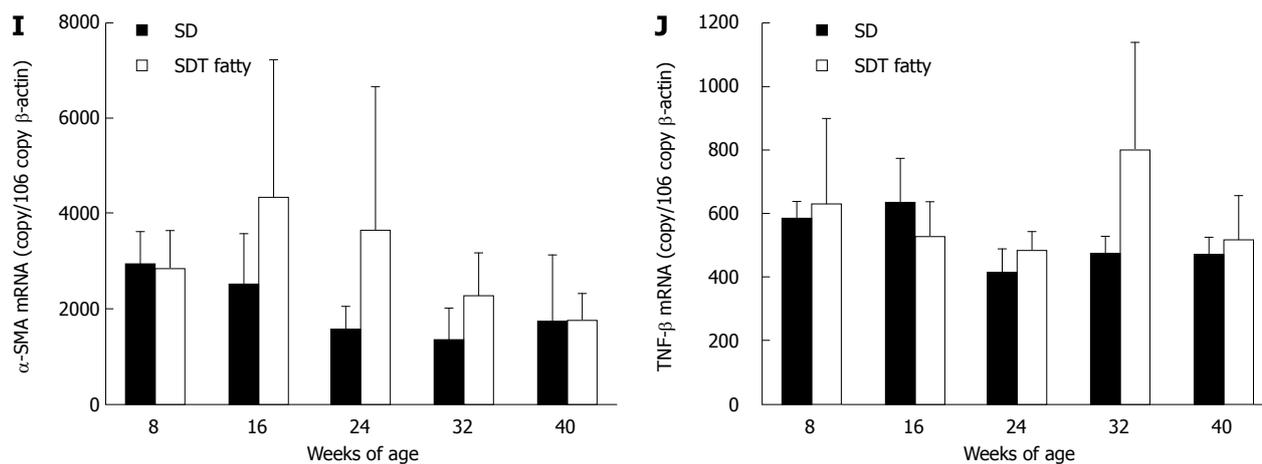
**Figure 3** Liver histopathology at 40 wk of age. A, C, E, G: SD rats; B, D, F, H: SDT fatty rats. A, B: Hematoxylin and eosin (HE); C, D: Sirius Red; E, F: Alpha-smooth muscle actin ( $\alpha$ -SMA); G, H: Toluidine blue staining and immunohistochemistry for ED-1.

changes of the liver in SDT fatty rats. In our previous study, we confirmed that female SDT rats only exhibit a fatty liver based on histopathology<sup>[16]</sup> but not fibrosis at 40 wk of age. Furthermore, male SDT rats, which are a severe type 2 diabetic model, do not exhibit a

fatty liver and liver fibrosis at 24 wk of age<sup>[20]</sup>.

SDT fatty rats showed hyperglycemia associated with increased body weight at 6 wk of age, and they exhibited severe hyperglycemia after 6 wk of age, which is considered to be due to a decrease in insulin





**Figure 4** Expression of genes related to triglyceride synthesis, triglyceride secretion, inflammation, and fibrosis in spontaneously diabetic Torii fatty rats and SD rats. A: SREBP-1c; B: FAS; C: ACC; D: DGAT2; E: MTP; F: TNF- $\alpha$ ; G: MCP-1; H: Collagen type 1; I:  $\alpha$ -SMA; J: TGF- $\beta$ . Data are shown as the mean  $\pm$  SD ( $n = 4-5$ ). <sup>a</sup> $P < 0.05$ , <sup>b</sup> $P < 0.01$ , vs SD rats. SDT: Spontaneously diabetic Torii; SD: Sprague-Dawley.

levels. Indeed, our previous study has shown that insulin levels in SDT fatty rats increase until 6 wk of age and gradually decrease thereafter, but the levels are significantly higher than those in SD rats until 20 wk of age ( $8.89 \pm 7.07$  ng/mL vs  $1.42 \pm 0.49$  ng/mL;  $P < 0.05$ ). From 20 to 40 wk of age, the insulin levels in SDT fatty rats are slightly greater than those in SD rats<sup>[14]</sup>. As expected, the hepatic TG content in SDT fatty rats was higher than those of SD rats, and it peaked at 16 wk of age. After 16 wk of age, it gradually decreased to the levels observed in the SD rats. The mRNA expression levels of MCP-1 and TNF- $\alpha$  at 16 wk of age were increased, suggesting that SDT fatty rats had accelerated liver injury and inflammation. The mRNA expression of collagen type 1 and  $\alpha$ -SMA tended to increase in SDT fatty rats. With regard to the pathophysiology, hepatic fibrosis was observed in SDT fatty rats at 32 wk of age. These findings indicated that female SDT fatty rats spontaneously develop nonalcoholic steatohepatitis-like hepatic lesions without dietary manipulation.

The hepatic steatosis in SDT fatty rats observed at 8 wk of age resulted from increased hepatic TG synthesis and decreased VLDL secretion, and histopathological findings, such as fatty changes in the hepatocytes, showed hepatic steatosis. Indeed, the hepatic FFA content, which is a component of TG, was also increased. The expression of genes related to lipogenesis, FAS, and ACC1 was increased, and the expression of MTP, which is a gene related to VLDL secretion, was decreased in SDT fatty rats at 8 wk of age. Furthermore, the FFA content in the liver was attributed to the degradation of serum lipoproteins and uptake of serum FFA. The hyperlipidemia in SDT fatty rats is considered to supply fatty acids to the liver.

In terms of the hepatic steatosis, SDT fatty rats are comparable to other rat strains at 8 wk of age<sup>[19,21]</sup>. In contrast, the decrease in the hepatic TG content after 24 wk of age may be based on the decreased

*de novo* lipogenesis. The mRNA expression of SREBP-1c, which regulates FAS and ACC1 mRNA expression, was decreased in SDT fatty rats after 16 wk of age. Some complex mechanisms involved in the decrease in SREBP-1c mRNA expression of liver should be considered. It is known that insulin activates the SREBP-1c promoter in liver, thereby increasing fatty acid and triglyceride synthesis<sup>[22]</sup>. A previous study has shown that the serum insulin levels are decreased in SDT fatty rats after 6 wk of age<sup>[14,16]</sup>. The reduction of SREBP-1c levels could contribute to the decrease in serum insulin levels. The reduction of hepatic steatosis has also been shown in Otsuka Long-Evans Tokushima fatty rats after 30 wk of age<sup>[21]</sup>. The change in hepatic steatosis is considered to be associated with a decrease in SREBP-1c expression<sup>[23]</sup>. The change in hepatic steatosis in SDT fatty rats may be partly due to decreased SREBP-1c expression based on the decreased serum insulin levels. Further study is required to confirm this mechanism.

The spontaneous development of steatohepatitis and hepatic fibrosis in SDT fatty rats could be explained by the increase in hepatic FFA. The excess FFA could serve as precursors for the production of lipid peroxidation<sup>[24]</sup>, which can activate hepatic stellate cells and thereby result in the development of fibrosis<sup>[25,26]</sup>. Indeed, an increase in  $\alpha$ -SMA level, which is an indicator of activated hepatic stellate cells, was observed. In addition to lipid peroxidation, CYP2E1 may be involved in hepatic fibrosis development in SDT fatty rats. It has been reported that fatty acids increase both CYP2E1 mRNA and protein levels in the liver of high-fat diet fed animals<sup>[27]</sup>. CYP2E1 produces high levels of reactive oxygen species<sup>[28,29]</sup>, which play an important role in the development of NASH and fibrosis<sup>[30]</sup>.

The difference in the pathophysiology of the liver between SDT fatty rats and other diabetic animal models remains unknown. However, the hepatic

vulnerability in SDT fatty rats could be explained by the sensitivity to inflammation. SDT rats, which are the strain of origin of SDT fatty rats, show hyperglycemia caused by age-dependent degenerative changes in pancreatic islets<sup>[31,32]</sup>. Treatment with telmisartan ameliorates hyperglycemia and hypoinsulinemia due to the reduction in oxidative stress caused by renin angiotensin system activation in SDT rats<sup>[33]</sup>, suggesting that a large amount of NO is produced by macrophages in the islets<sup>[34]</sup>. It has been suggested that SDT rats are under inflammatory conditions physiologically. In SDT fatty rats, ED-1 staining, which is a marker for activated macrophages, was increased in the liver at 40 wk of age. These results suggested that SDT fatty rats have increased inflammation as a steady condition in the liver. This state may enhance the stress sensitivity caused by lipid peroxidation, which induces hepatic fibrosis. Further study is needed to clarify the hepatic characteristics of SDT fatty rats. In addition, it is necessary to investigate whether SDT fatty rats develop hepatocarcinoma after 40 wk of age.

Compared with human NASH, SDT fatty rats have steatohepatitis accompanied by metabolic syndrome, including hyperglycemia, hyperinsulinemia, and obesity, which are similar features to those of human NASH. Human NASH is strongly associated with obesity, type 2 diabetes, and dyslipidemia<sup>[35,36]</sup>. In contrast, there is a difference between SDT fatty rats and human NASH. Pathophysiological features of NASH with fibrosis are observed in only female SDT fatty rats and not in male SDT fatty rats. Meanwhile, the prevalence of human NASH is common in men rather than women, which might be explained by sex hormones<sup>[37]</sup>. Further studies are needed to examine the molecular basis of this sex difference in SDT fatty rats.

In conclusion, female SDT fatty rats showed steatohepatitis (*e.g.*, increased plasma ALT and hepatic steatosis) associated with hyperglycemia and dyslipidemia at 8 wk of age. Thereafter, hepatic fibrosis began to be observed at 32 wk of age. These results demonstrated that female SDT fatty rats exhibit pathophysiological features of NASH in the absence of dietary manipulation. The precise mechanisms for developing NASH in humans remain unknown. However, female SDT fatty rats have the potential to become an important animal model of NASH with type 2 diabetes and obesity, a condition for which few models currently exist.

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## COMMENTS

### Background

Obesity and type 2 diabetes are well established risk factors for many chronic

disorders, such as non-alcoholic steatohepatitis (NASH). Approximately 5%-10% of NASH patients are diagnosed with cirrhosis and/or hepatocellular carcinoma within 10 to 20 years. Currently, no medicines are approved for the treatment of NASH. To understand the complicated features of the disease, animal models of non-alcoholic fatty liver disease (NAFLD) offer important information for therapeutic intervention. Currently, there are no animal models of NASH accompanied by essential features of clinical conditions, such as obesity and insulin resistance, without dietary manipulation.

### Research frontiers

Spontaneously diabetic Torii *Lep<sup>fa</sup>* (SDT fatty) rats show several metabolic syndrome features, including obesity, hyperglycemia, hyperlipidemia, and insulin resistance. However, the features of liver lesions in SDT fatty rats have not been reported in detail. In the present study, the pathophysiological changes in the liver of SDT fatty rats were examined over time.

### Innovations and breakthroughs

Many NASH studies have used animal models, such as diet-induced NASH models or mutant gene models, and these NASH animal models spontaneously progress to significant steatohepatitis accompanied by metabolic syndrome. This is the first study to show the time course of pathophysiological changes in the liver of SDT fatty rats. Furthermore, the *in vivo* studies suggest that female SDT fatty rats exhibit pathophysiological features of NASH in the absence of dietary manipulation.

### Applications

Female SDT fatty rats may be useful to identify the complicated molecular mechanisms of the disease. In addition, this model may be a suitable preclinical model to evaluate candidate drugs for NASH.

### Terminology

NAFLD is presently well recognized as the most common chronic liver disease in the Western world. NAFLD is a broad-spectrum liver disease that ranges from simple steatosis to NASH and cirrhosis. Approximately 20%-30% of NAFLD patients progress to NASH. Because NASH leads to serious diseases, such as cirrhosis of the liver and hepatocarcinoma, the development of therapeutic agents is desirable. The pathogenesis of NASH remains poorly defined, and effective pharmacological therapies have not been approved.

### Peer-review

This is a good study in which the authors examined the pathophysiological changes in the liver of SDT fatty rats over time. The results are interesting and suggest that female SDT fatty rats spontaneously exhibit pathophysiological features of NASH. Female SDT fatty rats have the potential to become a suitable preclinical model of NASH.

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