

Choroidal and Retinal Atrophy of Bietti Crystalline Dystrophy Patients
with CYP4V2 Mutations Compared to Retinitis Pigmentosa Patients with
EYS Mutations

Manabu Miyata, MD, PhD, Masayuki Hata, MD, Sotaro Ooto, MD, PhD, Ken Ogino,
MD, PhD, Norimoto Gotoh, MD, PhD, Satoshi Morooka, MD, PhD, Tomoko
Hasegawa, MD, Takako Hirashima, MD, Masako Sugahara, MD, Yoshimasa Kuroda,
MD, Kenji Yamashiro, MD, PhD, and Nagahisa Yoshimura, MD, PhD

Department of Ophthalmology and Visual Sciences, Kyoto University Graduate
School of Medicine, Kyoto, Japan

Corresponding author:

Manabu Miyata, MD, PhD
Department of Ophthalmology and Visual Sciences, Kyoto University Graduate
School of Medicine
Shogoin Kawahara-cho 54, Sakyo-ku, Kyoto 606-8507, Japan
TEL: +81-75-751-3248
FAX: +81-75-752-0933
Email: miyatam@kuhp.kyoto-u.ac.jp

Short title:

Choroidal and retinal atrophy of BCD

Conflict of Interest:

None declared.

Funding/Support:

This work was supported in part by the Innovative Techno-Hub for Integrated Medical
Bio-Imaging of the Project for Developing Innovation Systems, from the Ministry of
Education, Culture, Sports, Science and Technology (MEXT), Japan.

Financial Disclosures:

NY: Topcon Corporation (Financial Support), Nidek (Financial Support, Consultant),
Canon (Financial Support).

KEY WORDS

Bietti crystalline dystrophy; choroid; CYP4V2; EYS; optical coherence tomography; retinitis pigmentosa

BRIEF SUMMARY STATEMENT

We found severe atrophy in all layers of the retina, retinal pigment epithelium/Bruch's membrane, and choroid in the macular region in Bietti crystalline dystrophy patients with *CYP4V2* mutations, while only the retina was severely atrophic in retinitis pigmentosa patients with *EYS* mutations.

ABSTRACT

Purpose

To compare atrophy of the choroid and retina between Bietti crystalline dystrophy (BCD) patients and *EYS*-related retinitis pigmentosa (RP) patients with a similar degree of central visual field defects, age, and axial length (AL).

Methods

Nine eyes of nine BCD patients with *CYP4V2* mutations (BCD group) were examined. Moreover, we selected ten eyes of ten RP patients with *EYS* mutations matched for age, AL, and mean deviation (measured with the 10-2 SITA standard program; *EYS*-RP group), and ten eyes of ten normal volunteers matched for age and AL (control group). Macular thicknesses of the choroid and retina were measured via swept-source optical coherence tomography.

Results

The macular choroid was significantly thinner in the BCD group than in the *EYS*-RP and control groups, although the thickness did not significantly differ between the *EYS*-RP and control groups. The macular retina was significantly thinner in the BCD and *EYS*-RP groups than in the control group, although the thickness did not significantly differ between the BCD and *EYS*-RP groups at most sites.

Conclusion

BCD patients with *CYP4V2* mutations showed more severe macular choroid atrophy as compared to *EYS*-related RP patients. These different damage patterns suggest differences in choroidal expression between *CYP4V2* and *EYS*.

INTRODUCTION

1 Bietti crystalline dystrophy (BCD) is a rare retinal dystrophy characterized by the
2 presence of yellow and shiny deposits on the cornea and posterior pole of the retina,
3 along with progressive atrophy of the retina, choriocapillaris, and choroid.¹ A
4 long-term follow-up study showed that the formation of retinal crystals is followed by
5 gradual atrophy of the retinal pigment epithelium (RPE) and constriction of the visual
6 field.² BCD is caused by mutations of the *CYP4V2*.³ The phenotype of
7 *CYP4V2*-related BCD is highly variable⁴ and the mechanisms underlying the
8 pathological changes in BCD remain unclear because of the rarity of the disease;
9 therefore, no efficacious treatment is currently available.

10 The symptoms of BCD, including night blindness and progressive constriction
11 of the visual field, are similar to those of retinitis pigmentosa (RP). However, gene
12 expression patterns of *CYP4V2* and the causative genes for RP are reportedly
13 different. In particular, *CYP4V2* is expressed in the retina and RPE in the eyes.³
14 Although the RPE is generally thought to be the site of the primary pathological
15 abnormality in BCD patients with *CYP4V2* mutations, Nakano et al. showed weak
16 staining of ganglion cells and internal/external nuclear layers in the retina by
17 immunohistochemical staining.⁵ Their figure also suggested *CYP4V2* expression in
18 the choroid. In contrast, the human gene *EYS*, which is the most frequently mutated
19 gene in Japanese RP patients,^{6, 7} encodes a protein that is localized in the outer
20 segments of photoreceptors.⁸ Thus, the difference in expression patterns of the
21 causative genes is responsible for the degree and the timing of the damage to each
22 neuroretinal and choroidal layer.

23 Although the degenerative changes in RP mainly occur in photoreceptor cells,
24 Dhoot et al. reported that the macular choroidal thickness was significantly decreased
25 in patients with RP.⁹ The precise mechanism of choroidal thinning in RP is unclear.
26 However, several studies have shown that the death of RPE cells secondary to the
27 death of photoreceptor cells is implicated in choriocapillary atrophy.^{10, 11} In fact,
28 vascular endothelial growth factor (VEGF) is produced by the RPE and is necessary
29 for choroidal maintenance¹²; therefore, a lack of VEGF due to an RPE disorder may
30 play a role in choroidal thinning. Recently, Saatoci et al. demonstrated the thinning of
31 the retina and choroid in BCD.¹³ Considering the differences in the sites of expression
32 of the causative gene between BCD and *EYS*-related RP, we believe that the degree
33 and timing of the damage to each neuroretinal layer and choroid layer may differ
34 between these two diseases. In the present study, we compared the presence of
35 macular atrophy of the retina, RPE/Bruch's membrane (BM), and choroid between
36 BCD patients with *CYP4V2* mutations and *EYS*-RP patients at a similar disease
37 stage.

METHODS

This study was approved by the ethics committee of Kyoto University, Graduate School of Medicine (Kyoto, Japan) and adhered to the tenets of the Declaration of Helsinki. The nature of the study and the possible consequences of participation were explained to all candidates, and written informed consent was obtained from all participants.

Subjects

Consecutive patients with retinal degenerative disease or retinal dystrophy who visited the Department of Ophthalmology and Visual Sciences at Kyoto University Graduate School of Medicine (Kyoto, Japan) between January 2011 and October 2015 were recruited to participate in the study. All patients provided blood samples for the detection of gene mutations. All participants also underwent a comprehensive ophthalmologic examination, including measurement of best-corrected visual acuity (BCVA) with a Landolt C chart and axial length (AL) with an IOL Master (Carl Zeiss Meditec). All BCVA data were converted to the logarithm of the minimal angle of resolution (logMAR) for statistical analysis. Slit-lamp biomicroscopy, indirect ophthalmoscopy, color fundus photography (TRC-NW8F; Topcon Corp, Tokyo, Japan), fundus autofluorescence (FAF) with an Optos device (Optos PLC, Scotland, United Kingdom), spectral domain optical coherence tomography (SD-OCT, Spectralis; Heidelberg Engineering, Dossenheim, Germany), swept source optical coherence tomography (SS-OCT, Topcon Corp., Tokyo, Japan), mean deviation (MD) measurements with a Humphrey field analyzer (HFA; Carl Zeiss Meditec, Dublin, CA) with the 10-2 Swedish Interactive Threshold Algorithm standard program for the evaluation of macular sensitivity, and 30-Hz flicker electroretinography (ERG) were also performed. ERG results were recorded according to the International Society for Clinical Electrophysiology of Vision standard protocol recommended in 2008 with the LS-C (Mayo Co., Nagoya, Japan) and Neuropack MEB-2204 systems (Nihon Kohden, Tokyo, Japan).¹⁴ With regard to inclusion criteria, we enrolled BCD patients with *CYP4V2* mutations and with clear SD-OCT and SS-OCT images available. A clinical diagnosis of BCD was made based on the presence of characteristic fundus crystalline deposits, retino-choroidal dystrophy, and patchy RPE atrophy, whereas *CYP4V2* mutations were detected via Sanger sequencing. The exclusion criteria were as follows: missing necessary data and the presence of other eye diseases except for refractive errors, cataract, and pseudophakia. When both eyes in one patient met the criteria, one eye was randomly selected for analysis. We also examined Japanese RP patients with *EYS* mutations detected by next-generation

sequencing⁶ who were selected to be matched for age, AL, and MD among the consecutive patients, as well as Japanese volunteers with no eye disease except for refractive errors who were matched for age and AL. The BCVA of the volunteers was 20/20 or better.

SS-OCT measurement of choroidal and retinal thicknesses

For choroidal and retinal thickness measurements, we acquired horizontal and vertical B-scan images through the fovea with SS-OCT. The light source was a wavelength-sweeping laser with a tuning range of approximately 100 nm centered at 1050 nm, which yielded an axial resolution in tissue of 8 μ m. Trained examiners performed the SS-OCT examinations after pupil dilation. They achieved pupil centration during the scan by using an internal fixation target, which was confirmed through a built-in camera within the SS-OCT system. Choroidal and retinal thicknesses were measured with a built-in caliper tool. The outermost highly reflective retinal band comprised the RPE and BM.¹⁵ Retinal thickness was defined as the distance between the vitreoretinal interface and the outer border of the RPE/BM complex. Choroidal thickness was defined as the distance between the outer border of the RPE/BM complex and the choriocleral interface (Figure 1). Each thickness was manually measured at the subfovea and at 0.5, 1.0, and 1.5 mm from the center of the fovea in the superior, temporal, inferior, and nasal quadrants (a total of thirteen points) with built-in caliper tool software and AL adjustment system. Central retinal thickness and subfoveal choroidal thickness measurements were made on horizontal and vertical scans, and the mean of the two measurements was used as a representative value.

SD-OCT measurement of the RPE/BM thickness

By using a previously described method,¹⁶ we measured the thickness of the RPE/BM complex on horizontal and vertical B-scan images through the fovea, obtained with SD-OCT, as the axial resolution of SS-OCT (8 μ m) is higher than that of SD-OCT (4–6 μ m), which is adequate to measure a thin RPE/BM complex.¹⁷ At each location of interest on the retina, we acquired 100 SD-OCT images and averaged them to reduce speckle noise. Thereafter, we manually measured the thickness of the RPE/BM complex at the same subfoveal and macular sites described above with the built-in caliper tool software and AL adjustment system. The representative subfoveal value was calculated as the mean of the horizontal and vertical measurements.

Statistical analysis

Data are presented as the mean \pm standard deviation where applicable. One-way

analysis of variance was performed to compare the three groups. Corrections for multiple comparisons were made with a Tukey test. Comparisons of differences between the two groups, including age, AL, and MD matching, were performed using *t*-tests. We analyzed the correlations of the thickness values of the choroid, RPE/BM complex, and retina with age by using Pearson's rank correlation coefficient. The above statistical analyses were performed using SPSS version 21 (IBM, New York, NJ, USA). The cut-off value for subfoveal choroidal thickness, the best sensitivity–specificity balance, and the area under the receiver operating characteristic curve (AUROC) were calculated using MedCalc version 12 (MedCalc Software, Ostend, Belgium). A *P*-value of < .05 was considered statistically significant.

RESULTS

A total of nine eyes of nine Japanese BCD patients with *CYP4V2* mutations (five women and four men; BCD group); 10 eyes of 10 Japanese RP patients with *EYS* mutations who were matched for age, AL, and MD (four women and six men; *EYS*-RP group); and 10 healthy eyes of 10 Japanese volunteers (seven women and three men; control group) were included in this study. The characteristics of the study population are shown in Table 1 and clinical characteristics of the BCD group and *EYS*-RP group are shown in Table 2. The mean ages of the BCD group, *EYS*-RP group, and control group were not significantly different (BCD group vs *EYS*-RP group, *P* = .31; BCD group vs control group, *P* = .36; and *EYS*-RP group vs control group, *P* = .82). The mean AL was not significantly different between the groups (BCD group vs *EYS*-RP group, *P* = .92; BCD group vs control group, *P* = .80; and *EYS*-RP group vs control group, *P* = .93). The mean MD was not significantly different between the BCD group and *EYS*-RP group (*P* = .17).

Choroidal thickness

The mean subfoveal choroidal thicknesses in the BCD, *EYS*-RP, and control groups were 109.6 ± 30.3 , 251.8 ± 55.2 , and 285.7 ± 91.7 μm , respectively (Table 3). The macular choroid in the BCD group was significantly thinner than that in the control group at all sites and that in the *EYS*-RP group at all sites, except for the nasal site, 1.5 mm from the fovea. In contrast, there were no significant differences in the macular thickness of the choroid between the *EYS*-RP group and control group at all sites.

RPE/BM complex thickness

The mean subfoveal RPE/BM complex thicknesses in the BCD, *EYS*-RP, and control groups were 11.3 ± 1.6 , 17.9 ± 3.6 , and 23.5 ± 2.0 μm , respectively (Table 4). There

1 were significant differences in complex thickness among the three groups at all
2 thirteen sites. The RPE/BM complex was thinner in the BCD group than in the
3 *EYS*-RP group and thinner in *EYS*-RP than in the control group.
4
5

6 **Retinal thickness**

7 The mean central retinal thicknesses in the BCD, *EYS*-RP, and control groups were
8 117.9 ± 52.6 , 180.3 ± 44.5 , and 228.7 ± 22.4 μm , respectively (Table 5). Compared to
9 the control group, the macular retina was significantly thinner in the BCD and *EYS*-RP
10 groups at all sites. In contrast, the retinal thickness in the macular area of the BCD
11 and *EYS*-RP groups was significantly different at only three sites.
12
13
14
15

16 **Choroidal thickness uniformity in the BCD and *EYS*-RP groups**

17 In the *EYS*-RP group, the choroid was significantly thinner at the nasal site, 1.5 mm
18 from the fovea, than at the subfovea ($P < .01$) (Figure 2), and the choroid was
19 significantly thinner nasal to the fovea than temporal to the fovea. However, no such
20 difference was observed in the BCD group ($P = .29$). The choroidal thickness at 1.5
21 mm temporal to the fovea was not significantly different compared to the subfoveal
22 choroidal thickness both in the BCD and *EYS*-RP groups ($P = .77$ and $P = .66$,
23 respectively).
24
25
26
27
28
29

30 **Differentiation of the BCD and *EYS*-RP groups by thickness analysis**

31 When the cut-off value for the subfoveal choroidal, subfoveal RPE/BM complex, and
32 central retinal thicknesses were set at 156.5, 12, and 166 μm , respectively, we could
33 distinguish between the BCD and *EYS*-RP groups with a sensitivity of 100% and
34 specificity of 90%, with a sensitivity of 89% and specificity of 100%, and with a
35 sensitivity of 89% and specificity of 70%, respectively. With regard to the AUROC, the
36 best sensitivity–specificity balance achieved was 0.978, 0.967, and 0.833,
37 respectively.
38
39
40
41
42
43
44
45

46 **Correlation between age and thickness**

47 There were no significant correlations between age and the thickness of the
48 subfoveal choroid, subfoveal RPE/BM complex, and central retina in the BCD group
49 (choroid, $P = .60$; RPE/BM, .66; and retina, .91, respectively) or in the *EYS* group
50 (choroid, $P = .51$; RPE/BM, .09; and retina, .34, respectively).
51
52
53
54
55

56 **Differentiation of homozygotes and heterozygotes in the BCD group**

57 The mutations detected in the study patients are listed in Table 2. Among the nine
58 BCD patients, four had homozygous mutations and five had compound heterozygous
59
60
61
62
63
64
65

1 mutations. All patients had at least one c.802-8_810del17inGC mutation which is
2 known to cause deletion of exon 7. There were no significant differences in age (57.5
3 ± 6.6 vs 58.2 ± 4.7 years), MD (-30.9 ± 2.7 vs -28.8 ± 4.9 dB), subfoveal choroidal
4 thickness (96.4 ± 19.6 vs 120.1 ± 35.1 μ m), subfoveal RPE/BM complex thickness
5 (10.5 ± 0.4 vs 12.0 ± 1.9 μ m), and central retinal thickness (103.9 ± 40.2 vs $129.2 \pm$
6 62.9 μ m) between the cases with homozygous and compound heterozygous
7 mutations ($P = .22, .09, .15, .37$, and $.20$, respectively). However, there was a
8 significant difference in AL between them (24.4 ± 1.0 vs 24.1 ± 0.5 mm, $P = .003$).
9

14 Novel mutations in *CYP4V2*

16 We found two novel mutations (c.710C>A and c.1378T>C) in *CYP4V2* by identifying
17 the pathogenicity of the variants. These two variants were predicted as damaging by
18 silico prediction programs (SIFT and Polyphen2), and were not found in the Japanese
19 exome database (Human Genetic Variation Database,
20 <http://www.genome.med.kyoto-u.ac.jp/SnpDB/> [in the public domain]).
21
22
23
24

26 DISCUSSION

27 This study clearly showed that the macular choroid in the eyes of BCD patients with
28 *CYP4V2* mutations was significantly thinner than that in the eyes of RP patients with
29 *EYS* mutations at a similar stage of visual field damage. The BCD group also had a
30 thinner macular RPE/BM thickness as compared to the *EYS*-RP group. In contrast,
31 the thickness of the macular retina did not markedly differ between the eyes of BCD
32 patients with *CYP4V2* mutations and the eyes of RP patients with *EYS* mutations,
33 although the macular retina was significantly thinner in both the BCD group and
34 *EYS*-RP group than in the control group. These findings suggest that pathological
35 atrophy occurs simultaneously in the retina, RPE/BM, and choroid in BCD patients
36 with *CYP4V2* mutations. In the patients with *EYS*-RP, pathological atrophy occurs
37 initially at the retina, followed by the RPE/BM and choroid. We matched participants
38 according to MD values in the BCD and *EYS*-RP groups, and according to age and
39 AL, as these parameters are associated with choroidal thickness in normal eyes.¹⁸
40
41
42
43
44
45
46
47

48 Li et al. reported that *CYP4V2* is expressed in the retina and RPE.³ Another
49 previous report also suggested expression of *CYP4V2* in the choroid in their
50 immunohistochemistry figure.⁵ In contrast, *EYS* is expressed in the outer segments of
51 photoreceptors.⁸ The expression of *CYP4V2* and *EYS* in the retina would cause
52 retinal atrophy in the BCD and *EYS*-RP groups, respectively, consistent with our
53 results; in particular, retinal thickness was decreased in the BCD and *EYS*-RP groups
54 as compared to the control group. The expression of *CYP4V2* in the RPE/BM and
55
56
57
58
59
60
61
62
63
64
65

choroid could have led to the severe damage of the RPE/BM and choroid observed in the BCD group as compared to the *EYS*-RP and control groups.

A previous study reported that macular choroidal thickness was significantly decreased in patients with RP⁹, inconsistent with our findings. The mean subfoveal choroidal thickness was $251.8 \pm 55.2 \mu\text{m}$ in the *EYS*-RP group in the present study and $245.6 \pm 103 \mu\text{m}$ in the previous study, whereas the thickness of the controls was $285.7 \pm 91.7 \mu\text{m}$ in the present study and $337.8 \pm 109 \mu\text{m}$ in the previous study. The differences in mean age (54.0 ± 11.2 versus 40.6 ± 12.9 years) and the possible difference in AL (mean spherical equivalent of -1.45 ± 2.61 diopters in the previous study and mean AL of 24.33 ± 1.26 mm in the present study) could explain these differences. The clinical findings of RP are variable because of the variability in the causative genes.¹⁹ This study focused on *EYS* mutations because *EYS* is a major causative gene of RP not only in Japanese,^{6, 7} but also in Spanish,²⁰ British and Chinese,²¹ and Israeli and Palestinian populations.²² At present, the choroidal thickness in cases of RP with *EYS* mutations has not been thoroughly investigated. Hence, further studies investigating the choroidal thickness in *EYS*-RP at various disease stages are warranted.

Although BCD can be classified into early, intermediate, and advanced stages,^{23, 24} the choroidal thickness would not differ according to the stage and the choroid would become atrophic during the early stage. In the present study, the enrolled patients were at the early and intermediate stages and the mean subfoveal choroidal thickness of the BCD group was $107.5 \pm 29.3 \mu\text{m}$. A previous study reported that the mean subfoveal choroidal thickness was $95.37 \pm 55.93 \mu\text{m}$ in intermediate- and late-stage BCD patients; however, the authors did not confirm the presence of *CYP4V2* mutations.¹³ These values for the subfoveal choroidal thickness are substantially less than those in the controls. Furthermore, in the present study, case 4 (early stage) had a subfoveal choroidal thickness of $75 \mu\text{m}$ and case 7 (intermediate stage) had a subfoveal choroidal thickness of $81 \mu\text{m}$. These values are similar to the mean subfoveal thickness values of patients in the previous study at the intermediate and late stage ($95.37 \mu\text{m}$). In the present study, the cut-off value for the subfoveal choroidal thickness was set at $156.5 \mu\text{m}$, which enabled the discrimination between BCD with *CYP4V2* mutations and RP with *EYS* mutations (sensitivity of 100% and specificity of 90%). Therefore, it is important to monitor choroidal thickness, as it can be used as an index for distinguishing BCD patients from *EYS*-RP patients even during the early stages.

Halford et al. reported that the phenotype was highly variable in BCD patients, and that deletions of exon 7 were associated with more severe disease.⁴ In the present study, all BCD patients carried at least one mutation that caused the deletion

of exon 7; however, there was no significant difference in subfoveal choroidal atrophy, subfoveal RPE/BM complex atrophy, and central retinal atrophy between homozygotes and compound heterozygotes. The severity of the atrophy of the choroid, RPE/BM complex, and retina was similar between the patients who carried the c.802-8_810del17inGC mutation in the homozygous state and in the compound heterozygous state. However, we could not compare the severity between patients with deletion of exon 7 and with other mutations in the present study.

The distribution of the choroidal thickness may also help in distinguishing between BCD and RP with *EYS* mutations. The choroid was thinner in regions nasal to the fovea compared to regions temporal to the fovea in RP with *EYS* mutations; however, in BCD patients with *CYP4V2* mutations, the choroidal thickness became uniformly atrophic. Hence, longitudinal studies on the progression of choroidal atrophy in these patients are needed to investigate the causes.

Retinal and RPE sheet transplantation is being suggested as a potential treatment method for degenerative diseases such as age-related macular degeneration and RP.^{25, 26} However, choroidal degeneration develops at the early stage of BCD and sheet transplantation of the choroid, along with the retina and RPE, would be necessary in those cases. Although retinal atrophy was confirmed in the BCD group, we recently reported that the cone photoreceptor cell density remained for visual dysfunction in BCD.²⁷ Hence, sheet transplantation of the choroid and RPE might be sufficient for resolving BCD. Gene therapy is another option for BCD. In fact, gene therapy for severe retinal dystrophy has previously been reported,²⁸⁻³⁰ and would be beneficial for patients with *CYP4V2* mutations at any stage of BCD.

This study has some limitations, including the cross-sectional design. The choroid, RPE/BM, and retina were significantly thinner even in the early-stage BCD patients than in the controls in this study. However, the choroid was not significantly thinner in the *EYS*-PR group than in the control group. In the *EYS*-RP group, retinal atrophy is the primary change observed, followed by atrophy of the photoreceptor cells and RPE/BM; finally, atrophy of the choroid occurs. A longitudinal study evaluating the choroidal, RPE/BM complex, and retinal thicknesses will aid in understanding the time course of structural changes in BCD patients with *CYP4V2* mutations and RP patients with *EYS* mutations. Moreover, the thicknesses were manually measured, which could have introduced errors. However, most previous studies have used manual measurements, and the reliability of these methods is generally accepted. Third, all BCD patients carried the c.802-8_810del17inGC mutation, which was reported to be associated with more severe phenotype, although we did not select the BCD patients. This mutation is most commonly found in Japanese BCD patients.³¹ Hence, further study should be performed in BCD patients

without this specific mutation.

In conclusion, BCD patients with *CYP4V2* mutations showed severe atrophy of the macular choroid, compared to the macular choroidal atrophy in RP patients with *EYS* mutations. These different patterns in damage suggested differences in the choroidal expression between *CYP4V2* and *EYS*.

REFERENCES

1. Bietti G. Ueber familiaeres vorkommen von "retinitis punctata albescens" (verbunden mit "dystrophia marginalis cristallinea corneae"), glitzern des glaskoerpers und anderen degenerativen augenveraenderungen. Klin Monbl Augenheilkd. 1937;99:21.
2. Mansour AM, Uwaydat SH, Chan CC. Long-term follow-up in Bietti crystalline dystrophy. Eur J Ophthalmol 2007;17:680-682.
3. Li A, Jiao X, Munier FL, et al. Bietti crystalline corneoretinal dystrophy is caused by mutations in the novel gene CYP4V2. Am J Hum Genet 2004;74:817-826.
4. Halford S, Liew G, Mackay DS, et al. Detailed phenotypic and genotypic characterization of bietti crystalline dystrophy. Ophthalmology 2014;121:1174-1184.
5. Nakano M, Kelly EJ, Wiek C, et al. CYP4V2 in Bietti's crystalline dystrophy: ocular localization, metabolism of ω -3-polyunsaturated fatty acids, and functional deficit of the p.H331P variant. Mol Pharmacol 2012;82:679-686.
6. Oishi M, Oishi A, Gotoh N, et al. Comprehensive molecular diagnosis of a large cohort of Japanese retinitis pigmentosa and Usher syndrome patients by next-generation sequencing. Invest Ophthalmol Vis Sci 2014;55:7369-7375.
7. Arai Y, Maeda A, Hirami Y, et al. Retinitis pigmentosa with EYS mutations is the most prevalent inherited retinal dystrophy in Japanese populations. J Ophthalmol 2015;2015.
8. Abd El-Aziz MM, Barragan I, O'Driscoll CA, et al. EYS, encoding an ortholog of Drosophila spacemaker, is mutated in autosomal recessive retinitis pigmentosa. Nat Genet 2008;40:1285-1287.
9. Dhoot DS, Huo S, Yuan A, et al. Evaluation of choroidal thickness in retinitis pigmentosa using enhanced depth imaging optical coherence tomography. Br J Ophthalmol 2013;97:66-69.
10. Neuhardt T, May CA, Wilsch C, et al. Morphological changes of retinal pigment epithelium and choroid in rd-mice. Exp Eye Res 1999;68:75-83.
11. Korte GE, Reppucci V, Henkind P. RPE destruction causes choriocapillary atrophy. Invest Ophthalmol Vis Sci 1984;25:1135-1145.
12. Saint-Geniez M, Kurihara T, Sekiyama E, et al. An essential role for RPE-derived soluble VEGF in the maintenance of the choriocapillaris. Proc Natl Acad Sci U S A 2009;106:18751-18756.
13. Saatci AO, Doruk HC, Yaman A, Oner FH. Spectral domain optical coherence tomographic findings of bietti crystalline dystrophy. J Ophthalmol 2014;2014:739271.
14. Marmor MF, Fulton AB, Holder GE, et al. ISCEV Standard for full-field clinical

- electroretinography (2008 update). *Doc Ophthalmol* 2009;118:69-77.
15. Drexler W, Sattmann H, Hermann B, et al. Enhanced visualization of macular pathology with the use of ultrahigh-resolution optical coherence tomography. *Arch Ophthalmol* 2003;121:695-706.
 16. Karampelas M, Sim DA, Keane PA, et al. Evaluation of retinal pigment epithelium-Bruch's membrane complex thickness in dry age-related macular degeneration using optical coherence tomography. *Br J Ophthalmol* 2013;97:1256-1261.
 17. Wolf-Schnurrbusch UE, Ceklic L, Brinkmann CK, et al. Macular thickness measurements in healthy eyes using six different optical coherence tomography instrumental. *Invest Ophthalmol Vis Sci* 2009;50:3432-3437.
 18. Ikuno Y, Kawaguchi K, Nouchi T, Yasuno Y. Choroidal thickness in healthy Japanese subjects. *Invest Ophthalmol Vis Sci* 2010;51:2173-2176.
 19. Daiger SP, Sullivan LS, Bowne SJ. Genes and mutations causing retinitis pigmentosa. *Clin Genet* 2013;84:132-141.
 20. Barragán I, Borrego S, Pieras JI, et al. Mutation spectrum of EYS in Spanish patients with autosomal recessive retinitis pigmentosa. *Hum Mutat* 2010;31:E1772-E1800.
 21. Abd El-Aziz MM, O'Driscoll CA, Kaye RS, et al. Identification of novel mutations in the ortholog of Drosophila eyes shut gene (EYS) causing autosomal recessive retinitis pigmentosa. *Invest Ophthalmol Vis Sci* 2010;51:4266-4272.
 22. Bandah-Rozenfeld D, Littink KW, Ben-Yosef T, et al. Novel null mutations in the EYS gene are a frequent cause of autosomal recessive retinitis pigmentosa in the Israeli population. *Invest Ophthalmol Vis Sci* 2010;51:4387-4394.
 23. Yuzawa M, Mae Y, Matsui M. Bietti's crystalline retinopathy. *Ophthalmic Paediatr Genet* 1986;7:9-20.
 24. Mataftsi A, Zografos L, Millá E, et al. Bietti's crystalline corneoretinal dystrophy: a cross-sectional study. *Retina* 2004;24:416-426.
 25. Bertolotti E, Neri A, Camparini M, et al. Stem cells as source for retinal pigment epithelium transplantation. *Prog Retin Eye Res* 2014;42:130-144.
 26. Seiler MJ, Aramant RB. Cell replacement and visual restoration by retinal sheet transplants. *Prog Retin Eye Res* 2012;31:661-687.
 27. Miyata M, Ooto S, Ogino K, et al. Evaluation of photoreceptors in Bietti crystalline dystrophy with CYP4V2 mutations using adaptive optics scanning laser ophthalmoscopy. *Am J Ophthalmol* 2016;161:196-205.
 28. Bainbridge JW, Smith AJ, Barker SS, et al. Effect of gene therapy on visual function in Leber's congenital amaurosis. *N Engl J Med* 2008;358:2231-2239.
 29. Vasireddy V, Mills JA, Gaddameedi R, et al. AAV-mediated gene therapy for

choroideremia: preclinical studies in personalized models. PLoS One
2013;8:e61396.

30. MacLaren RE, Groppe M, Barnard AR, et al. Retinal gene therapy in patients with
choroideremia: initial findings from a phase 1/2 clinical trial. Lancet
2014;383:1129-1137.
31. Yokoi Y, Nakazawa M, Mizukoshi S, et al. Crystal deposits on the lens capsules in
Bietti crystalline corneoretinal dystrophy associated with a mutation in the
CYP4V2 gene. Acta Ophthal 2010;88:607-609.

FIGURE LEGENDS

Figure 1. Vertical swept source optical coherence tomography (SS-OCT) images and Heidelberg retina angiography images of eyes in Bietti crystalline dystrophy patients with *CYP4V2* mutations, retinitis pigmentosa patients with *EYS* mutations, and control subjects.

Left Top. Representative 30° Heidelberg retina angiography (HRA) infrared images of an eye of a patient with Bietti crystalline dystrophy with *CYP4V2* mutations (Case 2) Sporadic crystalline deposits can be observed. The dots represent the measurement points at different distances from the fovea.

Red, subfovea; green, 0.5 mm; yellow, 1.0 mm; blue, 1.5 mm

Left Bottom. Representative 30° HRA autofluorescence images of a Bietti crystalline dystrophy patient with *CYP4V2* mutations (Case 2)

Patchy atrophy of the retinal pigment epithelium has spread.

Right Top. Representative swept source optical coherence tomography (SS-OCT) images of an eye of a Bietti crystalline dystrophy patient with *CYP4V2* mutations (Case 2)

The thicknesses of the subfoveal choroid, subfoveal retinal pigment epithelium-Bruch's membrane (RPE-BM) complex, and central retina are 94, 12, and 166 μm , respectively. The left side is superior and right-side is inferior.

The summits of the triangles represent the measurement points at different distances from the fovea.

Red, subfovea; green, 0.5 mm; yellow, 1.0 mm; blue, 1.5 mm

Right Middle. Representative SS-OCT images of an eye of a retinitis pigmentosa patient with *EYS* mutations (Case 9)

The thicknesses of the subfoveal choroid, subfoveal RPE-BM complex, and central retina are 273, 14, and 195 μm , respectively.

Right Bottom. Representative SS-OCT images of a normal eye (56 year-old man; axial length, 24.69 mm)

The thicknesses of the subfoveal choroid, subfoveal RPE-BM complex, and central retina are 363, 23, and 227 μm , respectively.

Figure 2. Horizontal and vertical choroidal thickness differences according to specific sites between Bietti crystalline dystrophy patients with *CYP4V2* mutations and retinitis pigmentosa patients with *EYS* mutations

BCD group = Bietti crystalline dystrophy with *CYP4V2* mutations group

EYS-RP group = retinitis pigmentosa with *EYS* mutations group

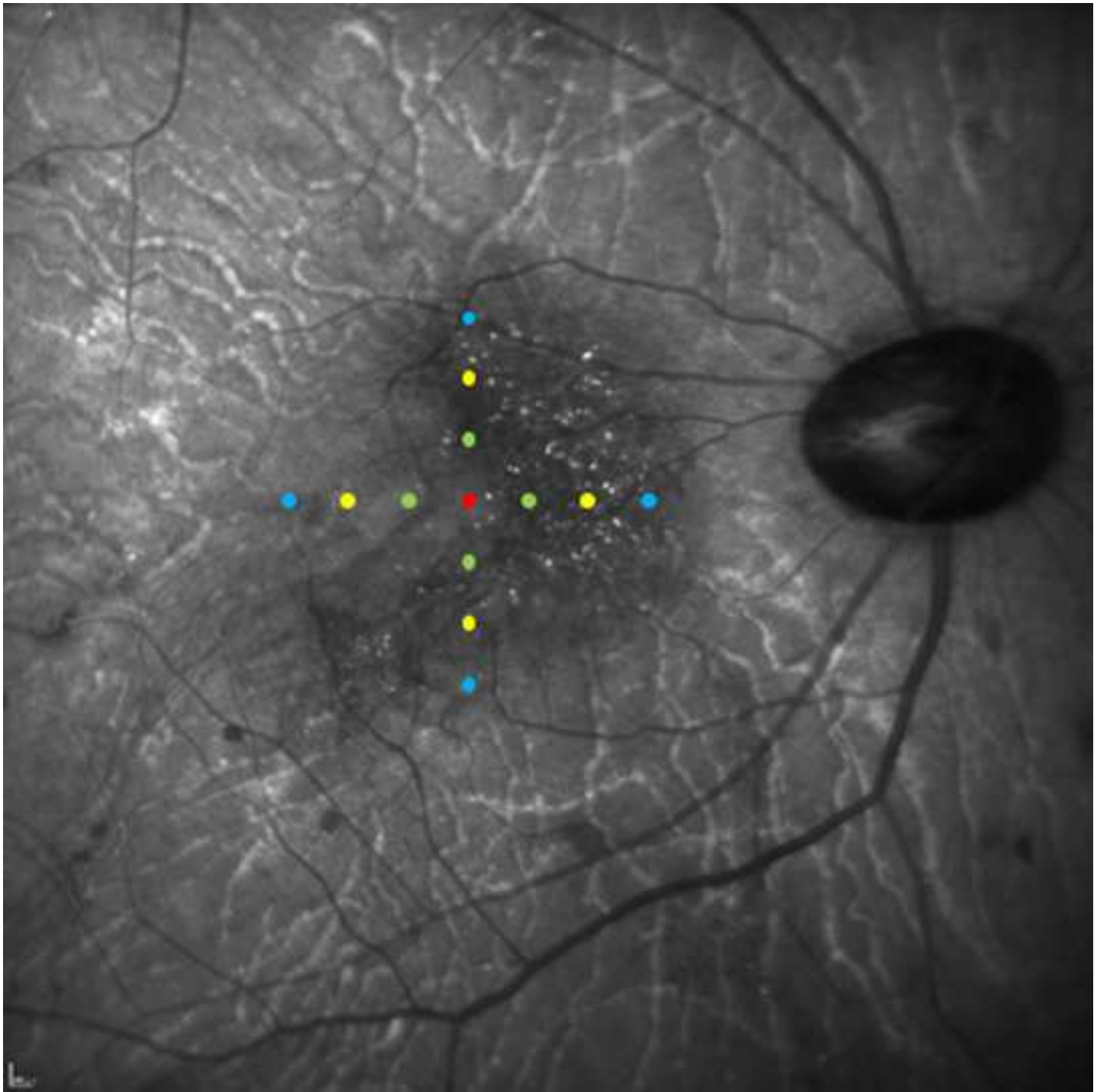
The bars indicate standard errors.

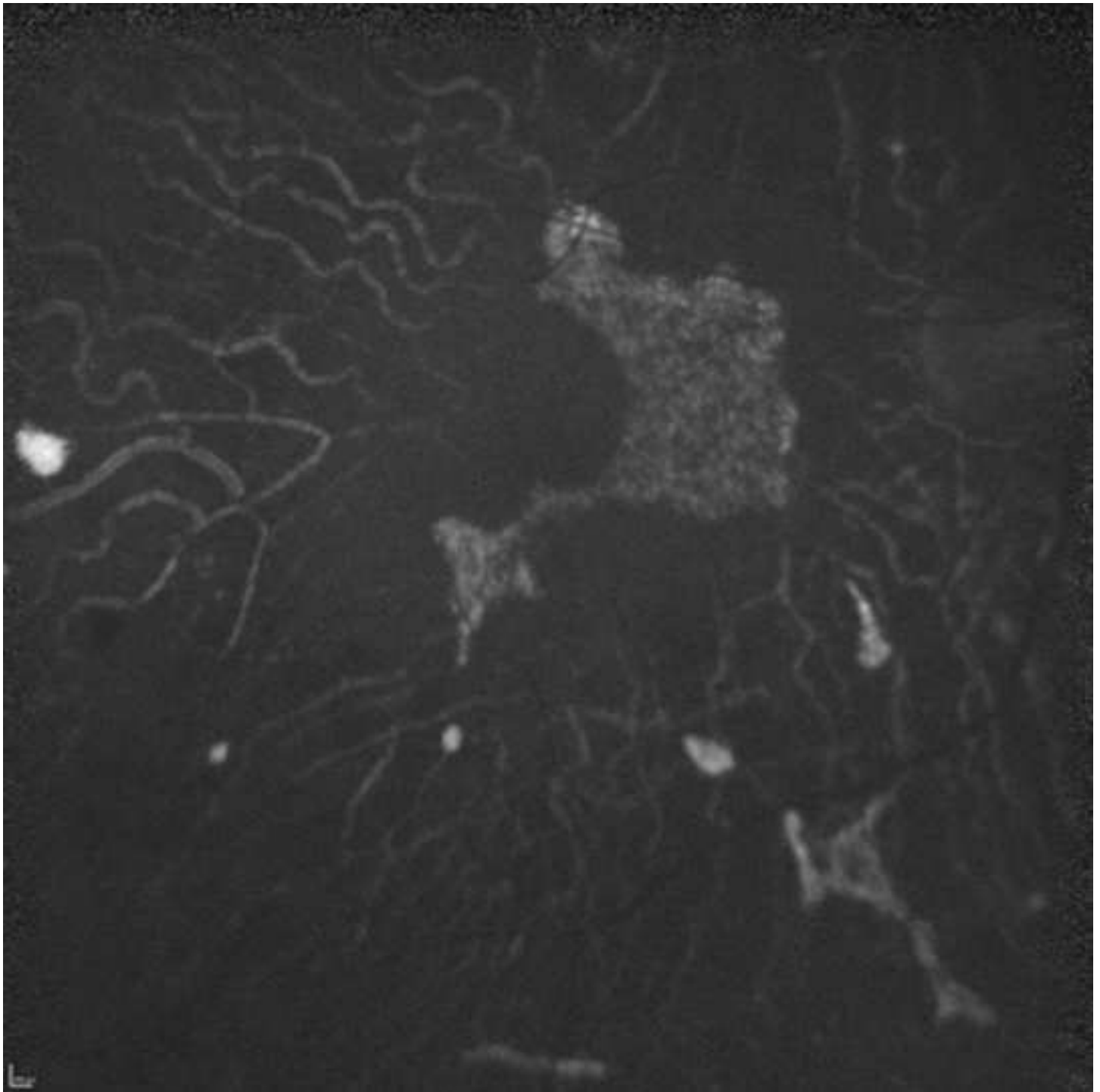
Top: Horizontal changes in the choroidal thickness

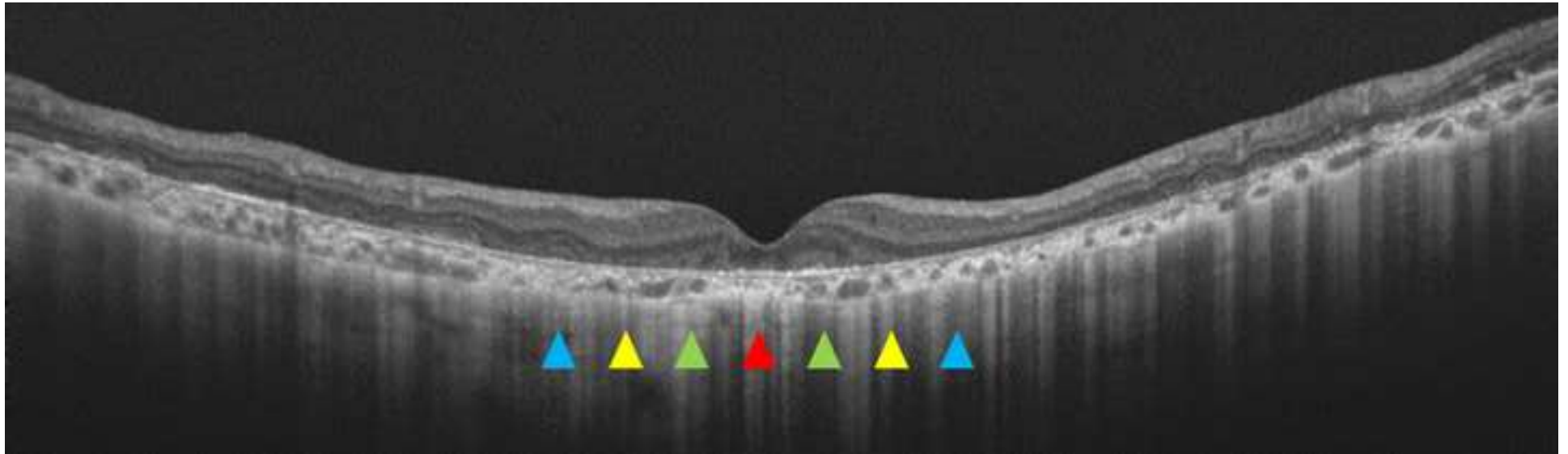
1 Although the choroidal thickness at each site is comparable in Bietti crystalline
2 dystrophy patients with *CYP4V2* mutations, the choroid in retinitis pigmentosa
3 patients with *EYS* mutations is thinner in regions nasal to the fovea than in regions
4 temporal to the fovea.
5

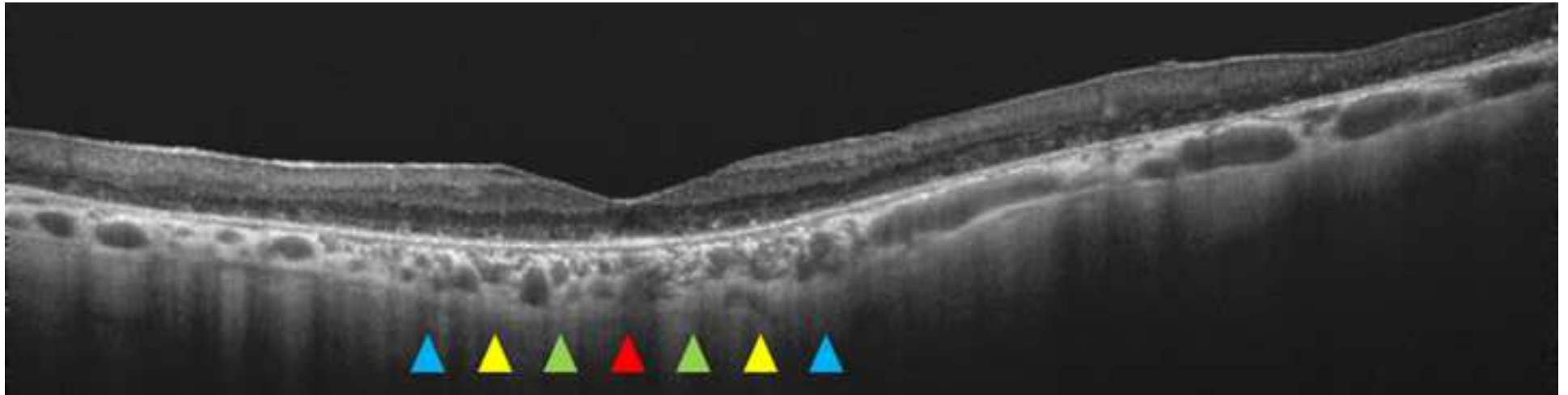
6 Bottom: Vertical changes in choroidal thickness
7

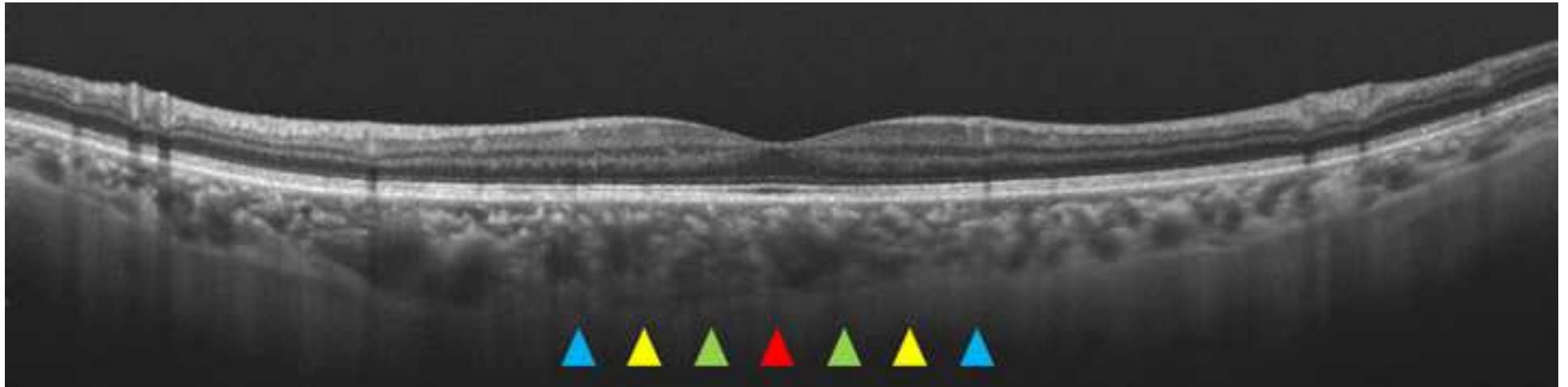
8 There is almost no difference in vertical choroidal thickness across the sites in these
9 groups.
10
11
12
13
14
15
16
17
18
19
20
21
22
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65

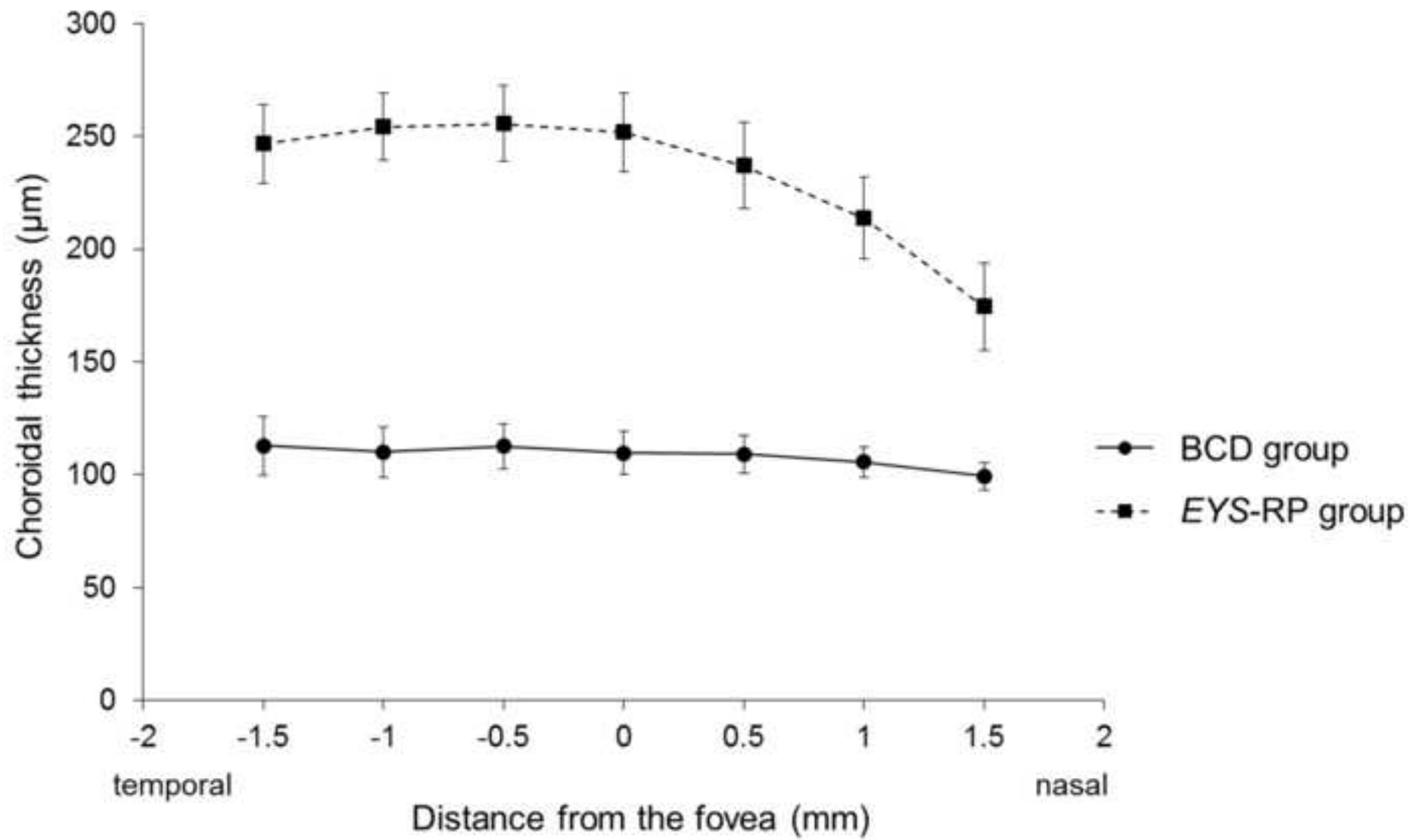












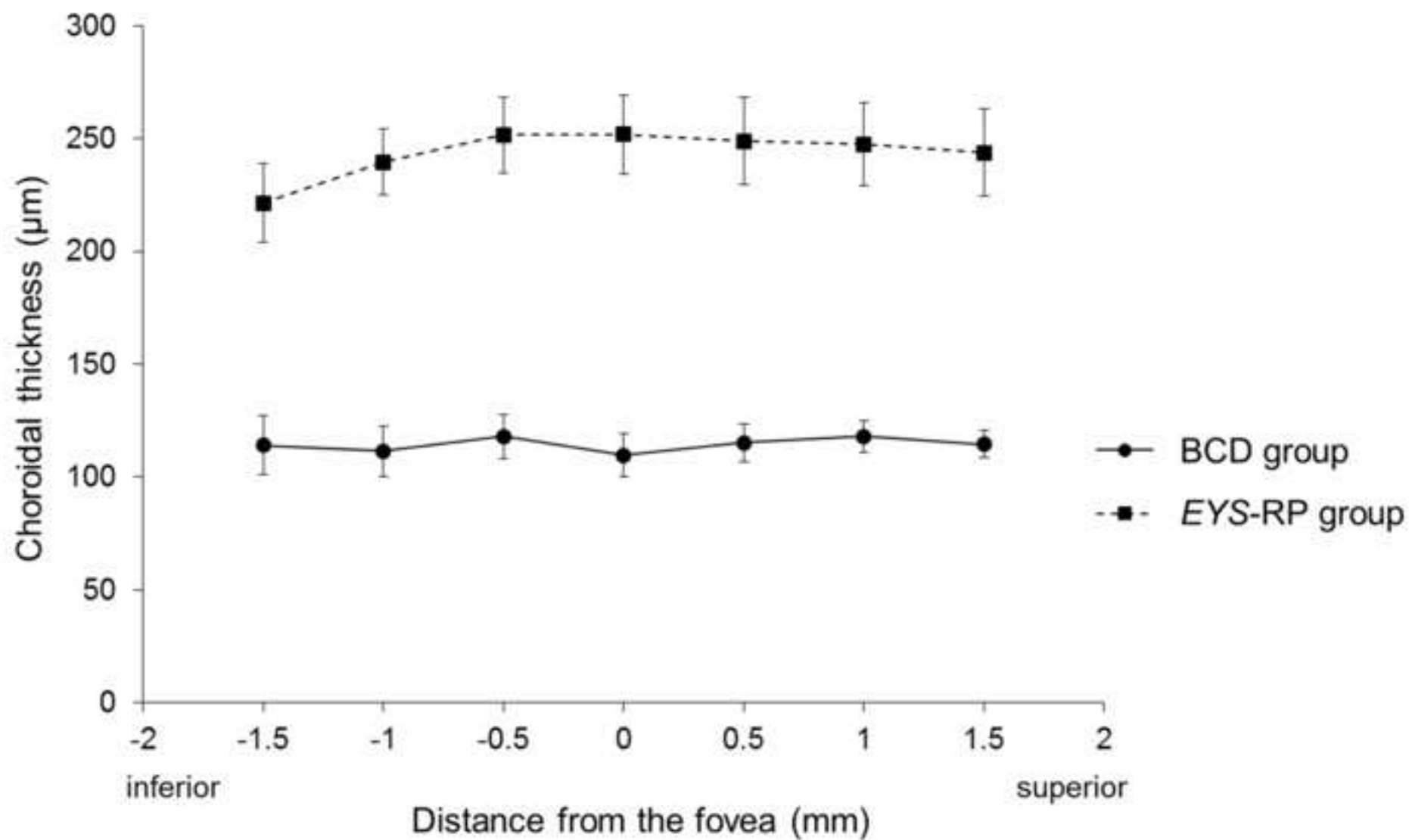


Table 1. Characteristics of the study population.

	BCD group	EYS-RP group	Control group
n, eyes (patients)	9 eyes (9 patients)	10 eyes (10 patients)	10 eyes (10 patients)
Female sex, eyes	5 eyes	4 eyes	7 eyes
Age (years)	57.9 ± 5.3	52.7 ± 14.1	54.0 ± 11.2
Visual acuity, logMAR	0.41 ± 0.37	0.73 ± 0.64	-0.10 ± 0.07
AL (mm)	24.21 ± 0.72	24.27 ± 1.70	24.33 ± 1.26
MD (dB)	-29.77 ± 4.01	-26.13 ± 6.66	

BCD group: Bietti crystalline dystrophy patients with CYP4V2 mutations

EYS-RP group: Retinitis pigmentosa patients with EYS mutations who were matched for age, AL, and MD with the BCD group.

Control group: Japanese volunteers with no eye disease, except for refractive errors, who were matched for age and AL with the BCD group.

Data are presented as mean ± standard deviation where applicable.

AL = axial length; logMAR = logarithm of the minimal angle of resolution; MD = mean deviation measurements using a Humphrey field analyzer (Carl Zeiss Meditec, Dublin, CA) with the 10-2 Swedish Interactive Threshold Algorithm standard program.

Table 2

Case	Age (years)	Sex	Eye	BCVA	AL (mm)	HFA10-2	ERG (cone flicker)		Mutation
						MD (dB)	Amplitude (μV)	Latency (ms)	
1	60	F	L	20/20	24.66	−27.59	91.75	25.2	c.802-8_810del17insGC / c.327+1G>A
2	58	F	L	20/25	24.52	−33.18	15.7	29.2	c.802-8_810del17insGC / c.1378T>C*
3	51	M	R	20/250	23.52	−34.82	7.39	34	c.802-8_810del17insGC / c.710C>A*
4	64	F	R	20/40	23.93	−24.25	67.50	30.2	c.802-8_810del17insGC / c.1226-6_1235del16
5	53	M	R	20/100	25.35	−32.26	25.71	28.4	c.802-8_810del17insGC homozygous
6	65	F	L	20/100	23.58	−30.62	extinguished		c.802-8_810del17insGC homozygous
7	51	F	R	20/66	25.12	−33.56	extinguished		c.802-8_810del17insGC homozygous
8	61	M	R	20/20	23.44	−27.21	6.64	37.8	c.802-8_810del17insGC homozygous
9	58	M	R	20/40	23.73	−24.40	39.75	28.0	c.802-8_810del17insGC / c.518T>G
EYS-RP group									
1	66	F	R	20/222	22.55	−33.36	extinguished		c.4957dupA / c.7048delT
2	49	F	R	20/33	24.89	−24.04	extinguished		c.5202_5203del / c.8379_8380insTGCA
3	45	F	R	20/100	23.08	−30.07	extinguished		c.4957dupA / c.8805C>A
4	79	F	R	20/222	23.37	−32.41	N.A.		c.4957dupA / c.8805C>A
5	34	M	R	20/29	28.03	−14.41	extinguished		c.4957dupA homozygous
6	51	M	L	20/200	26.2	−31.06	extinguished		c.9209T>C / c.4957dupA
7	34	M	L	20/13	24.24	−17.53	N.A.		c.4957dupA / c.5014C>T
8	64	M	R	20/222	23.21	−31.77	extinguished		c.7919G>A homozygous
9	56	M	L	20/1000	23.08	−24.78	extinguished		c.8805C>A / c.5014C>T
10	49	M	R	20/22	24.01	−21.91	extinguished		c.4957dupA / c.8805C>A

AL = axial length; BCD group = Bietti crystalline dystrophy with *CYP4V2* mutations group; BCVA = best-corrected visual acuity; ERG = electroretinography; EYS-RP group = retinitis pigmentosa with *EYS* mutations group; F = female; HFA10-2 = Humphrey field analyzer with the 10-2 SITA standard program; L = left eye; N.A.= not available; M = male; MD = mean deviation; R = right eye

* novel mutation

