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Citation
Scientific Reports (2017), 7

Issue Date
2017-07-12

URL
http://hdl.handle.net/2433/228161

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Type
Journal Article

Textversion
publisher

Kyoto University
Clinical relevance of reduced decorrelation signals in the diabetic inner choroid on optical coherence tomography angiography

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Diabetes induces lesions of the retinal and choroidal capillaries, which promote the pathogenesis of diabetic retinopathy (DR). The decorrelation signals in optical coherence tomography angiography (OCTA) represent the blood flow and vascular structure, and three-dimensional OCTA images enable individual capillary layers to be evaluated separately. The current study documented that en-face OCTA images revealed spots of flow void in the choriocapillaris layer in eyes with DR. Quantitative investigation demonstrated that non-flow areas within the central subfield (CSF) increased in eyes with more severe DR grades. The non-flow areas in the choriocapillaris layer were also associated with poorer visual acuity (VA) in all 108 eyes. A modest correlation was noted between the areas of flow void and poorer VA in 69 eyes without DME, whereas the non-flow areas were not related to VA or to CSF thickness measured by OCT in 39 eyes with DME. In 12 eyes with ischemic maculopathy, the choriocapillaris layer beneath the disrupted ellipsoid zone of the photoreceptor (EZ) had greater areas of flow void than did the area beneath an intact EZ. These data suggested that disrupted choroidal circulation has clinical relevance and contributes to the pathogenesis of DR.

Diabetic retinopathy (DR), which is a diabetic microangiopathy, often leads to severe visual impairment1. Therapeutic strategies, including anti-vascular endothelial growth factor (VEGF) therapy, photocoagulation, and vitrectomy, have been widely used to treat vision-threatening DR, e.g., proliferative diabetic retinopathy (PDR) and diabetic macular edema (DME). However, patients with diabetes often have poor visual prognosis2–4. Further investigations should elucidate the pathogenesis in the chorioretinal vessels and its effect on neuroglial dysfunction5, 6.

In the physiologic state, the inner and outer retinal layers are nourished via the retinal and choroidal vessels, respectively, and diabetes impairs the function and structure of these vessels. The choroidal vessels are composed of hierarchical structures that are represented by their layers. From the short posterior ciliary arteries, larger and smaller vessels in Haller’s layer and Sattler’s layer bifurcate into fine meshwork-like capillaries with small intercapillary spaces in the choriocapillaris layer7–9. The choroidal system nourishes the outer retinal layers in concert with the retinal pigment epithelium (RPE) and Bruch’s membrane10. A few histologic publications have documented the morphologic changes in the choroidal vasculature and periodic acid-Schiff positive deposits in the stroma and have proposed “diabetic choroidopathy” as another diabetic microangiopathy11, 12. In addition to the arteriosclerotic changes or vascular luminal narrowing in the larger choroidal vessels, capillary dropout in the choriocapillaris layer suggests ischemic changes in the outer retinal layers13. However, the in vivo relationship between the choriocapillaris and the outer retinal layers has not been investigated in diabetic eyes14.

Recent advances in fundus imaging have promoted the understanding of the pathogenesis in DR. Enhanced-depth imaging of spectral-domain optical coherence tomography (OCT) and swept-source OCT have particularly enabled both the qualitative and the quantitative in vivo evaluation of choroidal structure15, 16. Whether the quantified choroidal thickness is associated with DR severity or DME remains controversial17–24. Recent publications have documented several morphologic changes in the choroidal vasculature in Sattler’s layer or Haller’s layer in diabetic eyes25, 26. OCT angiography (OCTA), in which the decorrelation signals derive mainly from the movements of erythrocytes, delineates the three-dimensional vascular structure27–30. In healthy eyes, the
The areas of flow void in the inner choroid in eyes with moderate NPDR, severe NPDR, and PDR were significantly larger than those with no apparent retinopathy (Fig. 6A). On the ~10-μm-thick choroidal capillary slab images, the areas increased gradually according to the DR severity, and eyes with moderate NPDR, severe NPDR, and PDR had significantly larger areas of flow void than those with no apparent retinopathy (p = 0.032, p = 0.009, and p = 0.002, respectively) (Fig. 6B). We selected the non-flow areas in the 10-μm-thick slab images for further evaluation to investigate the clinical relevance of the flow void in the choriocapillaris layer. In 69 eyes without center-involved DME, the areas of flow void tended to increase as the DR severity progressed, and eyes with severe NPDR or PDR had larger non-flow areas than those with no apparent retinopathy (Fig. 6C). By contrast, we found no associations of the non-flow areas with systemic factors, i.e., age, HbA1c, diabetes duration, and systolic blood pressure.

Further quantification demonstrated that the areas of flow void did not differ among the central subfield (CSF) and the superior, nasal, temporal, and inferior subfields in the parafovea of the modified Early Treatment Diabetic Retinopathy Study (ETDRS) grid (3.25 ± 1.85 mm, 2.93 ± 1.69 mm, 2.91 ± 1.53 mm, 3.00 ± 1.76 mm, and 3.04 ± 1.53 mm, respectively; p = 1.000 for all comparisons) after the projection artifacts and shadow artifacts were carefully removed (Fig. 1).

**Clinical relevance of flow void in the choriocapillaris layer.** We evaluated how the areas of flow void in the CSF were associated with systemic and ocular parameters. We prepared the en-face OCTA images of choroidal capillary slab (~29 μm-thickness) according to the default setting (Fig. 6A). Eyes with severe NPDR or PDR had greater areas of flow void than those with no apparent retinopathy (Fig. 6A). On the ~10-μm-thick choriocapillaris slab images, the areas increased gradually according to the DR severity, and eyes with moderate NPDR, severe NPDR, and PDR had significantly larger areas of flow void than those with no apparent retinopathy (p = 0.032, p = 0.009, and p = 0.002, respectively) (Fig. 6B). We selected the non-flow areas in the 10-μm-thick slab images for further evaluation to investigate the clinical relevance of the flow void in the choriocapillaris layer. In 69 eyes without center-involved DME, the areas of flow void tended to increase as the DR severity progressed, and eyes with severe NPDR or PDR had larger non-flow areas than those with no apparent retinopathy (Fig. 6C). By contrast, we found no associations of the non-flow areas with systemic factors, i.e., age, hemoglobin A1c, diabetes duration, systemic hypertension, or dyslipidemia (data not shown).

Further statistical analyses showed that the logarithm of the minimum angle of resolution visual acuity (logMAR VA) was modestly associated with the non-flow areas within the CSF in either all 108 eyes or 69 eyes without center-involved DME (Fig. 7A,B). By contrast, no relationship was observed between the areas of flow void and logMAR VA in eyes with center-involved DME (Fig. 7C). No differences in the non-flow areas were noted between eyes with and without center-involved DME (p = 0.150), and the CSF thickness was not correlated with the non-flow areas in eyes with center-involved DME (Fig. 7D).

We compared the areas of flow void in the regions with and without intact ellipsoid zone of the photoreceptor (EZ) lines in 12 eyes with ischemic maculopathy among 108 eyes. Within the non-flow areas in the deep retinal capillaries, the regions without intact EZ lines had greater non-flow areas in the choriocapillaris layer than those with intact EZ lines (6.28 ± 3.34% vs. 2.00 ± 1.10%; p < 0.001) (Fig. 5).
contribute to retinal ischemia and concomitant expression of VEGF\textsuperscript{3,33}. Intriguingly, the non-flow areas in the choriocapillaris layer were associated with poorer VA and disrupted photoreceptor cells, thus indicating their clinical relevance and suggesting the pathological contribution of disrupted choroidal circulation to neuroglial impairment in DR.

Histological publications showed that the intercapillary spaces in the choriocapillaris layer in the macula were much smaller than those in the retina\textsuperscript{7,34}. Since the lateral resolution on en-face OCTA images was not high, we could not definitively recognize the choroidal capillaries but could recognize the retinal capillaries on 3 × 3-mm en-face OCTA images\textsuperscript{8}. This encouraged us to investigate the areas of flow void as an index of diabetes-induced dropout of capillaries in the choriocapillaris layer rather than the vascular structure per se\textsuperscript{11,13}. A choriocapillaris lobule in the macula with a diameter that ranges from dozens to hundreds of micrometers is fed by an arteriole,

**Figure 1.** Tiny dots of flow void in the choriocapillary layer in a 60-year-old patient with no apparent retinopathy. The en-face OCTA images in the superficial retinal layer (A) and the choriocapillaris layer (10 μm thick) (C) and structural OCT image at the RPE level (B). The corresponding B-scan images along the green arrow in panel A with (D,F) or without (E) the decorrelation signals (red). Red or green lines revealed the segmentation of individual layers. (G,H) The binary images of panels A and B using the global threshold in ImageJ. (I) The binary image of panel C using the mean decorrelation signal in the outer retina as the cutoff value. (J) The merged image of panel H with the inverted signal and panel G means the projection artifacts and/or shadow artifacts. (K) The areas of flow void in the choriocapillaris layer (black) after removing the artifacts. (L) The merged image of panels C (grayscale), (J) (green), and (K) with inverted signals (red). Scale bar = 500 μm.
and the dark spots of a similar size on OCTA images might represent disrupted blood flow in this unit. The non-flow areas with extended and undefined shapes might correspond to the histologic capillary dropout in the choriocapillaris layer.

OCTA showed that among several diseases with flow void in the inner choroid, geographic atrophy (GA) had significantly attenuated decorrelation signals in the choriocapillaris layer. Eyes with DR and GA appeared to share these characteristics in vascular morphology. By contrast, the boundary was indefinite, and the decorrelation signals were mildly attenuated in the most choroidal lesions in DR compared with GA. Theoretically, OCTA does not allow us to know the definite absence of in vivo blood flow, and the decorrelation signals represent the limited range of flow velocity. Further advances in fundus imaging technology should elucidate the structural and functional changes in choroidal vessels more definitively.

The areas of flow void were extended in the choriocapillaris layers as DR progressed. Systemic factors, including hyperglycemia and hypertension, might be common upstream regulators of both retinal and choroidal vascular lesions. Another possibility might be that VEGF from the retinas in DR increases Intercellular Adhesion Molecule (ICAM) expression, and concomitant leukostasis exacerbates capillary nonperfusion in the choriocapillaris layer. Alternatively, the circulatory disturbance in the choroid might contribute to DR progression. The outer retinal layers are nourished by the choroidal circulation in healthy eyes. Disrupted blood flow in the choroid would contribute to outer retinal ischemia, which would promote VEGF expression and subsequent DR progression. VEGF expression is increased mainly in ganglion cells, astrocytes, Müller cells, and RPE in diabetic retinas, and a future study should evaluate whether VEGF is up-regulated in the photoreceptors in eyes with disrupted choroidal circulation. Another explanation might be that diabetes-induced damage to the RPE, which contributes to the maintenance of capillaries in the choriocapillaris layer, might promote capillary dropout in the inner choroid.

A recent publication reported the association between deep retinal capillary nonperfusion and photoreceptor disruption in diabetic macular ischemia. We found that disrupted choroidal circulation is associated with photoreceptor damage and visual impairment within the areas of deep retinal capillary nonperfusion. A deficiency in oxygen or nutrients from the chorioretinal capillaries could contribute to photoreceptor damage. Three of 12 eyes had center-involved DME, which could also induce photoreceptor damage and be a confounding factor for the analyses in this study. Further study of a large cohort should be planned to elucidate the processes involved in the damage to photoreceptors and choroidal circulation.

Classical fundus imaging using fluorescent dyes delineated hypofluorescence in the choroidal flush phase of fluorescein angiography or the early phase of indocyanine green angiography in DR, suggesting disruption in choroidal blood flow. These modalities provide two-dimensional data, whereas three-dimensional OCTA images specifically depict the motion contrast images in the inner choroidal layer. Another modality, laser Doppler flowgraphy, allows the measurement of flow velocity, although OCTA images show a limited range of velocity. Comparative studies would improve the understanding of disrupted choroidal circulation in DR.

The current study had several limitations, e.g., its retrospective nature and the small number of cases. We carefully excluded the areas with projection artifacts from the retinal vessels and shadow cast by retinal lesions with higher OCT reflectivity; however, it is unclear how these artifacts might influence the non-flow areas in the inner choroidal layers. We had to recognize the limited generalizability because we used one of the major OCTA devices to image structures in Asian patients who contain higher amounts of pigment in the RPE and choroid. Another population-based study should be planned to compare the flow void between nondiabetic and diabetic...
subjects and to evaluate the reproducibility in other races, including other participants in whom systemic parameters are normalized, and using other devices.

In conclusion, we documented the lesions with attenuated decorrelation signals in the inner choroid in DR and their association with visual impairment and photoreceptor damage, suggesting the clinical relevance and pathophysiology of choroidal circulation in eyes with DR.

Methods

Patients. We retrospectively reviewed 108 consecutive eyes of 66 patients with DR who visited the Department of Ophthalmology of Kyoto University Hospital from February 2015 to August 2016. Eyes with DR were included when OCTA images of sufficient quality were acquired using Optovue RTVue XR Avanti (Optovue, Fremont, CA). The exclusion criteria were the presence of other chorioretinal diseases, glaucoma or ocular hypertension; an axial length shorter than 22.0 mm or longer than 26.0 mm; intraretinal or preretinal hemorrhages or hard exudates with diameters larger than one-third of the disc diameter within the central 3 × 3-mm square on the color fundus photograph; a history of any intervention for macular lesions; intraocular surgery other than cataract extraction; and cataract surgery within 6 months of study enrollment. We excluded eyes with a history of photocoagulation of the macular area because the decorrelation signals in the choriocapillaris layer were attenuated after this treatment49. To guarantee image quality, we also excluded eyes with a signal strength index (SSI) score of 60 or less50. Thirty-nine eyes had center-involved DME, which was determined by two-dimensional mapping constructed using the same dataset. According to the previously reported equation, the cutoff value for the CSF thickness was 299.52 μm on Optovue images, which corresponds to 250 μm on OCT images obtained by Stratus OCT (Carl Zeiss Meditec AG, Oberkochen, Germany)51,52. We thus determined that the eyes with a CSF thickness exceeding 299.52 μm had center-involved DME in the current study.

All research and measurements adhered to the tenets of the Declaration of Helsinki. The Kyoto University Graduate School and Faculty of Medicine Ethics Committee approved the study protocol. All participants provided written informed consent before inclusion in the study.
We performed a comprehensive ophthalmic examination including the color fundus photography (TRC-NW8F; Topcon Corp, Tokyo, Japan) and the measurement of the axial length using partial coherence interferometry (IOLMaster, Carl Zeiss Meditec AG). The best-corrected decimal VA was converted to logMAR VA.

OCTA images in the 3×3-mm square centered on the fovea were obtained using the Optovue RTVue XR Avanti (Fig. 1). Given the high A-scan speed of 70,000 scans/second and a light source of approximately 840 nm, this instrument delineates motion-dependent decorrelation signals using the split-spectrum amplitude decorrelation angiography algorithm. Briefly, three-dimensional scans are comprised of two consecutive B-scans (M-B frames) at a fixed position. The calculation of the decorrelation between the sequential images enables the detection of the motion of the blood cells and concomitant construction of a motion contrast AngioFlow image.

We applied segmentation processes to create the en-face OCTA images in the choroidal capillary slab, the retinal superficial and deep capillary plexus layers, and the avascular outer retina and the en-face structural OCT images in the RPE slab using the manufacturer’s software. The choroidal capillary slab images using the default OCTA. We performed a comprehensive ophthalmic examination including the color fundus photography (TRC-NW8F; Topcon Corp, Tokyo, Japan) and the measurement of the axial length using partial coherence interferometry (IOLMaster, Carl Zeiss Meditec AG). The best-corrected decimal VA was converted to logMAR VA.

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setting in the manufacturer’s software corresponded to the inner choroidal slab images extending from 31 to 59 μm posterior to the RPE-Bruch membrane complex segmentation. According to a publication documenting in vivo thicknesses in the choriocapillaris-Sattler’s layer complex and Haller’s layer, the choroidal capillary slab might correspond to parts of the choriocapillaris-Sattler’s layer complex in most eyes. Additionally, we prepared the ~10-μm-thick en-face images in the choriocapillaris layer (from 31 to 40 μm posterior to the RPE-Bruch membrane complex segmentation). The en-face images were used to evaluate EZ status and decorrelation signals in the choriocapillaris layer along the 10 gray lines. The images with and without decorrelation signals (red) along the blue arrow in panels G and H demonstrate that the EZ line is not intact in the central region (arrowheads). The magnified images of areas of flow void (green; K,M) and those without the artifacts (green; L,N) within the blue rectangle in panels G and H. The red lines correspond to the ‘not intact’ EZ line in panel I and partly colocalize with the areas of flow void. The blue lines correspond to the ‘intact’ EZ line in panel I and rarely colocalized with the non-flow areas. Scale bar = 500 μm.

**Figure 5.** Association between photoreceptor damage and flow void in the choriocapillaris layer in a 46-year-old patient with PDR. En-face OCTA images in the 10-μm-thick choriocapillaris layer (A) and the superficial (B) and deep (C) retinal capillary layers. (D) The merged image of the superficial and deep layers reveals ischemic maculopathy. The binary images of the flow void (E) or the artifacts (F). (G,H) The merged images of the deep capillary layer (panel C) and panel E or F with inverted signals (green). Both EZ status and decorrelation signals in the choriocapillaris layer were evaluated along the 10 gray lines. (I,J) The B-scan images with and without decorrelation signals (red) along the blue arrow in panels G and H demonstrate that the EZ line is not intact in the central region (arrowheads). The magnified images of areas of flow void (green; K,M) and those without the artifacts (green; L,N) within the blue rectangle in panels G and H. (K,L) The red lines correspond to the ‘not intact’ EZ line in panel I and partly colocalize with the areas of flow void. (M,N) The blue lines correspond to the ‘intact’ EZ line in panel I and rarely colocalized with the non-flow areas. Scale bar = 500 μm.
membrane complex) according to a recent publication. The corresponding structural OCT images were also created to assess shadow artifacts. The retinal superficial or deep slab OCTA images showed the superficial layer from the inner boundary 3 μm beneath the internal limiting membrane (ILM) to the outer boundary 15 μm beneath the inner plexiform layer (IPL) or the deep layer from the inner boundary 15 μm beneath the IPL to the outer boundary 70 μm beneath the IPL. The en-face images in the avascular outer retina were from the inner boundary 70 μm below the IPL to the outer boundary located 30 μm below the RPE reference. Given that most shadow artifacts from the intraretinal lesions are clearly depicted in the RPE, we created an en-face structural OCT slab (~28 μm thick) at the level of the RPE as described recently. These images were exported into ImageJ (NIH, Bethesda, MD) for image processing and analyses.

We then quantified the areas of flow void in the choroidal capillary slab in individual subfield of the modified ETDRS grid, including the central subfield (1 mm diameter) and the parafoveal quadrants (superior, inferior, nasal, and temporal subfields from 1 mm to 2.5 mm diameter), after the removal of the areas with projection artifacts and shadow artifacts as described recently. Histologic publications suggested that this instrument cannot detect the specific morphology of the normal vascular network in the choriocapillaris layer within the macular area. We thus evaluated the intensity of the decorrelation signals as the index of blood flow but not the vascular structure per se in the choriocapillaris layer. The preliminary study showed that the decorrelation signals are occasionally increased in the dilated capillaries around the nonperfused areas, which might incorrectly increase the average signal intensity. We therefore determined the mean decorrelation signals in the avascular outer retina as the cutoff value and measured the areas with decorrelation signals below the threshold to assess the disrupted blood flow.

We first calculated the mean levels of decorrelation signals in the avascular outer retina of every eye individually and used them as the individual threshold of the flow void in two en-face OCTA images of the choroidal capillary layer, i.e., the inner choroidal slab image using the default setting of the manufacturer’s software (from 31 to 59 μm posterior to the RPE-Bruch membrane complex segmentation) and the 10-μm-thick choriocapillaris slab image (from 31 to 40 μm posterior to the RPE-Bruch membrane complex) (Fig. 1). We further created the binary images of the en-face OCTA image in the retinal superficial layer and the structural OCT image in the RPE using the global threshold as described recently (Fig. 1G, H). We then removed the projection artifacts from the retinal superficial vessels and the shadow artifacts from the preretinal or intraretinal hyperreflective lesions.

Figure 6. The areas of flow void in the choroidal capillary OCTA slab in individual DR severity grades. The non-flow areas within the CSF in the 29-μm-thick choroid capillary slab of the default setting (A) or the 10-μm-thick choriocapillaris layer (B) increase gradually according to individual DR severity grades in all 108 eyes. Areas of flow void in the 10-μm-thick choriocapillaris layer in eyes without (C) and with (D) center-involved DME.
e.g., microaneurysms, hemorrhages and hard exudates (Fig. 1K), before we measured the areas of flow void and calculated the percentage of the non-flow areas in the total areas without artifacts.

We further investigated the relationship between photoreceptor damage and the non-flow areas in the choriocapillaris layer in eyes with ischemic maculopathy. We first determined ischemic maculopathy of grade 3 or 4 according to the modified methods described by the ETDRS Report Number 11.\(^{55}\) We merged the en-face OCTA images in the retinal superficial and deep capillary layers (Fig. 5D) using the colour merge function in the analysis plugin WCIF-ImageJ (http://www.uhnresearch.ca/facilities/wcif/imagej/). Ischemic maculopathy was defined on the merged OCTA images instead of FA images based on the agreement of two independent masked graders. We selected and further evaluated 12 eyes with the outline of the FAZ destroyed for one-half or more of the original circumference.\(^{55}\) A recent publication demonstrated the association between photoreceptor disruption and deep retinal capillary nonperfusion in diabetic eyes,\(^{45}\) prompting us to investigate the relationship between the EZ status and the flow void in the choriocapillaris layer within the nonperfused areas in the deep retinal capillary layer. We prepared the binary images of the non-flow areas in the choriocapillaris layer and artifacts (Fig. 5E,F). The regions without intact EZ lines were determined on the B-scan images along 10 transverse lines (17th, 47th, 77th, 107th, 137th, 167th, 197th, 227th, 257th, and 287th lines from the most superior line; Fig. 5G,I,K,L)\(^{14}\). The transverse length of the areas of flow void and those without artifacts were measured within the corresponding regions (Fig. 5K,L). We then summed the transverse lengths along all 10 lines and calculated the percentages of the areas of flow void. Similarly, we evaluated the non-flow areas in the regions with intact EZ lines (Fig. 5M,N).

**Figure 7.** Association between logMAR VA and the areas of flow void in the choriocapillaris layer. There is a modest correlation between logMAR VA and the non-flow areas within the CSF in all 108 eyes (A) and in the 69 eyes without center-involved DME (B). (C,D) The areas of flow void in the CSF are not associated with either logMAR VA or CSF thickness in the 39 eyes with center-involved DME.

### Statistical analysis.

The data are expressed as the mean ± SD. After the Kolmogorov–Smirnov test was applied to confirm that the dataset was normally distributed, the significant differences were evaluated using Student’s t-test or analysis of variance with Bonferroni correction, and the correlation was assessed using Pearson’s correlation coefficient.
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**Acknowledgements**
This work was partially funded by the Grant-in-Aid for Scientific Research of Japan Society for the Promotion of Science to T.M. (26462637).

**Author Contributions**
Y.D. wrote the manuscript; K.S., K.I., S.Y., M.F., and T.M. analyzed the data; K.S., K.I., S.Y., M.F., T.Y., Y.M., and T.M. prepared the figures; K.S., K.I., S.Y., and T.M. provided the conception and design of the research. K.S., K.I., S.Y., M.F., T.Y., Y.M., and T.M. edited and reviewed the manuscript. All authors accepted the final version of the manuscript.

**Additional Information**
Competing Interests: The authors declare that they have no competing interests.

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