Near-infrared long persistent luminescence of Er$^{3+}$ in garnet for the third bio-imaging window†

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By utilizing efficient persistent energy transfer from Ce$^{3+}$ to Er$^{3+}$, we have successfully developed a novel garnet persistent phosphor of Y$_3$Al$_2$Ga$_3$O$_{12}$ doped with Er$^{3+}$, Ce$^{3+}$, Cr$^{3+}$ ions (YAG:Er–Ce–Cr) exhibiting long (>10 h) near-infrared (NIR) persistent luminescence (PersL) in the broad range from 1450 nm to 1670 nm due to the typical Er$^{3+}$:4$I_{13/2}$ → 4$I_{15/2}$ transition in garnet. The NIR PersL bands of Er$^{3+}$ match well with the third bio-imaging window (NIR-III, approximately from 1500 nm to 1800 nm) and the response curve of InGaAs detectors. The photon emission rate ($8.33 \times 10^{17}$ cps Sr$^{-1}$ m$^{-2}$) of the YAG:Er–Ce–Cr persistent phosphor at 10 min after ceasing blue light illumination was over two times higher than that of the widely used ZnGa$_2$O$_4$:Cr$^{3+}$ deep-red persistent phosphor ($3.30 \times 10^{17}$ cps Sr$^{-1}$ m$^{-2}$). We also show the first PersL imaging by a commercial InGaAs camera monitoring Er$^{3+}$ emission indicating that this material can be a promising candidate for in vivo bio-imaging in the NIR-III window.

1. Introduction

Over 20 years have passed since the new generation green persistent phosphor SrAl$_2$O$_4$:Eu$^{2+}$–Dy$^{3+}$ (SAO:Eu–Dy) was developed and successfully commercialized for watch dials, luminous paints, safety guidance signs, etc.$^4$ Recently, the luminescence wavelength of persistent phosphors has extended from visible light to near-infrared (NIR),$^2$–$^6$ which is suitable for in vivo bio-imaging due to the reduced scattering and minimal absorption coefficient when compared with visible light, allowing imaging of deep tissues. Furthermore, persistent phosphors in the form of nano-particles charged by ultraviolet (UV) light (visible light in rare cases) before injecting into small animals can emit NIR persistent luminescence (PersL) for minutes to even hours without further in situ excitation. The exclusion of external illumination totally removes the autofluorescence as background noise, avoids the complicated background subtraction procedures and thus improves the signal-to-noise ratio (SNR) remarkably.$^7$–$^9$ This new bio-imaging technology has soon become quite attractive while the most reported emission region of persistent phosphors is located in the first biological window (NIR-I, 650–950 nm),$^4$–$^{13}$ partially due to the easy availability of Si-detectors that work well in this region.

Since the Rayleigh scattering (varies as $\lambda^{-4}$, here $\lambda$ is the wavelength) decreases with increasing wavelength, two more biological windows called the second (NIR-II, 1000–1350 nm)$^{14}$ and the third (NIR-III, approximately from 1500 nm to 1800 nm)$^{15}$–$^{17}$ ones are defined, which fall on either side of a strong water absorption band centered around 1450 nm. Compared with the NIR-I window, these two windows, especially the NIR-III window, give much lower scattering coefficient leading potentially to an improved resolution quality and deeper penetration depth.$^{18}$–$^{20}$ Moreover, thanks to the recent development of affordable InGaAs detectors that possess adequate sensitivity and high quantum efficiency in the wavelength region above 1000 nm and up to 1650 nm, the shift of the luminescence wavelength of the bio-imaging probe from the NIR-I to the NIR-III window is definitely demanded. However, even for NIR-to-NIR fluorescence bio-probes using real-time excited photoluminescence (PL) for in vivo imaging, only a few candidates are available for the NIR-II window including the single-walled carbon nanotubes (SWNTs)$^{16}$,$^{21}$ and heavy metal based semiconductor quantum dots (QDs)$^{16}$,$^{22}$ both of which suffer from their potential cytotoxicity for living beings.$^{15}$–$^{17}$ Therefore, the development of nontoxic and biocompatible luminescent materials emitting in the NIR-III window still remains a big challenge.

Due to the typical $4I_{13/2}$ → $4I_{15/2}$ transition at around 1.55 μm and several important transitions such as the $4S_{3/2}$ → $4I_{15/2}$ one in green,$^{23}$ the Er$^{3+}$ ion has already played a key role in applications such as eye-safe lasers,$^{24}$ optical amplifiers in fiber telecommunication$^{15}$,$^{26}$ and upconversion fluorescence$^{27}$,$^{28}$ for long years. On the other hand, Er$^{3+}$ is also considered as the most promising lanthanide ion whose emitting wavelength matches well with both the NIR-II window and the response curve of InGaAs detectors.$^{15}$–$^{17}$ Pan et al.$^{29}$ firstly observed the PersL from Er$^{3+}$ through the so-called persistent energy transfer (ET) from Eu$^{2+}$ in the well-known SAO:Eu–Dy persistent phosphor.

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However, compared with super long PersL (> 10 h) from Eu\textsuperscript{3+} in the green region, the PersL intensity and duration of Er\textsuperscript{3+} emission peaking at 1530 nm were much weaker and shorter (less than 10 min) after ceasing UV excitation. On the other hand, based on our previous work, in which a novel garnet persistent phosphor of Ce\textsuperscript{3+}Cr\textsuperscript{3+} co-doped Y\textsubscript{2}Al\textsubscript{5}Ga\textsubscript{3}O\textsubscript{12} (YAG:Ce–Cr) was developed and Cr\textsuperscript{3+} acts as an efficient electron trap with an ideal trap depth at \(x = 3\) for PersL of Ce\textsuperscript{3+} working at room temperature (RT),\textsuperscript{30–33} we successfully developed a persistent phosphor of Y\textsubscript{2}Al\textsubscript{5}Ga\textsubscript{3}O\textsubscript{12}:Nd\textsuperscript{3+},Ce\textsuperscript{3+},Cr\textsuperscript{3+} (YAG:Nd–Ce–Cr) emitting long PersL (> 10 h) at 880 nm, 1064 nm, and 1335 nm due to f–f transitions of Nd\textsuperscript{3+} through efficient persistent ET from Ce\textsuperscript{3+}.

It is also worth noting that, in addition to the Er\textsuperscript{3+} emission mainly composed of the C-band (1530–1565 nm) and the L-band (1565–1625 nm) usually observed in many hosts,\textsuperscript{35–40} it exhibits an extended U-band (1625–1675 nm) in garnet hosts owing to the large Stark splitting of the terminal \(^{4}\mathrm{I}_{15/2}\) level.\textsuperscript{41,42} In this paper, we give an example of how to realize Er\textsuperscript{3+} long PersL through an efficient persistent ET process from Ce\textsuperscript{3+} in a garnet host and show the first PersL imaging for the NIR-III window by an InGaAs camera.

2. Experimental section

YAG:Er–Ce–Cr, YAG:Ce–Er and YAG:Er ceramic phosphors with the compositions of \(\mathrm{Y}_{2.925}\mathrm{Ce}_{0.015}\mathrm{Er}_{0.06}\mathrm{Al}_{0.999}\mathrm{Cr}_{0.001}\mathrm{Ga}_{0.012}\), \(\mathrm{Y}_{2.925}\mathrm{Ce}_{0.015}\mathrm{Er}_{0.06}\mathrm{Al}_{0.999}\mathrm{Ga}_{0.012}\) and \(\mathrm{Y}_{2.925}\mathrm{Er}_{0.06}\mathrm{Al}_{0.001}\mathrm{Ga}_{0.012}\) respectively, were fabricated by a conventional solid-state reaction method. \(\mathrm{Y}_{2}\mathrm{O}_{3}\) (99.99%), \(\mathrm{Al}_{2}\mathrm{O}_{3}\) (99.99%), \(\mathrm{Ga}_{2}\mathrm{O}_{3}\) (99.99%), \(\mathrm{CeO}_{2}\) (99.99%), \(\mathrm{Er}_{2}\mathrm{O}_{3}\) (99.99%) and \(\mathrm{Cr}_{2}\mathrm{O}_{3}\) (99.9%) were used as raw materials. The starting powder was mixed by a ball milling method (Premium Line P-7, Fritsch Co. Ltd) with anhydrous alcohol for several hours. The mixed powder was dried at 80 °C for 36 h, compacted to form a ceramic green body (0.2 mm thickness) under uniaxial pressing of 50 MPa, and finally sintered at 1600 °C for 10 h in air. The YAG:Ce–Cr \(\left(\mathrm{Y}_{2.985}\mathrm{Ce}_{0.015}\mathrm{Al}_{0.999}\mathrm{Cr}_{0.001}\mathrm{Ga}_{0.012}\right)\) and YAG:Ce \(\left(\mathrm{Y}_{2.985}\mathrm{Ce}_{0.015}\mathrm{Al}_{0.001}\mathrm{Ga}_{0.012}\right)\) ceramic phosphors prepared by the same experimental procedure were used as reference samples. All the as-prepared samples were confirmed to be single phase (ICPDS: No. 089-6660) by XRD measurements (see Fig. S1, ESI†).

The diffuse reflectance spectrum of the YAG:Er ceramic sample was measured using a spectrophotometer (UV3600, Shimadzu) equipped with an integrating sphere. The lifetime measurements of the YAG:Ce and YAG:Ce–Er samples were made using a compact fluorescence lifetime spectrometer (Quantaurus-Tau-C11367, Hamamatsu Photonics & Co. Ltd) using a 405 nm picosecond pulsed LED as an excitation source. Photoluminescence (PL) spectra of the YAG:Ce, YAG:Er and YAG:Ce–Er ceramic samples by pumping with a 405 nm laser diode (LD) (SDL-405-LM-100T, Shanghai Dream Lasers Technology Co. Ltd) and the YAG:Ce–Cr and YAG:Er–Ce–Cr samples by pumping with a 442 nm LD (NDHBS10APA-E, Nichia Co. Ltd) were recorded in the range of 350–1670 nm. The PL spectra were measured using a monochromator (G250, Nikon) equipped with a Si photodiode (PD) detector (S-025-H, Electro-Optical System Inc.) from 350 to 1100 nm and an InGaAs PD detector (IGA-030-H, Electro-Optical System Inc.) from 1100 to 1670 nm (see the schematic illustration of the measurement setup in Fig. S2, ESI†). All the PL spectra were calibrated by using a standard halogen lamp (SCL-600, Labsphere). The persistent luminescence (PersL) spectrum of the YAG:Er–Ce–Cr sample was measured using a Si CCD spectrometer (QE65-Pro, Ocean Optics) from 350 to 1075 nm and an InGaAs CCD spectrometer (NIR-Quest512, Ocean Optics) from 1075 to 1700 nm connected with an optical fiber. All the PersL spectra were calibrated using the same halogen lamp. A 300 W Xe lamp (MAX-302, Asahi Spectra) with an UV mirror module (250–400 nm) was used as the excitation source for thermoluminescence (TL) two-dimensional (2D) plot measurements. The ceramic sample was set in a cryostat (Helitran LT3, Advanced Research Systems) to control the temperature and firstly illuminated by UV light at 100 K for 10 min, then heated up to 600 K at a rate of 10 K min\(^{-1}\) for 10 min after ceasing the illumination, and the TL signals were recorded using a PMT detector (R11041, Hamamatsu Photonics & Co. Ltd) covered with 475 nm short-cut and 650 nm long-cut filters to monitor Ce\textsuperscript{3+} emission and the same InGaAs PD detector covered with a 1000 nm short-cut filter to monitor the NIR emission of Er\textsuperscript{3+}. The same Si CCD and InGaAs CCD spectrometers were operated simultaneously with the TL measurement to monitor the emission spectra at different temperatures. All persistent luminescence decay curves of the YAG:Er–Ce–Cr sample after being excited for 5 min by the Xe lamp with a 460 nm band-pass filter were measured at 25 °C using the same PMT or InGaAs PD detectors (see the schematic illustration of the measurement setup in Fig. S3, ESI†). In order to monitor the Ce\textsuperscript{3+} emission, the PMT detector was covered with 475 nm short-cut and 650 nm long-cut filters to filter out all but the Ce\textsuperscript{3+} luminescence. Then the decay curves were calibrated to absolute luminance (in units of mcd m\(^{-2}\)) using a radiance meter (Glacier X, B&W Tek Inc.). In order to monitor the Er\textsuperscript{3+} luminescence, the InGaAs PD was covered with a 1000 nm short-cut filter to filter out all but the Er\textsuperscript{3+} luminescence. Then the decay curves were calibrated to the photon emission rate (in units of cps Sr\(^{-1}\) m\(^{-2}\)) using the same radian meter calculated by the absolute radian (in units of mW Sr\(^{-1}\) m\(^{-2}\)) divided by photon energy (photon energy: \(E = hc/\lambda\), where \(h\) is the Planck constant, \(c\) is the speed of light in vacuum and \(\lambda\) is the wavelength). The photographs of the YAG:Ce–Cr and YAG:Er–Ce–Cr samples after charging by a 455 nm LED lamp (M445L3, THORLABS, Inc.) for 5 min were taken using a digital camera (EOS kiss X5, Canon) for the visible light region, and the settings remained constant: exposure time – 1 s, ISO value – 1600, aperture value (F value) – 5.0. The photographs for the short-wave infrared (SWIR) region (~900–1700 nm) were taken using a bio-imaging machine (NIS-OPT, Shimadzu) equipped with an InGaAs camera (Xeva-1.7-320, TE3 cooling system down to 223 K) after charging by the same 455 nm LED for 5 min, and the integrating time was set to be 0.04 s.
3. Results and discussion

3.1. Efficient energy transfer from Ce\(^{3+}\) to Er\(^{3+}\) in the YAG\(_G\) host

Fig. 1(a)–(c) show the PL spectra of the YAG\(_G\):Ce, YAG\(_G\):Er and YAG\(_G\):Ce–Er samples under 405 nm laser excitation. Both the broad band emission from Ce\(^{3+}\):5d\(_1\) \(\rightarrow\) 4f peaking at 500 nm and sharp-line emission bands from Er\(^{3+}\):4S\(_{3/2}\) \(\rightarrow\) 4I\(_{15/2}\) peaking at 555 nm, the S\(_{3/2}\) \(\rightarrow\) I\(_{15/2}\) band peaking at 862 nm, and the \(4\text{I}_{13/2}\) \(\rightarrow\) \(4\text{I}_{15/2}\) band peaking at 1532 nm in the YAG\(_G\):Ce–Er sample are clearly observed. Especially for the \(4\text{I}_{13/2}\) \(\rightarrow\) \(4\text{I}_{15/2}\) transition, similar to that in the Y\(_3\)Al\(_5\)O\(_{12}\) (YA\(_G\)) host,\(^{40–44}\) broadly split bands ranging from 1450 to 1670 nm in the YAG\(_G\) host are also observed. On the other hand, the emission bands due to the Er\(^{3+}\):4I\(_{11/2}\) \(\rightarrow\) 4I\(_{15/2}\) transition peaking at 966 nm and 1013 nm are almost quenched in the YAG\(_G\):Ce–Er sample compared with that in the YAG\(_G\):Er sample. In the YAG\(_G\):Ce–Er sample, the extra decay pathways mainly from the 5d\(_1\) level of Ce\(^{3+}\) to the \(4\text{F}_{7/2}\) and \(2\text{H}_{11/2}\) levels of Er\(^{3+}\) are generated as shown in Fig. 1(d). The total decay rate in the co-doped sample is given by

\[
W_{\text{tot}} = A + W_{\text{MP}} + W_{\text{ET}}
\]

where \(A\) is the radiative rate, \(W_{\text{MP}}\) is the multi-phonon relaxation rate, and \(W_{\text{ET}}\) is the energy transfer rate. The ET efficiency is

\[
\eta_{\text{ET}} = \frac{W_{\text{ET}}}{A + W_{\text{MP}} + W_{\text{ET}}} = 1 - \frac{\tau_{\text{Ce,Er}}}{\tau_{\text{Ce}}}
\]

Thus, the efficiency can be estimated to be \(~64\%\) indicating that not only in YA\(_G\),\(^{43,44}\) but also in the YAG\(_G\) host, efficient ET can occur from Ce\(^{3+}\) to Er\(^{3+}\) (although energy transfer between Cr\(^{3+}\) and Er\(^{3+}\) also occurs, to simplify the mechanism in this paper, Cr\(^{3+}\) is assumed to act only as an electron trap due to its very low doping concentration).
3.2. PL and PersL spectra

Fig. 2(a) shows the PL spectra of the YAGG:Ce–Cr and YAGG:Er–Ce–Cr samples under blue laser (442 nm) excitation. The YAGG:Ce–Cr sample exhibits an intense emission band peaking at 505 nm due to the parity allowed 5d₁ → 4f transition of Ce³⁺.⁴⁶–⁴⁸ Besides, a weak emission band peaking at around 690 nm is attributed to the Cr³⁺:E⁵⁷(G) → A⁴⁹(E) transition (the so-called R-line). Comparing the PL spectrum of the Ce³⁺ emission intensity in the visible range and the YAGG:Ce–Cr sample with the diffuse reflectance of the Er³⁺ singly doped YAGG sample (YAGG:Er), the absorption bands (|11/₂, 2| → |3/₂, 4/₂, 5/₂, 7/₂, 2H₁₁/₂, 4S₃/₂) of Er³⁺ are largely overlapped with the emission band of Ce³⁺ indicating that the ET process from Ce³⁺ to Er³⁺ can efficiently occur. This is confirmed by the decrease of the Ce³⁺ emission intensity in the visible range and the presence of several sharp emission bands at around 555 nm, 862 nm and 1532 nm owing to the f–f transitions of Er³⁺:|4I₁₅/₂, 4I₁₃/₂, 4I₁₁/₂, |4I₉/₂, |4I₇/₂, |4I₅/₂, |4I₃/₂, |4I₁/₂⟩ → |4F₃/₂⟩ bands peaking at 966 nm and 1013 nm is also observed as in the YAGG:Ce–Er sample due to the cross-relaxation process mentioned in Section 3.1.

The PersL spectra of the YAGG:Er–Ce–Cr sample recorded at different times after ceasing blue light illumination are shown in Fig. 2(b). The persistent emission bands not only exhibit a broad band peaking at around 500 nm due to Ce³⁺:5d₁ → 4f transition but also intense sharp bands located in the NIR region (around 862 nm and 1532 nm) due to the f–f transitions of Er³⁺. Especially the PersL bands in the range of 1450–1670 nm match well with the NIR-III window. The identical shape of the PL and PersL spectra of the YAGG:Er–Ce–Cr sample suggests that the emission centers are the same under and after excitation. Since the efficient ET process from Ce³⁺ to Er³⁺ was confirmed in the YAGG:Ce–Er sample shown in Fig. 1(e), we assume that the efficient persistent ET also occurs in the YAGG:Er–Ce–Cr phosphor after ceasing the blue light illumination.

3.3. Persistent luminescence decay curves

The persistent luminescence decay curve monitoring the Ce³⁺ emission (475–650 nm) of the YAGG:Er–Ce–Cr sample after ceasing the blue light illumination is shown in Fig. 3(a), in which the decay curves of the standard YAGG:Ce–Cr ceramic phosphor and a compacted ceramic pellet made of the commercial SAO:Eu–Dy phosphor (LumiNova-GLL300FFS, Nemoto & Co. Ltd) under the same experimental condition are also plotted as references.⁴⁹ The luminescence values at 10 min after ceasing the excitation are 58 mcd m⁻² for YAGG:Er–Ce–Cr, 627 mcd m⁻² for YAGG:Ce–Cr, and 211 mcd m⁻² for SAO:Eu–Dy [see the photographs of the two samples after ceasing blue LED illumination in Fig. 3(c)]. Persistent luminescence duration to reach a luminescence value of 0.32 mcd m⁻² in the YAGG:Er–Ce–Cr sample is around 213 min, which is much shorter than that of the YAGG:Ce–Cr sample (about 808 min), due to the quenching effect of the green Ce³⁺ emission by the non-radiative persistent ET to Er³⁺ (note that the luminescence value 0.32 mcd m⁻² is the minimum value commonly used by the safety signage industry, about 100 times the sensitivity of the dark-adapted eye).⁴⁹

The persistent luminescence decay curve monitoring Er³⁺:NIR emission (> 1000 nm) of the YAGG:Er–Ce–Cr sample after ceasing the same blue light illumination is shown in Fig. 3(b), in which the decay curve of the standard ZnGa₂O₄:Cr³⁺ (ZGO:Cr, emitting wavelength peaking at 695 nm) sample under the same experimental condition is also plotted as a reference.⁵⁰ The NIR photon emission rate of the YAGG:Er–Ce–Cr sample for the NIR-III window at 10 min after ceasing the blue light excitation (8.33 × 10¹⁷ cps Sr⁻¹ m⁻²) is over two times higher than that of the widely used deep-red persistent phosphor ZGO:Cr (3.30 × 10¹⁷ cps Sr⁻¹ m⁻²) for the NIR-I window (summarized in Table 1), which indicates that this persistent phosphor exhibits excellent PersL in the NIR region by Er³⁺ as well as in the visible light region by Ce³⁺. In Fig. 3(d), we exhibit the first PersL imaging by a commercial InGaAs camera for the two garnet ceramic phosphors. Although the YAGG:Ce–Cr sample shows bright green PersL by a digital camera in the visible light region [see Fig. 3(c)], no signal can be captured by the InGaAs camera due to its lack of PersL in the short-wave infrared (SWIR, ~900–1700 nm) region. Since the InGaAs camera is only sensitive to the luminescence located in the SWIR region,
3.4. Thermoluminescence (TL) glow curves and 2D contour plots

Fig. 4(a) and (c) show the TL glow curves of the YAG:Ce–Cr and YAG:Er–Ce–Cr samples monitoring only Ce$^{3+}$ emission, respectively. Although the intensity of the Ce$^{3+}$ emission in the YAG:Er–Ce–Cr sample is much weaker than that in the YAG:Ce–Cr sample due to the persistent ET from Ce$^{3+}$ to Er$^{3+}$, both of the glow peaks are located at around 300 K. This suggests that PersL from Ce$^{3+}$ is originated from the same electron trap, Cr$^{3+}$, in both samples with the same trap depth or more precisely with the same trap distribution.$^{31,34}$ Therefore, the thermal activation energy which is required to clean up the electrons captured by the traps is the same.$^{31}$

For TL measurements monitoring the visible light region, the black-body radiation has nearly no effects on the TL read-out when the heating temperature is lower than 600 K. On the other hand, when monitoring NIR emission, the excessive effect of black-body radiation on the TL read-out must be taken into account.$^{52,53}$ Fig. 4(b) and (d) show the TL glow curves of the YAG:Ce–Cr and YAG:Er–Ce–Cr samples monitoring only NIR emission (>1000 nm), respectively. Since there is no PersL contribution from the YAG:Ce–Cr sample in the wavelength region over 1000 nm, only intense black-body radiation from the sample and partially from the measurement setup is observed. However, for the YAG:Er–Ce–Cr sample, the additional TL glow curve centered at around 300 K is also observed after subtracting the intense black-body radiation signal (blue-line by subtracting the black-dotted-curve from the red-dotted-line), which is ascribed to the PersL from Er$^{3+}$ in the NIR region.

Fig. 5 shows the TL two-dimensional (2D) contour plots of the YAG:Ce–Cr and YAG:Er–Ce–Cr samples in order to see what kind of emission bands contribute to the TL glow peak at different temperatures. From the contour plots of the YAG:Ce–Cr sample in Fig. 5(a) and (b), it can be seen that at increased temperatures, the TL spectrum is simply composed of two emission bands from Ce$^{3+}$ and Cr$^{3+}$, and no contributed
emission except strong black-body radiation in the NIR region (1300–1700 nm, above ~500 K) is observed. On the other hand, in the YAG:Er–Ce–Cr sample as shown in Fig. 5(c) and (d), besides the emission bands from Ce$^{3+}$ and Cr$^{3+}$, the NIR emission band of Er$^{3+}$ appears in the same temperature range, which agrees well with its PersL spectrum in Fig. 2(b). The intense TL glow peaks of both YAG:Ce–Cr and YAG:Er–Ce–Cr samples lie in almost the same temperature range (around 300 K), close to RT and body temperature of Mammalia (around 310 K). Since the TL peak temperature is correlated to the energy gap between

![Fig. 4](image1.png)

**Fig. 4** TL glow curves of the YAG:Ce–Cr ceramic sample monitored by the (a) PMT detector in the range of 475–650 nm, (b) InGaAs detector in the range of over 1000 nm and that of the YAG:Er–Ce–Cr ceramic sample monitored by the (c) PMT detector in the range of 475–650 nm, and (d) InGaAs detector in the range of over 1000 nm.

![Fig. 5](image2.png)

**Fig. 5** Wavelength–temperature (λ–T) contour plots of the (a), (b) YAG:Ce–Cr and (c), (d) YAG:Er–Ce–Cr ceramic samples.
the bottom of the conduction band and the electron trap; the identical glow temperature of the two samples indicates the same electron trapping and de-trapping process in both, where Cr\(^{3+}\) works as an efficient electron trap with ideal trap depth for long PersL in the host of Y\(_3\)Al\(_2\)Ga\(_3\)O\(_{12}\) (YAG) host.

3.5. Persistent luminescence mechanism

The persistent luminescence mechanism of the YAG:Er–Ce–Cr persistent phosphor is briefly explained by constructing the vacuum referred binding energy (VRBE) diagram composed of Ce\(^{3+}\), Er\(^{3+}\), Cr\(^{3+}\), conduction band (C.B.), and valence band (V.B.) energy levels in the Y\(_3\)Al\(_2\)Ga\(_3\)O\(_{12}\) host (see Fig. 6). When the YAG:Er–Ce–Cr sample is charged by blue light, the electron located at the ground state (2\(F\))\(_{5/2}\) of Ce\(^{3+}\) is promoted to the excited state of the lowest 5d energy level (5d\(_{1}\)). Since the energy gap between the bottom of C.B. and the 5d\(_{1}\) level is small, the excited electron can “jump” to C.B. with thermal assisted activation energy at RT and then be trapped by the electron trapping center (Cr\(^{3+}\))\(_{0}\). At that time, Ce\(^{3+}\) is photo-oxidized into Ce\(^{4+}\) or (Ce\(^{3+}\) + h\(^+\)) and Cr\(^{3+}\) changes into Cr\(^{2+}\) or (Cr\(^{3+}\) + e\(^-\)) after capturing one electron (process 3). Then the de-trapping process occurs with the thermal release of the captured electron from the Cr\(^{2+}\) (Cr\(^{3+}\) + e\(^-\)) trap, and finally the excited state of the Ce ion, (Ce\(^{3+}\))\(^*\), appears after capturing the released electron in the re-combination process (process 2). The radiative relaxation gives a broad band emission of Ce\(^{3+}\):5d\(_{1}\) \(\rightarrow\) 2\(F\)_\(^{5/2}\), 2\(F\)_\(^{7/2}\), and the resonant ET process takes place nearly at the same time to the Er\(^{3+}\) ion (process 3), which is followed by rapid multi-phonon relaxation down to the \(4S\)\(_{3/2}\) and \(4I\)\(_{13/2}\) excited levels leading to the sharp luminescence bands of Er\(^{3+}\): \(4S\)\(_{3/2}\) \(\rightarrow\) \(4I\)\(_{15/2}\) in the green and \(4I\)\(_{3/2}\) \(\rightarrow\) \(4I\)\(_{15/2}\) in the NIR region.

4. Conclusion

In summary, we have successfully developed a blue-light-chargeable NIR persistent phosphor (Y\(_3\)Al\(_2\)Ga\(_3\)O\(_{12}\):Er\(^{3+}\), Ce\(^{3+}\), Cr\(^{3+}\)) with long (> 10 h) persistent luminescence due to the typical Er\(^{3+}\):\(4I\)\(_{13/2}\) \(\rightarrow\) \(4I\)\(_{15/2}\) transition ranging from 1450 nm to 1670 nm in garnet. The NIR PersL bands from Er\(^{3+}\) through efficient persistent energy transfer from Ce\(^{3+}\) match well with the NIR-III window, and its photon emission rate \((8.33 \times 10^{17} \text{ cps Sr}^{-1} \text{ m}^{-2})\) at 10 min after ceasing blue light (460 nm) illumination was over two times higher than that of the widely used ZnGa\(_2\)O\(_4\):Cr\(^{3+}\) deep-red persistent phosphor \((3.30 \times 10^{17} \text{ cps Sr}^{-1} \text{ m}^{-2})\). We also showed the first persistent luminescence imaging with high intensity and long duration by a commercial InGaAs camera. In vivo bio-imaging in the NIR-III window with improved optical resolution quality and deep tissue penetration depth can be expected in the near future by using this material (in the form of nano-particles) as a functionalized bio-probe.

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