

SCIENTIFIC REPORTS



OPEN

Hes1 expression in mature neurons in the adult mouse brain is required for normal behaviors

Tadanobu Matsuzaki^{1,2}, Toru Yoshihara³, Toshiyuki Ohtsuka^{1,2,4} & Ryoichiro Kageyama^{1,2,4,5}

Hes1 regulates the maintenance and proliferation of neural stem/progenitor cells as an essential effector of the Notch signaling pathway. Although Notch signaling is also involved in the functions of mature neurons in learning and memory and in the risk factors for mental disorders such as schizophrenia and bipolar disorder, the *in-vivo* role of Hes1 in mature neurons remains unknown. Here, we found that Hes1 is expressed by subsets of both excitatory and inhibitory neurons in the adult mouse brain, and that Hes1 expression is induced by neuronal stimulation. Furthermore, inactivation of *Hes1* in excitatory neurons resulted in abnormal fear and anxiety behaviors concomitantly with higher neuronal excitability in the amygdala, while inactivation of *Hes1* in inhibitory neurons resulted in increased sociability and perseverative tendencies. These results indicated that Hes1 is functionally important for normal behaviors not only in excitatory neurons but also in inhibitory neurons in the adult brain.

Notch signaling regulates the maintenance and proliferation of neural stem/progenitor cells and glial versus neuronal differentiation in the developing nervous system¹. Recent studies suggest that Notch signaling is also involved in neuronal activity during learning and memory^{2–5} and in mental disorders such as schizophrenia and bipolar disorder^{6–10}. Indeed, the Notch intracellular domain (NICD), an active form of the Notch protein, is expressed by subsets of neurons in many regions of the brain, including the cerebral cortex and hippocampus, and inactivation of Notch signaling in the postnatal hippocampal neurons leads to memory defects^{4,5}. However, the mechanism by which Notch signaling regulates such neuronal activities remains unknown.

Notch is a transmembrane protein, which upon activation by its ligands, is cleaved at the transmembrane region, releasing NICD. NICD is then transferred into the nucleus, where it forms a complex with the DNA-binding factor Rbpj and the transcriptional co-activator Mastermind-like¹. This complex activates the expression of various downstream genes such as *Hes1* and *Hes5*, which encode basic helix-loop-helix transcriptional repressors¹. As an essential Notch signaling effector, Hes1 plays important roles in the maintenance and proliferation of neural stem/progenitor cells and glial versus neuronal differentiation in the developing nervous system^{11–13}. It was previously shown that Hes1 is also expressed by cultured neurons and controls the neuronal activity¹⁴. Hes1 represses the expression of GluR1, an AMPA receptor required for synaptic plasticity, and thereby inhibits GluR1-mediated calcium influx in cultured cortical neurons¹⁴. Furthermore, Hes1 antagonizes the effects of β -amyloid and increases the survival of cultured hippocampal neurons¹⁵. These results provide evidence that Hes1 regulates *in vitro* neuronal activity, but the *in vivo* functions of Hes1 in the adult brain still remain to be analyzed.

To understand the role of Hes1 in mature neurons in the adult brain, we examined Hes1 expression in the adult mouse brain. Furthermore, we genetically ablated the Hes1 gene in mouse neurons. Because conventional *Hes1* knock-out (KO) leads to premature death¹⁶, we generated *Hes1* conditional KO (cKO) mice by taking advantage of Cre/LoxP recombination system. We crossed *Hes1* flox mice with either *Math2*-Cre mice or *Gad2*-Cre mice, which express Cre recombinase in excitatory or inhibitory neurons, respectively. *Hes1* is therefore deleted in all excitatory neurons in the former cKO mice, and it is deleted in all inhibitory neurons in the latter cKO mice.

¹Institute for Frontier Life and Medical Sciences, Kyoto University, Kyoto, 606-8507, Japan. ²Graduate School of Medicine, Kyoto University, Kyoto, 606-8501, Japan. ³Medical Research Support Center, Graduate School of Medicine, Kyoto University, Kyoto, 606-8501, Japan. ⁴Graduate School of Biostudies, Kyoto University, Kyoto, 606-8501, Japan. ⁵Institute for Integrated Cell-Material Sciences (iCeMS), Kyoto University, Kyoto, 606-8501, Japan. Correspondence and requests for materials should be addressed to R.K. (email: rkageyam@infront.kyoto-u.ac.jp)

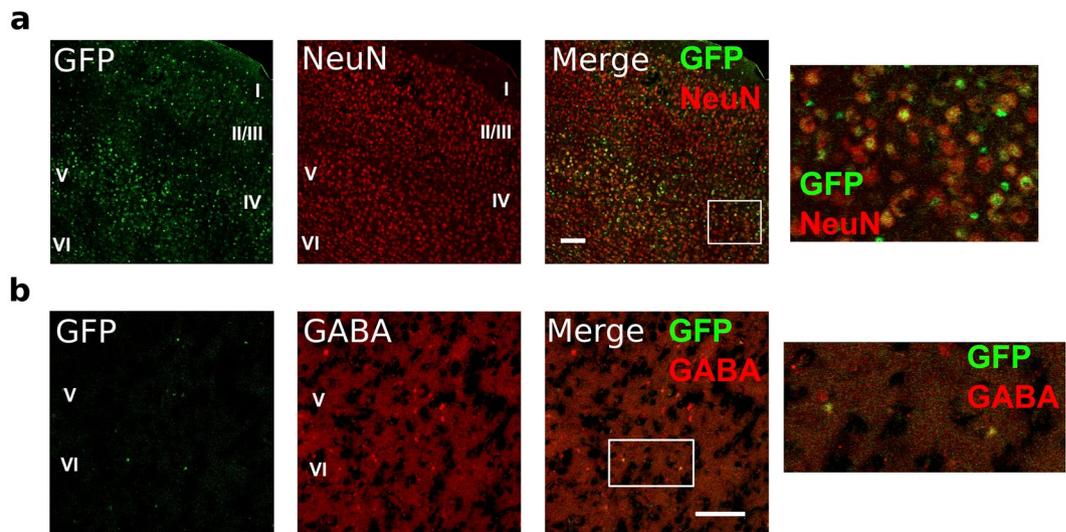


Figure 1. Hes1 expression in neurons in the adult mouse brain (10 weeks old). **(a)** Double immunofluorescence staining of GFP (green, with TSA amplification) and NeuN (red) in a coronal section of the primary sensory cortex of a Venus-Hes1 knock-in mouse. **(b)** Double immunofluorescence staining of GFP (green, without TSA amplification) and GABA (red) in a coronal section of the primary motor cortex of a Venus-Hes1 knock-in mouse. Boxed regions are enlarged on the right. Cortical layers I–VI are labeled. Scale bars = 100 μm .

We performed behavior analyses of these cKO mice and found that Hes1 is functionally important for normal behaviors not only in excitatory neurons but also in inhibitory neurons in the adult brain.

Results

Hes1 is expressed by both excitatory and inhibitory neurons. We first analyzed the expression pattern of Hes1 in the cerebral cortex of adult mice. To this end, we used Venus-Hes1 fusion knock-in mice, in which Venus (GFP variant) cDNA was knocked-in in-frame into the 5' region of the Hes1 gene so that a Venus-Hes1 fusion protein was expressed¹⁷. In these mice, Venus expression, detected with a GFP antibody, correlated very well with endogenous Hes1 (ref.¹⁷). In the cortex of adult mice, Hes1 was expressed at low levels in subsets (approximately 3–15%) of NeuN⁺ neurons (Fig. 1a).

We next tried to perform immunostaining for Venus-Hes1 and excitatory pyramidal neuron-specific markers, but it was technically difficult (data not shown), because Venus-Hes1 expression levels were very low in neurons. To overcome this difficulty, we adopted an alternative approach. *Venus-Hes1* floxed;*Hes1* floxed mice^{17,18} were crossed with *Math2-Cre* mice, in which Cre is expressed specifically in post-mitotic excitatory neurons from around embryonic day (E)11 (ref.¹⁹). In the resultant *Venus-Hes1* floxed;*Hes1* floxed;*Math2-Cre* mice, residual Venus expression occurred under the control of the endogenous *Hes1* promoter in Cre-positive cells (*Math2*-expressing excitatory neurons) (Supplementary Fig. S1a). Hes1 expression is normally kept at very low levels by negative feedback, in which Hes1 represses its own expression by directly binding to its promoter²⁰. However, because there was no Hes1 protein in these Cre-positive cells (Supplementary Fig. S1a), Venus protein should be overexpressed without negative feedback. Indeed, Venus protein was clearly expressed in many neurons in the cortex (Supplementary Fig. S1b) and the hippocampus (Supplementary Fig. S1c). Higher magnification showed that Venus was highly expressed in many pyramidal neurons in the cerebral cortex of adult mice (Supplementary Fig. S1d). Thus, Hes1 is expressed by *Math2*-expressing excitatory neurons *in vivo*, but is maintained at very low levels by negative feedback. Low levels of Hes1 were also detected in subsets (approximately 30%) of GABA⁺ inhibitory neurons in the cortex (Fig. 1b). These results indicated that Hes1 is expressed by subsets of both excitatory and inhibitory neurons in the adult mouse brain.

Hes1 expression is induced in neurons by stimulation. The Hes1 expression patterns in cortical neurons differed among mice, which led us to suspect that Hes1 expression is temporally dynamic and induced by certain stimuli. It is well known that immediate early genes (IEG), such as *Arc* and *c-Fos*, are expressed promptly after cell stimulation²¹. To test the possibility that Hes1 expression might be also induced by neuronal stimulation, we focused on the visual cortex and assessed whether Hes1 expression can be induced by visual stimulation. When mice were kept in the dark for three days, there were only few Hes1⁺ neurons in the visual cortex (Fig. 2a, upper panel). Furthermore, *Arc* and *c-Fos* were mostly negative in this region (Fig. 2a, upper panels). However, both *Arc* and *c-Fos* expression were robustly induced 2 hours after light stimulation (~70-fold and ~30-fold increase, respectively, in cell number) (Fig. 2a, lower panels), consistent with the previous reports^{22,23}. Hes1 expression was also induced at the same time point, although in fewer cells (~5-fold increase in cell number) (Fig. 2a).

We next performed electro-convulsive stimulation, another well-known method to induce IEG expression^{24,25}, to see whether broader stimulation of cortical neurons leads to broader expression of Hes1. Trans-cranial electric stimulation was administered to mice, who exhibited general seizures. Two hours after electric stimulation, in

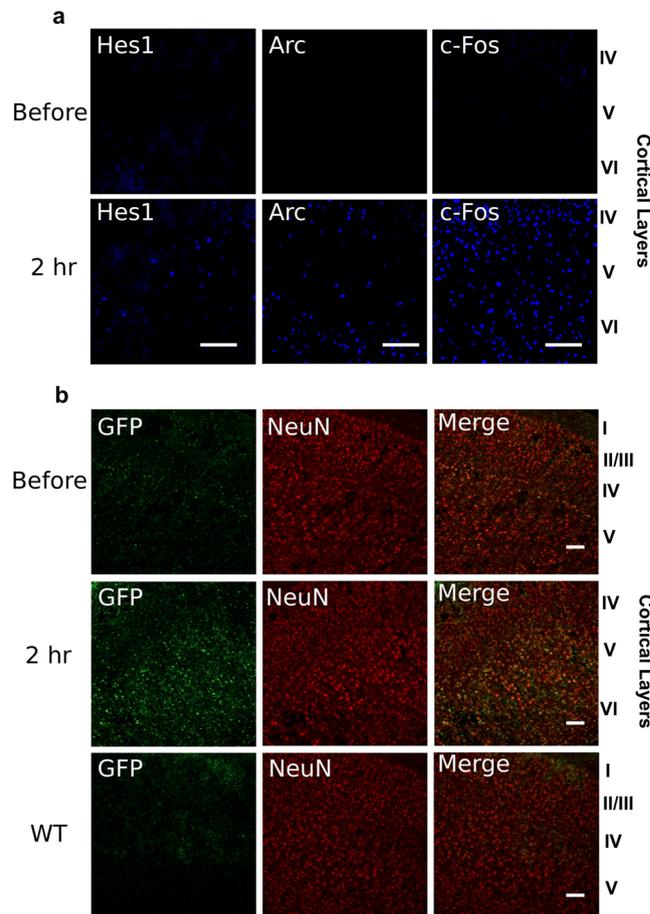


Figure 2. Induction of *Hes1* expression by neuronal stimulation. **(a)** Immunofluorescence staining for *Hes1* (left panels), *Arc* (middle panels), and *c-Fos* (right panels) in the primary visual cortex of wild-type mice before (upper panels) and 2 hours after (lower panels) visual stimulation. **(b)** Double immunofluorescence staining for GFP (left panels), NeuN (middle panels), and merged images (right panels) in the primary sensory cortex of Venus-*Hes1* knock-in mice before (upper panels) and 2 hours after (middle panels) electroconvulsive stimulation. Wild-type (WT) mice were shown as a negative control of immunostaining for GFP (lower panels). Cortical layers I–VI are labeled on the right. Scale bars = 100 μm .

addition to *Arc* and *c-Fos* expression (data not shown), *Hes1* expression was broadly induced in NeuN⁺ neurons in the cerebral cortex (~2-fold increase in cell number) (Fig. 2b). These results indicated that *Hes1* expression can be induced in neurons by stimulation.

Abnormal fear and anxiety in excitatory neuron-specific *Hes1* cKO mice. To analyze the function of *Hes1* in the adult brain, we deleted *Hes1* specifically from excitatory neurons. To this end, we mated the *Math2-Cre* driver line with the *Hes1*-floxed line, in which loxP sequences are located at the first intron and 3' UTR of the *Hes1* gene (Supplementary Fig. S1a)¹⁸. The resulting cKO (*Math2-Cre;Hes1* cKO) mice (10 weeks old) were then subjected to a series of behavioral analyses. These cKO mice displayed no significant difference in general health, spatial memory, working memory, sociability, depression-like behavior, or sensorimotor gating, compared to wild-type control mice (Supplementary Figs S2–S4). Furthermore, the freezing response at the baseline level was not significantly different between the wild type and cKO mice (Fig. 3a, baseline). However, we found that the cKO mice displayed abnormal behaviors in the following tests.

In the fear conditioning test, these cKO mice were moved to a chamber and conditioned to associate an auditory cue with a foot shock (see Methods). The next day, they were placed in the same chamber without an auditory cue (contextual recall test) or in a different chamber with an auditory cue (cued recall test). *Math2-Cre;Hes1* cKO mice exhibited no significant differences in freezing time in the first contextual recall test or in the cued recall test, compared to wild-type control mice (Fig. 3a). The second contextual recall and cued recall tests were performed six months later. While *Math2-Cre;Hes1* cKO mice still exhibited no significant difference in the second contextual recall test compared with wild-type control mice, they did exhibit significantly longer freezing time in the second cued recall test than did the wild-type control mice (Fig. 3a). These results indicated that the mutant mice exhibited abnormal retention of fear memory in an auditory cued test.

In an open field test, mice were placed in a bright open-field apparatus (see Methods). While the total traveled distance was not significantly different between *Math2-Cre;Hes1* cKO and wild-type control mice (Fig. 3b, upper panel), the cKO mice spent a significantly shorter time in the center area than did the wild-type control mice

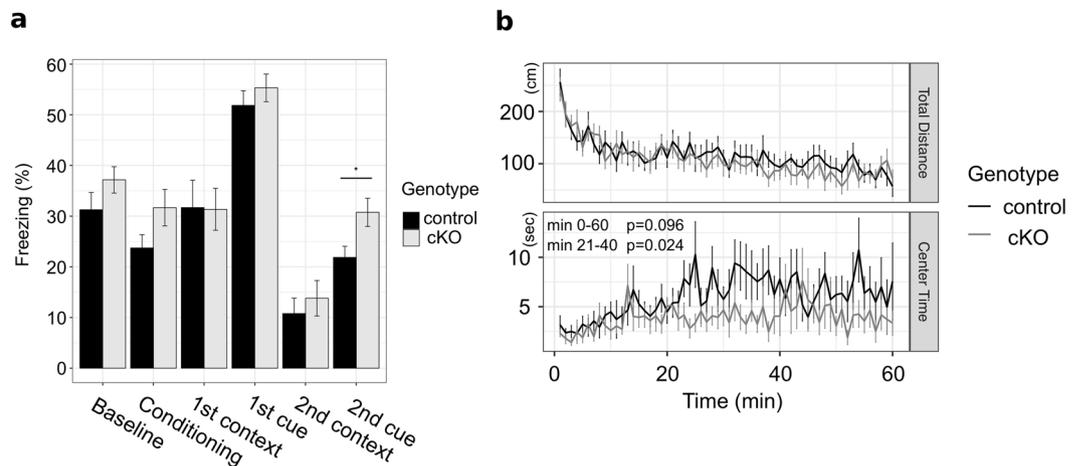


Figure 3. Inactivating *Hes1* in excitatory neurons leads to abnormal retention of fear memory and a slight increase in anxiety level. **(a)** Bar plots showing freezing time of wild-type control (black, $n = 17$) and *Math2-Cre;Hes1* cKO (gray, $n = 17$) mice during fear conditioning, context recall, and cued recall of the fear conditioning test as well as before fear conditioning (baseline). Interval between 1st and 2nd tests was 6 months. Data are presented as mean \pm SEM; * $p < 0.05$; two tailed Student's *t*-test. **(b)** Line plots showing total distance traveled and time spent in the center during the open field test for wild-type control (black, $n = 17$) and *Hes1* cKO (gray, $n = 17$) mice. Data are presented as mean \pm SEM; the *p*-values indicate a genotype effect in the two-way repeated measures ANOVA.

(Fig. 3b, lower panel), suggesting that they had a mild increase in anxiety levels. Thus, inactivation of *Hes1* in excitatory neurons led to abnormal fear and anxiety, suggesting that *Hes1* plays an important role in the function of post-mitotic excitatory neurons in the adult brain.

Higher excitability in the amygdala of excitatory neuron-specific *Hes1* cKO mice. Behavioral analyses suggested that *Math2-Cre;Hes1* cKO mice had increased fear retention and anxiety levels. To determine the brain regions responsible for these abnormal fear and anxiety behaviors, we compared the neuronal excitabilities of the cKO and wild-type control mice. In particular, we focused on the medial pre-frontal cortex (mPFC) and amygdala - two regions that are reported to be important for fear memory²⁶⁻³⁰. We examined *c-Fos* expression to visualize the neuronal excitability in these two regions after the second recall with auditory cue in the fear conditioning test described above. In the mPFC, there was no significant difference in *c-Fos* expression levels between *Math2-Cre;Hes1* cKO and wild-type control mice (Fig. 4a, upper panels, and b, upper panel). However, there were more *c-Fos*⁺ neurons in the basolateral amygdala (BLA) of *Math2-Cre;Hes1* cKO mice than in that of wild-type control mice (Fig. 4a, lower panels, and b, lower panel). We confirmed that there were some *Hes1*⁺ cells in the BLA of wild-type control mice but approximately 60% less in that of the cKO mice (Fig. 4c). These results suggest that *Hes1* may play an important role in the activity of mature neurons in the amygdala.

Increased sociability and perseveration in inhibitory neuron-specific *Hes1* cKO mice. We next examined the role of *Hes1* in inhibitory neurons in the adult brain. To this end, we crossed mice of the *Hes1*-floxed line with those of the *Gad2-Cre* driver line, in which *Cre* is expressed specifically in inhibitory post-mitotic neurons³¹. The resulting cKO (*Gad2-Cre;Hes1* cKO) mice (10 weeks old) were subjected to a series of behavioral analyses. These cKO mice were normal in general health, working memory, depression-like behavior, sensorimotor gating, fear, and anxiety (Supplementary Figs S5 and S6), but differed from wild-type control mice in the following tests.

In Crawley's social interaction test, a subject mouse was placed for habituation in the middle of a three-chambered apparatus, and then a stranger mouse was placed in the left chamber (see Methods). After a while, another stranger mouse was placed in the right chamber. We found that *Gad2-Cre;Hes1* cKO mice contacted both stranger mice more frequently and spent more time with these stranger mice than wild-type control mice (Fig. 5a), suggesting that the mutant mice showed enhanced social interactions with novel mice.

In the Barnes maze test, spatial memory was measured using a round table with 12 holes, one of which was an escape hole (see Methods). Mice went through training sessions to memorize the position of the escape hole. On day 1 (Test 1) and day 8 (Test 2) after the training sessions, probe tests were performed using the same table but without an escape hole. In Test 1, *Gad2-Cre;Hes1* cKO mice spent a longer time near the escape hole than wild-type control mice, but not in Test 2 (Fig. 5b), suggesting that the mutant mice showed perseveration on day 1.

Together, these results indicated that inactivation of *Hes1* in inhibitory neurons led to increased sociability and perseveration. Thus, *Hes1* is functionally important for normal behaviors not only in excitatory neurons but also in inhibitory neurons in the adult brain.

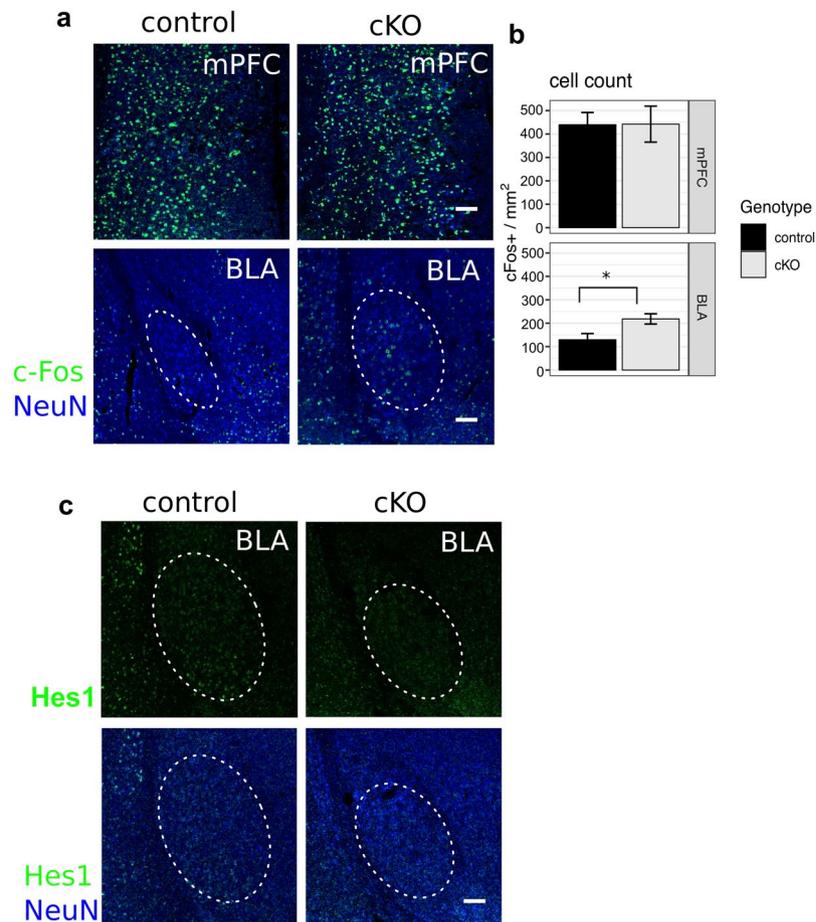


Figure 4. Cell excitability in the amygdala differs in excitatory neuron-specific Hes1 cKO mice. **(a)** Double immunofluorescence staining for c-Fos (green) and NeuN (blue) in the medial pre-frontal cortex (mPFC, upper panels) and basolateral amygdala (BLA, lower panels) of wild-type control and Math2-Cre;*Hes1* cKO mice. We first performed fear conditioning experiment, and then examined c-Fos expression 2 hours after the second cued recall test. Scale bars = 100 μ m. **(b)** Bar graphs showing the density of c-Fos-positive neurons in mPFC and BLA of wild-type control (black, n = 4) and Math2-Cre;*Hes1* cKO (gray, n = 4) mice. Data are presented as mean \pm SEM; *p < 0.05; two tailed Student's *t*-test. **(c)** Double immunofluorescence staining for Hes1 (green) and NeuN (blue) in the BLA of wild-type control and Hes1 cKO mice 2 hours after the auditory cue. Scale bar = 100 μ m.

Discussion

Hes1 is a well-known effector of Notch signaling that regulates the maintenance and proliferation of neural stem/progenitor cells in the developing nervous system. In this study, we first showed that Hes1 is also expressed by both excitatory and inhibitory mature neurons in the adult mouse brain, and that Hes1 expression is induced by neuronal stimulation *in vivo*. Furthermore, inactivation of *Hes1* in excitatory or inhibitory neurons in the mouse brain led to abnormal behaviors, indicating that Hes1 is functionally important in both excitatory and inhibitory neurons *in vivo*. However, it remains to be determined whether Hes1 expression is controlled by Notch signaling in these neurons. Hes1 expression might be regulated by other signaling pathways. For example, cAMP and Ca²⁺ signaling molecules activate CREB transcription factor, which plays an important role in neuronal plasticity and protection³². CREB induces Hes1 expression in neural stem/progenitor cells³³, which raises the possibility that cAMP and Ca²⁺ signaling can also regulate Hes1 expression in neurons, and therefore Hes1 expression might be regulated by various signaling molecules in neurons.

Inactivation of *Hes1* in excitatory neurons of adult mice led to abnormal fear and anxiety behaviors concomitantly with higher neuronal excitability in the amygdala. It was previously reported that Hes1 overexpression in cultured neurons increased GABAergic input¹⁴; therefore, inactivation of *Hes1* may decrease GABAergic input and increase neuronal excitability in the amygdala. In addition, because Hes1 represses GluR1 expression¹⁴, inactivation of *Hes1* may increase GluR1-mediated calcium influx and thereby enhance the excitability of neurons. Further study is required to elucidate the mechanism of how the excitability changes in the amygdala in the absence of Hes1 in excitatory neurons, and whether the altered excitability observed in the amygdala is directly or indirectly involved in the behavioral phenotype. The increased anxiety level observed in the open field test was not captured in other analyses, such as the light-dark transition test (Supplementary Fig. S3a,b) and elevated-plus

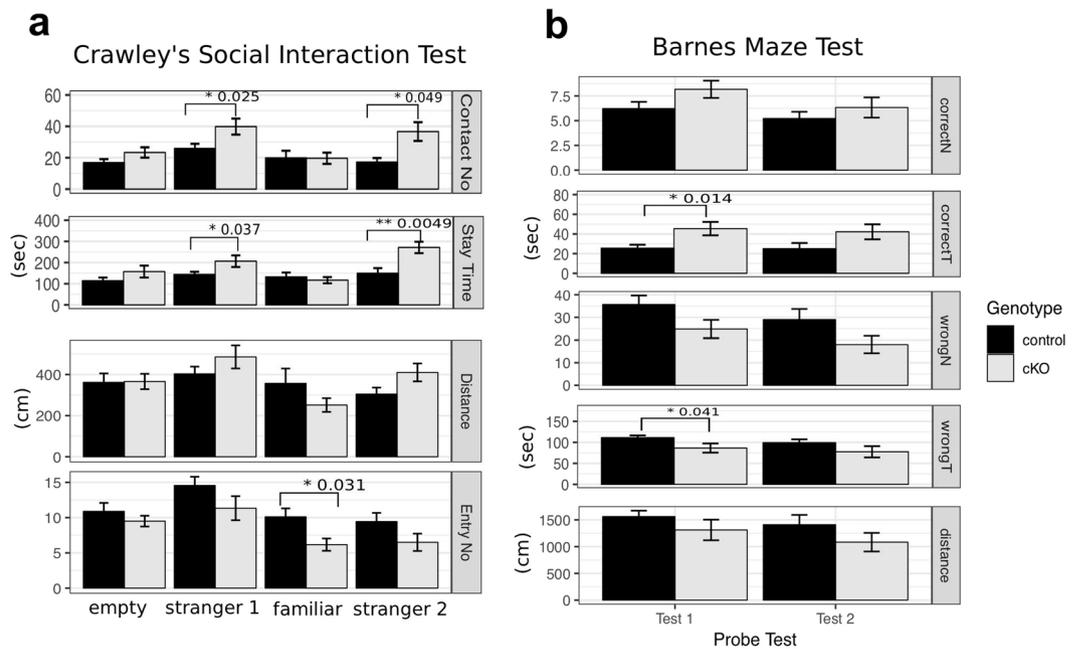


Figure 5. Inhibitory neuron-specific inactivation of *Hes1* leads to enhanced sociability and perseveration. **(a)** Crawley's social interaction test. The number of contacts (Contact No), time spent in each compartment (Stay Time), distance traveled in each compartment (Distance), and number of entries into each compartment (Entry No) were recorded for wild-type control ($n = 9$) and *Gad2-Cre;Hes1* cKO ($n = 6$) mice. Data are presented as mean \pm SEM; * $p < 0.05$, ** $p < 0.01$; two tailed Student's *t*-test. **(b)** Barnes maze test. The number of visits to the correct hole (correctN) and wrong holes (wrongN), time spent in the correct hole (correctT) and wrong holes (wrongT), and total distance traveled (distance) were measured for wild-type control (black, $n = 9$) and *Gad2-Cre;Hes1* cKO (gray, $n = 6$) mice. Data are presented as mean \pm SEM; * $p < 0.05$; two tailed Student's *t*-test.

maze test (Supplementary Fig. S4e). Thus, the effect of *Hes1* on anxiety is relatively mild, if any, and increased anxiety might not be caused directly by *Hes1* inactivation but rather may be secondary to strongly retained fear memory.

Inactivation of *Hes1* in excitatory neurons led to abnormal retention of fear memory, which is similar to the symptom of posttraumatic stress disorder (PTSD). It is known that even when exposed to the same traumatic stress, some people develop symptoms of PTSD while others do not³⁴. This difference may result from different individual characteristics, such as sex, genetics, prior trauma, or age, and *Hes1* cKO mice possibly have a predisposition for PTSD. PTSD is a chronic and debilitating disorder, whose pathology is not yet fully understood³⁵. A high comorbidity of PTSD with other diseases is making it difficult to accurately understand its pathogenesis. This fundamental lack of understanding of PTSD makes it harder to select or develop effective drugs for this disease. Empirical pharmacological therapies include selective serotonin reuptake inhibitors, which are partially effective³⁶. So far, plausible explanations of PTSD pathogenesis consist of monoamine-based and glutamate-based models³⁵. Our mice exhibited increased fear memory after we tampered with their excitatory neurons, which may agree with the latter model. Further analysis of *Hes1* function in excitatory neurons might shed light on the pathophysiology of PTSD and lead to innovative treatment of the disease. The mouse line we created mainly exhibited enhanced fear memory, which we believe may be useful in elucidating the core pathology of the disease.

We also found that inactivating *Hes1* in inhibitory neurons leads to enhanced sociability and perseveration behavior. The social interaction test and the second probe test of the Barnes maze test also demonstrated a similar tendency, although the difference was not significant. In addition, the cKO mice demonstrated slightly higher stereotypic activity during the first five minutes ($p = 0.054$) of the open field test (Supplementary Fig. S5e), suggesting that these mice may be hyperattentive. One plausible explanation for these phenotypes is that the hyperattentive state urges the mice to explore stranger mice more carefully, as observed in Crawley's social interaction test, and to perseverate on their memory, as observed in Barnes maze test. Together, our results indicated that *Hes1* expressed in both excitatory and inhibitory neurons plays an important role in the adult brain functions and suggested that two types of *Hes1* cKO mice described here might be useful for analyzing various neurological disorders.

Methods

Animals. All animals were handled in accordance with the Kyoto University Guide for the Care and Use of Laboratory Animals. The experimental protocols were approved by the Experimental Animal Committee of Institute for Frontier Life and Medical Sciences, Kyoto University. Mice were housed in a temperature-controlled environment under a 12h light/dark cycle, with food and water provided *ad libitum*. Generation of *Hes1* floxed (*Hes1*f) mice

and Hes1-Venus mice were described previously^{17,18}. Math2-Cre and Gad2-Cre mice^{19,31} were obtained from Dr. Klaus-Armin Nave and Jackson Laboratory (JAX stock #28867), respectively. Excitatory neuron-specific *Hes1* cKO mice were obtained by crossing Hes1f/Hes1f mice with Hes1f/Hes1f; Math2-Cre mice. Inhibitory neuron-specific *Hes1* cKO mice were obtained by crossing Hes1f/Hes1f mice with Hes1f/Hes1f; Gad2-Cre mice.

Genotyping protocol. For genotyping polymerase chain reaction (PCR), tissues lysed with proteinase K were put into reaction with Emerald Amp master mix (Takara Bio, Japan) using specific primer sets for each genotype. Primer sets used for each genotype were as follows:

Hes1-floxed (224 bp for WT, 272 bp for floxed mice)
 F: 5'-CAGCCAGTGTCAACACGACACCGGACAAAC-3'
 R: 5'-TGCCCTTCGCCTCTTCTCCATGATA-3'
 Math2-Cre (525 bp)
 F: 5'-GAGTCCTGGAATCAGTCTTTTTTC-3'
 R: 5'-CCGCATAACCAGTCAAACAG-3'
 Gad2-Cre (176 bp)
 F: 5'-CACTGCATTCTAGTTGTGGTTTG-3'
 R: 5'-AACAGTTTGATGAGTGAGGTGA-3'
 Venus-Hes1 (336 bp)
 F: 5'-GTAAACGGCCACAAGTTCAGCGTGTC-3'
 R: 5'-GTCCTCCTGAAGTCGATGCCCTTCAG-3'

Light stimulation and electroconvulsive stimulation. For visual stimulation experiment, each mouse was first kept in a dark cage for 3 consecutive days and then placed in a bright cage with stripe patterns on the wall for 2 hours, after which mice were sacrificed. Control group was kept in dark cages for 3 days and then sacrificed with minimum light exposure.

For electroconvulsive stimulation experiment, each mouse was administered with 0.1 ms, 15 mA square-wave pulse at 100 Hz for 1 second through electrodes placed on both ears using *in vivo* electroporator (CUY21EDIT, BEX Co., Ltd). Mice that showed tonic-clonic convulsion were considered successful and sacrificed 2 hours after stimulation.

Tissue preparation and immunohistochemistry. Mice were treated intraperitoneally with pentobarbital, and perfused transcardially with 30 ml of PBS and 30 ml of 4% paraformaldehyde (PFA)/PBS (pH 7.2). Brains were post fixed in the 4% PFA/PBS overnight at 4 °C and then cryoprotected for 5 hr in 15% sucrose/TBS and 48 h in 30% sucrose/TBS. Brains were embedded in OCT compound (Sakura Finetek), and frozen at -80 °C. Free-floating brain sections were made at 40 µm thickness with freezing microtome (Leica CM 1950). Sections were incubated in TBS containing 3% H₂O₂ for 15 min, 0.5% Triton X-100/TBS for 30 min, 3% normal donkey serum (NDS) and 0.1% Triton X-100/TBS for 2 h at room temperature, then with primary antibodies, rabbit anti-GFP (1:1000, Invitrogen A11122), chicken anti-GFP (1:1000, Abcam ab13970), mouse anti-NeuN (1:200, Merck Millipore MAB377), rabbit anti-Hes1 (1:1000, ref.³⁷), rabbit anti-Arc (1:500, Synaptic Systems 156003), and rabbit anti-c-Fos (1:1000, Synaptic Systems 226003) diluted in 0.1% Triton X-100/TBS containing 3% NGS for 3 days at 4 °C. After incubation with secondary antibodies (HRP-donkey anti-rabbit IgG (1:500; GE Healthcare), Alexa 488 or Alexa 594 (1:400, invitrogen) for 2 h at room temperature, sections were reacted with TSA Fluorescein staining solution (Perkin Elmer) for 15 min at room temperature, mounted with Fluormount-G (Southern Biotech) and photographed with laser-scanning confocal microscopy (LSM510, Zeiss).

Behavioral analysis. Behavioral analyses were performed, as previously described³⁸. Mice were genotyped at the time of weaning and group-housed with four mice per cage, two control and two cKO littermates. All the behavioral analysis was performed between 9:00 a.m. and 5:00 p.m.

General health and neurological screening. Motor function was assessed with wire hang test and grip strength test. In a wire hang test, each mouse was placed on a cage top, which was then inverted to make the mouse hang on to the cage top. Latency of each mouse to fall was measured. In a grip strength test, strength was measured by pulling the tail of each mouse while its front paw grabbing the measurement apparatus.

Barnes maze. Spatial memory was measured using Barnes circular maze (1.0 m in diameter, 12 holes, O'Hara & Co., Ltd., Tokyo, Japan). Following a habituation phase, each mouse went through 15 training sessions, each lasting for 5 minutes, to memorize the position of one escape hole, which was consistent for a given mouse, but was randomized across mice. After finishing training sessions, 2 probe tests were performed with 7 days interval, each test lasting for 3 minutes. During a training session, the total distance, number of visits around correct and wrong holes, and latency to enter correct hole were recorded. During a probe test, the total distance, number of visits and time spent around correct and wrong holes were recorded.

Social interaction test. To assess social behavior, a pair of mice was placed in the same cage for 10 minutes. Pairs were chosen with following criteria: same genotype, housed in different cages, and less than 10% difference of body weight. The total duration of contacts, total duration of active contacts, mean duration of contacts, number of total contacts, and total distance traveled were recorded. Active contact was defined as a contact, which occurred after moving speed faster than 10.0 cm/sec.

Crawley's social interaction test. Social cognition was assessed using a three-chambered apparatus (TimeCSI for Three-chambered social interaction system O'Hara & Co., Ltd., Tokyo, Japan). First, a subject mouse was placed in the center chamber with the other chambers empty for habituation, and then a stranger mouse was placed in the left chamber to assess general sociability. After that, another yet stranger mouse was placed in the right chamber to assess social novelty recognition. The number of contacts, stay time, distance traveled, and number of entries were recorded for each session.

Elevated plus maze. Anxiety level was assessed using elevated plus maze, which has one closed arm and one open arm. Each mouse was first placed at the center of the maze and behavior was monitored for 10 minutes. Time spent in each arm, total distance traveled, and number of entries in each arm were recorded.

Light-dark transition test. Light-dark transition test were performed using an apparatus, which consists of a bright chamber (390 lux) and a dark chamber (2 lux), and mice were allowed to move freely between the two chambers. Time spent in each chamber, total distance traveled, and latency to enter bright chamber were recorded.

Prepulse inhibition. Sensorimotor gating function was measured by prepulse inhibition test. A 120 dB white noise was used to evoke startle response, and the inhibition of response by preceding prepulse (70 dB, 75 dB, 80 dB, 85 dB) was measured. In addition, a startle response to a single noise (90 dB, 100 dB, 110 dB, 120 dB) was measured to assess auditory and motor function as acoustic startle response (ASR).

Porsolt forced swim test. Depression-like behavior was assessed using cylindrical aquarium. Each mouse was placed in the aquarium to swim freely and the behavior was monitored for 10 minutes. The experiment was done once a day for two consecutive days. Immobile time and distance traveled were measured.

Rotarod test. Locomotor activity was assessed using apparatus with a rotating rod (UGO BASILE, VA, Italy). Each mouse was placed on top of rod, which rotates at gradually accelerating rate. Latency of the mouse to fall was recorded.

Y-maze test. Working memory was assessed with a Y-shaped maze, each arm placed at 120 degrees angle from adjacent arms. Each mouse was first placed at the center of the maze and the following behavior was monitored for 5 minutes. The number of entries into arms, alternation rate, and total distance traveled were recorded.

Tail suspension test. In a tail suspension test, each mouse was hanged by its tail for 10 minutes, and the behavior is monitored. Immobile time was recorded.

Open field test. Locomotor activity was measured using an open-field test. Each mouse was placed in the corner of the open-field apparatus (40 × 40 × 30 cm; Accusan Instruments, Columbus, OH). The test chamber was illuminated at 100 lux. The total distance traveled, vertical activity (rearing measured by counting the number of photo beam interruptions), time spent in the center area, and beam-break counts for stereotypic behaviors were recorded. The center area was defined as 1 cm away from the walls. Data were collected for 120 minutes.

Fear conditioning test. Fear memory was assessed with a fear conditioning test. Each mouse was conditioned to associate an auditory cue (55 dB white noise for 30 seconds) with a foot shock (0.35 mA shock for 2 seconds), by administering 3 pairs of an auditory cue and a foot shock. Recall tests were done the next day and certain days after conditioning. In a contextual recall test, each mouse was placed in the same chamber as conditioning but without foot shocks, and behavior was monitored for 5 minutes. In a cued recall test, each mouse was placed in the different chamber as conditioning, and after 3 minutes of observation, the auditory cue was presented for another 3 minutes. Freezing time was recorded in each session.

Statistical analysis. Statistical analyses were performed using the computing environment R (R Development Core Team). Statistical differences between two groups were analyzed using two-tailed Student's t-test, and the differences between the two groups at different time points were analyzed using two-way repeated measures ANOVA. P values less than 0.05 were considered to be significant.

References

- Shimojo, H., Maeda, Y., Ohtsuka, T. & Kageyama, R. Dynamic Notch signaling in neural progenitor cells in *Cortical Development: Neural Diversity and Neocortical Organization* (eds Kageyama, R. & Yamamori, T.) 1–17 (Springer, 2013).
- Costa, R. M., Honjo, T. & Silva, A. J. Learning and memory deficits in Notch mutant mice. *Cur. Biol.* **13**, 1348–1354 (2003).
- Wang, Y. *et al.* Involvement of Notch signaling in hippocampal synaptic plasticity. *Proc. Natl. Acad. Sci. USA* **101**, 9458–9462 (2004).
- Saura, C. A. *et al.* Loss of presenilin function causes impairments of memory and synaptic plasticity followed by age-dependent neurodegeneration. *Neuron* **42**, 23–36 (2004).
- Alberi, L. *et al.* Activity-Induced Notch Signaling in Neurons Requires *Arc/Arg3.1* and Is Essential for Synaptic Plasticity in Hippocampal Networks. *Neuron* **69**, 437–444 (2011).
- Wei, J. & Hemmings, G. P. The NOTCH4 locus is associated with susceptibility to schizophrenia. *Nat. Genet.* **25**, 376–377 (2000).
- Gregorio, S. P. *et al.* Analysis of coding-polymorphisms in NOTCH-related genes reveals NUMBL poly-glutamine repeat to be associated with schizophrenia in Brazilian and Danish subjects. *Schizophrenia Res.* **88**, 275–282 (2006).
- Stefansson, H. *et al.* Common variants conferring risk of schizophrenia. *Nature* **460**, 744–747 (2009).
- Aukrust, P., Andreassen, O. A. & Ueland, T. Up-regulation of NOTCH4 gene expression in bipolar disorder. *Am. J. Psychiatry* **169**, 1292–1300 (2012).
- Hoseth, E. Z. *et al.* Attenuated Notch signaling in schizophrenia and bipolar disorder. *Sci. Rep.* **8**, 5349 (2018).
- Ohtsuka, T. *et al.* Hes1 and Hes5 as notch effectors in mammalian neuronal differentiation. *EMBO J.* **18**, 2196–2207 (1999).
- Hatakeyama, J. *et al.* Hes genes regulate size, shape and histogenesis of the nervous system by control of the timing of neural stem cell differentiation. *Development* **131**, 5539–5550 (2004).

13. Imayoshi, I. & Kageyama, R. bHLH factors in self-renewal, multipotency, and fate choice of neural progenitor cells. *Neuron* **82**, 9–23 (2014).
14. Lin, C. H. & Lee, E. H. Y. JNK1 inhibits GluR1 expression and GluR1-mediated calcium influx through phosphorylation and stabilization of Hes-1. *J. Neurosci.* **32**, 1826–1846 (2012).
15. Chacón, P. J. & Rodríguez-Tébar, A. Increased expression of the homologue of enhance-of-split 1 protects neurons from beta amyloid neurotoxicity and hints at an alternative role for transforming growth factor beta1 as a neuroprotector. *Alzheimers Res. Ther.* **4**, 31 (2012).
16. Ishibashi, M. *et al.* Targeted disruption of mammalian hairy and Enhancer of split homolog-1 (HES-1) leads to up-regulation of neural helix-loop-helix factors, premature neurogenesis and severe neural tube defects. *Genes Dev.* **9**, 3136–3148 (1995).
17. Imayoshi, I. *et al.* Oscillatory control of factors determining multipotency and fate in mouse neural progenitors. *Science* **342**, 1203–1208 (2013).
18. Imayoshi, I., Shimogori, T., Ohtsuka, T. & Kageyama, R. Hes1 genes and neurogenin regulate non-neural versus neural fate specification in the dorsal telencephalic midline. *Development* **135**, 2531–2541 (2008).
19. Goebbels, S. *et al.* Genetic targeting of principal neurons in neocortex and hippocampus of NEX-Cre mice. *Genesis* **44**, 611–621 (2006).
20. Hirata, H. *et al.* Oscillatory expression of the bHLH factor Hes1 regulated by a negative feedback loop. *Science* **298**, 840–843 (2002).
21. Okuno, H. Regulation and function of immediate-early genes in the brain: beyond neuronal activity markers. *Neurosci. Res.* **69**, 175–186 (2011).
22. Tagawa, Y., Kondo, P. O., Majdan, M. & Shatz, C. J. Multiple periods of functional ocular dominance plasticity in mouse visual cortex. *Nat. Neurosci.* **8**, 380–388 (2005).
23. Wang, K. H. *et al.* *In vivo* two-photon imaging reveals a role of arc in enhancing orientation specificity in visual cortex. *Cell* **126**, 389–402 (2006).
24. Winston, S. M., Hayward, M. D., Nestler, E. J. & Duman, R. S. Chronic electroconvulsive seizures down-regulate expression of the immediate-early genes c-fos and c-jun in rat cerebral cortex. *J. Neurochem.* **54**, 1920–1925 (1990).
25. D'Costa, A., Breese, C. R., Boyd, R. L., Booze, R. M. & Sonntag, W. E. Attenuation of Fos-like immunoreactivity induced by a single electroconvulsive shock in brains of aging mice. *Brain. Res.* **567**, 204–211 (1991).
26. Lai, C. S., Franke, T. F. & Gan, W. B. Opposite effects of fear conditioning and extinction on dendritic spine remodelling. *Nature* **483**, 87–91 (2012).
27. Giustino, T. F. & Maren, S. The Role of the Medial Prefrontal Cortex in the Conditioning and Extinction of Fear. *Front. Behav. Neurosci.* **9**, 298 (2015).
28. Vidal-Gonzalez, I., Vidal-Gonzalez, B., Rauch, S. L. & Quirk, G. J. Microstimulation reveals opposing influences of prelimbic and infra limbic cortex on the expression of conditioned fear. *Learn. Mem.* **13**, 728–733 (2006).
29. Pape, H. C. & Pare, D. Plastic synaptic networks of the amygdala for the acquisition, expression, and extinction of conditioned fear. *Physiol. Rev.* **90**, 419–463 (2010).
30. Quirk, G. J. & Mueller, D. Neural Mechanism of extinction learning and retrieval. *Neuropsychopharmacology* **33**, 56–72 (2008).
31. Taniguchi, H. *et al.* A resource of Cre driver lines for genetic targeting of GABAergic neurons in cerebral cortex. *Neuron* **71**, 995–1013 (2011).
32. Sakamoto, K., Karelina, K. & Obrietan, K. CREB: a multifaceted regulator of neuronal plasticity and protection. *J. Neurochem.* **116**, 1–9 (2011).
33. Fusco, S. *et al.* A CREB-Sirt-Hes1 circuitry mediates neural stem cell response to glucose availability. *Cell Rep.* **14**, 1195–1205.
34. Abdallah, C. G. *et al.* The Neurobiology and Pharmacotherapy of Posttraumatic Stress Disorder. *Annu. Rev. Pharmacol. Toxicol.* **59**, 171–189 (2019).
35. Pitman, R. K. *et al.* Biological studies of post-traumatic stress disorder. *Nat. Rev. Neurosci.* **13**, 769–787 (2012).
36. Davidson, J. R. Pharmacologic treatment of acute and chronic stress following trauma: 2006. *J. Clin. Psychiatry.* **67**, 34–39 (2006).
37. Kobayashi, T. *et al.* The cyclic gene Hes1 contributes to diverse differentiation responses of embryonic stem cells. *Genes Dev.* **23**, 1870–1875 (2009).
38. Imayoshi, I. *et al.* Roles of continuous neurogenesis in the structural and functional integrity of the adult forebrain. *Nat. Neurosci.* **11**, 1153–1161 (2008).

Acknowledgements

We thank Klaus-Armin Nave for the Math2/Nex-Cre mouse strain and Satsuki Kitano, Hitoshi Miyachi, Mariko Hayashi, and Yoko Horiguchi for technical assistance. This work was supported by Grant-in-Aid for Scientific Research on Innovative Areas (16H06480, R.K.) from Ministry of Education, Culture, Sports, Science, and Technology (MEXT), Japan and AMED-CREST (JP18gm1110002, R.K.) from Japan Agency for Medical Research and Development.

Author Contributions

T.M. and T.Y. conducted the experiments. T.M., T.O. and R.K. designed the experiments. T.M. and R.K. wrote the manuscript.

Additional Information

Supplementary information accompanies this paper at <https://doi.org/10.1038/s41598-019-44698-y>.

Competing Interests: The authors declare no competing interests.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2019