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Human Genome Variation (2015), 2

2015-06-11

http://hdl.handle.net/2433/245649

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Journal Article

publisher

Kyoto University
A novel germline mutation in a patient with nevoid basal cell carcinoma syndrome showing cystic lesion in the lung

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Nevoid basal cell carcinoma syndrome (NBCCS), also called Gorlin–Goltz syndrome, is a rare multisystemic disease inherited in an autosomal-dominant manner. Historically, it was first reported in 1894 for a patient with multiple basocellular carcinomas.¹ In 1960, Gorlin and Goltz established a classical triad that characterizes this syndrome: multiple basocellular epithelomas, keratocysts in the jaws and bifid ribs.² Its estimated incidence varies from 1 in 57,000 to 256,000 among various studies, with a male-to-female ratio of 1:1.³ The pathogenesis of NBCCS is attributed to abnormalities linked to the long arm of chromosome 9 (q22.3-q31). Various kinds of mutations in the patched homologue 1 (PTCH1) gene, a tumor-suppressor gene, are considered as its molecular origin.⁴ PTCH1 has an important role for embryonic structuring and cell cycle, and thus its genetic alteration can cause a key event for the development of the disease. It is important to genetically diagnose the disease to prevent its fatal consequences such as multiple skin cancers and other malignant tumors.

A 19-year-old female with the first episode of a primary spontaneous pneumothorax was referred to our hospital. Her left lung was completely collapsed in chest X-ray, although she had almost no respiratory symptoms. She was a non-smoker without any history of pulmonary diseases. She was born at her parents ages in early thirty, and there is no other cases of NBCCS in her family. She had undergone surgery for four times because of keratocystic odontogenic tumor (formerly called odontogenic keratocyst) in the upper jaw. Despite chest tube drainage for 5 days, air leakage persisted and thoracoscopic lung resection was selected. Intraoperatively we found pleural adhesion at the superlateral segment of the left lower lobe. The visceral pleura of the left lung appeared generally edematous. A cystic lesion was found at the lowest part of the dorsobasal segment of the left lower lobe and was resected with a stapler. Interestingly, the surface of the lesion appeared unusually thick for a bulla with an appearance similar to other parts of the visceral pleura (Figure 1a). Macroscopically, the cystic lesion had smooth surface and unevenly thickened wall (Figure 1b,c). There remained persisted air leakage for 7 days postoperatively and eventually she underwent rethoracotomy; since then there has been no evidence of relapse to date for more than 20 months. We diagnosed her as NBCCS because she fulfilled two major criteria for NBCCS: bilamellar calcification of the falx cerebri and keratocystic odontogenic tumor (Supplementary Table 1).⁵

Sections of 4 μm in thickness were used for hematoxylin and eosin staining and immunocytochemical analysis (Figure 2). For immunostaining, antibodies were used to detect alpha-smooth muscle actin and desmin (Dako corporation, Code No. N1584 and N1526, respectively). The detection was on the basis of the Labeled Streptavidin Biotin method (Bench Mark GX VENTANA).

The study was approved by a local ethical committee and informed consent was obtained from the patient and parents. Genomic DNA was extracted from peripheral blood leukocytes with a standard method using proteinaseK and phenol. Nine pairs of oligonucleotide primers were designed for the amplification of the 23 exons of the PTCH1 gene (Supplementary Table 2). PCR was performed using PrimeSTAR Max DNA Polymerase (Takara Bio, Shiga, Japan) according to the manufacturer’s protocol. PCR products were resolved on a 1% agarose gel and purified with the Wizard SV Gel and PCR Clean-Up System (Promega, Tokyo, Japan). Sequencing of PCR products was performed on an ABI 373A automated sequencer (Life Technologies). The sequences obtained were aligned with a genomic sequence (NG_007664) as well as a full-length complementary DNA sequence of PTCH1 (NM_000264.3). To confirm the results of direct sequencing, PCR products with P3F and P3R covering exon 4 to exon 8 were cloned into pGEM-T Easy vector (Promega) and sequenced with T7 and SP6 vector primers. The exon numbers and the annotation of the detected mutation were given on the basis of the mRNA sequence of NM_000264.3 according to the recommendation of the Human Genome Variation Society (http://www.hgvs.org/mutnomen/refsseq.html).

The histological examination of the cystic lesion revealed the followings. The cysts were lined by columnar cells with no atypia (Figure 2a), and underlying stroma was mainly composed of loose connective tissue with inflammatory cells (Figure 2b) and the

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Received 11 October 2014; revised 12 February 2015; accepted 5 March 2015

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Figure 1. Intraoperative finding of the pleura and cystic lesion (a). Gross appearance of the resected cystic lesion with smooth surface (b) and the cut section with unevenly thickened cyst wall (c). An orthopantomogram showing cystic lesions in the mandibular region (white arrows) (d). Brain-computed tomography showing intracranial ectopic calcifications of the falx cerebri (e). L, left; R, right.

Figure 2. Histological appearance of the resected cystic lesion. The cyst lumen was covered by columnar epithelium (a). The underlying stroma was mainly composed of loose connective tissue with inflammatory cells (b). Many bundles of bland spindle cells are identified (c). The spindle cells are positive for alpha-smooth muscle actin (d) and desmin (e). These bundles surrounded the cyst lumen, which resembles respiratory tract tissue (f).
dense bundles of bland spindle cells (Figure 2c). They were confirmed to be smooth muscle cells using immunohistochemistry (Figure 2d,e). These structures resembled respiratory tract tissue, and the cyst was covered with fibrous thickening visceral pleura (Figure 2f). These findings were not the histological feature in the pneumothorax; therefore, hamartoma-like lesion or reactive change possibly associated with NBCCS should be considered.

We have undertaken a genetic analysis of the PTCH1 gene using direct sequencing. Two genetic alterations were identified in PTCH1 of the patient. The first was located in exon 8 and was an insertion of T nucleotide (c.1207dupT) in one of the two alleles, leading to a premature truncation of the PTCH1 protein with 33 aberrant amino acids in its C terminus (Figure 3a). This mutation was localized in the first extracellular loop of the PTCH1 protein carrying 12 transmembrane domains. The insertion was confirmed by sequencing each allele using the cloned PCR products. We also examined whether this insertion was observed in her parents’ genome. As the mutation was not present in either of her parents, it was a de novo germ-line mutation specific to the patient (Figure 3a). The second genetic alteration, a T to C transition that leads to an amino-acid substitution from leucine to proline, was found in exon 23 for one of the two alleles (Figure 3b). The genetic variant was registered in dbSNP as rs357564, and the frequency of C allele was reported as 0.384 in the Japanese population of the International HapMap Project. It was observed in both parents but not in the patient. To our knowledge, there was only one case of a NBCCS patient with large congenital cyst reported in a family consisting of four NBCCS cases.7 The patient showed similar clinical manifestations related to the disease as shown in Supplementary Table 1. In our case, the four patients were most likely to share the same genetic mutation. The reason for such a cyst in only one of the four cases was speculated as either an additional manifestation of NBCCS or simply a coincidental finding in one case.

Sequencing analysis of the whole exons of PTCH1 identified a novel single-nucleotide insertion in c.1207dupT resulting in a frameshift at the amino acid. Most of the abnormal mRNAs were reported to be degraded via a mechanism called nonsense-mediated mRNA decay. In the present case, however, a truncated PTCH1 protein with 33 aberrant amino acids at its carboxyl terminus is likely to be synthesized and caused disease phenotypes. The PTCH protein has 12 transmembrane domains and two large extracellular loops.8 PTCH1 is involved in the hedgehog (HH) pathway and served as a receptor for the HH ligand.9 Two extracellular loops are coded in exons 2–9 and exons 15–18 in PTCH1,10 and the second extracellular loop of PTCH1 works as receptor for the HH ligand.11 HH signaling is a key regulator of embryonic development and tumorigenesis controlling cellular proliferation. Therefore, truncated PTCH1 protein because of this insertion in exon 8 could result in an aberrant cell cycle progression and neoplastic growth.12

Figure 3. Sequencing analysis of the PTCH1 gene. (a) Direct sequencing results using PCR products covering exon 8 of the PTCH1 gene in the patient and parents. An insertion of a T nucleotide c.1207dupT causing a frameshift is shown by an arrow. Allele-specific sequencing results of cloned PCR products of the patient with a vector primer are also shown. As cloned PCR fragments were sequenced with reverse primer relative to gene orientation, electropherograms were inverted to match their direction with the other results. (b) Direct sequencing results of a T to C transition identified in exon 23 (indicated by an arrow).
HGV DATABASE
The relevant data from this Data Report are hosted at the Human Genome Variation Database at http://dx.doi.org/10.6084/m9.figshare.hgv.584.

ACKNOWLEDGEMENTS
We thank Junpei Takagi and Jiro Kitamura for critical reading of the manuscript. This research is in part supported by grants-in-aid from Research on rare and intractable diseases, the Ministry of Health, Labor, and Welfare of Japan and from the Ministry of Education, Culture, Sports, Science, and Technology of Japan.

COMPETING INTERESTS
The authors declare no conflict of interest.

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Supplemental Information for this article can be found on the Human Genome Variation website (http://www.nature.com/hgv).