



Simplification of circadian rhythm measurement using species-independent time-indicated genes

Yusuke Tanigaki^{a,*}, Takanobu Higashi^b, Kotaro Takayama^c, Atsushi J. Nagano^d, Mie N. Honjo^e, Ayumi Tezuka^d, Mari Kamitani^d, Hirokazu Fukuda^{a,f}

^a Department of Mechanical Engineering, Graduate School of Engineering, Osaka Prefecture University, Sakai, Japan

^b Department of Applied Life Sciences, Graduate School of Life and Environmental Sciences, Osaka Prefecture University, Sakai, Japan

^c Faculty of Agriculture, National University Corporation Ehime University, Matsuyama, Japan

^d Faculty of Agriculture, Ryukoku University, Otsu, Japan

^e Center for Ecological Research, Kyoto University, Otsu, Japan

^f JST PRESTO, Kawaguchi, Japan

ARTICLE INFO

Keywords:

Circadian rhythms
Homologous genes
Tomato
Lettuce
Arabidopsis thaliana

ABSTRACT

The circadian rhythm varies among species, and the distribution of common circadian rhythm-related genes in plants is not yet clearly understood. In this study, we analyzed the transcriptome data from plants of three different species (*Solanum lycopersicum*, *Arabidopsis thaliana*, and *Lactuca sativa*) and their circadian rhythms. Homology of the gene sequences was analyzed. Thirty genes containing time information were found to be common among the three species studied and were used to measure the circadian rhythm. Because 22 of these 30 genes were associated with photosynthesis, we suggest that light control could be used to regulate the circadian rhythm. Currently, a high-cost transcriptome analysis is required for the measurement of circadian rhythm; however, our results showed that it was possible to reduce the number of target genes to 30 and, hence, to reduce the cost of the analysis. Our findings will enable easier estimation of circadian time, which, in turn, will facilitate environmental control for plant growth through better control of circadian time, thus facilitating better crop management practice.

1. Introduction

Almost all organisms have an approximately 24-h biological rhythm, the so-called circadian rhythm that controls various biological phenomena, such as the opening and closing of stomata, flower initiation, and gene expression [1]. Moreover, circadian rhythms affect plant metabolic pathways by controlling the underlying plant hormones [2]. Crop quality strongly depends on the metabolites produced by complex metabolic pathways; thus, a circadian rhythm is an important determinant of crop quality parameters, such as sugar concentration and nutrient content [3]. Therefore, circadian rhythms may be used for controlling plant growth.

Previous studies have shown that a circadian rhythm can be controlled by controlling the external environment [4,5]. However, circadian rhythms have not been fully elucidated, and a circadian rhythm-based technology of wide applicability in agriculture has not been developed yet. One form of control over the circadian rhythm involves clock genes, which have been found to occur in *Arabidopsis thaliana*.

These genes were *Circadian clock-associated 1/Late elongated hypocotyl (CCA1/LHY)*, *Timing of chlorophyll a/b-binding protein (CAB) expression 1 (TOC1)*, and *Pseudo-response regulators 5, 7, and 9 (PRR5, PRR7, and PRR9)*.

The circadian rhythm cycle is formed mainly by the transfer feedback (negative feedback loop) of *CCA1/LHY* and *TOC1*, which show cyclic expression fluctuation for 24 h. [6]. Many genes exhibit a 24-h cycle because they are regulated by the expression cycle of *CCA1/LHY* and *TOC1*. However, *CCA1* and *LHY* are not common to all species. For example, tomato (*Solanum lycopersicum*) and barley (*Hordeum vulgare*) have the *LHY* gene, but not the *CCA1* gene [7]. In addition, the central clock genes are not highly conserved. Therefore, when attempting to control the circadian rhythm, rather than modifying genes, it is more effective to control the cycle generated by the circadian rhythm. For example, lettuce growth rate has been shown to increase by synchronizing its photoperiod with the circadian rhythm cycle [8], a phenomenon called circadian resonance [8,9].

Chlorophyll fluorescence may be used as a method for gathering

* Corresponding author.

E-mail address: yu.tanigaki@me.osakafu-u.ac.jp (Y. Tanigaki).

biological information, such as photosynthesis potential and plant susceptibility to stress [10], and can be measured to assess biological rhythms [11,12]. However, this method does not detect the clock-gene behavior. In contrast, the molecular timetable method (MTM) has been effectively used to calculate the circadian rhythms in animals from their gene expression patterns [13]. In this method, the transcriptome data are acquired continuously in chronological order, and the expression levels at the same sampling time-points are arranged for each expression pattern. Additionally, circadian time is measured from the peak (molecular peak time) of the scatter plot. In previous studies, we demonstrated that MTM can also be used in plants cultivated under different conditions [14–16], and, although expensive, time can be accurately estimated by using hundreds of periodic genes in the time-series transcriptome analysis. Nonetheless, the circadian time estimation cannot be performed frequently and is difficult to use in industrial, agricultural, or other applications because only a high-cost method that involves the acquisition of transcriptome data is available for the accurate measurement of a circadian rhythm.

In the present study, we aimed to develop a circadian clock technology for use in agriculture. The accurate measurement of a circadian clock requires comprehensive determination of gene expression levels, but the cost is abnormally high for agriculture. Therefore, it is necessary to reduce the cost by conducting a precisely targeted analysis. We focused on the gene expression cycle to identify the circadian rhythm-related genes common to the three species under study. We described a detailed method for identifying these common genes and characterizing their behavior. By successfully estimating the internal circadian time with these common genes, our study demonstrates a method for easily understanding the characteristics of a circadian rhythm, regardless of the species studied. This will be useful for developing highly productive farming strategies.

2. Materials and methods

2.1. Plant material

Three plant species were used in these experiments: tomato (*Solanum lycopersicum*, cultivars ‘Taian-kichijitsu’ and ‘Momotaro-peace’), lettuce (*Lactuca sativa* L. ‘Frill Ice’), and *A. thaliana* accession Col-0. Tomato was cultivated in a sunlight-type plant factory (Ehime University, Japan), and tomato seedlings were transplanted into rockwool cubes (Grodan Delta, GRODAN, Roermond, the Netherlands) in August 2013. The rockwool cubes were watered using a nutrient solution [17]. Twenty slabs were set in each line. The sampling of ‘Taian-kichijitsu’ tomato was carried out in January 2014, June 2016, and April 2017. The tomato plants sampled in June 2016 were diseased (probably powdery mildew). The sampling of ‘Momotaro-peace’ tomato was carried out in November 2017. The temperature, relative humidity, and level of insolation are shown in Supplementary Data 1. The winter, spring, diseased, and autumn tomatoes were sampled in January 2014, June 2016, April 2017, and November 2017, respectively.

Arabidopsis thaliana was seeded on aseptic 40-mm diameter dishes containing 4 mL of Murashige and Skoog medium (2% (w/v) sucrose) and solidified with 2 g L⁻¹ gellan gum. The *A. thaliana* and lettuce seeds were placed on a urethane sponge containing water and incubated for 1 (lettuce) or 2 (*A. thaliana*) weeks under fluorescent light [photosynthetic photon flux density (PPFD) = 250–450 μmol m⁻² s⁻¹] at 22 °C with a 12-h light/12-h dark (LD) photoperiod. Thereafter, the seedlings were transplanted and grown in a deep-flow hydroponic system in a fully controlled plant factory.

For transplant, a panel with a planting hole and a root growth area at a water depth of 90 mm was used (885 mm × 590 mm × 30 mm; M Hydroponic Research, Co. Ltd, Aichi, Japan). Seedlings with each urethane sponge were transplanted into the hole, and the panels were installed in the cultivation beds. The pump circulating the solution was run at a flow of 10–15 L min⁻¹. The cultivation medium was Otsuka

House No. 1 and 2 (N:P₂O₅:K₂O:CaO:MgO = 10:8:27:0:4 and 11:0:0:23:0; Otsuka Chemical, Co. Ltd, Osaka, Japan). The pH and electrical conductivity (EC) of the nutrient solution were set at 6.0 and 2.0, respectively. The growing conditions in the plant factory were as follows: temperature 22 °C, relative humidity (RH) 50%, and CO₂ concentration 1000 μmol mol⁻¹. The light environment was either LL (continuous light) or LD (LED; R:G:B = 120:40:40; total PPFD = 180–220 μmol m⁻² s⁻¹). Illumination was set as LD and was switched to LL 12 days after transplant.

2.2. RNA extraction and sequence analysis

Tomato leaves of the fifth branch from the tip were sampled every 2 h for 2 days (winter and diseased tomatoes, n = 1, spring and autumn tomatoes, n = 3), sliced into 1-cm² segments, and stored at 0 °C in RNAlater solution (Qiagen, Hilden, Germany). The RNA of ‘Momotaro-peace’ tomato was mixed with equal amounts of the RNA extracted from the samples for further use. Lettuce leaves grown under LL and LD conditions and *A. thaliana* leaves were sampled every 2 h for 2 days at 13 after transplant (n = 1). The sampled leaves were frozen in liquid nitrogen and stored at –80 °C. Total RNA was extracted from each sample using the Agilent Plant RNA Isolation Mini Kit (Agilent Technologies, Santa Clara, CA, USA), according to the manufacturer’s instructions.

Total RNA extracted was detected using a Bioanalyzer (Agilent Technologies). In the process of preparing the RNA sequencing library, the total RNA extracted was mixed with ERCC RNA Spike-In control mixes (Life Technologies, Carlsbad, CA, USA). Poly(A) RNA was isolated using oligo(dT)25 Dynabeads (Invitrogen, Carlsbad, CA, USA), according to the method of Wang et al. [18]. The sequencing libraries of lettuce, *A. thaliana*, winter tomato, and LL tomato were prepared according to the method of Nagano et al. [19], and the sequences were analyzed using a HiSeq 2000 sequencer (single-end, 50-bp reads; Illumina, San Diego, CA, USA). The sequencing libraries of spring and autumn tomatoes were prepared according to the method of Kamitani et al. [20], and the sequences were analyzed using a HiSeq 2500 sequencer (single-end, 50-bp reads; Illumina). The reads obtained were registered in the DNA Databank of Japan (DDBJ) database (<http://trace.ddbj.nig.ac.jp/DRASearch>) under the following accession numbers: [DRA004542](#) (LL lettuce), [DRA004561](#) (LD lettuce), [DRA005748](#) (*A. thaliana*), [DRA003530](#) (winter tomato), [DRA003528](#) (LL tomato), [DRA004568](#) (diseased tomato), [DRA007095](#) (spring tomato), and [DRA007076](#) (autumn tomato).

2.3. Transcriptome analysis

Quality control of the clean reads obtained was performed using FastQC (<http://www.bioinformatics.babraham.ac.uk/projects/fastqc/>). The reference sequences for the mapping of tomato and lettuce were obtained from the National Center for Biotechnology Information (NCBI; <http://www.ncbi.nlm.nih.gov/>) database. The reference sequences for *A. thaliana* were obtained from The Arabidopsis Information Resource (TAIR; <https://www.arabidopsis.org/>) database. Mapping was performed by using Bowtie 2 (ver.2.3) [21] and quantification by using the RNA-Seq by Expectation Maximization software (RSEM; <http://deweylab.biostat.wisc.edu/rsem/>) [22]. Both software programs were run with default settings. The expression data were normalized to reads per kilo-base million (RPKM) values.

2.4. Periodicity analysis and circadian time estimation

We used the MTM [13,14] to detect the periodicity genes or contigs of tomato, lettuce, and *A. thaliana*. In this method, the time-series gene expression data were fitted to the cosine curve of the 24-h cycle, and the periodically expressed genes were selected from the correlation. The genes with a coefficient of determination (R²) value ≥ 0.7 were selected

as periodically expressed genes (time-indicated genes, TiGs). The selected TiGs were arranged for each expression peak-time, and the expression level of each gene was extracted at each sampling time-point to prepare a scatter plot. The peak of this scatter plot was the circadian time at the sampling time-point. The accuracy of these time estimates was evaluated by correlation between the cosine curve of the scatter plot obtained and the same length of time as the natural period of the target plant. A heatmap was generated using z-score normalization values transformed from RPKM values and R software.

2.5. Search for common genes

Gene homology was determined using the basic local alignment search tool (BLAST) analysis (tBLASTx). The homology between tomato and *A. thaliana* was determined using PSI-BLAST, which was executed with *descriptions* set to 5000, *interactions* set to 2000, and *alignments* set to 1000. Other BLAST parameters were set to default values. The gene with the highest e-value was selected as the homologous gene. All homologous TiGs were linked to *A. thaliana* using BLAST (including PSI-BLAST), and common genes were selected.

2.6. Functional analysis

The TAIR ID (*A. thaliana*) was used for classification. The genes were annotated based on Gene Ontology (GO) terms (<https://geneontology.org/>). The Kyoto Encyclopedia of Genes and Genomes (KEGG) (<https://www.genome.jp/kegg/kegg.ja.html>) was used for metabolic pathway analysis. Localization was determined based on the Cell eFP Browser (http://bar.utoronto.ca/cell_efp/cgi-bin/cell_efp.cgi).

3. Results

The result of quality check showed that the Q30 quality scores of all sample reads were $\geq 80\%$. However, this result was excluded from the analysis because the final sample on the last day of the June tomato sampling had a low-quality sequence read. The clean reads (approximately 90% or more for *A. thaliana*, 80% or more for tomato, and 90% or more for lettuce) were mapped to the reference sequences for each obtained sequence. Each normalized expression level data is provided as supplemental data (Supplementary data 2–8). Lettuce showed 1710 and 1095 contigs (TiGs; herein, we include contigs as genes) with $R^2 \geq 0.7$ in LL and LD conditions, respectively. Of these, 614 TiGs

overlapped (Fig. 1). *Arabidopsis thaliana* showed 923 TiGs with $R^2 \geq 0.7$ in LL condition. The winter and diseased tomatoes showed 2518 and 628 TiGs with $R^2 \geq 0.7$, respectively. In lettuce, 614 TiGs were homologous with 556 genes of winter tomato and 439 genes of *A. thaliana*, according to the BLAST results. In winter tomato, 556 genes were homologous with 402 genes of *A. thaliana*, according to the PSI-BLAST results (Fig. 1A). In *A. thaliana*, 439 genes were homologous with 402 genes of winter tomato, according to the PSI-BLAST results, and 923 TiGs were homologous with 818 tomato genes (number of genes duplicated in winter and diseased tomatoes) and 751 TiGs (contigs) of lettuce (Fig. 1B). In winter tomato, 2518 TiGs were homologous with 1538 contigs of lettuce and 1798 genes of *A. thaliana* (Fig. 1C). Lastly, in diseased tomato, 628 TiGs were homologous with 443 contigs of lettuce and 482 genes of *A. thaliana* (Fig. 1D).

These homologous TiGs appeared to convey time information; however, whether they indicated circadian rhythm was unclear. Therefore, we selected them to estimate circadian time. A clear scatter diagram was obtained with the base TiGs (e.g., 614 lettuce genes listed in Supplementary Data 1), which rendered an average correlation value with the cosine curve at each sampling time-point of approximately 0.8 and allowed accurate estimation of the circadian time involved (Fig. 1). However, the scatter plots of homologous genes showed blurred curves. Moreover, to estimate circadian time, the average correlation value with the cosine curve for all plant species combined was approximately 0.5, which permitted a certain degree of accuracy (Supplementary Data 9). At the time estimated from the homologous genes, the estimation error was within the 2-h sampling interval for all except seven (of 325) time points. (Fig. 2).

The ability to estimate circadian time with homologous TiGs across species implies that they might encode information indicating the internal time derived from the circadian rhythm. Furthermore, these genes appeared to be common to all three species studied. Based on the results of the MTM ($R^2 \geq 0.7$), we selected 30 genes from the three plant species studied (Table 1). These 30 genes did not include *CCA1*, nor did they include *PRRs* or *CO* (genes related to clock function), but did include *TOC1* and *GI*. Additionally, according to the information available in the TAIR database, the products of 22 of these 30 genes were localized in the chloroplast. In contrast, only five were localized in the nucleus, where the circadian clock was formed.

We annotated these 30 genes by function using GO terms based on the accession numbers of *A. thaliana*. The 30 genes were classified into three broad groups: carbohydrate metabolism, light response-related,

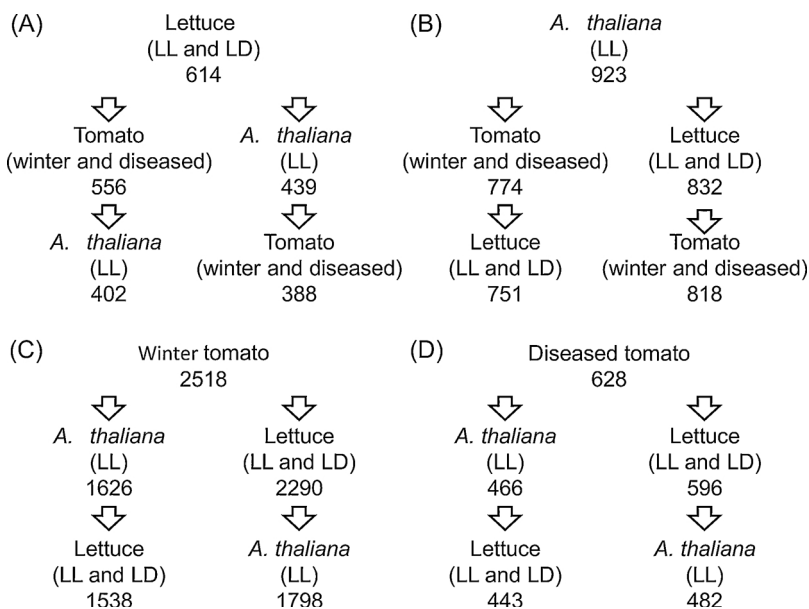


Fig. 1. Homolog search of periodically expressed time-indicated genes (TiGs).

(A) Lettuce [light and light (LL) and light and dark (LD)], (B) *Arabidopsis thaliana*, (C) Winter tomato, (D) Diseased tomato. Numbers under the plant name indicate the number of homologous genes. All of these results showed a $R^2 \geq 0.7$ for the cosine curve as fitted using the molecular timetable method (MTM).

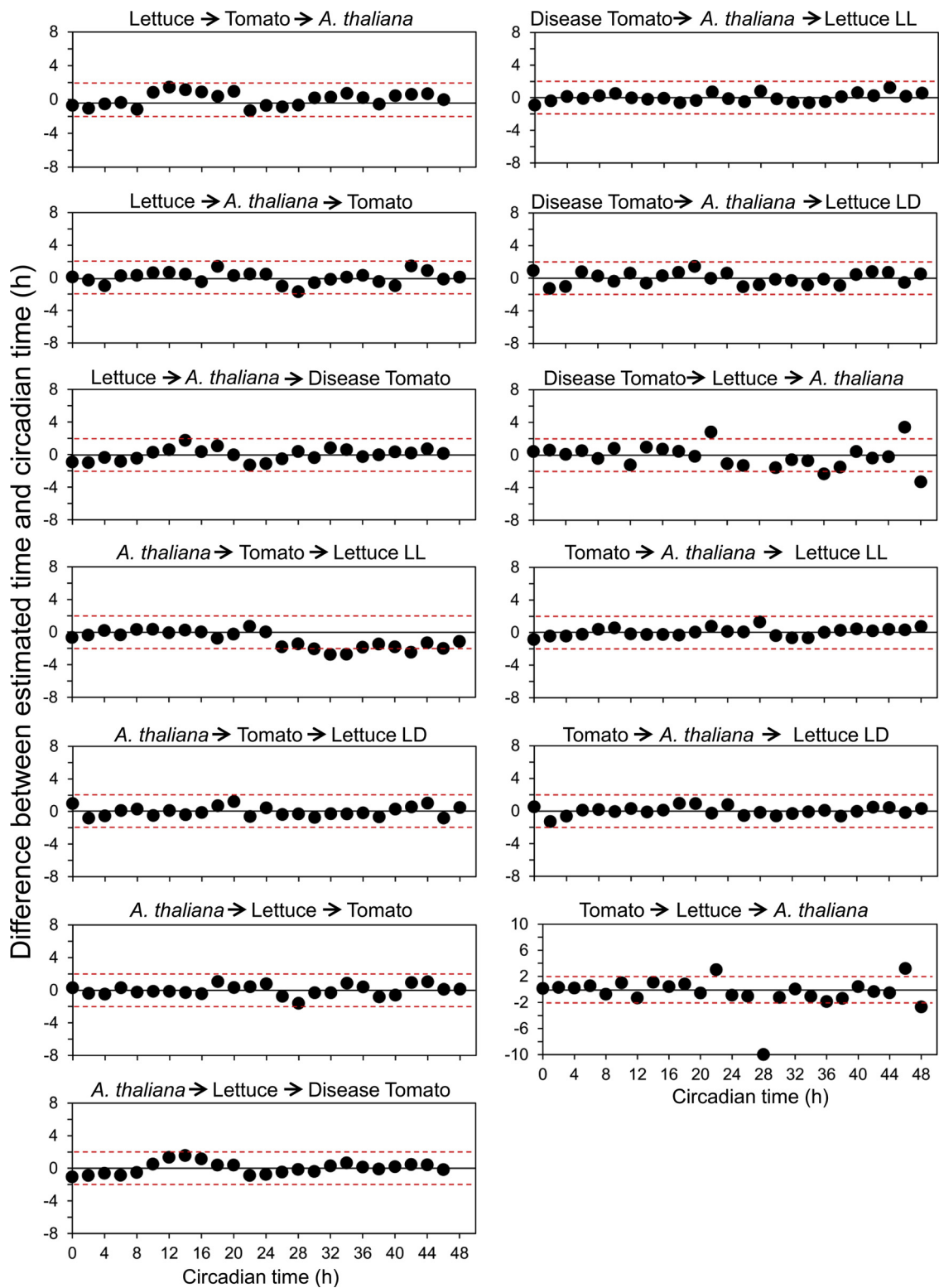


Fig. 2. Time estimation using time-indicated gene (TiG) homology.

Black dots indicate the estimated circadian time. The vertical axis in each figure shows the difference between the estimated time and the circadian time. Red dotted line indicates an error of ± 2 h. The plant names in the figure indicate the order of homology search. All of these results showed a $R^2 \geq 0.7$ for the cosine curve as fitted using the molecular timetable method (MTM).

and others (Fig. 3). The categories of these groups indicated that these 30 genes were related to light. However, there was no common characteristic sequence in the upstream regions and sequences of these common 30 genes.

We analyzed the expression fluctuation of these 30 genes. Although

the expression pattern of these 30 genes was slightly different depending on the species, the expression was cyclically fluctuated (Fig. 4). Next, we analyzed the peak times of these 30 genes. The circadian time was calculated from the correlation between the expression pattern of TiGs and the cosine curve. Thus, the period that can be estimated

Table 1
Annotation of 30 genes in *Arabidopsis thaliana*.

Accession No.	Description	Pathway
AT4G26850.1	GDP-L-Galactose Phosphorylase	Ascorbate and aldarate metabolism
AT4G25700.1	Beta Carotenoid Hydroxylase 1	Carotenoid biosynthesis
AT1G22770.1	GIGANTEA	Circadian rhythm
AT5G61380.1	Timing Of CAB Expression 1	Circadian rhythm
AT4G38970.1	Fructose-Bisphosphate Aldolase 2	Glycolysis/Gluconeogenesis
AT1G31330.1	Photosystem I Subunit F	Photosynthesis
AT4G12800.1	Photosystem I Subunit L	Photosynthesis
AT1G15820.1	Light-Harvesting Complex Photosystem II Subunit 6	Photosynthesis - antenna proteins
AT3G47470.1	Light-Harvesting Chlorophyll-Protein Complex I Subunit A4	Photosynthesis - antenna proteins
AT3G54890.1	Photosystem I Light-Harvesting Complex Gene 1	Photosynthesis - antenna proteins
AT4G10340.1	Light-Harvesting Complex Of Photosystem II 5	Photosynthesis - antenna proteins
AT5G54270.1	Light-Harvesting Chlorophyll B-Binding Protein 3	Photosynthesis - antenna proteins
AT1G44446.1	Chlorophyll A Oxygenase	Porphyrin and chlorophyll metabolism
AT1G74470.1	Pyridine Nucleotide-disulfide Oxidoreductase family protein	Porphyrin and chlorophyll metabolism
AT3G56940.1	Copper Response Defect 1	Porphyrin and chlorophyll metabolism
AT5G13630.1	Genomes Uncoupled 5	Porphyrin and chlorophyll metabolism
AT2G40840.1	Disproportionating Enzyme 2	Starch and sucrose metabolism
AT3G29320.1	Alpha-Glucan Phosphorylase 1	Starch and sucrose metabolism
AT3G46970.1	Alpha-Glucan Phosphorylase 2	Starch and sucrose metabolism
AT4G17090.1	Beta-Amylase 3	Starch and sucrose metabolism
AT1G06040.1	Salt Tolerance protein	
AT2G15890.1	Maternal Effect Embryo Arrest 14	
AT2G28840.1	E3 Ubiquitin-protein Ligase XB3 Ortholog	
AT2G41250.1	Haloacid Dehalogenase-like Hydrolase (HAD) superfamily protein	
AT2G42750.1	DNA J Protein C77	
AT3G56290.1	Potassium Transporter	
AT5G18130.1	Transmembrane protein	
AT5G23240.1	DNA J Protein C76	
AT5G35970.1	P-loop Containing Nucleoside Triphosphate Hydrolases superfamily protein	
AT5G67370.1	Conserved In The Green Lineage And Diatoms 27	

Gene description is based on the description of these genes in *A. thaliana*. The notation of the pathway is based on the Kyoto Encyclopedia of Genes and Genomes (KEGG) database.

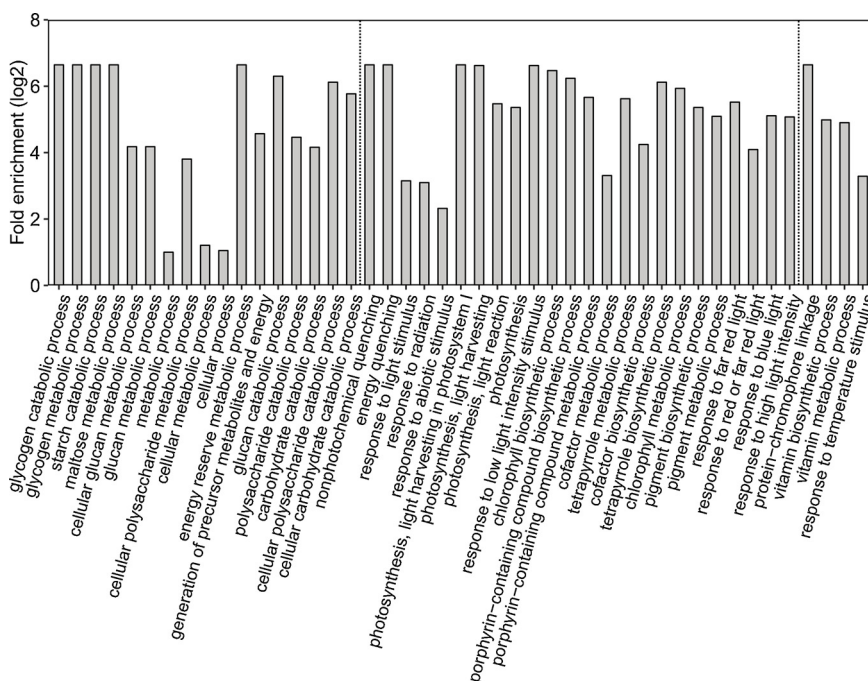


Fig. 3. Gene Ontology (GO) enrichment analysis of 30 homologous genes.

The figure shows parent terms with false discovery rate (FDR) ≤ 0.05 in biological process. The GO analysis was based on the *Arabidopsis thaliana* data. The complete data of cellular component, molecular function, and biological process are presented in Supplementary data 10. The vertical axis indicates the \log_2 value of the fold-enrichment value. The left side delimited by a broken line shows the energy metabolism category. The middle part divided by the broken line shows the light response category. The right side separated by a broken line shows other categories.

decreases with increasing similarity among the expression patterns of these 30 genes. Further, we confirmed the variation in the expression peak time of these 30 genes. *Arabidopsis thaliana* showed a distinctly biased expression peak, but the 30 genes showed expression peaks from morning to night in all species (Fig. 5).

Finally, we confirmed that circadian time estimation is possible using these 30 genes. Using the selected 30 genes from lettuce LL,

lettuce LD, ‘Taian-kichijitsu’ tomato, diseased ‘Taian-kichijitsu’ tomato, and *A. thaliana*, the circadian time was estimated with an accuracy of ± 2 h of the sampling interval (Fig. 6). In *A. thaliana* and autumn tomato, the estimation accuracy decreased after 24:00 in circadian time. In addition, it was possible to accurately estimate the time even in spring (‘Taian-kichijitsu’) and autumn (‘Momotaro-peace’) tomatoes, which were not used for selecting these 30 genes (Fig. 6F, G).

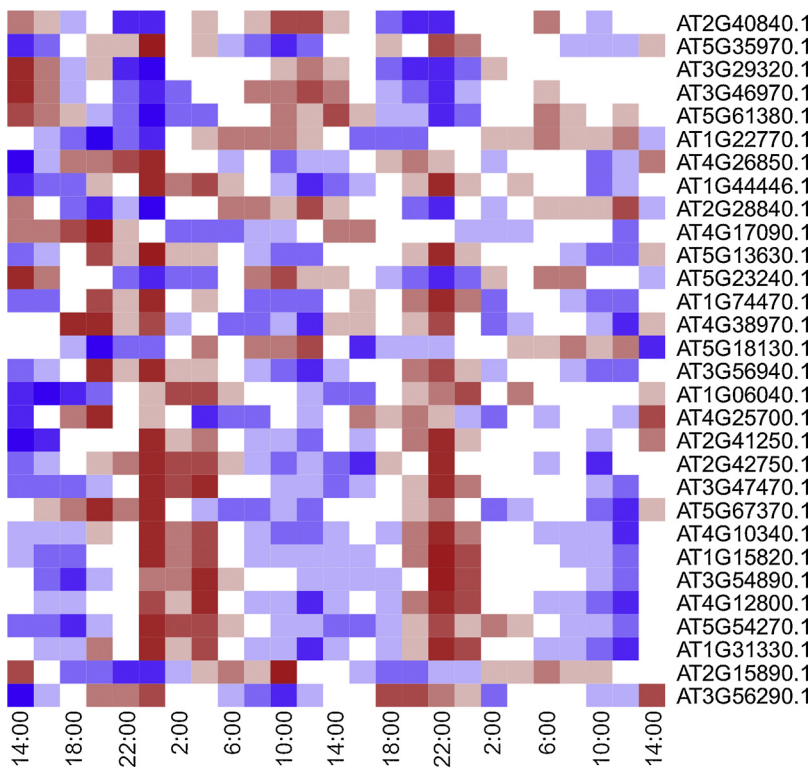


Fig. 4. Expression pattern analysis.

The gene expression values were represented as Z-scores. The color shows the data obtained in 12 steps. (A) Winter tomato. (B) Diseased tomato. (C) Lettuce [light and dark condition (LD)]. (D) Lettuce [light and light condition (LL)]. (E) *Arabidopsis thaliana*. The order of genes in each heatmap represents the relationship of homologs.

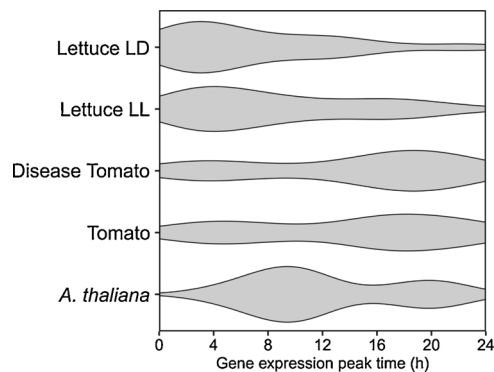


Fig. 5. Variation in the expression peak time of 30 homologous genes. The violin plot shows the variation in the expression peak time of 30 homologous genes. The vertical axis labels indicate the plant in which the expression peak of 30 genes was analyzed.

4. Discussion

In the present study, we focused on the circadian rhythm-related periodicity of gene expression and gene commonality among tomato, lettuce, and *A. thaliana*. We found 30 common genes that were successfully used to estimate the circadian rhythm duration. We analyzed the homology between the periodicity of gene expression calculated by MTM and genes of other plant species. Since homology analysis was performed twice, the number of final homologous genes was greatly reduced, as compared with the initial number of genes (Fig. 1). Nevertheless, these homologous genes were able to estimate the circadian time to a sufficient level of accuracy. This indicated that we had selected highly conserved genes that contained circadian time information by selecting the initial periodic gene and setting the R^2 of periodicity strictly to ≥ 0.7 .

However, the accuracy of estimation tended to decrease as the circadian time exceeded 24:00 (Fig. 6). This was probably due to the use of LL samples for selecting common genes. It is well known that the

attenuation of the amplitude and the deviation of the free-run cycle are caused by exposure to continuous light [4,23]. In addition, diseased tomato plants existed among the samples, and circadian clocks are known to effectively cope with pathogens by periodically expressing the genes of plant hormones and disease response pathways [24,25]. We previously reported that disease response-hormones (salicylic acid, abscisic acid, ethylene, and jasmonic acid) are under the influence of the circadian rhythm [2]. Therefore, we believe that our data from the diseased samples might explain the decline observed in the accuracy of estimation. Research on the effects of infection and disease response on the body clock is underway. Previous studies have reported that plant immunity is closely related to the circadian rhythm [26]. From these studies, we believe that the difference in responsiveness of the circadian clock between opportunistic and severe infections is evident.

The fact that 628 genes in diseased tomato plants showed a circadian cycle indicates that the circadian clock behavior is at the center of daily activities. Circadian clocks do not show periodicity during cell differentiation. Particularly in plants, circadian clocks are reset in the apical tissues of the roots and surrounding cells during lateral root formation [27]. Moreover, the circadian clock is known to form a phase again as it triggers the auxin signal for new cell differentiation [28]. For this reason, our data may suggest that when events, such as disease, occur, the circadian clock is maintained, but with gene expression. It has been reported that is the cycle of *Opuntia ficus-indica* for change in the expression of *CCA1* and *TOC1* in circadian clocks differs by as much as 12 h [29]. In addition, Nose et al. [30] reported that circadian clock formation and its expression network are different between angiosperms and gymnosperms. Moreover, Hayama et al. [31] have reported a high possibility of the existence of two types of clock in *Pharbitis nil*. Therefore, *CCA1* and *TOC1* were considered not necessarily related to each other. Our results also suggest a similar possibility. However, we were unable to find a characteristic common sequence in the upstream regions of these 30 genes.

Even in the tomato depicted in Fig. 1A, the number of common genes decreased to 30, despite the presence of 388 homologous genes. This is probably because we used the data from LL and diseased tomatoes. In addition, Higashi et al. [15] observed that the number of

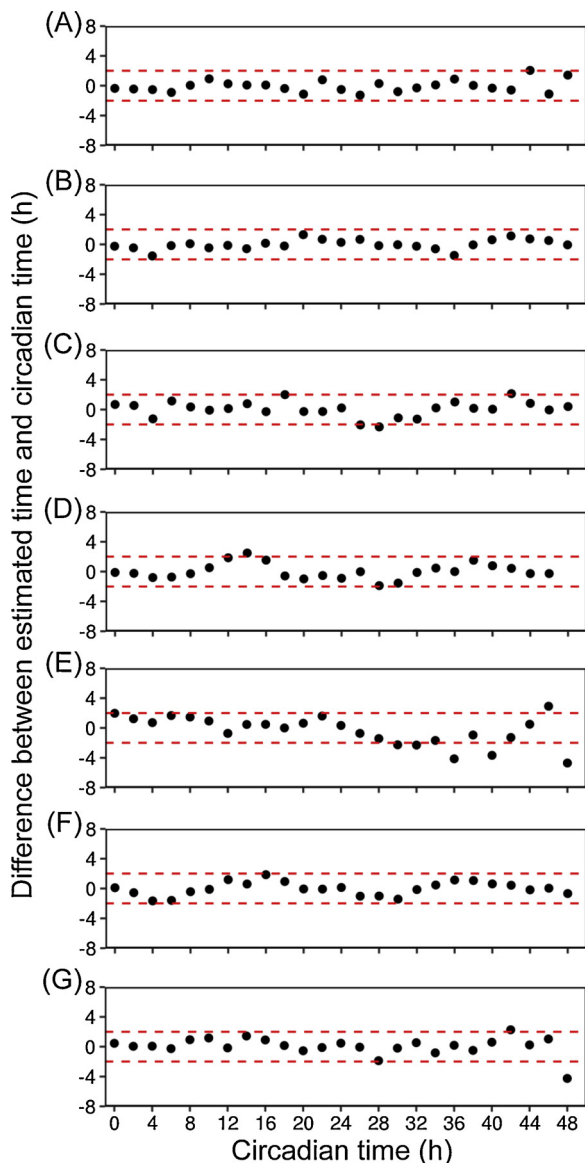


Fig. 6. Circadian time estimation based on 30 homologous genes. (A) Lettuce, light and light cycle (LL). (B) Lettuce, light and dark cycle (LD). (C) Winter tomato. (D) Diseased tomato. (E) *Arabidopsis thaliana*. (F) Spring tomato. (G) Autumn tomato. The vertical axis in each figure shows the difference between the estimated time and the circadian time. The red dotted line indicates an error of ± 2 h. All of these results showed a $R^2 \geq 0.7$ for the cosine curve as fitted using the molecular timetable method (MTM)

periodically expressed genes decreased by 80% because of the differences in light conditions (LL or LD), even for the same species. Moreover, we confirmed that the periodic genes were reduced by approximately 80%, regardless of the presence or absence of disease in tomato (data not shown). We believe that the selected 30 genes encode substantial information on circadian rhythms. Nevertheless, *CCA1/LHY* was not included among those 30 genes.

Plant circadian rhythms are known to differ between shoots and roots [32,33], and *CCA1* does not inhibit the expression of other genes in the roots [34]. Further, it has been demonstrated that this difference in circadian rhythms between shoots and roots is synchronized by weak binding between cells [35,36]. Therefore, the importance of clock genes is believed to vary, depending on tissues and cells. In addition, unlike *A. thaliana*, both, tomato and *H. vulgare* have *LHY*, but not *CCA1* [7]. In contrast, *TOC1* exists in many plant species, including tomato and *H. vulgare*. These findings support the idea of species-specific importance

of different clock genes. In addition, owing to the likely existence of two clocks in *Pharbitis nil* [31], the circadian clock system of plants might require the presence of multiple clocks. The known clock involving *CCA1* and *TOC1* cannot clearly explain the circadian rhythm of a plant circadian clock. Therefore, it might be necessary to analyze *CCA1* and *TOC1* separately to fully understand their influence on the circadian clock.

We found that 22 of the 30 genes were located in chloroplasts. In addition, five genes were localized in the nucleus. Furthermore, we analyzed data under different light conditions; thus, we considered that 22 genes were not affected by a simple light and light cycle. Therefore, among these 22 genes, a gene was thought to play an important role in circadian rhythm. Wilson and Connolly [37] reported that Fe signals from chloroplasts regulate the circadian clock. Among the 22 chloroplast-localized genes, some were related to Fe signaling, but we could not clarify their direct association with signal transduction. On the other hand, Lu et al. [26] summarized the relationship between light, immunity, and circadian rhythm. They separated the relationship between a simple light response and circadian rhythm and discussed the relationship between circadian rhythm and immunity. The 22 genes from the present study might be involved in the physiological functions related to immune response and circadian clock.

We found 30 common genes from the transcriptome data of tomato (including diseased plants), lettuce, and *A. thaliana* grown under different conditions, including continuous light. We reduced the number of genes to be analyzed to 30, which made the analysis possible on a small scale. Even if we compare the transcriptomes of different species simply by real-time quantitative PCR analysis, this can considerably reduce the cost (by at least 90%) and time required to complete the analysis.

5. Conclusion

Contrary to the brief summary of the plant circadian rhythm that was thought heretofore, our results indicate a new circadian clock [31], likely comprising of multiple systems. Furthermore, we considered that the 30 common genes that normally show circadian rhythms – even under continuous light – are at, or close to, the core of the plant circadian clock. Therefore, the behavior of these 30 genes is a clear indicator of the persistence of a plant circadian clock and can provide stable support for carrying out transduction studies of other plant circadian clocks. Furthermore, in agriculture, the reduction of the number of genes in this study makes accurate measurement of the circadian clock rhythm more practical. A number of phenotyping techniques have been developed for use in agriculture, and we believe that integrating these techniques with the measurement of the circadian clock rhythm would enable us to accurately determine the state of plant growth.

Author contributions

YT, TH, and HF designed the study.

YT performed the data analysis.

AN, MH, and AT prepared the RNA-Seq library.

AN, AT, and MK devised a method of library construction.

YT and HF wrote the manuscript.

KT assisted in the sampling of tomato in the sunlight-type plant factory.

All authors discussed the results and implications and commented on the manuscript.

Funding

This work was supported, in part, by Grants-in-Aid for Scientific Research (nos. 25712029 and 16H05011) and Precursory Research for Embryonic Science and Technology (no. JPMJPR1504) of the Japan Science and Technology Agency (to HF).

Declaration of Competing Interest

None.

CRediT authorship contribution statement

Yusuke Tanigaki: Conceptualization, Writing - original draft, Writing - review & editing, Formal analysis, Data curation, Methodology. **Takanobu Higashi:** Conceptualization, Methodology. **Kotaro Takayama:** Methodology. **Atsushi J. Nagano:** Methodology. **Mie N. Honjo:** Methodology. **Ayumi Tezuka:** Methodology. **Mari Kamitani:** Methodology. **Hirokazu Fukuda:** Conceptualization, Writing - original draft, Writing - review & editing.

Acknowledgments

The authors are grateful to Dr. Noriko Takahashi for plant cultivation and to Professor Hiroshi Kudoh for RNA-Seq. The present study was supported by the Joint Usage /Research program of Center for Ecological Research, Kyoto University.

Appendix A. Supplementary data

Supplementary material related to this article can be found, in the online version, at doi:<https://doi.org/10.1016/j.cpb.2019.100118>.

References

- [1] S. Barak, E.M. Tobin, R.M. Green, C. Andronis, S. Sugano, All in good time: the arabidopsis circadian clock, *Trends Plant Sci.* 5 (2000) 517–522, [https://doi.org/10.1016/S1360-1385\(00\)01785-4](https://doi.org/10.1016/S1360-1385(00)01785-4).
- [2] Y. Tanigaki, T. Higashi, K. Takayama, A.J. Nagano, M.N. Honjo, H. Fukuda, Transcriptome analysis of plant hormone-related tomato (*Solanum lycopersicum*) genes in a sunlight-type plant factory, *PLoS One* 10 (2015) e0143412, <https://doi.org/10.1371/journal.pone.0143412>.
- [3] J.A. Kim, H.S. Kim, S.H. Choi, J.Y. Jang, M.J. Jeong, S.I. Lee, The importance of the circadian clock in regulating plant metabolism, *Int. J. Mol. Sci.* 18 (2017) 2680, <https://doi.org/10.3390/ijms18122680>.
- [4] H. Fukuda, H. Murase, I.T. Tokuda, Controlling circadian rhythms by dark-pulse perturbations in *Arabidopsis thaliana*, *Sci. Rep.* 3 (2013) 1533, <https://doi.org/10.1038/srep01533>.
- [5] N. Seki, K. Ukai, T. Higashi, H. Fukuda, Entrainment of cellular circadian rhythms in *Lactuca sativa* L. Leaf by spatially controlled illuminations, *J. Biosens. Bioelectron.* 6 (2015) 186, <https://doi.org/10.4172/2155-6210.1000186>.
- [6] M.J. Haydon, L.J. Bell, A.A.R. Webb, Interactions between plant circadian clocks and solute transport, *J. Exp. Bot.* 62 (2011) 2333–2348, <https://doi.org/10.1093/jxb/err040>.
- [7] C.P.G. Calixto, R. Waugh, J.W.S. Brown, Evolutionary relationships among barley and *Arabidopsis* core circadian clock and clock-associated genes, *J. Mol. Evol.* 80 (2015) 108–119, <https://doi.org/10.1007/s00239-015-9665-0>.
- [8] H. Fukuda, T. Ichino, T. Kondo, H. Murase, Early diagnosis of productivity through a clock gene promoter activity using a luciferase bioluminescence assay in *Arabidopsis thaliana*, *Environ. Control Biol.* 2 (2011) 51–60, <https://doi.org/10.2525/ecb.49.51>.
- [9] A.N. Dodd, N. Salathia, A. Hall, E. Kevei, R. Toth, F. Nagy, J.M. Hibberd, A.J. Millar, A.A. Webb, Plant circadian clocks increase photosynthesis, growth, survival, and competitive advantage, *Science* 309 (2005) 630–633, <https://doi.org/10.1126/science.1115581>.
- [10] K. Takayama, R. Hirota, N. Takahashi, H. Nishina, S. Arima, K. Yamamoto, Y. Sakai, H. Okada, Development of chlorophyll fluorescence imaging robot for practical use in commercial greenhouse, *Acta Hort.* 1037 (2014) 671–676, <https://doi.org/10.17660/ActaHortic.2014.1037.86>.
- [11] P.D. Gould, P. Diaz, C. Hogben, J. Kusakina, R. Salem, J. Hartwell, A. Hall, Delayed fluorescence as a universal tool for the measurement of circadian rhythms in higher plants, *Plant J.* 5 (2009) 893–901, <https://doi.org/10.1111/j.1365-313X.2009.03819.x>.
- [12] S. Moriyuki, H. Fukuda, High-throughput growth prediction for *Lactuca sativa* L. Seedlings using chlorophyll fluorescence in a plant factory with artificial lighting, *Front. Plant Sci.* 7 (2016) 394, <https://doi.org/10.3389/fpls.2016.00394>.
- [13] H.R. Ueda, W. Chen, Y. Minami, S. Honma, K. Honma, M. Iino, S. Hashimoto, Molecular-timetable methods for detection of body time and rhythm disorders from single-time-point genome-wide expression profiles, *Proc. Natl. Acad. Sci. U. S. A.* 101 (2004) 11227–11232, <https://doi.org/10.1073/pnas.0401882101>.
- [14] T. Higashi, Y. Tanigaki, K. Takayama, A.J. Nagano, M.N. Honjo, H. Fukuda, Detection of diurnal variation of tomato transcriptome through the molecular timetable method in a sunlight-type plant factory, *Front. Plant Sci.* 7 (2016) 87, <https://doi.org/10.3389/fpls.2016.00087>.
- [15] T. Higashi, K. Aoki, A.J. Nagano, M.N. Honjo, H. Fukuda, Circadian oscillation of the lettuce transcriptome under constant light and light–dark conditions, *Front. Plant Sci.* 7 (2016) 1114, <https://doi.org/10.3389/fpls.2016.01114>.
- [16] M. Takeoka, T. Higashi, M.N. Honjo, A. Tezuka, A.J. Nagano, Y. Tanigaki, H. Fukuda, Estimation of the circadian phase by oscillatory analysis of the transcriptome in plants, *Environ. Control Biol.* 2 (2018) 67–72, <https://doi.org/10.2525/ecb.56.67>.
- [17] C. Sonneveld, A Method for Calculating the Composition of Nutrient Solutions for Soilless Cultures, 2nd ed., Glasshouse Crops Research and Experiment Station, Naaldwijk, 1985.
- [18] L. Wang, Y. Si, L.K. Dedow, Y. Shao, P. Liu, T.P. Brutnell, A low-cost library construction protocol and data analysis pipeline for illumina-based strand-specific multiplex RNA-Seq, *PLoS One* 6 (2011) e26426, <https://doi.org/10.1371/journal.pone.0026426>.
- [19] A.J. Nagano, M.N. Honjo, M. Mihara, M. Sato, H. Kudoh, Detection of plant viruses in natural environments by using RNA-Seq, *Methods Mol. Biol.* 1236 (2015) 89–98, https://doi.org/10.1007/978-1-4939-1743-3_8.
- [20] M. Kamitani, M. Kashima, A. Tezuka, A.J. Nagano, High-throughput RNA-Seq analysis on plant response under fluctuating temperature conditions, *bioRxiv* (2018), <https://doi.org/10.1101/463596>.
- [21] B. Langmead, S.L. Salzberg, Fast gapped-read alignment with Bowtie 2, *Nat. Methods* 9 (2012) 357–359, <https://doi.org/10.1038/nmeth.1923>.
- [22] B. Li, C.N. Dewey, RSEM: accurate transcript quantification from RNA-Seq data with or without a reference genome, *BMC Bioinf.* 12 (2011) 323, <https://doi.org/10.1186/1471-2105-12-323>.
- [23] E. Yakir, M. Hassidim, N. Melamed-Book, D. Hilman, I. Kron, R.M. Green, Cell autonomous and cell-type specific circadian rhythms in *Arabidopsis*, *Plant J.* 68 (2011) 520–531, <https://doi.org/10.1111/j.1365-313X.2011.04707.x>.
- [24] O. Windram, P. Madhou, S. McHattie, C. Hill, R. Hickman, E. Cooke, D.J. Jenkins, C.A. Penfold, L. Baxter, E. Breeze, S.J. Kiddle, J. Rhodes, S. Atwell, D.J. Kliebenstein, Y.-s. Kim, O. Stegle, K. Borgwardt, C. Zhang, A. Tabrett, R. Legaie, J. Moore, B. Finkenstadt, D.L. Wild, A. Mead, D. Rand, J. Beynon, S. Ott, V. Buchanan-Wollaston, K.J. Denby, *Arabidopsis* defense against *Botrytis cinerea*: chronology and regulation deciphered by high-resolution temporal transcriptomic analysis, *Plant Cell* 24 (2012), <https://doi.org/10.1105/tpc.112.102046> 3530–357.
- [25] G. Jander, Timely plant defenses protect against caterpillar herbivory, *Proc. Natl. Acad. Sci. U. S. A.* 12 (2012) 4343–4344, <https://doi.org/10.1073/pnas.1201443109>.
- [26] H. Lu, C.R. McClung, C. Zhang, Tick tock: circadian regulation of plant innate immunity, *Annu. Rev. Phytopathol.* 55 (2017) 287–311, <https://doi.org/10.1146/aanurev-phyto-080516-035451>.
- [27] H. Fukuda, K. Ukai, T. Oyama, Self-arrangement of cellular circadian rhythms through phase-resetting in plant roots, *Phys. Rev. E* 86 (2012) 041917, <https://doi.org/10.1103/PhysRevE.86.041917>.
- [28] U. Voß, M.H. Wilson, K. Kenobi, P.D. Gould, F.C. Robertson, W.A. Peer, M. Lucas, K. Swarup, I. Casimiro, T.J. Holman, D.M. Wells, B. Péret, T. Goh, H. Fukaki, T.C. Hodgman, L. Laplace, K.J. Halliday, K. Ljung, A.S. Murphy, A.J. Hall, A.A. Webb, M.J. Bennett, The circadian clock rephases during lateral root organ initiation in *Arabidopsis thaliana*, *Nat. Commun.* 6 (2015) 7641, <https://doi.org/10.1038/ncomms8641>.
- [29] I. Mallona, M. Egea-Cortines, J. Weiss, Conserved and divergent rhythms of crasulacean acid metabolism-related and core clock gene expression in the cactus *Opuntia ficus-indica*, *Plant Physiol.* 156 (2011) 1978–1989, <https://doi.org/10.1104/pp.111.179275>.
- [30] M. Nose, A. Watanabe, Clock genes and diurnal transcriptome dynamics in summer and winter in the gymnosperm Japanese cedar (*Cryptomeria japonica* (L.f.) D. Don), *BMC Plant Biol.* 14 (2014) 308, <https://doi.org/10.1186/s12870-014-0308-1>.
- [31] R. Hayama, T. Mizoguchi, G. Coupland, Differential effects of light-to-dark transitions on phase setting in circadian expression among clock-controlled genes in *Pharbitis nil*, *Plant Signal. Behav.* 13 (2018) e1473686, <https://doi.org/10.1080/15592324.2018.1473686>.
- [32] P. Más, M.J. Yanovsky, Time for circadian rhythms: plants get synchronized, *Curr. Opin. Plant Biol.* 12 (2009) 574–579, <https://doi.org/10.1016/j.cpb.2009.07.010>.
- [33] N. Takahashi, Y. Hirata, K. Aihara, P. Mas, A hierarchical multi-oscillator network orchestrates the *Arabidopsis* circadian system, *Cell* 163 (2015) 148–159, <https://doi.org/10.1016/j.cell.2015.08.062>.
- [34] A.B. James, J.A. Monreal, G.A. Nimmo, C.L. Kelly, P. Herzyk, G.I. Jenkins, H.G. Nimmo, The circadian clock in *Arabidopsis* roots is a simplified slave version of the clock in shoots, *Science* 322 (2008) 1832–1835, <https://doi.org/10.1126/science.1161403>.
- [35] H. Fukuda, N. Nakamichi, M. Hisatsune, H. Murase, T. Mizuno, Synchronization of plant circadian oscillators with a phase delay effect of the vein network, *Phys. Rev. Lett.* 99 (2007) 098102, <https://doi.org/10.1103/PhysRevLett.99.098102>.
- [36] B. Wenden, D.L.K. Toner, S.K. Hodge, R. Grima, A.J. Millar, Spontaneous spatiotemporal waves of gene expression from biological clocks in the leaf, *Proc. Natl. Acad. Sci. U. S. A.* 109 (2012) 6757–6762, <https://doi.org/10.1073/pnas.1118814109>.
- [37] G.T. Wilson, E.L. Connolly, Running a little late: chloroplast Fe status and the circadian clock, *EMBO J.* 32 (2013) 490–492, <https://doi.org/10.1038/emboj.2013.14>.