

1 **Drug monitoring for mycophenolic acid in graft-versus-host disease prophylaxis in cord**
2 **blood transplantation**

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26 Running head: Drug monitoring for mycophenolic acid in CBT

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29 What is already known about this subject: Low level of mycophenolic acid after allogeneic
30 hematopoietic stem cell transplantation is associated with high incidence of acute
31 graft-versus-host disease.

32 What this study adds: Low level of mycophenolic acid after cord blood transplantation is
33 associated with high incidence of acute graft-versus-host disease and human herpesvirus 6
34 reactivation, whereas high level of mycophenolic acid is associated with high incidence of
35 sepsis before engraftment.

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38

39 **Abstract**

40 Aims: We performed the retrospective analysis to clarify the significance of drug
41 monitoring for mycophenolic acid (MPA), the active form of mycophenolate mofetil (MMF),
42 in prophylaxis for graft-versus-host disease (GVHD) in cord blood transplantation (CBT).

43 Methods: We retrospectively analyzed the data of 46 patients who underwent first CBT and
44 received GVHD prophylaxis with tacrolimus plus MMF. MPA levels were measured on days
45 7 and 21, and 24-hour areas under the curve (AUC_{0-24}) were estimated.

46 Results: The engraftment and 3-year overall survival rates of all patients were 94% and
47 78%, respectively. The cumulative incidence of sepsis before engraftment was higher in
48 patients with AUC_{0-24} on day 7 of $> 60 \mu\text{g h/mL}$ than in other patients (33% vs. 6%, $p = 0.02$).
49 The cumulative incidence of grade II–IV acute GVHD was higher in patients with AUC_{0-24}
50 on day 21 of $\leq 30 \mu\text{g h/mL}$ than in other patients (80% vs. 50%, $p = 0.04$). The cumulative
51 incidence of human herpesvirus 6 reactivation was higher in patients with AUC_{0-24} on day 21
52 of $\leq 48 \mu\text{g h/mL}$ (median) than in other patients (50% vs. 19%, $p = 0.03$).

53 Conclusions: Blood level of MPA was associated with the risks of acute GVHD and
54 infection. A prospective trial evaluating the benefit of personalized MMF dosing using MPA
55 levels is needed.

56

57

58 **Introduction**

59 Graft-versus-host disease (GVHD) is a major cause of morbidity and mortality after
60 allogeneic hematopoietic stem cell transplantation (allo-SCT). The combination of
61 calcineurin inhibitors (CNIs) with short-course methotrexate (MTX) after allo-SCT has been
62 standard GVHD prophylaxis for more than 20 years.[1, 2] Following the successful use of
63 mycophenolate mofetil (MMF) in the United States and Europe, the use of MMF with CNI in
64 cord blood transplantation (CBT) has recently increased in Japan to promote neutrophil
65 recovery.[3, 4] Wide interpatient variability in blood levels of mycophenolic acid (MPA), the
66 active form of MMF, has been reported even after administering the same dose of MMF.[5, 6]
67 In kidney transplantation, therapeutic monitoring of MPA is reported to be important, with
68 recommended target ranges for areas under the curve (AUCs) of 30–60 $\mu\text{g h/mL}$. AUCs <30
69 $\mu\text{g h/mL}$ are associated with transplanted kidney rejection, whereas AUCs >60 $\mu\text{g h/mL}$ are
70 associated with infection and myelosuppression.[6, 7] Similarly, there are some reports on
71 drug monitoring for MPA in bone marrow or peripheral blood stem cell transplantation.[5,
72 8-11] However, significance of drug monitoring for MPA in CBT was not fully
73 elucidated.[12] In 2015, we reported that a low blood AUC of MPA on day 21 was associated
74 with a high incidence of acute GVHD in the analysis of 24 patients who received CBT.[13]
75 Based on this previous result, we started to increase the dose of MMF in patients with low

76 MPA AUCs on day 7. In 2018, the number of patients increased sufficiently to perform
77 univariate and multivariate analyses, and the observation times became long enough to
78 evaluate the 3-year overall survival (OS). This retrospective analysis was performed to clarify
79 the association between MPA AUCs and CBT outcomes.

80

81 **Methods**

82 *Data collection*

83 We retrospectively analyzed the data of 46 patients who underwent their first CBT and
84 received GVHD prophylaxis with tacrolimus plus MMF in our institute between October
85 2011 and July 2016. Tacrolimus was continuously and intravenously administered from day
86 -1. Blood levels of tacrolimus were measured at least three times per week and were
87 maintained at 12–15 ng/mL. After neutrophil engraftment, tacrolimus was orally administered,
88 with trough levels maintained at 5–10 ng/mL. The patients received 10 mg/kg of oral MMF
89 every 8 hours from days -1 to 30 as inpatient treatment. Blood samples were collected
90 immediately before and 1, 2, and 4 hours after the morning administration of MMF on days 7
91 and 21. Total MPA levels in the plasma were measured using the enzyme multiplied
92 immunoassay technique (EMIT)[14, 15] or the Roche Total Mycophenolic Acid® assay[16]
93 (C₀, C₁, C₂, and C₄, respectively), and C₈ (implying the trough level of the next
94 administration) was assumed to be equal to the C₀ values based on the results of previous

95 studies.[9, 11, 17, 18] The 8-hour area under the curve (AUC_{0-8}) was determined using the
96 linear trapezoidal method, and the 24-hour area under the curve (AUC_{0-24}) was calculated as
97 three times the AUC_{0-8} . This method is frequently used in the setting of MMF administration
98 three times daily (every 8 hours)[9, 18] because MPA levels peak within 2 hours from
99 administration and decrease linearly after 4 hours.[17, 19] After 2014, the dose of MMF for
100 patients with MPA AUC_{0-24} of $<30 \mu\text{g h/mL}$ on day 7 was increased. The dose of those
101 patients from days 8 to 30 was calculated as follows: [the dose of MMF before day 7] \times
102 $40/[the AUC_{0-24} \text{ on day 7}]$. We analyzed the effect of MPA AUC_{0-24} on transplant outcomes.
103 The cut-off values of AUC_{0-24} were set to 30 and $60 \mu\text{g h/mL}$, which were the same as the
104 values used for kidney transplantation.[6] We also set the cut-off value as the median. To
105 evaluate human herpesvirus 6 (HHV-6) and cytomegalovirus (CMV) reactivation, peripheral
106 blood was collected at least once weekly until 90 days after transplantation. The HHV-6 DNA
107 copy number was measured using real-time polymerase chain reaction methods, as described
108 previously.[20] The present study was approved by the institutional ethics committee of the
109 Graduate School of Medicine, Kyoto University. All patients provided consent to participate
110 in the present study.

111

112 *Definition*

113 OS was defined as the time from transplantation to death, and patients who remained alive

114 at the time of the last follow-up were censored. Complete haematological remission (CR) was
115 defined based on morphology for all the patients, except for those with lymphoma; for
116 lymphoma patients, it was based on positron emission tomography-computed tomographic
117 (PET-CT) imaging.[21, 22] The relapse date of patients who did not achieve CR before and
118 after stem cell transplantation (SCT) was defined as day 1. The patients were divided into two
119 groups according to the conditioning regimen: myeloablative conditioning (MAC) or
120 reduced-intensity conditioning (RIC). MAC and RIC were defined as proposed by Giralt et
121 al.[23] and Bacigalupo et al,[24] respectively. Pre-engraftment immune reaction (PIR) was
122 characterized by the presence of at least three of the following symptoms with no direct
123 consequences of infection or adverse effects of medication six or more days before
124 engraftment, as described previously: a high fever ($>38.5^{\circ}\text{C}$), skin eruptions, body weight
125 gain greater than 5% of baseline, or peripheral edema.[25, 26] HHV-6 reactivation was
126 defined as plasma HHV-6 DNA level $\geq 10^4$ copies/mL. CMV reactivation was defined as the
127 detection of at least two CMV pp65 antigen-positive cells per 50,000 leucocytes.

128

129 *Data analysis*

130 Descriptive statistics were used to summarize the variables related to patient demographics
131 and transplantation characteristics. The probability of the OS time was estimated according to
132 the Kaplan–Meier method.[27] To evaluate the influences of confounding factors on OS,

133 log-rank tests were used for the univariate analysis.[28] The competing risk regression model
134 was used for the multivariate analysis of acute GVHD.[29] The cumulative incidences of
135 engraftment, relapse, and GVHD were calculated, and death without events were considered
136 a competing risk. Landmark analysis was performed to evaluate the effect of MPA level on
137 days 7 and 21, with the landmark day set to days 7 and 21. Mann–Whitney *U* tests were used
138 as non-parametric tests for two groups. Correlation of AUC and single point concentration
139 was assessed by Spearman’s correlation coefficient test.[30] The results are expressed as
140 hazard ratios (HRs) and their 95% confidence intervals (CIs). All tests were two sided, and
141 *P*-values <0.05 were considered to indicate statistical significance. All statistical analyses
142 were performed using Stata (version 13.0, Stata Corporation) and EZR (Saitama Medical
143 Center, Jichi Medical University), a graphical user interface for R (The R Foundation for
144 Statistical Computing, version 2.3.0).[31]

145 **Results**

146 *Patient characteristics*

147 The median observation time of the survivors was 1,352 (range: 103–2,487) days. The
148 median age was 50 (range: 20–66) years. The percentages of patients diagnosed with acute
149 myeloid leukemia, acute lymphoid leukemia, myelodysplastic syndrome, and malignant
150 lymphoma were 43%, 13%, 15%, and 17%, respectively. The percentage of patients with CR
151 was 52%. Fifty-two percent of patients received MAC and 93% received total body
152 irradiation (TBI) (Table 1).

153

154 *Outcomes of all patients*

155 The cumulative incidence of neutrophil engraftment was 94%. A median of 25 (range: 11–
156 43) days was required for neutrophil engraftment (Figure 1A). Engraftment failure was seen
157 in 3 patients, and they had risks for engraftment failure; one patient had high tumor burden,
158 another patient received weak conditioning (fludarabine 125 mg/m² plus melphalan 80 mg/m²
159 without TBI) and the other patient transplanted the minimum count of CD34⁺ cells
160 (0.18×10⁵/kg) in this cohort. The cumulative incidences of PIR, grade II–IV acute GVHD,
161 grade III–IV acute GVHD, limited chronic GVHD, and extensive chronic GVHD were 30%,
162 63%, 11%, 15%, and 13%, respectively. The 3-year OS, cumulative incidence of relapse, and
163 non-relapse mortality rates were 78% (95% CI: 62–87%), 20% (95% CI: 10–33%), and 11%

164 (95% CI: 4–22%), respectively. After excluding two patients with aplastic anemia, we
165 divided patients with respect to disease status at SCT. The 3-year OS, cumulative incidence
166 of relapse, and non-relapse mortality rates of patients in CR (n = 24) were 82%, 22%, and
167 13%, (95% CI: 59–93%, 8–41%, and 3–30%), respectively. The 3-year OS, cumulative
168 incidence of relapse, and non-relapse mortality rates of patients in non-CR (n = 20) were 70%
169 (95% CI: 45–85%), 20% (95% CI: 6–40%), and 10% (95% CI: 2–28%), respectively (Figure
170 1B, C, D). Next, we divided the patients with respect to the number of human leucocyte
171 antigen (HLA) mismatches for graft-versus-host direction. The cumulative incidence of grade
172 II–IV acute GVHD in patients who underwent CBT with more than two HLA antigen
173 mismatches in the HLA-A, B, C, and DRB1 loci (n = 15) was 80% (95% CI: 46–94%), which
174 was significantly higher than that of the other patients (48%, 95% CI: 30–65%, n = 31) ($p =$
175 0.02) (Figure 1E). The cumulative incidence of grade II–IV acute GVHD in patients who
176 underwent CBT with more than two HLA allele mismatches in the HLA-A, B, C, and DRB1
177 loci (n = 24) was 75% (95% CI: 51–88%), which was significantly higher than that of the
178 other patients (41%, 95% CI: 20–61%, n = 22) ($p = 0.02$) (Figure 1F).

179 *Impact of MPA AUCs on outcomes*

180 None of the cases discontinued MMF administration, except for one case of death before
181 day 30. No patients experienced MMF-related severe toxicities such as diarrhea and
182 myelosuppression. The AUC_{0-24} of MPA on day 7 (MPA7) was $49.4 \pm 24.7 \mu\text{g h/mL}$ (mean \pm

183 standard deviation). There was no association between MPA7 and the incidence of PIR,
184 engraftment rate, engraftment speed or acute GVHD after day 7 (data not shown). In the
185 landmark analysis on day 7, the rate of sepsis between day 7 and engraftment in patients with
186 MPA7 >60 $\mu\text{g h/mL}$ was 33% (95% CI: 9–60%), which was higher than that in the other
187 patients (6%, 95% CI: 1–18%) ($p = 0.02$) (Figure 2A). The AUC_{0-24} of MPA on day 21
188 (MPA21) was $47.7 \pm 20.3 \mu\text{g h/mL}$. After excluding three cases of graft failure, we divided
189 patients with respect to MPA21. In the landmark analysis on day 21, the cumulative incidence
190 of grade II–IV acute GVHD after day 21 in patients with MPA21 $\leq 30 \mu\text{g h/mL}$ was 80%
191 (95% CI: 34–96%), which was higher than that in the other patients (50%, 95% CI: 31–67%)
192 ($p = 0.04$) (Figure 2B). In the landmark analysis on day 21, the cumulative incidence of
193 HHV-6 reactivation after day 21 in patients with MPA21 $\leq 48 \mu\text{g h/mL}$ (median) was 50%
194 (95% CI: 28–68%), which was higher than that in the other patients (19%, 95% CI: 6–38%)
195 ($p = 0.03$) (Figure 2C). Only two patients developed HHV-6 encephalitis, and there was no
196 significant association between the development of HHV-6 encephalitis and the blood level
197 of MPA (data not shown). In the analysis of OS, the cumulative incidence of relapse, chronic
198 GVHD, and reactivation of cytomegalovirus, no significant association was found with the
199 blood level of MPA (data not shown).

200 Next, we surveyed the causes of low MPA levels. The MPA21 of patients with diarrhea on
201 day 21 was significantly lower than that of patients without diarrhea (35.8 vs. 54.5 $\mu\text{g h/mL}$,

202 $p = 0.006$) (Figure 3A). Patients with serum albumin levels <3 g/dL on day 21 tended to have
203 low MPA21 (37.8 vs. 53.8 $\mu\text{g h/mL}$, $p = 0.10$) (Figure 3B).

204 We assessed the efficacy of the dose escalation of MMF. After 2014, the MPA7 of eight
205 patients was <30 $\mu\text{g h/mL}$ and the MMF dose was escalated after day 7. In these patients, the
206 MPA21 of six patients was higher than their MPA7; the MPA 21 of one patient who had
207 moderate diarrhea was less than MPA7 and one patient died before day 21 (Figure 4A). The
208 cumulative incidence of grade II–IV acute GVHD of these eight patients was 43%, which
209 was lower than the incidence of all patients. The MPA7 of 13 patients was 30–60 $\mu\text{g h/mL}$
210 and the dose of MMF was not escalated after day 7. Among these patients, nine and four
211 patients had MPA21 less and more than their MPA7s, respectively (Figure 4B). The
212 cumulative incidence of grade II–IV acute GVHD of these 13 patients was 78%, which was
213 higher than the incidence of all patients.

214

215 *Correlation between AUC_{0-24} and single point concentration*

216 To analyze whether we could reduce the number of blood collections, we investigated the
217 correlation between AUC_{0-24} and single point concentration of MPA. We found that there was
218 stronger correlation between AUC_{0-24} and C_2 (Correlation coefficient = 0.737, $p = 8.69 \times 10^{-17}$),
219 compared with C_0 , C_1 and C_4 (Supplemental Figure 1A-D). However, there was no
220 significant association between C_2 on day 7 and the incidence of sepsis before engraftment,

221 and between C₂ on day 21 and the incidence of grade II-IV acute GVHD or HHV-6
222 reactivation (data not shown).

223

224 *Univariate and multivariate analyses for grade II–IV acute GVHD*

225 After excluding the patients with graft failure and the patients who developed grade II-IV
226 acute GVHD before day 21, we performed univariate and multivariate analyses for grade II–
227 IV acute GVHD after day 21. In the univariate analysis, HLA allele mismatch, non-CR at
228 SCT, and MPA level were associated with a high incidence of grade II–IV acute GVHD
229 (Table 2). Multivariate analysis revealed that non-CR at SCT was associated with a high
230 incidence of grade II–IV acute GVHD (Table 3).

231

232

233 **Discussion**

234 In the present study, GVHD prophylaxis with tacrolimus and MMF enabled a high
235 incidence of neutrophil engraftment and prolonged OS compared to those reported in a
236 nationwide survey in Japan.[4, 32] A previous report indicated that 3,000 mg/day of MMF
237 was associated with delayed neutrophil engraftment in CBT,[33] but there was no evidence of
238 an association between the dose of MMF and the rate of neutrophil engraftment. TBI is
239 reportedly necessary for enhanced engraftment in CBT.[34] We administered MMF at 30
240 mg/kg/day, and 93% of patients received 3–12 Gy of TBI, which may have improved the
241 incidence of neutrophil engraftment. Randomized studies are required to determine the
242 optimal dose of MMF and the best conditioning regimen.

243 The cumulative incidence of grade II–IV acute GVHD in the present study was higher than
244 that in previous reports.[4, 32] The main reason for this result might be the high percentage of
245 patients who underwent CBT with more than two antigen or allele mismatches in the HLA-A,
246 B, C, and DRB1 loci. In Japan, HLA mismatches are generally evaluated with HLA-A, B,
247 and DRB1 antigens, and fewer than three mismatches were permitted;[35] our institute
248 evaluated HLA mismatches in the same way. Our results suggest that HLA mismatch in CBT
249 should be evaluated with the HLA-A, B, C, and DRB1 alleles to avoid acute GVHD.

250 In the present study, we observed that the blood levels of MPA fluctuated widely among
251 patients after CBT. The kinetics of MMF are influenced by gastrointestinal damage caused by

252 conditioning, loss of appetite, decreased oral intake, and other drug interactions.[36] Change
253 from oral to intravenous MMF may be one strategy to stabilize blood level;[37] however,
254 intravenous MMF is not available in Japan.

255 The appropriate level of MPA in CBT is still not confirmed. We suggest that the AUC_{0-24}
256 on day 7 should be $<60 \mu\text{g h/mL}$ to avoid sepsis before engraftment and that the AUC_{0-24} on
257 day 21 should be $>30 \mu\text{g h/mL}$ to prevent acute GVHD. This range is similar to the adequate
258 AUC_{0-24} used in kidney transplantation.[6, 7]

259 CBT is one risk factor of HHV-6 encephalitis,[38] and epidemiologic evidence indicates
260 the association between HHV-6 reactivation and acute GVHD.[39] In the present study, the
261 cumulative incidence of HHV-6 reactivation was higher in patients with AUC_{0-24} on day 21 of
262 $\leq 48 \mu\text{g h/mL}$ (median) than in the other patients. Therefore, maintaining a high level of MPA
263 and suppressing graft-versus-host reaction may prevent HHV-6 reactivation.

264 Based on these results, we hypothesize that MPA level should be changed before and after
265 engraftment. In kidney transplantation, high MPA level is associated with myelosuppression
266 and infection.[7] Similarly, MPA level in CBT should not be high to prevent bacterial
267 infection before engraftment. After engraftment, alloreactive T cells become activated and
268 excessive activation of alloreactive T cells leads to acute GVHD and delays immune
269 reconstitution. Acute GVHD and delay of immune recovery cause HHV-6 reactivation.[39,
270 40] Moreover, treatment for GVHD also favors HHV-6 reactivation, as described with other

271 herpesviruses.[41] Therefore, MPA level should be high to suppress alloreactive T cells and
272 prevent acute GVHD and HHV-6 reactivation. These are hypotheses and should be further
273 explored.

274 After 2014, we escalated the dose of MMF of patients with low blood levels of MPA on
275 day 7 and succeeded in reducing the incidence of grade II–IV acute GVHD in those patients.
276 However, the levels on day 21 in some patients without dose escalation fell below the levels
277 measured on day 7 and many of them developed grade II–IV acute GVHD. More frequent
278 monitoring of MPA and dose adjustment of MMF is necessary, especially for patients with
279 diarrhea or poor nutrition, whose blood levels of MPA tend to be low.

280 We focused on AUC and collected blood from patients 4 times per day. To proceed to
281 multicenter study, we analyzed whether we could make the measurement simpler. As a result,
282 there was no significant association between single point concentration and the incidence of
283 acute GVHD or infection. Thus, AUC is a better marker to predict the risk of acute GVHD
284 and infection compared with single point concentration, and 4 times blood collection per day
285 is recommended.

286 Multivariate analysis revealed that non-CR at SCT was associated with the development of
287 grade II–IV acute GVHD, while a low MPA AUC₀₋₂₄ on day 21 was not. In some patients
288 with non-CR at SCT, the physicians decided to maintain low levels of MPA, expecting a
289 graft-versus-tumor effect of cord blood. This was the limitation of this small-scale

290 retrospective study.

291 The present study also had additional limitations. We measured whole-blood levels of MPA.
292 Levels of unbound form of MPA should be measured to evaluate the direct effect of MMF on
293 lymphocytes.[42] Moreover, we did not survey the single-nucleotide polymorphism (SNP)
294 for the genes coding inosine-5-monophosphate dehydrogenase (IMPDH). IMPDH is an
295 enzyme inhibited by MPA; SNPs in those genes influence the activity of IMPDH and
296 suppress lymphocyte proliferation.[43-45] In addition, our results should be interpreted with
297 caution because of the small sample size and the heterogeneous patient and transplant
298 backgrounds.

299 In conclusion, blood AUCs of MPA affected the incidence of grade II–IV acute GVHD,
300 sepsis before engraftment, and HHV-6 reactivation in CBT. A multicenter prospective study is
301 required to clarify the appropriate range of MPA AUCs.

302

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307 **Conflicts of interest statement**

308 The authors declare no conflicts of interest associated with this manuscript.

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311 **Data availability statement**

312 The data are not publicly available due to privacy or ethical restrictions.

313 **Author contributions**

314 HM, JK, YA and TK designed the research, organized the project, and drafted the
315 manuscript. HM and JK gathered the data and performed the statistical analysis. TS, MH, TY,
316 KY, KM, and ATK contributed to the data analysis and writing of the manuscript. All authors
317 approved the final version of the manuscript for submission for publication.

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477

479 **Table 1. Patient characteristics**

480

		Number (%) or Median (Range)
Age at SCT (years)		50 (20–66)
Sex	M/F	29 (63) / 17 (37)
Disease	AML/MDS	20 (43) / 17 (37)
	ALL	6 (13)
	NHL	8 (17)
	CML/MPN	1 (2) / 1 (2)
	ATL	1 (2)
	AA	2 (4)
PS at SCT	0/1	23 (50) / 14 (30)
	2–3	9 (20)
Serum Albumin at SCT (g/dL)		3.2 (1.9-4.2)
HLA Mismatch	0/1/2	1 (2) / 11 (24) / 10 (22)
(A, B, C, and DRB1 alleles)	3/4/5	11 (24) / 11 (24) / 2 (4)
Disease Status at SCT	CR/PR	24 (52) / 2 (4)
	Others	20 (43)
Days from Diagnosis to SCT	< 90	6 (13)
	90–180	14 (30)
	>180	26 (57)
Conditioning	MAC	24 (52)
	RIC	22 (48)
TBI	0	3 (7)
	3–4	21 (46)
	10–12	22 (48)
Total Nuclear Cell Count (×10 ⁷ /kg)		2.47 (1.94–4.27)
CD34 ⁺ Cell Count (×10 ⁵ /kg)		0.57 (0.18–1.91)

481

482 Abbreviations: SCT: stem cell transplantation, M: male, F: female, AML: acute myeloid

483 leukemia, MDS: myelodysplastic syndrome, ALL: acute lymphoblastic leukemia, NHL:

484 non-Hodgkin lymphoma, CML: chronic myeloid leukemia, MPN: myeloproliferative
485 neoplasm, ATL: adult T cell leukemia/lymphoma, AA: aplastic anemia, PS: performance
486 status, HLA: human leucocyte antigen, CR: complete remission, PR: partial remission, MAC:
487 myeloablative conditioning, RIC: reduced-intensity conditioning, TBI: total body irradiation.
488

489 **Table 2. Univariate analysis for grade II–IV acute GVHD**

Variables	Number	HR	95% CI	<i>P</i>
Age at SCT (years)				
<50	20	1		
≥50	20	1.55	0.72–3.35	0.27
PS at SCT				
0–1	34	1		
2–4	6	0.56	0.16–1.90	0.35
HCT-CI at SCT				
0–1	31	1		
>2	9	0.6	0.22–1.63	0.31
Sex mismatch				
Match, M to F	34	1		
F to M	6	0.99	0.61–1.62	0.98
HLA allele mismatch				
0–2	20	1		
3–5	20	2.26	1.05–4.86	0.04
Disease status at SCT				
CR	22	1		
Non-CR	18	1.29	1.16–1.45	<0.001
Days from diagnosis to SCT				
0–180	16	1		
>180	24	1.01	0.58–1.75	0.98
Conditioning				
MAC	22	1		
RIC	18	0.92	0.41–2.07	0.85
TBI				
No	2	1		
Yes	38	2.41	0.31–18.55	0.40
AUC ₀₋₂₄ of MPA on day 21				
≤30 µg h/mL	10	1		
>30 µg h/mL	30	0.38	0.15–0.91	0.03
CD34 ⁺ cell count				
≤ 0.57×10 ⁵ /kg		1		
> 0.57×10 ⁵ /kg		1.27	0.62–2.60	0.51

490

491 Abbreviations: GVHD: graft-versus-host disease, HR: hazard ratio, CI: confidence interval,
492 SCT: stem cell transplantation, PS: performance status, HCT-CI: comorbidity index of
493 hematopoietic cell transplantation, M: male, F: female, HLA: human leucocyte antigen, CR:
494 complete remission, MAC: myeloablative conditioning, RIC: reduced-intensity conditioning,
495 TBI: total body irradiation, AUC₀₋₂₄: 24-hour area under the curve, MPA: mycophenolic acid.

496 **Table 3. Multivariate analysis for grade II–IV acute GVHD**

497

Variables	Number	HR	95% CI	<i>P</i>
HLA allele mismatch				
0–2	20	1		
3–5	20	2.14	0.96–4.79	0.06
Disease status at SCT				
CR	22	1		
Non-CR	18	1.26	1.07–1.48	0.006
AUC ₀₋₂₄ of MPA on day 21				
≤30 µg h/mL	10	1		
>30 µg h/mL	30	0.66	0.24–1.78	0.41

498

499 Abbreviations: GVHD: graft-versus-host disease, HR: hazard ratio, CI: confidence interval,

500 HLA: human leucocyte antigen, SCT: stem cell transplantation, CR: complete remission,

501 AUC₀₋₂₄: 24-hour area under the curve, MPA: mycophenolic acid.

502

503 **Figure legends**

504 **Figure 1. Use of mycophenolate mofetil enabled high neutrophil engraftment and**
505 **feasible overall survival rates.**

506 (A) Cumulative incidence of engraftment. (B) Kaplan–Meier curves of overall survival of
507 patients in CR and non-CR at transplantation. (C) Cumulative incidence of relapse and (D)
508 non-relapse mortality of patients in CR and non-CR. (E) Cumulative incidence of grade II–IV
509 graft-versus-host disease and the number of HLA antigen mismatches (A, B, C, and DRB1).
510 (F) Cumulative incidence of grade II–IV graft-versus-host disease and the number of HLA
511 allele mismatches (A, B, C and DRB1).

512 Abbreviations: CR: complete remission. HLA: human leukocyte antigen.

513

514 **Figure 2. AUC₀₋₂₄ of MPA affected the incidence of sepsis, grade II–IV acute GVHD, and**
515 **reactivation of human herpesvirus 6.**

516 (A) AUC₀₋₂₄ on day 7 and cumulative incidence of sepsis before neutrophil engraftment. (B)
517 AUC₀₋₂₄ on day 21 and cumulative incidence of grade II–IV graft-versus-host disease. (C)
518 AUC₀₋₂₄ on day 21 and cumulative incidence of reactivation of human herpesvirus 6.

519 Abbreviations: AUC₀₋₂₄: 24-hour area under the curve, MPA: mycophenolic acid, GVHD:
520 graft-versus-host disease.

521 **Figure 3. Diarrhea and poor nutrition decreased AUC₀₋₂₄ of MPA on day 21.**

522 (A) Diarrhea and box plot of AUC₀₋₂₄ on day 21. (B) Serum albumin levels and box plot of
523 AUC₀₋₂₄ on day 21. Abbreviations: AUC₀₋₂₄: 24-hour area under the curve, MPA:
524 mycophenolic acid.

525

526 **Figure 4. Dose escalation for low AUC₀₋₂₄ on day 7 increased the AUC₀₋₂₄ on day 21.**

527 (A) AUC₀₋₂₄ of MPA on days 7 and 21 among patients with MPA₇ (AUC₀₋₂₄ of MPA on day
528 7) <30 µg h/mL. The MMF dose of these patients was escalated after day 7. (B) AUC₀₋₂₄ on
529 days 7 and 21 of patients with MPA₇ of 30–60 µg h/mL. The MMF dose of these patients
530 was not escalated after day 7.

531 Abbreviations: AUC₀₋₂₄: 24-hour area under the curve, MPA: mycophenolic acid, MMF:
532 mycophenolate mofetil.

533

534 **Supplemental Figure 1. Correlation between AUC₀₋₂₄ and single point concentration.**

535 Correlation between AUC₀₋₂₄ and (A) C₀, (B) C₁, (C) C₂ and (D) C₄.

536 Abbreviations: AUC₀₋₂₄: 24-hour area under the curve. *R*: correlation coefficient.

537