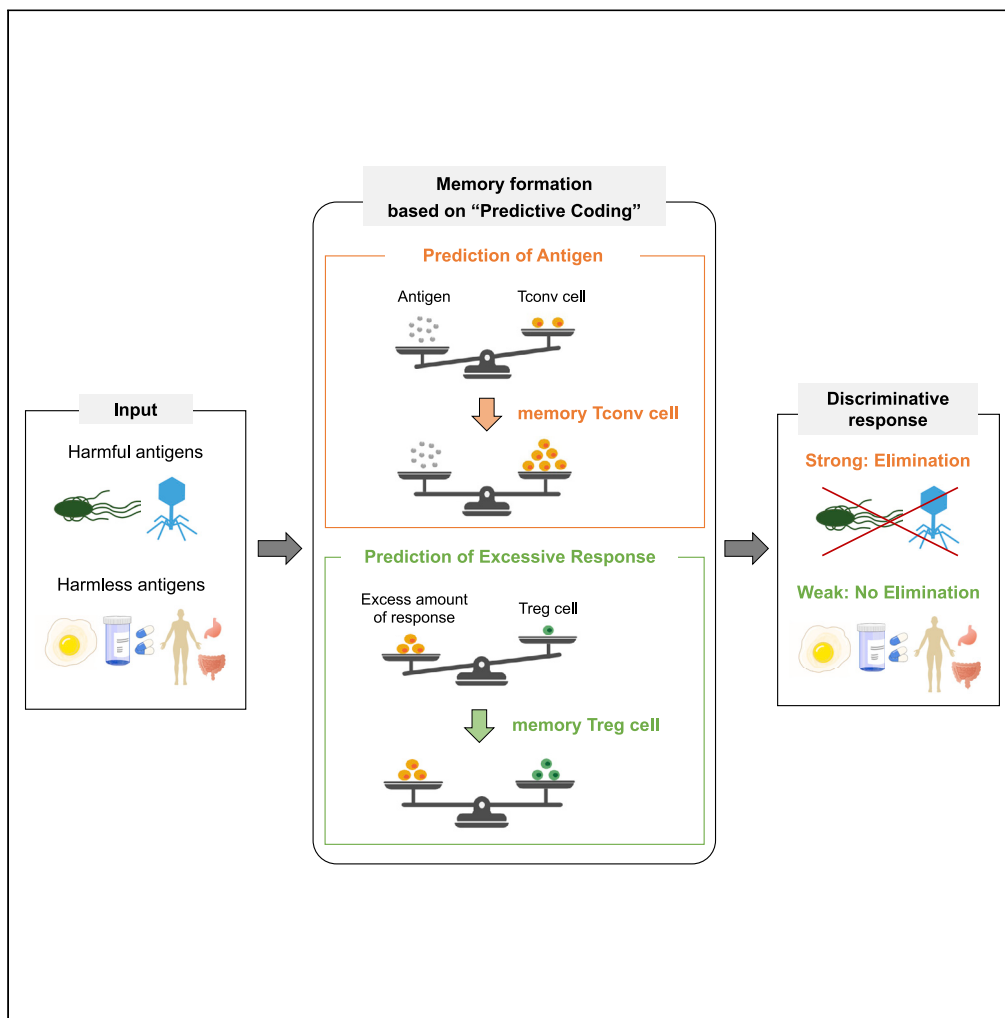## Article

Adaptive discrimination between harmful and harmless antigens in the immune system by predictive coding

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Highlights

Mathematical model of immune response by T-cell population dynamics was developed

Immunological memory is hypothesized to be formed based on predictive coding

Predictive coding mechanism led to discrimination between harmful/harmless antigens

The model also reproduced the onset and therapy of allergy

Article

Adaptive discrimination between harmful and harmless antigens in the immune system by predictive coding

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SUMMARY

The immune system discriminates between harmful and harmless antigens based on past experiences; however, the underlying mechanism is largely unknown. From the viewpoint of machine learning, the learning system predicts the observation and updates the prediction based on prediction error, a process known as "predictive coding." Here, we modeled the population dynamics of T cells by adopting the concept of predictive coding; conventional and regulatory T cells predict the antigen concentration and excessive immune response, respectively. Their prediction error signals, possibly via cytokines, induce their differentiation to memory T cells. Through numerical simulations, we found that the immune system identifies antigen risks depending on the concentration and input rapidness of the antigen. Further, our model reproduced history-dependent discrimination, as in allergy onset and subsequent therapy. Taken together, this study provided a novel framework to improve our understanding of how the immune system adaptively learns the risks of diverse antigens.

INTRODUCTION

The immune system faces the challenge of identifying unknown risks of diverse antigens and inducing proper immune responses. For harmful antigens, such as pathogens, the immune system induces strong immune responses for their elimination, whereas, for harmless antigens, such as food and self-antigens, it does not lead to strong responses to prevent unnecessary inflammation. Thus, the immune system should discriminate between harmful and harmless antigens appropriately. Defects in this discrimination induce immune diseases, including allergies and autoimmune diseases.[1,2] However, the mechanism by which the immune system distinguishes between harmful and harmless antigens upon exposure to numerous antigens remains to be understood. This study aimed to explore this field through computational modeling of T-cell population dynamics and we first introduced into immunology the concept that the immune system predicts its environment using predictive coding.

The central organizers of adaptive immunity are T cells, each of which expresses different T-cell receptors (TCRs) to specifically recognize antigens presented by antigen-presenting cells, such as dendritic cells (DCs).[3–5] Through the process of T-cell differentiation, the cells responsive to self-antigens are eliminated[6–8]; however, there still remain those that are specific not only to harmful antigens but also to harmless ones. Namely, such T cells have no way of knowing whether the antigen is harmful or harmless. Nevertheless, the immune system responds strongly to harmful foreign antigens but not to harmless ones. Therefore, we focused on the fact that the antigen specificity of T cells cannot explain the mechanism by which the immune system discriminates between harmful and harmless antigens.

The immune response is organized by the population dynamics of various cell types (Figure 1A). It is initiated by antigen-presenting cells, such as DCs, which take in antigens and present them to T cells. Naive T (T_{naive}) cells, with TCRs on their surface, recognize specific antigens presented by DCs. T_{naive} cells then differentiate into various types of T cells, such as conventional T (T_{conv}) cells and regulatory T (T_{reg}) cells, depending on cytokines, such as interleukins (ILs), in their microenvironment.[9] T_{conv} and T_{reg} cells play distinct roles in immune responses; T_{conv} cells, including T-helper (Th) 1, Th2, Th17 cells, accelerate immune responses, leading to the elimination of antigens by activating downstream cells, such as B cells and killer T cells,[9–11] whereas T_{reg} cells work as a brake for immune responses via the regulation of DCs and

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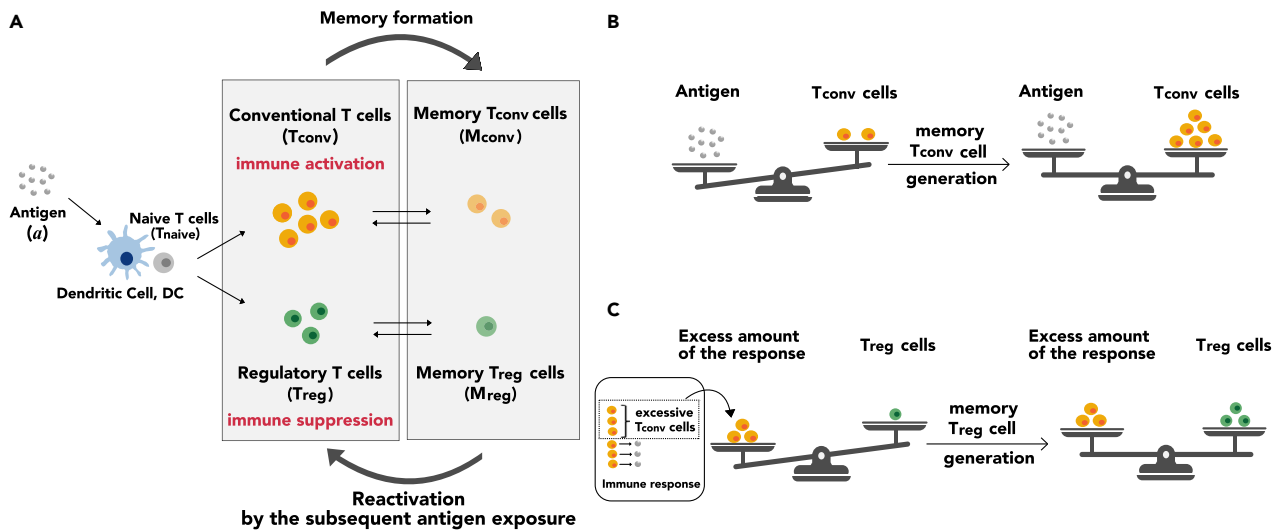https://doi.org/10.1016/j.isci.2022.105754

Figure 1. Scheme of the predictive immune memory model

(A) Population dynamics model of T cells in response to antigen input. The model includes the differentiation of T_{naive} cells into T_{conv} and T_{reg} cells by antigen-presenting cells such as dendritic cells (DCs), the differentiation of T_{conv} and T_{reg} cells into memory T cells, and the reactivation of memory T cells into T cells upon subsequent exposure to antigens.

(B and C) Predictive coding-based immunological memory formation. (B) Generation of memory T_{conv} cells. Memory T_{conv} cells are generated based on the prediction error $e_c|a - m_c T_{conv}|_+$. In other words, the production of memory T_{conv} cells is induced when the concentration of antigens is excessive compared to that of T_{conv} cells in order to efficiently eliminate antigens. (C) Generation of memory T_{reg} cells. Memory T_{reg} cells are generated based on the prediction error $e_r|g(T_{conv}) - a - m_r T_{reg}|_+$. In other words, the production of memory T_{reg} cells is induced when the excess amount of response, evaluated by the difference between the intensity of T_{conv} cell activation ($g(T_{conv})$) and antigen concentration, is larger than the concentration of T_{reg} cells in order to prevent unnecessary inflammation.

suppressive cytokines.[12,13] Note that this kind of T_{reg} cells is called induced T_{reg} cells, and their differentiation pathway is different from that of naturally occurring T_{reg} cells, which specifically respond to self-antigens. Following the immune response, most of the induced T_{conv} and T_{reg} cells are removed by apoptosis[14,15]; however, a small population of them differentiates into memory T cells, namely memory T_{conv} cells and memory T_{reg} cells, and they persist in the body for a long time.[16,17] Upon subsequent encounters with the same antigen, memory T cells are rapidly activated, resulting in more efficient responses.[18,19] This is termed immunological memory. Thus, the combinatorial dynamics of various types of T cells determine the intensity of the immune response. In other words, the discrimination between harmful and harmless antigens must be achieved at the level of T-cell population dynamics.

Discrimination between harmful and harmless antigens for each antigen is not always constant and varies in antigen experience-dependent manner. A prominent example is the onset and therapy of allergy, which is defined as an excessive response to harmless antigens, including pollen and mites. Although allergens, defined as substances that cause allergy, are initially regarded as harmless in our body, response to them can intensify upon repeated exposures, leading to allergic symptoms. Such a change in responsiveness indicates that the immune discrimination of allergens can change from harmless to harmful. Furthermore, allergic symptoms, the immune responses to allergens, can be weakened by allergen immunotherapy,[20–22] in which a small amount of allergen extract (not enough to cause symptoms) is repeatedly administered to the patients; after the therapy, allergic symptoms do not occur even when patients are exposed to large amounts of the allergen. This means that discrimination can be reversed from harmful to harmless through allergen immunotherapy. Thus, the immune system adaptively changes discrimination depending on the temporal history of antigens. Experimentally and clinically, allergen immunotherapy has been reported to induce regulatory cell populations, such as T_{reg} cells, and suppressive cytokines, such as IL-10.[23–25] However, the mechanisms by which immune discrimination is adaptively updated by antigen experience largely remain unclear.

The immune system can be viewed as an adaptive learning system that updates the discrimination of antigen risk. To induce the most appropriate responses, the immune system needs to predict and prepare for the subsequent invasion of antigens by the formation of memory cells. From the perspective of the machine

learning theory, a more accurate prediction is achieved by repeated observation and prediction, in which the prediction is updated based on prediction error, which is the difference between observation and prediction. This concept, called "predictive coding," was originally proposed in neuroscience[26] and has been widely accepted as a guiding principle for understanding learning systems, such as brain and artificial intelligence.[27–29] In this study, we adopted this concept to understand the immune system as a learning system. We hypothesized that T_{conv} and T_{reg} cells predict the risk of antigens and excessive response, respectively, and their predictions can be updated by prediction errors via the production of memory T cells.

Based on the idea of predictive coding, this study aimed to address how the immune system discriminates between harmful and harmless antigens and how it changes its response depending on the history of antigens. We developed a mathematical model of antigen-induced T-cell population dynamics named "the predictive immune memory model." By simulating the model, we demonstrated that the immune system can discriminate between harmful and harmless antigens using the predictive coding mechanism in an antigen concentration- and input rapidness-dependent manner. The model also demonstrated antigen history-dependent immune discrimination, as seen in the onset and therapy of allergy. Furthermore, we found that the dose-response of T-cell activation does not affect the outcome of allergen immunotherapy but changes its persistence upon additional higher exposure to allergens.

RESULTS

Mathematical model for T-cell population dynamics

To examine how the immune system discriminates between harmful and harmless antigens at the level of the T-cell population, we developed a mathematical model for the population dynamics of T cells and named it "the predictive immune memory model" (Figure 1A). The model consists of T_{conv}, T_{reg}, and their memory cells. T_{conv} and T_{reg} cells are generated by the differentiation of T_{naive} cells, activation of memory T cells, and their proliferation, as shown later in discussion.

$$\frac{d}{dt}T_{conv} = -d_c T_{conv} + \frac{D_c}{1 + s_r T_{reg}}T_{conv} + k_c T_{naive}a + w_c M_{conv}a - E_c T_{conv}, \quad \text{(Equation 1)}$$

$$\frac{d}{dt}T_{reg} = -d_r T_{reg} + \frac{D_r}{1 + s_c T_{conv}}T_{reg} + k_r T_{naive}a + w_r M_{reg}a - E_r T_{reg}, \quad \text{(Equation 2)}$$

where T_{conv} and T_{reg} represent the populations of T_{conv} and T_{reg} cells, respectively; M_{conv} and M_{reg} represent the populations of memory T_{conv} and memory T_{reg} cells, respectively; T_{naive} indicates a positive constant which represents the population of T_{naive} cells; a represents the concentration of antigen input; d_i, k_i, and w_i ($i \in \{c, r\}$) indicate the rates of death due to apoptosis, differentiation from T_{naive} cells, and production of T cells from memory T cells, respectively. In addition, the second terms represent the proliferation of T_{conv} and T_{reg} cells, which are inhibited by each other through some possible mechanisms, such as the competition for limited sources of cytokines (IL-2) and contact with DCs,[30–32] where D_i and s_i ($i \in \{c, r\}$) represents proliferation rate of each T cell itself and the rate of suppression to the counterparts, respectively. The fifth terms represent the decrease of T cells by their differentiation into memory T cells, as described later in discussion. Memory T cells differentiate from T_{conv} and T_{reg} cells as

$$\frac{d}{dt}M_{conv} = -d_{mc}M_{conv} + E_c T_{conv}, \quad \text{(Equation 3)}$$

$$\frac{d}{dt}M_{reg} = -d_{mr}M_{reg} + E_r T_{reg}, \quad \text{(Equation 4)}$$

where d_{mc} and d_{mr} indicate the death rates of memory T_{conv} and memory T_{reg} cells, respectively. We regarded their death rates as zero in the time span of our simulations due to the longevity of memory T cells ($d_{mc} = d_{mr} = 0$). Note that E_c and E_r are not constant parameters but are situation-dependent, following the idea of predictive coding (see the next section for details). In this model, we defined the intensity of response R, which is positively and negatively regulated by T_{conv} and T_{reg} cells, respectively, as

$$\frac{d}{dt}R = -(r_0 + r_s T_{reg})R + r_a T_{conv}, \quad \text{(Equation 5)}$$

where r_0 indicates a positive constant, which causes the convergence of R to zero in the absence of T_{conv} and T_{reg} cells; r_a, and r_s indicate the activation rates by T_{conv} cells and suppression rates by T_{reg} cells, respectively. Although we artificially defined the intensity R, we can biologically interpret r_0 as the amounts of other types of T_{reg} cells called naturally occurring T_{reg} cells, which possibly contribute to the suppression of excessive inflammation, and r_a and r_s could correspond to the amount of cytokines from T_{conv} cells which activate the response and those from T_{reg} cells which suppress the response, respectively.

Predictive coding scheme

We have introduced the concept of predictive coding under the hypothesis that the immune system predicts the level of antigen exposure and its consequent inflammation in an antigen experience-dependent manner. More specifically, the predictive coding scheme states that T_{conv} and T_{reg} cells are predictors of the antigen amount and excess amount of immune response, respectively, and that their predictions are updated based on prediction errors via the formation of memory T_{conv} and memory T_{reg} cells.

Since T_{conv} cells are the control center to achieve antigen elimination by inducing downstream reactions, they must be adequately controlled depending on the change in antigen concentration; when the concentration of antigens is excessive compared to that of T_{conv} cells, more T_{conv} cells need to be generated to completely eliminate the antigens in our hypothesis (Figure 1B). Accordingly, the production rate of memory T_{conv} cells can be described by

$$E_c = e_c |a - m_c T_{conv}|_+, \qquad \text{(Equation 6)}$$

where e_c and m_c indicate positive constants and $|x|_+$ represents ramp function (i.e., $|x|_+ = 0$ $(x < 0)$, x $(x \geqq 0)$). Note that E_c is the prediction error of antigen concentration, since a and $m_c T_{conv}$ represent the observation and prediction of the antigen concentration, respectively. Thus, memory T_{conv} cells are upregulated by the prediction error E_c (Equation 3).

On the other hand, T_{reg} cells play an important role in the prevention of excessive immune responses. Thus, their amount should be regulated based on the intensity of the response; when the excess amount of the immune response is larger than the concentration of T_{reg} cells, more T_{reg} cells need to be generated to suppress the excessive immune responses in our hypothesis (Figure 1C). Therefore, the production rate of memory T_{reg} cells can be described by

$$E_r = e_r |f(T_{conv}, a) - m_r T_{reg}|_+, \qquad \text{(Equation 7)}$$

where e_r and m_r indicate positive constants, and $f(T_{conv}, a) = g(T_{conv}) - a$ represents the excess amount of the immune response compared to antigen concentration. Here, we assumed that T_{reg} cells evaluated the level of T_{conv} cell activation by $g(T_{conv}) = A_{max} T_{conv}/(T_{conv} + K)$, where A_{max} and K indicate positive constants. Note that E_r is the prediction error of the excess amount of immune response, since $f(T_{conv}, a)$ and $m_r T_{reg}$ represent the observation and prediction of the excess amount of immune response, respectively. Thus, memory T_{reg} cells were upregulated by the prediction error E_r (Equation 4). Notably, we hypothesized that the generation of memory T cells (not T cells) reflects the calculation of the prediction error since memory T cells rather than T cells remain for a long time serving as immunological memory.

As an implementation of memory formation based on predictive coding, we assumed that the calculation of predictive coding can be achieved by cytokines. Cytokines secreted from immune cells determine their differentiation and proliferation under communication across various types of immune cells.[9,33] In this study, we regarded cytokines as the medium for transmitting this quantitative information. Specifically, the amounts of T_{conv} and T_{reg} cells can be coded by the concentration of cytokines secreted by themselves, whereas the amounts of antigens can be coded by cytokines secreted from antigen-presenting cells, such as DCs and macrophages. Based on these information-carrying cytokines, we hypothesized that the information obtained from each kind of cytokines is integrated into T cells and that prediction errors are computed through intracellular signal transduction in T cells. The parameters used in the numerical simulations are provided in Table S1. Although all of the parameters were just our assumption, we validated them by the parameter sensitivity analysis (Figures S1 and S2).

Concentration-dependent discrimination between harmful and harmless antigens

To examine the difference between harmful and harmless antigens for the immune system, we focused on the effect of antigen concentration on the immune response. We simulated the model with high and low
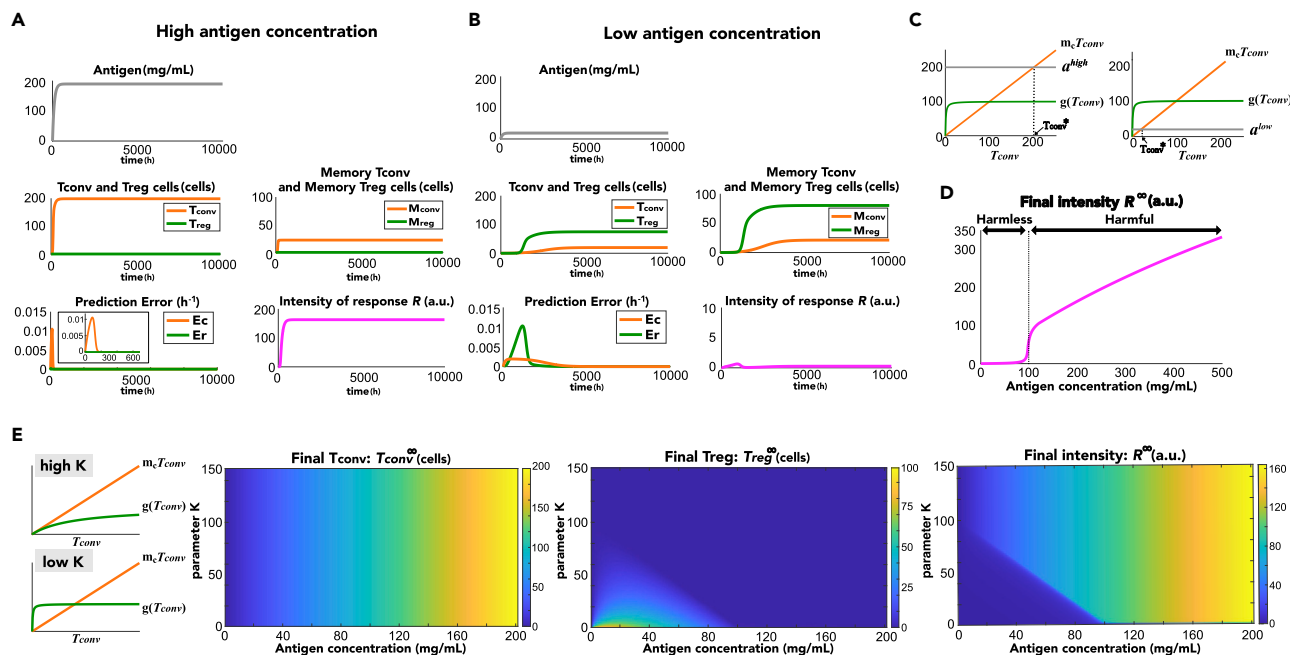
Figure 2. Antigen concentration-dependent immune discrimination

(A and B) Immune responses simulated with the exposure of (A) high and (B) low concentration of antigens. The inset shows an enlarged view of the prediction error in the early phase.

(C) Diagram for predictive coding-based memory formation. Antigen concentration is predicted by T_{conv} cells (orange line). When antigen concentration is observed (gray line), positive prediction error ($a - m_c T_{conv}$) increases memory T_{conv} cells, until T_{conv} converges to T_{conv}^* at the intersection of orange and gray lines. On the other hand, T_{reg} cells are assumed to predict the excess amount of immune response. The green line indicates the intensity of T_{conv} cell activation evaluated by T_{reg} cells. The excess amount of immune response is evaluated by the difference between this intensity of T_{conv} cell activation and observed antigen concentration. When antigen concentration is observed (gray line), positive prediction error between observed excess amount of immune response ($g(T_{conv}) - a$) and its prediction ($m_r T_{reg}$) increases memory T_{reg} cells, until the prediction error converges to zero. The left and right panels show the diagram for the case in which the observed antigen concentrations are high and low, respectively. Notably, different formulations of T_{conv} cell activation evaluated in memory T_{conv} and T_{reg} cell formation (orange and green lines, respectively) is necessary to the accumulation of memory T_{reg} cells under the exposure of low concentration of antigens.

(D) Change in intensity of immune responses depending on antigen concentrations. Convergence value of the intensity R is plotted upon the steady exposure to each antigen concentration.

(E) Steady-state responses of T_{conv} and T_{reg} populations and the immune intensity R depending on antigen concentration and a parameter K in $g(T_{conv})$. Left panels visually represent how the parameter K in $g(T_{conv})$ changes the diagram for predictive coding-based memory formation as in (C), where we can regard $m_c T_{conv}$ (orange line) and $g(T_{conv})$ (green line) as the evaluation of T_{conv} cell activation in memory T_{conv} and memory T_{reg} cell generation, respectively.

concentrations of antigen input (Figures 2A and 2B) and found that the steady exposure of high and low concentrations of antigens caused more accumulation of memory T_{conv} and memory T_{reg} cells, respectively. At high antigen concentrations (Figure 2A), memory T_{conv} cells were generated until the prediction error $e_c |a - m_c T_{conv}|_+$ was minimized to zero. Memory T_{reg} cells were not generated, since the prediction error $e_r |g(T_{conv}) - a - m_r T_{reg}|_+$ was always zero (left panel in Figure 2C). Therefore, the intensity of immune response R converged to a high level. On the other hand, at low antigen concentrations (Figure 2B), memory T_{conv} cells were produced, similar to the exposure of high antigen concentration. Memory T_{reg} cells were generated more since the prediction error $e_r |g(T_{conv}) - a - m_r T_{reg}|_+$ was positive, and the generation of memory T_{reg} cells continued until the prediction error was minimized to zero (right panel in Figure 2C). Therefore, the intensity of immune response was low. To summarize the immune responses depending on antigen concentrations, there was threshold of the antigen concentration ($a \fallingdotseq 100$), indicating that immune responses were specifically suppressed under low concentration of antigen exposures (Figure 2D). These results suggested that the immune system with predictive coding discriminates between harmful and harmless antigens based on antigen concentration.

Next, we examined the conditions necessary for the immune system to properly distinguish between harmful and harmless antigens depending on their concentration. We performed antigen concentration-dependent simulations by varying the parameter K, which regulated the T_{reg} cell-estimated level of
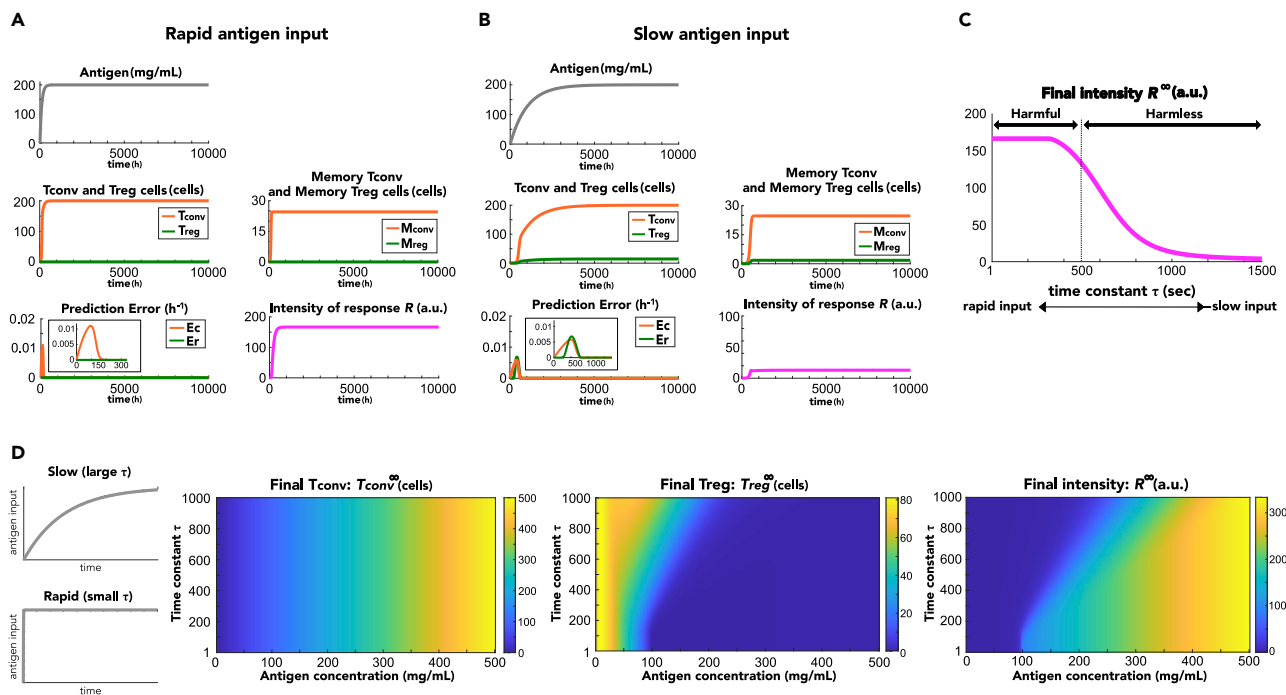
Figure 3. Antigen input rapidness-dependent immune discrimination

(A and B) Immune responses to (A) rapid and (B) slow inputs of high antigen concentration. Antigens were administered as $a(t) = a_0(1 - e^{-t/\tau})$, where τ indicates the time constant. Insets show an enlarged view of the prediction error in the early phase.

(C) Change in intensity of immune responses depending on time constants of antigen administration. Convergence value of the intensity R is plotted at each time constant. High antigen concentrations ($a_0 = 200$) were administered.

(D) Steady-state response of T_{conv} and T_{reg} populations and the immune intensity R depending on antigen concentration a_0 and time constant τ. Left panels visually represent how time constant τ affects rapidness of antigen inputs.

T_{conv} cell activation in memory T_{reg} generation (left panels in Figure 2E). We found that antigen discrimination could be achieved only with low K, in which immune responses were specifically suppressed under low concentration of antigen exposures (Figure 2E). Because T_{reg} cells underestimated and overestimated the level of T_{conv} cell activation in low antigen concentration with high and low K, respectively (left panels in Figure 2E), this result suggested that suppressive immune responses at low concentration of antigens can be achieved by the overestimation of T_{conv} cell activation in memory T_{reg} generation compared to the estimation in memory T_{conv} generation at low antigen concentration. Note that the relative values of K and m_c strictly determine whether antigen concentration-dependent discrimination is achieved since the overestimation of T_{conv} cell activation is defined by the relative estimation of T_{conv} cell activation in memory T_{conv} cell generation and that in memory T_{reg} cell generation. This also means that once the overestimation of T_{conv} cell activation in memory T_{reg} cell generation is satisfied by K and m_c, antigen concentration-dependent discrimination (seen in Figure 2D) can be robustly achieved without depending on other parameters, which we also verified by the parameter sensitivity analysis (Figures S1 and S2).

Input rapidness-dependent discrimination between harmful and harmless antigens

We focused on the rapidness of antigen input (the speed of antigen input) as another possible factor for discrimination between harmful and harmless antigens. We simulated the model in response to antigen inputs with different time constants (Figures 3A and 3B). Similar to that in Figure 2A, the intensity of immune response was high upon rapid exposure to high concentrations of antigens (Figure 3A). However, when the concentration of antigens increased slowly, eventually reaching a high concentration, the intensity of the immune response became weaker (Figure 3B). This was because the slowly increasing antigen input enabled the immune system to have a longer experience of low antigen concentration before reaching a high concentration, which caused a positive prediction error in memory T_{reg} cell generation followed by the production of memory T_{reg} cells.

Next, we examined input rapidness-dependent immune responses under exposure to the same high concentration of antigens with different input time constants and found that there was a threshold of time constant for discrimination between harmful and harmless antigens ($\tau \fallingdotseq 500$) (Figure 3C). This result showed that even when the final concentration was high, the immune system could recognize the antigens with slow input as harmless. To summarize the results, we examined the immune responses depending on both the antigen concentration and its input rapidness (Figure 3D). We found that low concentrations of antigens induced suppressive responses independent of their input rapidness. On the other hand, high concentrations of antigens induced responses with different intensities depending on their input rapidness; the immune system caused strong responses to rapidly increasing antigens while it caused suppressive responses to slowly increasing antigens. In addition, we examined immune discrimination with a time delay in memory formation (see the STAR Methods section) and demonstrated that the discrimination between harmful and harmless antigens based on the antigen concentration and its input rapidness was similarly achieved with a time delay in memory formation (Figures S3 and S4). Together, these results suggested that the immune system discriminates between harmful and harmless antigens based on their input rapidness as well as their concentration.

History-dependent discrimination between harmful and harmless antigens

Discrimination between harmful and harmless antigens is not invariable throughout our life span, in other words, discrimination can change depending on experiences of antigen exposure: antigen history. For example, at the onset of allergy, discrimination of the same antigen changes from harmless to harmful, whereas its discrimination can be reversed by allergen immunotherapy. To examine the mechanism of antigen history-dependent changes in immune discrimination, we simulated the immune responses to successive but different patterns of antigen exposure (Figure 4A). Specifically, we applied rapid exposure to high concentrations of antigens inducing allergy, followed by exposure to low concentrations of antigens, as allergen immunotherapy, and subsequently, rapid exposure to high concentrations of antigens again. The final input was provided to examine the effect of allergen immunotherapy.

After the first exposure to high concentrations of antigens, a strong immune response was induced due to the positive prediction error in memory T_{conv} cell generation, as shown in antigen concentration- and input rapidness-dependent discrimination. Upon exposure to a low concentration of antigens thereafter, more T_{conv} cells were produced than T_{reg} cells at the initiation of therapy due to the accumulated memory T_{conv} cells. In contrast, T_{reg} cells were gradually generated since a low concentration of antigens achieved a positive prediction error in memory T_{reg} cell generation. Accordingly, exposure to low concentrations of antigens for a certain period of time enabled the accumulation of memory T_{reg} cells. Therefore, even when the immune system was exposed to high concentrations of antigens again, more T_{reg} cells were generated, and the intensity of the immune response became weak. This result indicated that the simulation successfully reproduced the immune response at the onset of allergy and the effect of allergen immunotherapy. In summary, the immune system could discriminate between harmful and harmless antigens upon the first exposure to antigens in an antigen concentration- and input rapidness-dependent manner. Furthermore, the discrimination could adaptively change due to memory formation, based on predictive coding, in an antigen history-dependent manner.

Furthermore, we examined how therapeutic strategies influence the effect of allergen immunotherapy by evaluating the ratio of maximum intensity R in response to antigen input after therapy to that before therapy. We found that allergen immunotherapy was effective only when a low antigen dose was administered for the therapy (Figure 4B), which is consistent with the accumulation of memory T_{reg} cells at low antigen concentrations, as shown in Figure 2. Additionally, we examined the effect of both antigen concentration and input rapidness in allergen immunotherapy on the therapeutic effect (Figure 4C) and found that a low concentration and/or slow input enabled effective allergen immunotherapy. These findings could explain the validity of therapeutic strategies currently used in numerous clinical settings where antigen administration is initiated at a low dose and then gradually increased in the early phases of allergen immunotherapy with the aim of avoiding allergic symptoms during the therapy.[34–36]

The property of T-cell activation affects history-dependent discrimination

So far, the model has assumed that T-cell activation is linearly associated with antigen concentration. However, antigen concentration-dependent of T-cell activation might follow various types of activation patterns since the difference in ligands and its consequent difference in binding properties to TCRs largely affect the
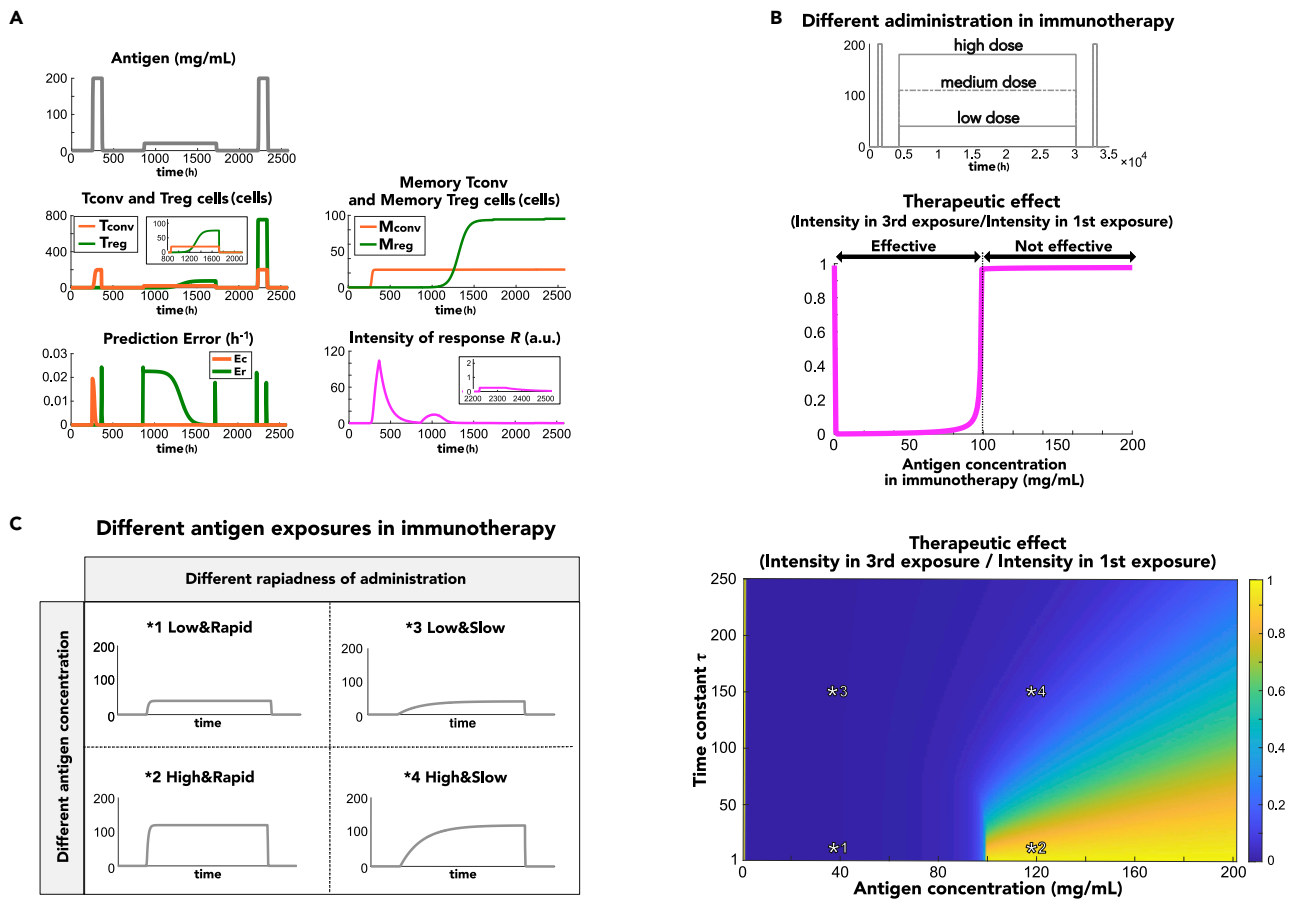
Figure 4. Antigen history-dependent immune discrimination in allergen immunotherapy

(A) Temporal change in immune responses to a series of different antigen inputs. The first antigen input was high enough for the induction of allergy, the second one was applied for allergen immunotherapy, and the third one was for checking the therapeutic effect. Insets show an enlarged view of the populations of T_{conv} and T_{reg} cells during allergen immunotherapy and the intensity of the response during the third antigen input.

(B) Therapeutic effects depending on antigen concentration administered during allergen immunotherapy. Therapeutic effect was evaluated by the ratio of maximum immune intensity R in the third antigen input to that in the first antigen input. A ratio smaller than one indicates the success of the therapy. Upper panel shows the schedule of antigen inputs with different doses in allergen immunotherapy.

(C) Effect of allergen immunotherapy depending on antigen concentration and rapidness of antigen inputs during allergen immunotherapy. The left diagram shows examples of antigen input in allergen immunotherapy with different concentrations and rapidness, corresponding to asterisks in the right panel.

T-cell activation potency.[37–39] Thus, we examined the effect of the dose-response pattern of T-cell activation on immune discrimination (see the STAR Methods section). Here, we simulated the model with three types of dose-response curves (linear, sigmoidal, and step-like curves) for both T_{conv} and T_{reg} cells (Figures 5A, 5D, and 5G). We found that different dose-response types of T-cell activation induced different accumulation patterns of memory T_{conv} cells depending on antigen concentrations (top panels in Figures 5B, 5E, and 5H). In the case of the linear dose-response curve, memory T_{conv} cells accumulated to an approximately constant value, independent of antigen concentration, while it transiently peaked and then constantly increased with the antigen concentration in cases of the sigmoidal and step-like curves. In contrast, the three dose-response types of T-cell activation did not show a critical difference in the accumulation of memory T_{reg} cells (middle panels in Figures 5B, 5E, and 5H), and antigen concentration-dependent discrimination was achieved in all dose-response types (bottom panels in Figures 5B, 5E, and 5H). Thus, these results indicated that the types of dose-response, or the properties of T-cell activation largely affected memory T_{conv} cell accumulation.

Next, we examined the effect of dose-response types of T-cell activation on history-dependent discrimination by simulating allergen immunotherapy, as shown in Figure 4 (Figures 5C, 5F, and 5I). We found that
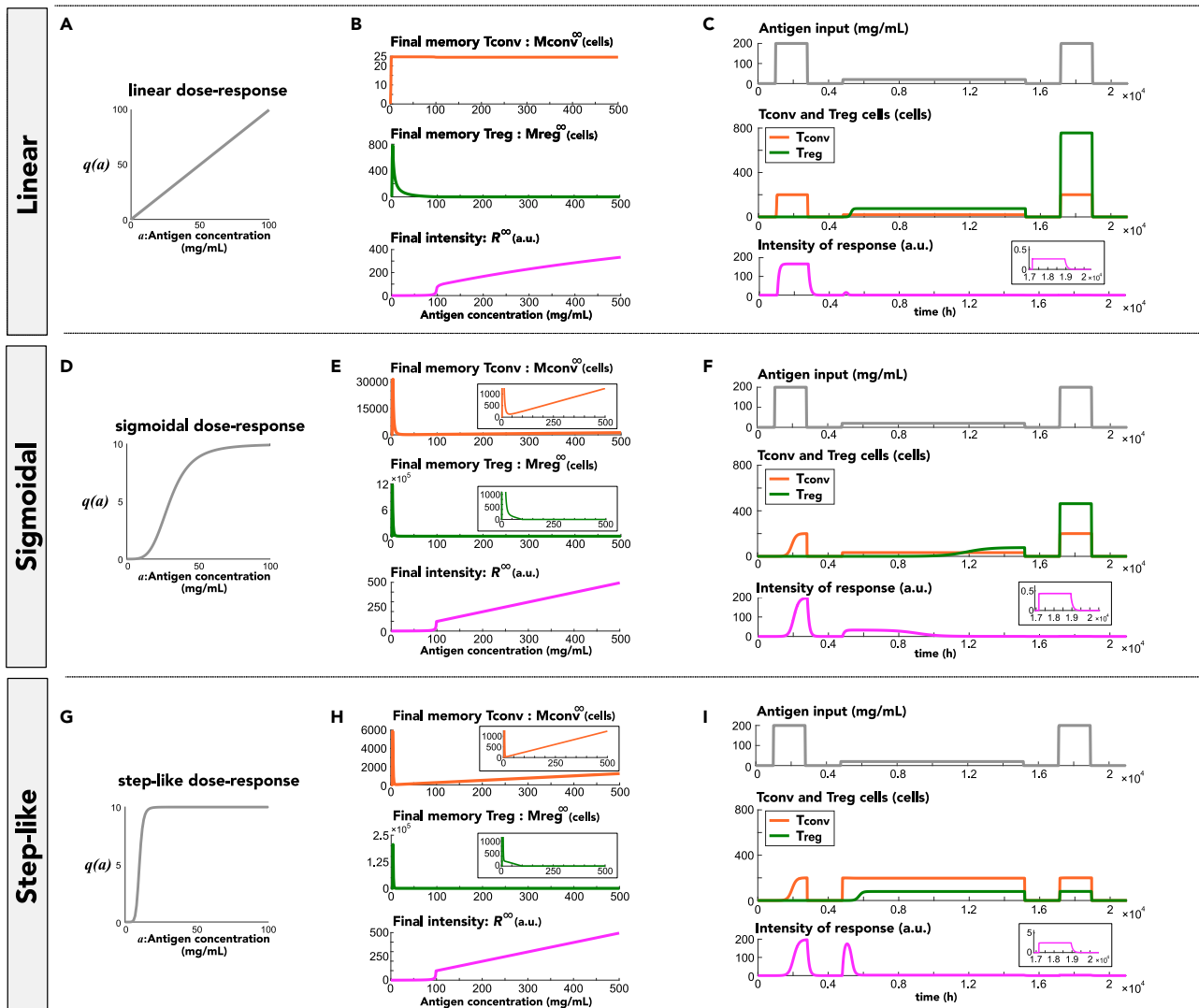
Figure 5. Robustness of allergen immunotherapy effect against the property of T-cell activation

(A, D, and G) Three types of dose-responses of T-cell activation to antigen concentrations. $q(a)$ represents the dose-response curves of T_{conv} and T_{reg} cell activation.

(B, E, and H) Change in memory T_{conv} and memory T_{reg} cell accumulation and the intensity of immune responses, depending on antigen concentrations. The convergence values of memory T_{conv} and memory T_{reg} cell populations and the intensity are plotted upon steady exposure to each antigen concentration. Insets in (E) and (H) show an enlarged view of memory T_{conv} and memory T_{reg} cell accumulation.

(C, F, and I) Temporal change in immune responses before, during, and after the allergen immunotherapy (as in Figure 4). Insets show an enlarged view of the intensity of the responses during the third antigen input.

allergen immunotherapy was successfully achieved in all three dose-responses, since the administration of low concentrations of antigens caused a positive prediction error in memory T_{reg} cell generation (details in Figures S5–S7). However, in the step-like dose-response, the intensity of the response was high at the initiation of allergen immunotherapy due to the production of T_{conv} cells from memory T_{conv} cells, which did not depend on the antigen concentration above the threshold. This implied that some patients with allergy with a step-like dose-response might exhibit allergic symptoms at the early stage of therapy.

From a clinical viewpoint, it is important to discern whether the effect of therapy is persistently maintained against subsequent exposures to antigens. To examine the persistence of the therapeutic effect, we considered the case where, after therapy, patients were exposed to an additional higher concentration of antigens followed by a subsequent lower concentration of antigens (top panels in Figures S5–S7). The

final antigen input was applied to quantify the persistence of the therapeutic effect. We found that allergen immunotherapy was effective in all combinations of dose-response types for T_{conv} and T_{reg} cells (Figures S5–S8A). However, its effect was persistently maintained against additional exposures to higher concentrations of antigens only when the dose-response type of T_{conv} cells was linear (Figures S5–S8B). These results indicated that the long-term effect of allergen immunotherapy can be determined by the dose-response type of T_{conv} cell activation. This may explain the heterogeneous effects of allergen immunotherapy across patients, as seen in some cases where patients demonstrated allergic symptoms again after discontinuing allergen immunotherapy.[34,35]

DISCUSSION

In order to understand how the immune system discriminates between harmful and harmless antigens despite their diversity, we developed a generalized model that does not assume any prior information on whether each antigen is harmful or harmless. We assumed predictive coding in T-cell population dynamics, by which we first introduced into immunology the concept that the immune system predicts its environment. Specifically, we developed a mathematical model of T-cell population dynamics under the hypothesis that T_{conv} and T_{reg} cells are predictors of the risk of antigens and excessive immune response, respectively, and their responses are regulated by prediction errors via memory T-cell generation. This predictive immune memory model led to both antigen concentration- and input rapidness-dependent discrimination between harmful and harmless antigens. In addition, our model showed that such discrimination can change in an antigen history-dependent manner, as seen in the onset of allergy and its subsequent therapy. To the best of our knowledge, this is the first learning system-based model of discrimination between harmful and harmless antigens by the immune system facing diverse antigens. Furthermore, it could be possible to validate our model in the future through the quantification of T cells with each TCR using single-cell RNA sequencing techniques.

Phenomenologically, harmful antigens usually originate from bacteria and viruses and show a rapid exponential increase in their population once they invade the body. In contrast, harmless antigens, such as food, do not sharply increase in amount inside the body but are expected to change gradually over time. Such distinct characteristics of harmful and harmless antigens can be distinguished by antigen concentration- and input rapidness-dependent discrimination (Figures 2 and 3). Clinically, immune discrimination for the same antigen is known to change over time; for example, the onset of allergy due to exposure to high concentrations of antigens and its remission through allergen immunotherapy. This can be represented by antigen history-dependent discrimination (Figure 4).

In this study, we introduced various types of T-cell activation dose-responses into the model based on the fact that the difference in ligands and its consequent difference in binding properties to TCRs largely affect T-cell activation potency.[37–39] Our results showed that the dose-response types of T-cell activation influenced antigen history-dependent changes in the immune response, as seen in allergen immunotherapy and subsequent recurrence (Figures 5, S5, S6, S7, and S8). Overall, these results suggested that the various dose-responses of T-cell activation cause heterogeneity in the immune responses of individuals and/or types of antigens. In fact, some patients with allergy acquire persistent remission of the symptoms by allergen immunotherapy, while others exhibit the symptoms again despite therapy.[34,35] Moreover, allergens, such as food and bee venom, sometimes induce lethal symptoms, while others, such as pollens, rarely do so.[40]

Our model is a minimal model that describes essential immune processes at the level of T cells, including antigen presentation by DCs, differentiation from T_{naive} cells to T cells, reactivation of memory T cells to T cells, and memory formation. Although there are several subtypes of T_{conv} cells, such as Th1, Th2, and Th17, which induce different downstream responses, we integrated these subtypes into a single T_{conv} cell population, since all T_{conv} cell subtypes have almost the same role in terms of the elimination of target antigens via different mechanisms.

Downstream of T_{conv} and T_{reg} cells, various types of cells are involved, such as killer T cells, B cells, macrophages, neutrophils, eosinophils, basophils, natural killer T cells, and mast cells. Although we need to consider these various immune cells to discuss the whole immune response, in principle, each subtype of T_{conv} cells facilitates the activation of these downstream cells, while T_{reg} cells suppress the response,[9,11] and global activity of these downstream T cells and cytokines should determine the intensity of response.

Therefore, our model simply assumed that the intensity of immune responses can be evaluated only by the amounts of T_{conv} and T_{reg} cells.

Some immune cell populations that eliminate antigens, such as killer T cells, T_{conv} cells, B cells, and natural killer T cells, are known to persist in the body for a long time, preparing for a second infection following the first antigen experience by natural infection and vaccination.[41] Furthermore, previous studies have extensively studied whether regulatory immune cell subsets, such as T_{reg} cells, generate memory populations after antigen exposure.[42] Due to the lack of memory-specific phenotypic markers for the identification of these populations, it remains controversial whether distinct memory subsets contribute to the persistence of immunosuppressive effects.[43,44] However, several studies have defined memory T_{reg} cells and revealed their characteristics as memory populations.[15,45] Hence, our model included the memory T_{reg} population as one of the possible implementations of regulatory memory formation.

In this study, we assumed memory T-cell production based on predictive coding. For implementation, we regarded cytokines as the media for transmitting quantitative information. Specifically, the amounts of T_{conv} and T_{reg} cells could be coded by the concentration of cytokines secreted by themselves, whereas the amount of antigens could be coded by the concentration of cytokines secreted from antigen-presenting cells, such as DCs and macrophages. Based on such information-carrying cytokines, we hypothesized that prediction errors in predictive coding can be computed through intracellular signal transduction in T cells. Similar to our hypothesis, this type of quantitative function of cytokines has recently become a point of focus, although qualitative molecular discoveries have been traditionally explored, such as the identification of previously unknown cytokines and potential T_{conv} cell subsets. For instance, various experimental and computational studies revealed that immune cell activation was controlled by cell density via cytokines.[32,46-50] This phenomenon is called quorum sensing and was originally proposed in bacterial cells[51,52] and then adopted to elucidate immune dynamics.[53-55] Our hypothesis that memory formation based on the calculation of T-cell populations can be achieved by cytokines is consistent with the concept of quorum sensing in terms of cell density-dependent induction of responses achieved by cytokines. Notably, it was also suggested that T_{conv} cell density regulated the rate of memory differentiation.[56] Furthermore, we introduced the idea that the information on cell densities was integrated into T cells because it is possible that T cells sensitive to various cytokines can integrate signals from them. To validate this hypothesis, however, it is necessary to quantify the time series of T-cell populations with each TCR and cytokines in future experiments.

Appropriate immune responses to each antigen kind have traditionally been assumed to be achieved at the single-cell level due to the antigen specificity of TCRs on each T cell. In addition, it has been suggested that the antigens themselves can determine the responses, which is referred to as the "danger theory."[57-59] The theory states that T cells are activated only in the presence of danger signals, such as pathogen-associated molecular patterns (PAMPs),[60] because they upregulate the expression of costimulatory molecules on antigen-presenting cells. However, this kind of antigen-type-dependent immune response does not explain the temporal change of immune discrimination (i.e., immune activation by and tolerance to the same antigen). It also does not explain the immune responses to harmless antigens, as seen in allergy and autoimmune diseases. In addition, some studies examining the quorum sensing mechanism suggested that the state of the cell population level, such as their densities and distributions, has a more important role in regulating immune responses than distinct antigen properties.[53-55] Therefore, our model hypothesized that memory T cells were generated based on the calculation of antigen concentrations and T-cell populations. This hypothesis is also based on the concept of immune regulation, which does not premise prior information on the risk of antigens and their own properties.

Several studies have reported computational models of immune dynamics. Different models of T-cell population dynamics have focused on allergen immunotherapy. In one model, allergen immunotherapy was represented by prolonged activation of T_{reg} cells with a large time constant.[61] In another model, the effect of allergen immunotherapy was represented by a transition from a Th2 cell-dominant state to a T_{reg} cell-dominant state.[62] However, the effect of the therapy spontaneously disappeared after antigen elimination due to the absence of explicit T-cell memory.

Immune discrimination had earlier been assessed by various mathematical models. Sontag modeled the interaction between T-cell population and antigens, such as pathogens and tumor cells[63]; this study

revealed immune discrimination based on dynamic features of antigen presentation, such as the growth rate of antigens. In addition, Pradeu et al. proposed the discontinuity theory stating that discontinuous (sudden or intermittent) exposures to antigens induce vigorous immune responses, whereas progressive and persistent exposures induce weak responses.[64] The findings of these studies were consistent with our results in terms of immune discrimination being independent of antigen type; however, they lacked immunological memory formation.

Here, we developed a minimal model of immune discrimination, by which we showed a possible mechanism of immune discrimination based on universal information about all antigen types, such as their concentration and input rapidness, and demonstrated temporal changes based on the history of antigen exposures. However, our current model considered antigen-induced responses of T cells that are specific to only one kind of antigens, and it did not include antigens that undergo self-renewal and can be eliminated by the immune system, such as pathogens. In previous studies, Domínguez-Hüttinger et al. and Christodoulides et al. have focused on the onset and therapy of atopic dermatitis and developed a mathematical model describing the interaction of pathogens, skin barrier integrity, and the innate/adaptive immune system.[65,66] They revealed different phenotypes in patients derived from certain parameters (genetic risks) and suggested an effective treatment strategy based on the optimal control theory. To precisely describe immune discrimination for self-proliferating antigens, we should introduce antigen proliferation into our current model and consider its interaction with the immune system, which would potentially enable us to understand more complex immune responses, such as in the case of atopic dermatitis with immunological memory formation.

Finally, our model would also enable us to address how immune responses change throughout our life, as seen in the hygiene hypothesis. This hypothesis states that an unhygienic experience (experience of numerous infections) during early childhood prevents allergic diseases; on the contrary, hygienic environments raise their risk.[67] The authenticity of this hypothesis is still controversial, but it suggests that antigen discrimination can be influenced by all previous exposures to multiple antigens, that is, personal hygiene. Our results on history-dependent discrimination, where immune responses to the same antigen input can be weakened by a certain antigen experience, is consistent with the hygiene hypothesis. Thus, our model might explain the difference in allergic risks based on individual antigen experiences. However, our current minimal model did not describe immune responses under multiple kinds of antigens based on the idea that responses to each antigen are dominantly determined by cells specific to each antigen, although immune cells specific to other antigens possibly contribute to the response under multiple types of antigens. Therefore, to validate the hygiene hypothesis, we need to expand our model into a form that is able to examine immune responses toward multiple antigens.

Limitations of the study

The mathematical model developed in this study (the predictive immune memory model) is a minimal model that describes essential immune processes at the level of T cells. For simplicity, we only modeled the response to only one kind of antigens. In addition, antigen inputs are completely external input and antigens do not proliferate and they are not eliminated by immune responses in the current model. Thus, to examine more complex immune responses, such as atopic dermatitis and hygiene hypothesis, we need to expand our model into the model that describes responses to multiple kinds of antigens and the dynamics of antigens (their proliferation and elimination by immune responses).

STAR★METHODS

Detailed methods are provided in the online version of this paper and include the following:

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AUTHOR CONTRIBUTIONS

K.Y. and H.N. conceived the project and developed the method. K.Y. implemented the model simulation. K.Y. and H.N. wrote the article.

DECLARATION OF INTERESTS

The authors declare no competing interests.

REFERENCES

1. Akdis, M., and Akdis, C.A. (2014). Mechanisms of allergen-specific immunotherapy: multiple suppressor factors at work in immune tolerance to allergens. J. Allergy Clin. Immunol. *133*, 621–631. https://doi.org/10.1016/j.jaci.2013.12.1088.

2. Buckley, C.D., and McGettrick, H.M. (2018). Leukocyte trafficking between stromal compartments: lessons from rheumatoid arthritis. Nat. Rev. Rheumatol. *14*, 476–487. https://doi.org/10.1038/s41584-018-0042-4.

3. Evavold, B.D., and Allen, P.M. (1991). Separation of IL-4 production from Th cell proliferation by an altered T cell receptor ligand. Science *252*, 1308–1310. https://doi.org/10.1126/science.1833816.

4. Dustin, M.L. (2008). T-cell activation through immunological synapses and kinapses. Immunol. Rev. *221*, 77–89. https://doi.org/10.1111/j.1600-065X.2008.00589.x.

5. Monks, C.R., Freiberg, B.A., Kupfer, H., Sciaky, N., and Kupfer, A. (1998). Three-dimensional segregation of supramolecular activation clusters in T cells. Nature *395*, 82–86. https://doi.org/10.1038/25764.

6. Kappler, J.W., Roehm, N., and Marrack, P. (1987). T cell tolerance by clonal elimination in the thymus. Cell *49*, 273–280. https://doi.org/10.1016/0092-8674(87)90568-X.

7. Kisielow, P., Blüthmann, H., Staerz, U.D., Steinmetz, M., and Von Boehmer, H. (1988). Tolerance in T-cell-receptor transgenic mice involves deletion of nonmature CD4+8+ thymocytes. Nature *333*, 742–746. https://doi.org/10.1038/333742a0.

8. Anderson, M.S., Venanzi, E.S., Klein, L., Chen, Z., Berzins, S.P., Turley, S.J., Von Boehmer, H., Bronson, R., Dierich, A., Benoist, C., et al. (2002). Projection of an immunological self shadow within the thymus by the aire protein. Science *298*, 1395–1401. https://doi.org/10.1126/science.1075958.

9. Luckheeram, R.V., Zhou, R., Verma, A.D., and Xia, B. (2012). CD4 +T cells: differentiation and functions. Clin. Dev. Immunol. *2012*, 925135. https://doi.org/10.1155/2012/925135.

10. Crotty, S. (2011). Follicular helper CD4 T cells (T FH). Annu. Rev. Immunol. *29*, 621–663. https://doi.org/10.1146/annurev-immunol-031210-101400.

11. Zhu, J., and Paul, W.E. (2008). CD4 T cells: fates, functions, and faults. Blood *112*, 1557–1569. https://doi.org/10.1182/blood-2008-05-078154.

12. Vignali, D.A.A., Collison, L.W., and Workman, C.J. (2008). How regulatory T cells work. Nat. Rev. Immunol. *8*, 523–532. https://doi.org/10.1038/nri2343.

13. Sakaguchi, S., Yamaguchi, T., Nomura, T., and Ono, M. (2008). Regulatory T cells and immune tolerance. Cell *133*, 775–787. https://doi.org/10.1016/j.cell.2008.05.009.

14. Chowdhury, D., and Lieberman, J. (2008). Death by a thousand cuts: granzyme pathways of programmed cell death. Annu. Rev. Immunol. *26*, 389–420. https://doi.org/10.1146/annurev.immunol.26.021607.090404.

15. Brincks, E.L., Roberts, A.D., Cookenham, T., Sell, S., Kohlmeier, J.E., Blackman, M.A., and Woodland, D.L. (2013). Antigen-specific memory regulatory CD4 + Foxp3 + T cells control memory responses to influenza virus infection. J. Immunol. *190*, 3438–3446. https://doi.org/10.4049/jimmunol.1203140.

16. Gasper, D.J., Tejera, M.M., and Suresh, M. (2014). CD4 T-cell memory generation and maintenance. Crit. Rev. Immunol. *34*, 121–146. https://doi.org/10.1615/CritRevImmunol.2014010373.

17. Kaech, S.M., Wherry, E.J., and Ahmed, R. (2002). Effector and memory T-cell differentiation: implications for vaccine development. Nat. Rev. Immunol. *2*, 251–262. https://doi.org/10.1038/nri778.

18. Harrington, L.E., Janowski, K.M., Oliver, J.R., Zajac, A.J., and Weaver, C.T. (2008). Memory CD4 T cells emerge from effector T-cell progenitors. Nature *452*, 356–360. https://doi.org/10.1038/nature06672.

19. Garcia, S., DiSanto, J., and Stockinger, B. (1999). Following the development of a CD4

T cell response in vivo: from activation to memory formation. Immunity *11*, 163–171. https://doi.org/10.1016/S1074-7613(00)80091-6.

20. Canonica, G.W., Cox, L., Pawankar, R., Baena-Cagnani, C.E., Blaiss, M., Bonini, S., Bousquet, J., Calderón, M., Compalati, E., Durham, S.R., et al. (2014). Sublingual immunotherapy: world Allergy Organization position paper 2013 update. World Allergy Organ. J. *7*, 6. https://doi.org/10.1186/1939-4551-7-6.

21. Noon, L. (1911). Prophylactic inoculation against hay fever. Lancet *177*, 1572–1573. https://doi.org/10.1016/S0140-6736(00)78276-6.

22. Pfaar, O., Bachert, C., Bufe, A., Buhl, R., Ebner, C., Eng, P., Friedrichs, F., Fuchs, T., Hamelmann, E., Hartwig-Bade, D., et al. (2014). Guideline on allergen-specific immunotherapy in IgE-mediated allergic diseases. Allergo J. Int. *23*, 282–319. https://doi.org/10.1007/s40629-014-0032-2.

23. Böhm, L., Maxeiner, J., Meyer-Martin, H., Reuter, S., Finotto, S., Klein, M., Schild, H., Schmitt, E., Bopp, T., and Taube, C. (2015). IL-10 and regulatory T cells cooperate in allergen-specific immunotherapy to ameliorate allergic asthma. J. Immunol. *194*, 887–897. https://doi.org/10.4049/jimmunol.1401612.

24. Shamji, M.H., and Durham, S.R. (2011). Mechanisms of immunotherapy to aeroallergens. Clin. Exp. Allergy *41*, 1235–1246. https://doi.org/10.1111/j.1365-2222.2011.03804.x.

25. Radulovic, S., Jacobson, M.R., Durham, S.R., and Nouri-Aria, K.T. (2008). Grass pollen immunotherapy induces Foxp3-expressing CD4+CD25+ cells in the nasal mucosa. J. Allergy Clin. Immunol. *121*, 1467–1472.e1. https://doi.org/10.1016/j.jaci.2008.03.013.

26. Rao, R.P., and Ballard, D.H. (1999). Predictive coding in the visual cortex: a functional interpretation of some extra-classical receptive-field effects. Nat. Neurosci. *2*, 79–87. https://doi.org/10.1038/4580.

27. Friston, K. (2010). The free-energy principle: a unified brain theory? Nat. Rev. Neurosci. *11*, 127–138. https://doi.org/10.1038/nrn2787.

28. Friston, K., and Kiebel, S. (2009). Predictive coding under the free-energy principle. Philos. Trans. R. Soc. Lond. B Biol. Sci. *364*, 1211–1221. https://doi.org/10.1098/rstb.2008.0300.

29. Friston, K.J., Daunizeau, J., Kilner, J., and Kiebel, S.J. (2010). Action and behavior: a free-energy formulation. Biol. Cybern. *102*, 227–260. https://doi.org/10.1007/s00422-010-0364-z.

30. Tadokoro, C.E., Shakhar, G., Shen, S., Ding, Y., Lino, A.C., Maraver, A., Lafaille, J.J., and Dustin, M.L. (2006). Regulatory T cells inhibit stable contacts between CD4+ T cells and dendritic cells in vivo. J. Exp. Med. *203*, 505–511. https://doi.org/10.1084/jem.20050783.

31. Busse, D., De La Rosa, M., Hobiger, K., Thurley, K., Flossdorf, M., Scheffold, A., and Höfer, T. (2010). Competing feedback loops shape IL-2 signaling between helper and regulatory T lymphocytes in cellular microenvironments. Proc. Natl. Acad. Sci. USA *107*, 3058–3063. https://doi.org/10.1073/pnas.0812851107.

32. Feinerman, O., Jentsch, G., Tkach, K.E., Coward, J.W., Hathorn, M.M., Sneddon, M.W., Emonet, T., Smith, K.A., and Altan-Bonnet, G. (2010). Single-cell quantification of IL-2 response by effector and regulatory T cells reveals critical plasticity in immune response. Mol. Syst. Biol. *6*, 437. https://doi.org/10.1038/msb.2010.90.

33. Turner, M.D., Nedjai, B., Hurst, T., and Pennington, D.J. (2014). Cytokines and chemokines: at the crossroads of cell signalling and inflammatory disease. Biochim. Biophys. Acta *1843*, 2563–2582. https://doi.org/10.1016/j.bbamcr.2014.05.014.

34. Sturm, G.J., Varga, E.M., Roberts, G., Mosbech, H., Bilò, M.B., Akdis, C.A., Antolín-Amérigo, D., Cichocka-Jarosz, E., Gawlik, R., Jakob, T., et al. (2018). EAACI guidelines on allergen immunotherapy: hymenoptera venom allergy. Allergy *73*, 744–764. https://doi.org/10.1111/all.13262.

35. Cox, L., Nelson, H., Lockey, R., Calabria, C., Chacko, T., Finegold, I., Nelson, M., Weber, R., Bernstein, D.I., Blessing-Moore, J., et al. (2011). Allergen immunotherapy: a practice parameter third update. J. Allergy Clin. Immunol. *127*, S1–55. https://doi.org/10.1016/j.jaci.2010.09.034.

36. Barni, S., Liccioli, G., Sarti, L., Giovannini, M., Novembre, E., and Mori, F. (2020). Immunoglobulin E (IgE)-mediated food allergy in children: epidemiology, pathogenesis, diagnosis, prevention, and management. Medicina *56*, 111. https://doi.org/10.3390/medicina56030111.

37. Aleksic, M., Dushek, O., Zhang, H., Shenderov, E., Chen, J.L., Cerundolo, V., Coombs, D., and van der Merwe, P.A. (2010). Dependence of T Cell antigen recognition on T cell receptor-peptide MHC confinement time. Immunity *32*, 163–174. https://doi.org/10.1016/j.immuni.2009.11.013.

38. Čemerski, S., Das, J., Locasale, J., Arnold, P., Giurisato, E., Markiewicz, M.A., Fremont, D., Allen, P.M., Chakraborty, A.K., and Shaw, A.S. (2007). The stimulatory potency of T cell antigens is influenced by the formation of the immunological synapse. Immunity *26*, 345–355. https://doi.org/10.1016/j.immuni.2007.01.013.

39. Evavold, B.D., Sloan-Lancaster, J., and Allen, P.M. (1993). Tickling the TCR: selective T-cell functions stimulated by altered peptide ligands. Immunol. Today *14*, 602–609. https://doi.org/10.1016/0167-5699(93)90200-5.

40. Turner, P.J., Jerschow, E., Umasunthar, T., Lin, R., Campbell, D.E., and Boyle, R.J. (2017). Fatal anaphylaxis: mortality rate and risk factors. J. Allergy Clin. Immunol. Pract. *5*, 1169–1178. https://doi.org/10.1016/j.jaip.2017.06.031.

41. Ratajczak, W., Niedźwiedzka-Rystwej, P., Tokarz-Deptuła, B., and Deptuła, W. (2018). Immunological memory cells. Cent. Eur. J. Immunol. *43*, 194–203. https://doi.org/10.5114/ceji.2018.77390.

42. Rosenblum, M.D., Way, S.S., and Abbas, A.K. (2016). Regulatory T cell memory. Nat. Rev. Immunol. *16*, 90–101. https://doi.org/10.1038/nri.2015.1.

43. Bianchi, D.W., Zickwolf, G.K., Weil, G.J., Sylvester, S., and Demaria, M.A. (1996). Male fetal progenitor cells persist in maternal blood for as long as 27 years postpartum. Proc. Natl. Acad. Sci. USA *93*, 705–708. https://doi.org/10.1073/pnas.93.2.705.

44. Nelson, J.L. (2012). The otherness of self: microchimerism in health and disease. Trends Immunol. *33*, 421–427. https://doi.org/10.1016/j.it.2012.03.002.

45. Sanchez, A.M., Zhu, J., Huang, X., and Yang, Y. (2012). The development and function of memory regulatory T cells after acute viral infections. J. Immunol. *189*, 2805–2814. https://doi.org/10.4049/jimmunol.1200645.

46. O'Garra, A., Gabryšová, L., and Spits, H. (2011). Quantitative events determine the differentiation and function of helper T cells. Nat. Immunol. *12*, 288–294. https://doi.org/10.1038/ni.2003.

47. Montaudouin, C., Anson, M., Hao, Y., Duncker, S.V., Fernandez, T., Gaudin, E., Ehrenstein, M., Kerr, W.G., Colle, J.-H., Bruhns, P., et al. (2013). Quorum sensing contributes to activated IgM-secreting B cell homeostasis. J. Immunol. *190*, 106–114. https://doi.org/10.4049/jimmunol.1200907.

48. Shalek, A.K., Satija, R., Shuga, J., Trombetta, J.J., Gennert, D., Lu, D., Chen, P., Gertner, R.S., Gaublomme, J.T., Yosef, N., et al. (2014). Single-cell RNA-seq reveals dynamic paracrine control of cellular variation. Nature *510*, 363–369. https://doi.org/10.1038/nature13437.

49. Burroughs, N.J., Oliveira, B.M.P.M., Pinto, A.A., and Sequeira, H.J.T. (2008). Sensibility of the quorum growth thresholds controlling local immune responses. Math. Comput. Model. *47*, 714–725. https://doi.org/10.1016/j.mcm.2007.06.007.

50. Schrom, E.C., Levin, S.A., and Graham, A.L. (2020). Quorum sensing via dynamic cytokine signaling comprehensively explains divergent patterns of effector choice among helper T cells. PLoS Comput. Biol. *16*, e1008051. https://doi.org/10.1371/journal.pcbi.1008051.

51. Fuqua, W.C., Winans, S.C., and Greenberg, E.P. (1994). Quorum sensing in bacteria: the LuxR-LuxI family of cell density- responsive transcriptional regulators. J. Bacteriol. *176*, 269–275. https://doi.org/10.1128/jb.176.2.269-275.1994.

52. Waters, C.M., and Bassler, B.L. (2005). Quorum sensing: cell-to-cell communication in bacteria. Annu. Rev. Cell Dev. Biol. *21*, 319–346. https://doi.org/10.1146/annurev.cellbio.21.012704.131001.

53. Antonioli, L., Blandizzi, C., Pacher, P., Guilliams, M., and Haskó, G. (2018). Quorum sensing in the immune system. Nat. Rev. Immunol. *18*, 537–538. https://doi.org/10.1038/s41577-018-0040-4.

54. Perié, L., Aru, J., Kourilsky, P., and Slotine, J.J. (2013). Does a quorum sensing mechanism direct the behavior of immune cells? C. R. Biol. *336*, 13–16. https://doi.org/10.1016/j.crvi.2013.01.006.

55. Al-Yassin, G.A., and Bretscher, P.A. (2018). Does T cell activation require a quorum of lymphocytes? J. Immunol. *201*, 2855–2861. https://doi.org/10.4049/jimmunol.1800805.

56. Polonsky, M., Rimer, J., Kern-Perets, A., Zaretsky, I., Miller, S., Bornstein, C., David, E., Kopelman, N.M., Stelzer, G., Porat, Z., et al. (2018). Induction of CD4 T cell memory by local cellular collectivity. Science *360*, eaaj1853. https://doi.org/10.1126/science.aaj1853.

57. Matzinger, P. (1994). Tolerance, danger, and the extended family. Annu. Rev. Immunol. *12*, 991–1045. https://doi.org/10.1146/annurev.iy.12.040194.005015.

58. Pradeu, T., and Cooper, E.L. (2012). The danger theory: 20 years later. Front. Immunol. *3*, 287. https://doi.org/10.3389/fimmu.2012.00287.

59. Matzinger, P. (2002). The danger model: a renewed sense of self. Science *296*, 301–305. https://doi.org/10.1126/science.1071059.

60. Janeway, C.A. (1989). Approaching the asymptote? Evolution and revolution in immunology. Cold Spring Harb. Symp. Quant. Biol. *54 Pt 1*, 1–13. https://doi.org/10.1101/sqb.1989.054.01.003.

61. Hara, A., and Iwasa, Y. (2017). When is allergen immunotherapy effective? J. Theor. Biol. *425*, 23–42. https://doi.org/10.1016/j.jtbi.2017.04.030.

62. Gross, F., Metzner, G., and Behn, U. (2011). Mathematical modeling of allergy and specific immunotherapy: Th1-Th2-Treg interactions. J. Theor. Biol. *269*, 70–78. https://doi.org/10.1016/j.jtbi.2010.10.013.

63. Sontag, E.D. (2017). A dynamic model of immune responses to antigen presentation predicts different regions of tumor or pathogen elimination. Cell Syst. *4*, 231–241.e11. https://doi.org/10.1016/j.cels.2016.12.003.

64. Pradeu, T., Jaeger, S., and Vivier, E. (2013). The speed of change: towards a discontinuity theory of immunity? Nat. Rev. Immunol. *13*, 764–769. https://doi.org/10.1038/nri3521.

65. Domínguez-Hüttinger, E., Christodoulides, P., Miyauchi, K., Irvine, A.D., Okada-Hatakeyama, M., Kubo, M., and Tanaka, R.J. (2017). Mathematical modeling of atopic dermatitis reveals "double-switch" mechanisms underlying 4 common disease phenotypes. J. Allergy Clin. Immunol. *139*, 1861–1872.e7. https://doi.org/10.1016/j.jaci.2016.10.026.

66. Christodoulides, P., Hirata, Y., Domínguez-Hüttinger, E., Danby, S.G., Cork, M.J., Williams, H.C., Aihara, K., and Tanaka, R.J. (2017). Computational design of treatment strategies for proactive therapy on atopic dermatitis using optimal control theory. Philos. Trans. A Math. Phys. Eng. Sci. *375*, 20160285. https://doi.org/10.1098/rsta.2016.0285.

67. Yazdanbakhsh, M., Kremsner, P.G., and Van Ree, R. (2002). Immunology: allergy, parasites, and the hygiene hypothesis. Science *296*, 490–494. https://doi.org/10.1126/science.296.5567.490.

STAR★METHODS

KEY RESOURCES TABLE

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Software and algorithms		
MATLAB	https://jp.mathworks.com/products/matlab.html	RRID: SCR:001622 Version: 9.10.0.1602886 (R2021a)
Simulation codes	N/A	https://github.com/kyoshido1213/Simulations-of-Predictive-immune-memory-model

RESOURCE AVAILABILITY

Lead contact

Further information and requests should be directed to and will be fulfilled by the lead contact, Honda Naoki (nhonda@hiroshima-u.ac.jp).

Materials availability

The study did not generate any new materials.

Data and code availability

No datasets were generated and analyzed during the current study. The predictive immune memory model was simulated using Matlab (R2021a) on GitHub (https://github.com/kyoshido1213/Simulations-of-Predictive-immune-memory-model). The parameters used in the simulation are provided in Table S1.

EXPERIMENTAL MODEL AND SUBJECT DETAILS

This study did not use experimental models and subjects.

METHOD DETAILS

T cell population dynamics with different properties of T cell activation

The model was extended to include the effect of the dose-responses of T cell activation on the immune response as

$$\frac{d}{dt}T_{conv} = -d_c T_{conv} + \frac{D_c}{1+s_r T_{reg}}T_{conv} + k_c\, T_{naive}\, q_c(a) + w_c\, M_{conv}\, q_c(a) - E_c T_{conv},$$

$$\frac{d}{dt}T_{reg} = -d_r T_{reg} + \frac{D_r}{1+s_c T_{conv}}T_{reg} + k_r\, T_{naive}\, q_r(a) + w_r\, M_{reg}\, q_r(a) - E_r T_{reg},$$

where $q_c(a)$ and $q_r(a)$ represent the dose-responses of T_{conv} and T_{reg} cells, respectively, as described by the linear and Hill equation:

$$q_i(a) = \begin{cases} a\,, & when\ linear\ dose - response \\ Q_i\,\dfrac{a^{h_i}}{K_i^{h_i}+a^{h_i}}, & when\ sigmoidal\ and\ step - like\ dose - response \end{cases}$$

where Q_i, K_i and h_i ($i \in \{c,r\}$) indicate the amplitude, half-maximal effective antigen concentration, and Hill coefficient, respectively. Here, we considered three types of dose-response curves (linear, sigmoidal, and step-like). In the linear dose-response curve, the same equations were applied as Equations 1 and 2. In the sigmoidal and step-like dose-response curves, the Hill equation was applied, where $(h_i, Q_i, K_i) = (4, 10, 30)$ and $(8, 10, 10)$, respectively. The same dose-response types were applied for both T_{conv} and T_{reg} cells in Figure 5 ($q_c(a) = q_r(a)$), whereas different types of dose-response were applied in Figures S5–S8 ($q_c(a) = q_r(a)$ or $q_c(a) \neq q_r(a)$).

T cell population dynamics with time delay in memory formation

The model was extended to include the effect of time delay in memory T cell formation.

$$\frac{d}{dt}M_{conv} = -d_{mc}M_{conv}(t) + E_c T_{conv}\left(t - \tau_{delay}\right),$$

$$\frac{d}{dt}M_{reg} = -d_{mr}M_{reg}(t) + E_r T_{reg}\left(t - \tau_{delay}\right),$$

where the prediction error is calculated based on the concentration of antigens and T cells at time $t - \tau_{delay}$:

$$E_c = e_c\left|a\left(t - \tau_{delay}\right) - m_c T_{conv}\left(t - \tau_{delay}\right)\right|_{+},$$

$$E_r = e_r\left|f\left(T_{conv}\left(t - \tau_{delay}\right), a\left(t - \tau_{delay}\right)\right) - m_r T_{reg}\left(t - \tau_{delay}\right)\right|_{+},$$

Biologically, it is a possibility that memory T cell generation based on the calculation of prediction error takes more time compared to T cell population dynamics stimulated by antigens.

QUANTIFICATION AND STATISTICAL ANALYSIS

This study did not include statistical analysis and quantification.

ADDITIONAL RESOURCES

This study did not generate and contributed to a new website/forum, and it is not part of a clinical trial.