

1 An effective and practical method of net settings in rearing tank to suppress hypermelanosis in Japanese

2 flounder

3

4 Tsuyoshi Onoyama¹, Toshiyuki Yamada², and Masatomo Tagawa¹

5

6 ¹Division of Applied Biosciences, Graduate School of Agriculture, Kyoto University, Kitashirakawa,

7 Sakyo, Kyoto 606-8502, Japan

8 ²Nagasaki Prefectural Institute of Fisheries, Taira, Nagasaki, Nagasaki 851-2213, Japan

9

10 *Corresponding author: Masatomo TAGAWA

11 Tel: 81-075-753-6221; Fax: 81-075-753-6229

12

13

14 Email addresses of all authors

15 onoyama.tsuyoshi.84m@st.kyoto-u.ac.jp (Tsuyoshi Onoyama)

16 yamadat@pref.nagasaki.lg.jp (Toshiyuki Yamada)

17 tagawa.masatomo.8m@kyoto-u.ac.jp (Masatomo Tagawa)

18

19 Abstract

20 In Japanese flounder aquaculture, hypermelanosis occurs widely on the blind side. Rearing flounders in a
21 net-lined tank was recently reported to prevent hypermelanosis. To effectively apply this method to larger
22 tanks for aquaculture farming, the net setting method was examined. Juvenile flounders without darkened
23 areas on the blind side (total length TL=13 cm) were selected, and reared for 6 months (TL = 32 cm). In
24 the control tank without a net, the median value of darkened area ratio (darkened area / blind side area) was
25 46%. By only covering the tank bottom with slack-net, darkened area ratio was suppressed to 8%, less than
26 1/5 of that in the control tank. at the end of experiment Bottom coverage ratios of 0%, 10%, 30%, 50%, and
27 100% revealed a negative correlation between bottom net coverage and darkened area ratio. In this
28 experiment, the darkening area in the tank with 50% bottom net coverage reduced to 1/3 of the control.
29 Although the occurrence of hypermelanosis differs depending on the production lot, these results are
30 expected to be a reference to select the suitable net size to meet the level of clearness of the blind side.

31

32 Keywords: Color abnormality · Hypermelanosis · Flatfish · Japanese flounder aquaculture · Bottom net ·
33 Covering ratio · Net-lined tank

34

35

36

37 Introduction

38 Japanese flounder *Paralichthys olivaceus* belongs to Pleuronectiformes, a large group of Heterosomata,
39 and has a bilaterally asymmetrical body. The left side (ocular side) has two eyes, and the right side (blind
40 side) has no eyes. On the ocular side, melanophores and xanthophores of the adult type appear after the
41 normal completion of eye relocation, while they do not appear on the blind side. Consequently, the ocular
42 side appears brown, and the blind side appears white (Seikai et al. 1987; Nakamura et al. 2010).

43 Japanese flounder is an important species in Japanese and Korean aquaculture and Japanese coastal
44 fisheries. Notably, the aquaculture production of the species was 2,186 t, which is approximately 1/4 of the
45 total yield in Japan in 2017 (Statistical survey of seawater fishery production 2018). Therefore, Japanese
46 flounder is a species that has been successfully used in aquaculture production at the industrial level.

47 However, there are various unsolved problems in Japanese flounder aquaculture. One of the problems is
48 color abnormality on the blind side; the white skin of the blind side gradually turns dark after the normal
49 completion of eye relocation (see reviews, Aritaki and Seikai 2017; Tagawa 2017). This phenomenon is
50 called hypermelanosis, and darkened flounders are sold at lower prices than normal (Kaji et al. 1999; Aritaki
51 2004). Therefore, establishing a method for preventing hypermelanosis is required, and various basic and
52 applied studies have been conducted. For example, the contribution of chromatophore-related hormones,
53 the stimulating effect of melanophore-stimulating hormone (Yamanome et al. 2007b; Kang and Kim 2012,
54 2015; Matsuda et al. 2018a), and the suppressing effect of melanin-concentrating hormone (Yamanome et

55 al. 2005, 2007a; Kang and Kim 2012, 2013a) have been clarified. Higher rearing density (Fukunaga 1998)
56 and cortisol (Matsuda et al. 2018b), a stress-responsive hormone, also increase hypermelanosis. Vitamin A
57 (Miwa and Yamano 1990; Tarui et al. 2006) and D (Haga et al. 2004) have been shown to stimulate
58 hypermelanosis. Although the control mechanisms of hypermelanosis have been gradually clarified, a
59 practical method that has strong reproducibility and easy applicability to aquaculture is still lacking.

60 Interestingly, the light-colored bottom of rearing tanks has been reported to decrease or delay
61 hypermelanosis (Amiya et al. 2005; Yamanome et al. 2005, 2007a; Nakata et al. 2017). However, in Kang
62 and Kim (2013a, 2013b), the light-colored bottom of rearing tanks did not suppress hypermelanosis.
63 Therefore, the effectiveness of the light color could be unstable depending on unknown factors. On the
64 other hand, rearing flounders in tanks with bottom sand can prevent hypermelanosis (Seikai 1991; Iwata
65 and Kikuchi 1998; Kang and Kim 2012, 2013b; Isojima et al. 2013a, 2014). As shown by several
66 independent groups, bottom sand strongly prevents hypermelanosis with higher reproducibility. However,
67 such a method of rearing has not been applied in aquaculture farm because the bottom sand is an obstacle
68 for tank cleaning, and uncleaned particles, remnant food, and excretes are deleterious to water and bottom
69 quality, as well as to fish health. Among various characteristics of bottom sand, Nakata et al. (2017) pointed
70 out the importance of undulation of the bottom of the tank for the prevention of hypermelanosis, and further
71 demonstrated that rearing juveniles in net-lined tanks also effectively prevented hypermelanosis
72 “expansion.” Mizutani et al. (2020), using flounders before the first appearance of hypermelanosis, proved

73 that rearing in net-lined tanks was also effective in preventing the “appearance” of hypermelanosis. From
74 these two studies, a net-lined tank is a strong candidate for a practical method for the prevention of
75 hypermelanosis.

76 Much larger tanks (several kL or more) than experimental tanks (mostly less than 1 kL) are used in the
77 aquaculture farm of Japanese flounder. It is difficult to install a net that covers the entire inner surface of a
78 large tank. In addition, tank cleaning is not easy because of the difficulty to remove the large net without
79 damaging the juvenile flounders inside of the net. Therefore, we conducted experiment to find a better way
80 of net settings that is easier to install and therefore more practical for larger tanks, and determine the suitable
81 net size.

82

83 **Materials and methods**

84 **Fish rearing**

85 The rearing experiment was conducted at the Nagasaki Prefectural Institute of Fisheries. Juvenile Japanese
86 flounders (total length, TL, = 12.82 ± 1.59 cm, mean \pm SD) without hypermelanosis or negligible
87 hypermelanosis (<1% of the darkened area ratio, see below in detail) on the blind side were selected and
88 kindly provided by a private hatchery (Ootawa Shubyo, Saikai, Nagasaki, Japan). They were randomly
89 allocated to seven experimental tanks (500 L circular tank, bottom diameter = 97.5 cm, transparent
90 polycarbonate) with the following characteristics: (1) net-lined tank (net-lined); (2) 100% coverage bottom-

91 net without slack (100% tight net); (3) 100% coverage bottom-net with slack (100% loose net); (4) 50%
92 coverage bottom-net with slack (50% loose net); (5) 30% coverage bottom net with slack (30% loose net);
93 (6) 10% coverage bottom-net with slack (10% loose net); and (7) without net (control).

94 In treatment (1) net-lined tank, white net (mesh size 12 mm, Russell knitting, polyethylene, Dionet; Dio
95 Chemicals Co., Ltd., Tokyo, Japan) was processed into a pouch shape of a size approximately similar to the
96 tank, and loosely set inside the tank, consequently covering the entire inner surface of the tank with
97 undulated net material. In treatment (2) 100% tight net tank, a framed round net (diameter = 97.5 cm, the
98 same size as the bottom of the tank, consequently covering 100% of the tank bottom) was made by fixing
99 the same net material without undulation to a circular white frame (fiber-reinforced plastics, $\text{\O} = 7$ mm,
100 GF-7; Yamaten Co., Ltd., Osaka, Japan), and further fixed to the tank bottom. In treatment (3) 100% loose
101 net tank, a framed round net of the same size was made, but with 10% larger net material, resulting in an
102 undulated net surface with approximately 5 cm bumping. In treatments (4) 50% loose net, (5) 30% loose
103 net, and (6) 10% loose net tanks, a similar undulated framed net was made, but with smaller round frames
104 (diameter = 68.9 cm, 53.4 cm and 30.8 cm, respectively) in order to adjust the bottom covering ratio [= $\frac{\text{area of the framed net}}{\text{bottom area of the tank}} \times 100$]
105 to 50, 30 and 10%, respectively. All framed nets
106 were fixed on the bottom with a silicone sealant (Bus Bond Q Clear, Konishi Co., Ltd., Osaka, Japan) to
107 prevent floating up from the bottom.

108 All rearing tanks were placed indoors and only received indirect and weak sunlight, even during the

109 daytime (max 10 lx; measured about 20 cm above the water surface). The rearing tanks were placed on a
110 white Styrofoam sheet, and the side walls of the tanks were covered with the same Styrofoam sheet on the
111 outside. Since the rearing tanks were transparent, the entire inner surface of the tank was white including
112 the area that uncovered by white net. On April 6, 2018, 30 juveniles were stocked in each experimental
113 tank. Ultraviolet-sterilized natural seawater was supplied at a rate of ten rotations per day, and an air stone
114 was set at the center of each tank. The water temperatures at the beginning and end of the experiment were
115 19.5°C and 22.7°C, respectively. The lowest and highest water temperatures during the experiment were
116 18.0°C (April 28, 2018) and 28.5°C (September 7, 2018), respectively.

117 Flounders were fed commercial pellets (initially Hirame EPF-2 and later Hirame EPF-3 according to fish
118 size, Marubeni Nisshin Feed Co., Ltd., Tokyo, Japan) every morning. Starting from April 27, the total body
119 weight of all flounders in each experimental tank was weighed once a month, and the pellet (0.7-3% of the
120 total body weight of the tank) was fed for the next month. Diet was not supplied from one day before
121 measurement and taking photographs, but was supplied in the evening after the manipulation. Remnant diet
122 and excretes in tanks were discarded once a day before feeding with a syphon tube. On October 26, 2018,
123 the rearing experiment was finished.

124

125 Photography and image analyses

126 Approximately once a month, all flounders in each experimental tank were anesthetized with 200 ppm 2-

127 phenoxyethanol, and photographs of the blind side were individually taken with a ruler using a digital
128 camera (α 550, DSLR-550, Sony Marketing Inc., Tokyo, Lens; AF 50/2.8 macro, MINOLTA Co., Ltd.,
129 Osaka, Japan, Photographic conditions; ISO = 200, aperture = 22 for 0 month and 6.3-8 for others, shutter
130 speed = 1/125 for 0 month and 1/2-1/15 for others). Using ImageJ (National Institute of Health, USA,
131 <https://imagej.nih.gov/ij/>. Accessed 4/6/2021), standard length (from the tip of the lower jaw to the posterior
132 end of the hypural), head length (to the posterior end of the operculum), body depth (maximum length in
133 the dorsal-ventral axis), blind side area (excluding fins), and darkened area of the blind side were measured
134 manually from the pictures of the blind side. Although the hypural itself was not visible on the pictures,
135 standard length could be determined with sufficient accuracy for the purpose of this study.

136 The darkened area ratio was calculated as follows:

$$137 \quad (\text{darkened area ratio}) = (\text{darkened area}) / (\text{blind side area}).$$

138 The fish coverage ratio of the tank was calculated as follows:

$$139 \quad (\text{fish coverage ratio})$$

$$140 \quad = (\text{sum of blind side area of all individuals in each tank}) / (\text{bottom area of the tank})$$

141 Because single value of fish coverage ratio is available from one tank, statistical analysis was not carried
142 out for this value.

143 To determine the timing suitable for comparison among groups, we first examined the time course
144 changes in the darkened area ratio in the following two tanks: control as the most heavily darkened, and

145 30% loose net tank as mildly darkened. The lightly darkened group (net-lined tank) was not examined at
146 this step because almost no darkening was observed even at the end of the experiment. In addition, changes
147 in the standard length and fish coverage ratio were examined in the two tanks.

148

149 Blood sampling and cortisol measurement

150 After taking the final photographs at the end of the experiment, blood samples were collected from fish in
151 the control tank and net-lined tank, from the tail vein using a heparinized syringe under anesthesia with 200
152 ppm 2-phenoxyethanol. To minimize the influence of stress-induced increase of cortisol on the comparison
153 between the tanks, blood samples were collected from three individuals at once (scooped and anesthetized
154 together) from each tank, and alternately from control and net-lined tanks. Blood samples were then
155 centrifuged, and the separated plasma was frozen at -20°C, and the cortisol concentration was measured by
156 a specific radioimmunoassay after diethyl ether extraction and carbon tetrachloride washing following the
157 methods of Hiroi et al. (1997).

158

159 Statistical analysis

160 All statistical analyses were performed using EZR (Kanda 2013; available at
161 <https://www.jichi.ac.jp/saitama-sct/> Accessed 4/6/2021, Saitama Medical Center, Jichi Medical University,
162 Saitama, Japan), which is a graphical user interface for R (R Core Team 2014). The significance level was

163 set at $p < 0.05$. First, the normality of data was confirmed using the Shapiro-Wilk test. When normality was
164 not rejected, the equality of variance of the data was confirmed using the Bartlett test. When both normality
165 of the data and equality of variance were not rejected, the statistical differences among the averages of the
166 data values were parametrically examined by one-way analysis of variance followed by the Tukey method.
167 When either normality or equality of the variance was rejected, the statistical differences among the
168 averages of the data values were non-parametrically examined by the Mann-Whitney U test (two groups)
169 or Kruskal-Wallis test followed by the Steel-Dwass method (three groups or more). For the survival of
170 juveniles, statistical significance was tested by the Fisher's exact test with significance level adjusted by
171 the Bonferroni method.

172

173 Results

174 Time course changes in the darkened area ratio and body size

175 To determine the timing suitable for comparison among groups, we first examined the time course changes
176 in the darkened area ratio in control and 30% loose net tank as described in materials and methods section.
177 Although the darkened area ratio was higher in the control tank, as expected, the value continued to increase
178 and did not saturate even at the end of the sixth month in both tanks (Fig. 1a). As shown in the figure, the
179 30% loose net tank showed a significantly lower darkened area ratio consistently after 4 months (Mann-
180 Whitney U test, $p < 0.05$, $n = 29-30$). In addition, the first appearance of the significantly darkened area looked

181 similar in the two tanks; almost no darkened area in the first month, appeared in the second month, and
182 significantly increased in the third month. The standard length (Fig. 1b) and fish coverage ratio (Fig. 1c)
183 increased similarly between the two tanks, with slightly higher values in the control tank. For standard
184 length, statistical differences were detected at three time points (Mann-Whitney *U* test, $p < 0.05$, $n = 29-30$).

185

186 Comparison of survival, body size and body shape at the sixth month

187 As basic information, survival of the juveniles, body size and body shape were compared among
188 treatments. The survival of juveniles at the sixth month were not statistically different among experimental
189 tanks (Fisher's exact test, $p > 0.05$, net-lined; $n = 29$, 100% tight net; $n = 29$, 100% loose net; $n = 30$, 50% loose
190 net; $n = 28$, 30% loose net; $n = 29$, 10% loose net; control; $n = 30$).

191 For the standard length of all tanks, as shown in Figure 2a, there was no statistical difference at the sixth
192 month (Kruskal-Wallis test, $p > 0.05$, $n = 28-30$). However, statistical differences were detected in head length
193 ratio (100% loose net < 30% loose net, Fig. 2b) and body depth/standard length (100% tight net < net lined,
194 30% loose net, 10% loose net and control, Fig. 2c) (Tukey method, $p < 0.05$, $n = 28-30$).

195

196 Comparisons of the darkened area ratio at the sixth month

197 At first, to find a better way of net settings the effects of the net-setting method are examined (Figure 3
198 and 4). The darkened area ratios in the tank with 100% tight net (median = 14%) and 100% loose net (8%)

199 were significantly lower than, and approximately 1/3 and 1/5, respectively, of the control tank (46%) (Steel-
200 Dwass method, $p < 0.05$, $n = 28-30$). However, these values were significantly higher, approximately three
201 times and two times higher, respectively, than those in the net lined tank (4%) (Steel-Dwass method, $p < 0.05$,
202 $n = 28-30$). Although the difference of darkened area ratio between the net-lined tank and the 100% loose
203 net was statistically significant, the blind side appearance of 50% individuals was not much different by the
204 naked eye (Fig. 4).

205 Because a strong suppression effect was observed in the tank with a loose net on the tank bottom, next,
206 using loose net, we examined the effect of the bottom covering ratio on the darkened area ratio at the sixth
207 month (Figs. 5 and 6). There was a clear dose-response relationship; a smaller darkened area ratio was
208 attained in the tank with a higher bottom covering ratio.

209

210 Plasma cortisol concentration in control tank and net-lined tank at the sixth month

211 To examine the possible contribution of stress-cortisol axis to the hypermelanosis prevention by the net,
212 plasma cortisol concentration was measured in the juveniles in control tank and net-lined tank. However,
213 there was no significant difference in the plasma cortisol concentration between the control tank and net-
214 lined tank (Fig. 7, Mann-Whitney U test, $p > 0.05$, $n = 29$ and 28, respectively).

215

216 **Discussion**

217 Although the occurrence of the hypermelanosis is known to differ among the production lot of
218 flounders, loosely installing net material covering only 50% of the bottom area significantly suppressed
219 hypermelanosis, as low as 1/3 of the control tank (ordinary flat bottom tank), in this specific experiment.
220 In addition, a strong negative correlation was found between bottom net coverage ratio with slack-net and
221 darkened area ratio. Because the juvenile flounders of about 13 cm in standard length without
222 hypermelanosis were selectively used in this experiment, there is a limitation of the applicability of results,
223 individual values of net size and darkened area ratio, for example. But effectiveness of loosely-installed
224 bottom net, together with the negative correlation between bottom net size and severity of hypermelanosis,
225 were clearly indicated in the present study.

226

227 Changes in darkened area ratio and body size during experiment

228 This study was the first trial to examine the time course of the appearance and expansion of
229 hypermelanosis using a net material. During the planning of the experiment, we expected that the increase
230 in the darkened area ratio would stop in the second month, because stasis of darkening expansion was
231 previously observed at about 60-80 days (Seikai 1991) or two months (Isojima et al. 2013b) of rearing
232 without bottom sand. Although the experimental period was extended to six months, the longest possible
233 period due to the restriction of experimental equipment, the darkening area expansion did not stop until the
234 sixth month in this study. Therefore, comparisons among experimental groups were conducted using the

235 data obtained in the sixth month. Between preceding studies and the present experiments, various factors
236 are different: the origin of flounder juveniles, initial size, and experimental season. At present, it is unclear
237 why the hypermelanosis expansion did not stop even after sixth months. It would be helpful to understand
238 the control mechanism of darkening expansion by further accumulating the information.

239 On the other hand, the appearance timing of the darkened area were similar between the control tank and
240 the 30% loose net tank. This result indicates that the effect of installing a net material does not delay the
241 appearance timing, but reduces the expansion speed. In addition, in the second month, the fish coverage
242 ratio of both tanks increased from approximately 0.3 to approximately 0.5. This is reasonable because the
243 growth speed was similar between the two tanks. Although we cannot exclude the possible “spontaneous”
244 appearance of hypermelanosis according to time, the appearance of darkened area is possibly “triggered”
245 by the fish coverage ratio of over 0.3-0.4, at least in the individuals used in the present experiment.

246 As shown in Fig. 1, the darkened area ratio at zero months was statistically different; average, not median,
247 was higher in the 30% loose tank. This was probably due to unexpected deviations in the random allocation
248 of individuals. Since all the individuals at zero months had smaller values of the darkened area ratio (less
249 than 1%), it is highly possible that this difference affected the comparison among treatments at the sixth
250 month.

251 The speculated mechanism(s) of the appearance and expansion of darkened area during the experiment
252 is 1) increase in melanin contents on non-pigmented cells on the skin, 2) increase in number of

253 melanophores potentially differentiated from melanoblasts, 3) both of them. However, we did not examine
254 the changes in melanin contents and number of melanophores on the blind side skin. Future studies on these
255 points will provide fundamental understanding on the staining type hypermelanosis.

256 Comparison of standard length and body shape at the sixth month

257 Nakata et al. (2017) and Mizutani et al. (2020) reported that the standard length of flounders reared in net-
258 lined tanks was significantly smaller than that in control tanks. From the results of the first experiment by
259 Mizutani et al. (2020), they attributed the reduced growth to the lower food availability due to the
260 inaccessible pellets between the net and the tank bottom, because pellets of the sinking type were offered,
261 and some of them sink through the mesh before consumption. However, in the second experiment, although
262 floating-type pellets were used, similar results were obtained. Therefore, Mizutani et al. (2020) suggested
263 the presence of other possible factors for reduced growth in net-lined tanks. In addition, body
264 depth/standard length in the net-lined tank was significantly lower than that in the control tank and was
265 closer to that of wild-caught flounders in Mizutani et al. (2020).

266 In the present study, we used floating-type pellets; however, there was no significant difference in the
267 standard length among the seven experimental tanks. In addition, regarding body depth/standard length and
268 head length ratio, although statistical differences were observed among several tanks, we could not find any
269 simple and logical explanations for the results. Therefore, to reduce growth and alter body shape, further
270 research is needed to clarify the effect of net-lined tanks, starting with the confirmation of the

271 reproducibility of the phenomena.

272

273 Comparisons of the darkened area ratio at the sixth month

274 In this study, the darkening suppression effect was first compared among various methods of net setting,
275 including a net-lined tank (covering the entire inner surface of the tank), in which the prevention effect has
276 been proven in our previous studies (Nakata et al. 2017; Mizutani et al. 2020), and a bottom net, in which
277 the setting of the net is expected to be easier than the net-lining when applied to much larger tanks of actual
278 aquaculture scene.

279 The median darkened area ratio was 4% in the net-lined tank, which was the smallest among all the
280 experimental tanks. The value was 9% in tanks with a 100% loose net, though larger than net-lined, but
281 significantly less than, and only approximately 1/5 of the control tank (46%). In the net-lined tank, it is
282 clear that the bottom portion of the net has a major effect, and the vertical portion has a minor effect on the
283 prevention of hypermelanosis. In our observation, some individuals occasionally locate themselves on the
284 side wall of the tank, attaching themselves vertically. Therefore, for fish in the net-lined tank, there is a
285 higher probability of being able to contact the net material.

286 The tension of the bottom net is another contributing factor. Although there was no significant difference,
287 the loose net (9%) had a slightly stronger effect than tight nets (14%). Together with the smaller variance
288 of the darkening area ratio, it is better to loosely set the bottom net, giving undulation on the net surface.

289 This conclusion is reasonable because the undulated surface increases the contact area between the blind
290 side of the flounder and the net, and therefore effectively prevents hypermelanosis (Nakata et al., 2017). As
291 shown in the photograph, although the net-lined tank seems to be the best, the 100% loose net has practically
292 comparable prevention effect.

293 For the effect of the bottom covering ratio on the darkened area ratio, we initially expected the presence
294 of a net size of “necessary and sufficient”, in a range less than 100% bottom coverage. However, as shown
295 in Figure 5, the higher the coverage, the lower the ratio of the darkening area. Therefore, for application to
296 actual aquaculture scenarios, it is necessary to first know the required clean level of the blind side from the
297 aquaculture farm. Next, based on the required clean level in terms of the darkened area ratio, the suitable
298 bottom covering rate by the net can be proposed. Although it seems difficult to decrease the darkened area
299 ratio of the “darkest” fish, a 50% loose net may be a candidate for practical application in aquaculture
300 (Fig.6).

301 Among the factors contributing for hypermelanosis suppression, tank color (Amiya et al. 2005;
302 Yamanome et al. 2005, 2007a; Nakata et al. 2017) and undulation of the bottom (Nakata et al. 2017) are
303 important. In this experiment, to have a similar color condition among tanks, the experimental tanks were
304 transparent and covered with white Styrofoam sheet, and color of the net was also white. In addition, to
305 have a similar undulation among different size nets, all loose nets were made with net materials having
306 10% larger size than their frames. Therefore, color condition of the tank and degree of undulation of the net

307 are expected to be similar among experimental tanks, and the difference among the tanks are expected to
308 be due to the difference of the net size. In this experiment, all darkened areas on the blind side were
309 measured together without considering the location. Hypermelanosis preventing effect of net-lined tank
310 was different depending on the location, minor on the darkened area at the base of pectoral fin and stronger
311 on those at the base of dorsal and anal fins (Mizutani et al., 2020). As shown in figure 6, similar tendency
312 was found in this experiment. Because the darkened areas at the base of dorsal and anal fins are larger than
313 those at other locations, it is possible that loose net on the tank bottom exerted its hypermelanosis-
314 preventing effect mainly through these areas.

315 Because the bases of dorsal and anal fins have different chance to contact with bottom substrate between
316 flat bottom and loose net, contact stimulation is a candidate as a direct inhibitor of the hypermelanosis in
317 net tanks, as suggested for the undulated bottom by Nakata et al. (2017). This idea could be supported by
318 the discussion on the relationship between the net size and the fish coverage rate (Online Resource 1).
319 Staying time of individual juveniles on the net and on the flat surface possibly affects the effectiveness of
320 hypermelanosis prevention but was not examined in this experiment. Therefore, this point should be
321 clarified in future. If the contact stimulation is the main inhibitor of the hypermelanosis, the prevention
322 effectiveness of the loose net may not be limited for circular tanks used in this experiment, and probably
323 also effective for larger rectangular concrete tanks. This point should be examined in the next step for the
324 application of loose net in aquaculture farms at industrial scale. More detailed mechanisms of

325 hypermelanosis prevention by loose net should be examined in future.

326

327 Blood cortisol levels of the fish reared in the control tank and net-lined tank at the sixth month

328 Cortisol is secreted by stress (Pickering and Pottinger 1989). Addition of cortisol in rearing water promotes

329 hypermelanosis in spotted halibut (Yamada et al. 2011) and cortisol supplementation in diet also promotes

330 hypermelanosis in Japanese flounder (Matsuda et al. 2018b). Therefore, in tanks with net materials on the

331 bottom, one of the possible mechanisms for hypermelanosis prevention is the lowered cortisol level caused

332 by possible stress reduction due to the comfortable environment. However, as shown in Figure 7, there was

333 no significant difference in the blood cortisol concentration at the end of experiment between the most

334 heavily darkened group (control tank) and the least darkened group (net-lined tank). This finding does not

335 support the contribution of stress-cortisol axis to the hypermelanosis prevention by the net. We could not

336 find information on the contribution of upstream regulatory hormone of cortisol, adrenocorticotropic

337 hormone (ACTH), for hypermelanosis. But in the two papers on the enhancing effect of cortisol on

338 hypermelanosis (Yamada et al. 2011; Matsuda et al. 2018b), ACTH levels are expected to be decreased

339 because of negative feedback. Hypermelanosis enhancing effect of ACTH should be minor compared to

340 cortisol.

341 It was better to collect blood samples several times during the experiment to cover the progression period

342 of hypermelanosis, not one time at the end of experiment. However, to avoid additional stress and damage

343 to juveniles, and not to affect the result of hypermelanosis, blood was collected only at the end of the
344 experiment. In order to examine the presence or absence of cortisol contribution on spontaneously
345 progressing hypermelanosis, further experiment is required.

346

347 In conclusion, this study demonstrated that loosely set bottom nets, an easier method to be applied to larger
348 tanks, can prevent hypermelanosis with effectiveness comparable to that of a net-lined tank. In addition,
349 covering 50% of the bottom significantly reduced the darkening area at approximately 1/3 of the ordinary
350 flat bottom tank in this experiment. When lesser darkening is required, the use of a larger bottom net with
351 a higher coverage ratio is recommended. For effective hypermelanosis prevention at the industrial scale,
352 the use of a rearing tank with a loosely set bottom net is a strong alternative for introducing bottom sand
353 for flounder aquaculture.

354

355 Acknowledgements

356 We would like to thank Ootawa Shubyo, Saikai City, Nagasaki Prefecture, for kindly providing the juvenile
357 Japanese flounder. We acknowledge the Radioisotope Research Center, Kyoto University, for the technical
358 support in the radioisotope experiments. We would also like to thank the members of the Laboratory of
359 Marine Stock-Enhancement Biology, Kyoto University, for their invaluable discussions and encouragement
360 throughout the course of the study. This work was supported by JSPS KAKENHI Grant number 19K06237

361 to M.T.

362

363

364 References

365 Amiya N, Amano M, Takahashi A, Yamanome T, Kawauchi H, Yamamori K (2005) Effects of tank

366 color on melanin-concentrating hormone levels in the brain, pituitary gland, and plasma of the barfin

367 flounder as revealed by a newly developed time-resolved fluoroimmunoassay. *Gen Comp Endocrinol*

368 143:251–256

369 Aritaki M (2004) Occurrence of ambicolored individuals in hatcheries of Japanese flounder, and

370 questionnaire survey for their market price. In: Fukunaga T, Shiozawa S, Tsuzaki T (eds) *Stock*

371 *enhancement technique series 10. Factor and prevention of color anomaly on blind side in Japanese*

372 *flounder*. Fisheries Research Agency, Tokyo, pp 135–139 (in Japanese)

373 Aritaki M, Seikai T (2017) Reducing malformations in body-coloration and eye-position in

374 *Pleuronectiformes* by controlling their developmental speed. In: Aritaki M, Tagawa M, Soyano K (eds)

375 *Morphological abnormality in teleosts - involvement of rearing environments in body coloration and*

376 *skeletal development*, pp 63–71 (in Japanese)

377 Fukunaga T (1998) Present status of technique to prevent occurrence of hypermelanosis on the blind

378 side of juvenile Japanese flounder. In: Association Japan Sea Farming (ed) *A textbook for*

379 understanding basic theory XII. Japan Sea Farming Association, Tokyo, pp 1–46 (in Japanese)

380 Haga Y, Takeuchi T, Murayama Y, Ohta K, Fukunaga T (2004) Vitamin D3 compounds induce
381 hypermelanosis on the blind side and vertebral deformity in juvenile Japanese flounder *Paralichthys*
382 *olivaceus*. *Fish Sci* 70:59–67

383 Hiroi J, Sakakura Y, Tagawa M, Seikai T, Tanaka M (1997) Developmental changes in low-salinity
384 tolerance and responses of prolactin, cortisol and thyroid hormones to low-salinity environment in
385 larvae and juveniles of Japanese flounder, *Paralichthys olivaceus*. *Zool Sci* 14:987–992

386 Isojima T, Makino N, Miyama Y, Tagawa M (2014) Effects of time and duration of rearing with bottom
387 sand on the occurrence and expansion of staining-type hypermelanosis in the Japanese flounder
388 *Paralichthys olivaceus*. *Fish Sci* 80:785–794

389 Isojima T, Makino N, Takakusagi M, Tagawa M (2013b) Progression of staining-type hypermelanosis
390 on the blind side in normally metamorphosed juveniles and pigmentation progression in pseudoalbino
391 juveniles of the Japanese flounder *Paralichthys olivaceus* using individual identification. *Fish Sci*
392 79:787–797

393 Isojima T, Tsuji H, Masuda R, Tagawa M (2013a) Formation process of staining-type hypermelanosis
394 in Japanese flounder juveniles revealed by examination of chromatophores and scales. *Fish Sci*
395 79:231–242

396 Iwata N, Kikuchi K (1998) Effects of sandy substrate and light on hypermelanosis of the blind side in

397 cultured Japanese flounder *Paralichthys olivaceus*. Environ Biol Fishes 52:291–297

398 Kaji S, Fukunaga T (1999) Results of a questionnaire on the recent status of seed production and
399 market price of recaptured Japanese flounder *Paralichthys olivaceus* showing abnormal coloration.
400 Saibai Giken 27:67–101 (in Japanese)

401 Kanda Y (2013) Investigation of the freely available easy-to-use software “EZR” for medical statistics.
402 Bone Marrow Transplant 48:452–458

403 Kang DY, Kim HC (2012) Relevance of environmental factors and physiological pigment hormones
404 to blind-side hypermelanosis in the cultured flounder, *Paralichthys olivaceus*. Aquaculture 356–
405 357:14–21

406 Kang DY, Kim HC (2013a) Influence of density and background color to stress response, appetite,
407 growth, and blind-side hypermelanosis of flounder, *Paralichthys olivaceus*. Fish Physiol Biochem
408 39:221–232

409 Kang DY, Kim HC (2013b) Importance of bottom type and background color for growth and blind-
410 side hypermelanosis of the olive flounder, *Paralichthys olivaceus*. Aquac Eng 57:1–8

411 Kang DY, Kim HC (2015) Functional relevance of three proopiomelanocortin (POMC) genes in
412 darkening camouflage, blind-side hypermelanosis, and appetite of *Paralichthys olivaceus*. Comp
413 Biochem Physiol B 179:44–56

414 Miwa S, Yamano K (1999) Retinoic acid stimulates development of adult-type chromatophores in the

415 flounder. *J Exp Zool* 284; 317-324

416 Matsuda N, Kasagi S, Nakamaru T, Masuda R, Takahashi A, Tagawa M (2018a) Left-right
417 pigmentation pattern of Japanese flounder corresponds to expression levels of melanocortin receptors
418 (MC1R and MC5R), but not to agouti signaling protein 1 (ASIP1) expression. *Gen Comp Endocrinol*
419 262:90–98

420 Matsuda N, Yamamoto I, Masuda R, Tagawa M (2018b) Cortisol promotes staining-type
421 hypermelanosis in juvenile Japanese flounder. *Aquaculture* 497:147–154

422 Mizutani K, Yamada T, Suzuki KW, Masuda R, Nakata K, Tagawa M (2020) Prevention of
423 hypermelanosis by rearing Japanese flounder *Paralichthys olivaceus* in net-lined tanks. *Fish Sci*
424 86:127–136

425 Nakamura M, Seikai T, Aritaki M, Masuda R, Tanaka M, Tagawa M (2010) Dual appearance of
426 xanthophores, and ontogenetic changes in other pigment cells during early development of Japanese
427 flounder *Paralichthys olivaceus*. *Fish Sci* 76:243–250

428 Nakata K, Yamamoto I, Miyama Y, Nakamaru T, Masuda R, Tagawa M (2017) Undulated flooring in
429 the rearing tank decreases hypermelanosis in Japanese flounder *Paralichthys olivaceus*. *Fish Sci*
430 83:1027–1035

431 Pickering AD, Pottinger TG (1989) Stress responses and disease resistance in salmonid fish: Effects
432 of chronic elevation of plasma cortisol. *Fish Physiol Biochem* 7:253–258

433 R Core Team (2014) R: A language and environment for statistical computing. R Foundation for
434 Statistical Computing, Vienna, Austria. <http://www.R-project.org/>. Accessed 4/6/2021

435 Seikai T (1991) Influences of fluorescent light irradiation, ocular side pigmentation, and source of
436 fishes on the blind side pigmentation in the young Japanese flounder, *Paralichthys olivaceus*. *Aquac*
437 *Sci* 39:173–180 (in Japanese with English abstract)

438 Seikai T, Matsumoto J, Shimozaki M, Oikawa A, Akiyama T (1987) An association of melanophores
439 appearing at metamorphosis as vehicles of asymmetric skin color formation with pigment anomalies
440 developed under hatchery conditions in the Japanese flounder, *Paralichthys olivaceus*. *Pigment Cell*
441 *Res* 1:143–151

442 Statistical survey of seawater fishery production (2018).
443 http://www.maff.go.jp/j/tokei/kouhyou/kaimen_gyosei/index.html. Accessed 4/6/2021

444 Tagawa M (2017) Possible involvement of endocrine systems in body-coloration and eye-position
445 abnormalities in Pleuronectiformes. In: Aritaki M, Tagawa M, Soyano K (eds) *Morphological*
446 *abnormality in teleosts - involvement of rearing environments in body coloration and skeletal*
447 *development*, pp 96–104 (in Japanese)

448 Tarui F, Haga Y, Ohta K, Shima Y, Takeuchi T (2006) Effect of *Artemia* nauplii enriched with vitamin
449 A palmitate on hypermelanosis on the blind side in juvenile Japanese flounder *Paralichthys olivaceus*.
450 *Fish Sci* 72:256–262

451 Yamada T, Donai H, Okauchi M, Tagawa M, Araki K (2011) Induction of ambicoloration by
452 exogenous cortisol during metamorphosis of spotted halibut *Verasper variegatus*. *Comp Biochem*
453 *Physiol B* 160; 174-180

454 Yamanome T, Amano M, Takahashi A (2005) White background reduces the occurrence of staining,
455 activates melanin-concentrating hormone and promotes somatic growth in barfin flounder.
456 *Aquaculture* 244:323–329

457 Yamanome T, Amano M, Amiya N, Takahashi A (2007a) Hypermelanosis on the blind side of Japanese
458 flounder *Paralichthys olivaceus* is diminished by rearing in a white tank. *Fish Sci* 73:466–468

459 Yamanome T, Chiba H, Takahashi A (2007b) Melanocyte-stimulating hormone facilitates
460 hypermelanosis on the non-eyed side of the barfin flounder, a pleuronectiform fish. *Aquaculture*
461 270:505–511

462

463

464 **Figure captions**

465 **Fig. 1 Time course changes in darkened area ratio (a), standard length (b), and fish coverage ratio**
466 **(c)**

467 Open circles, 30% loose net; closed circle, control. Data are presented as median values with 25th and 75th
468 percentiles. Asterisks indicate statistical difference between the groups (Mann-Whitney *U* test, $p < 0.05$,

469 $n=29 - 30$)

470

471 **Fig. 2 Comparisons of standard length (a), head length ratio (b), and body depth / standard length**
472 **(c), at the sixth month**

473 The upper and lower ends of the vertical bars show the maximum and minimum values, respectively. The
474 top and bottom of the boxes show the 75 and 25% values, respectively, and the horizontal line in the box
475 shows the median value. The different lowercase letters indicate statistical difference between the groups
476 (Kruskal-Wallis test, $p<0.05$, $n=28 - 30$)

477

478 **Fig. 3 Comparison of darkened area ratio among different net settings at sixth month**

479 Different letters indicate statistical significance among groups (Steel-Dwass method, $p<0.05$, $n=28 - 30$).
480 The upper and lower ends of the vertical bars show the maximum and minimum values, respectively. The
481 top and bottom of the boxes show the 75 and 25% values, respectively, and the horizontal line in the box
482 shows the median value

483

484 **Fig. 4 Blind side photographs of flounders reared in tanks with different net settings at the sixth**
485 **month**

486 The upper photographs are individuals with 75% values, the middle with median, and the lower with 25%

487 values in each tank

488

489 **Fig. 5 Comparison of darkened area ratio among tanks having different bottom covering ratio with**
490 **net at the sixth month**

491 Different letters indicate statistical significance between groups (Steel-Dwass method, $p < 0.05$, $n = 28 - 30$).

492 The upper and lower ends of the vertical bars show the maximum and minimum values, respectively. The

493 top and bottom of the boxes show the 75 and 25% values, respectively, and the horizontal line in the box

494 shows the median value. Dots represent individual values

495

496 **Fig. 6 Blind side photographs of flounders reared in tanks having different bottom covering ratio**
497 **with net at the sixth month**

498 The upper photographs are individuals with 75% values, the middle with median, and the lower with 25%

499 values in each tank

500

501 **Fig. 7 Comparison of plasma cortisol concentrations between control and net-lined tanks at the sixth**
502 **month**

503 There was no statistical difference (Mann-Whitney U test, $p = 0.08$; control: $n = 29$, net-lined: $n = 28$). The

504 upper and lower ends of the vertical bars show the maximum and minimum values, respectively. The top

505 and bottom of the boxes show the 75 and 25% values, respectively, and the horizontal line in the box shows

506 the median value

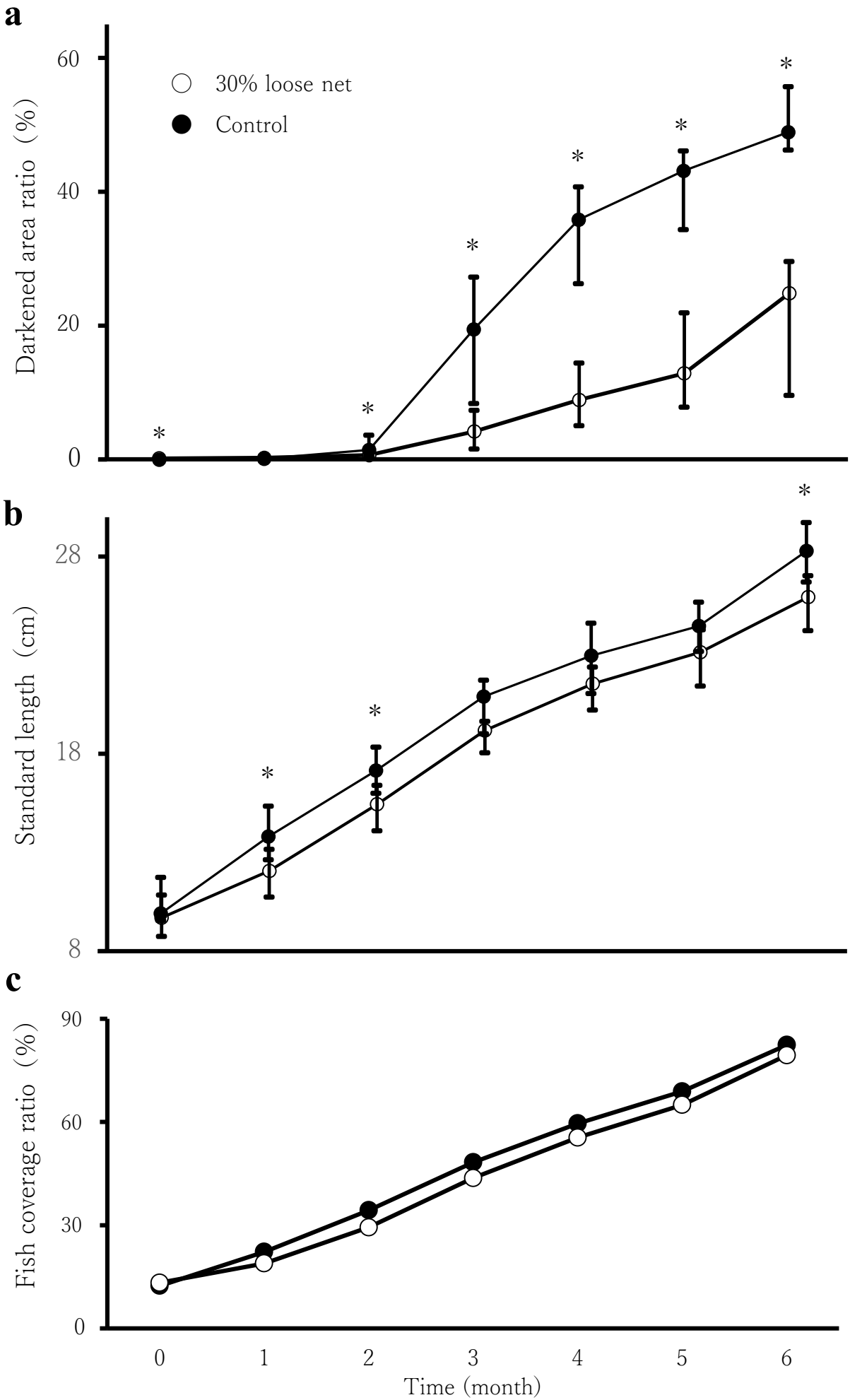
Fig. 1

Fig. 2

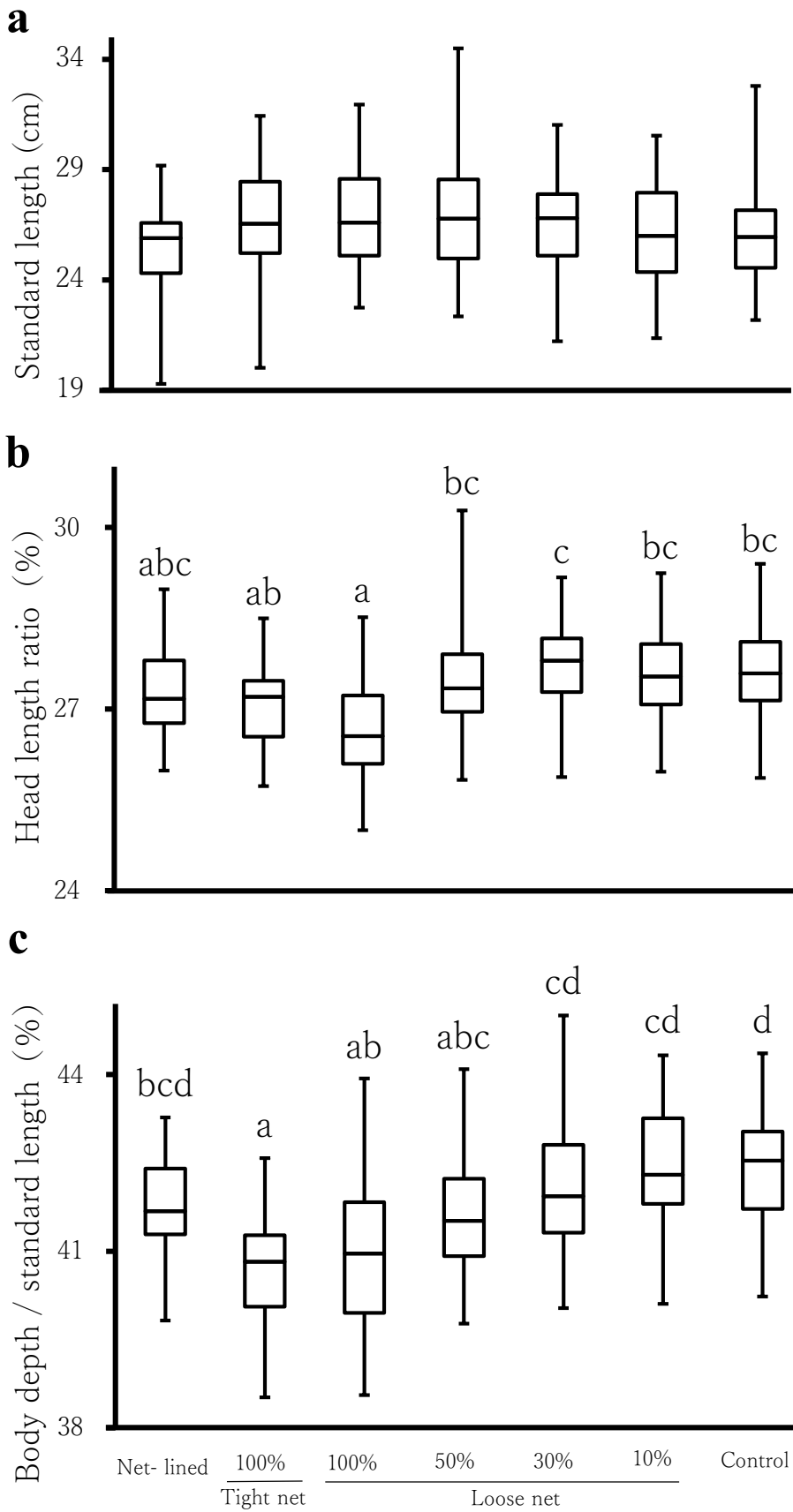


Fig. 3

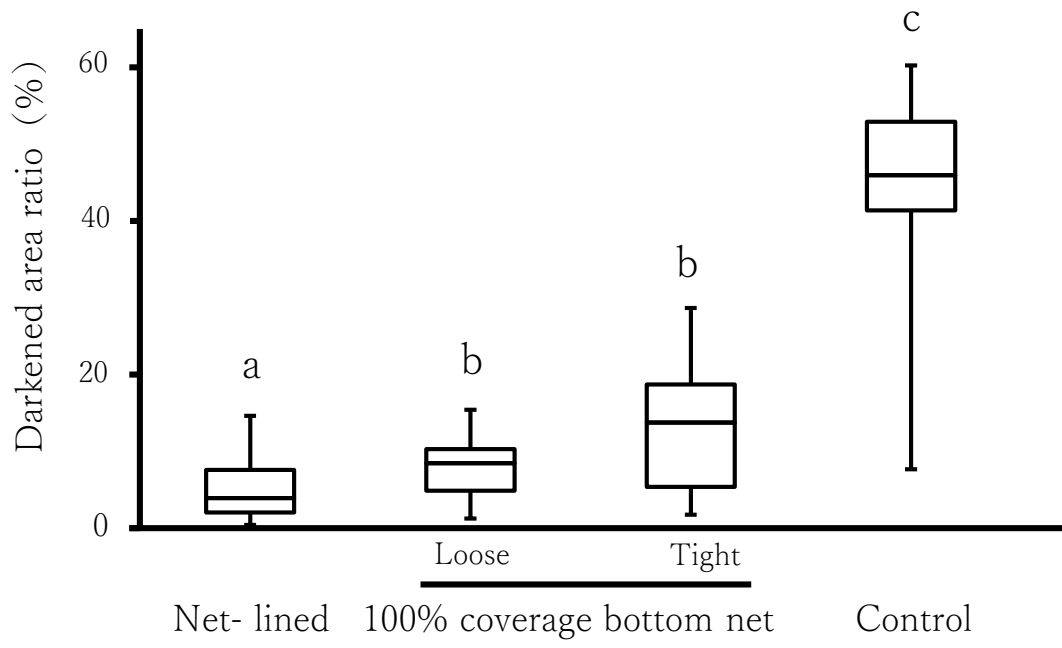
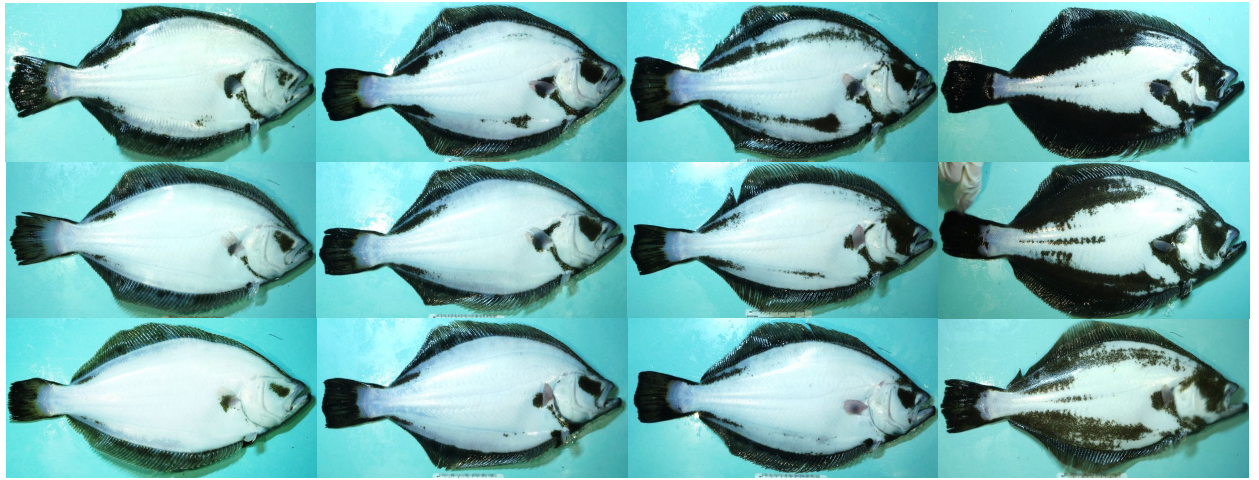


Fig. 4



Net-lined

Loose

Tight

100% coverage bottom net

Control

Fig. 5

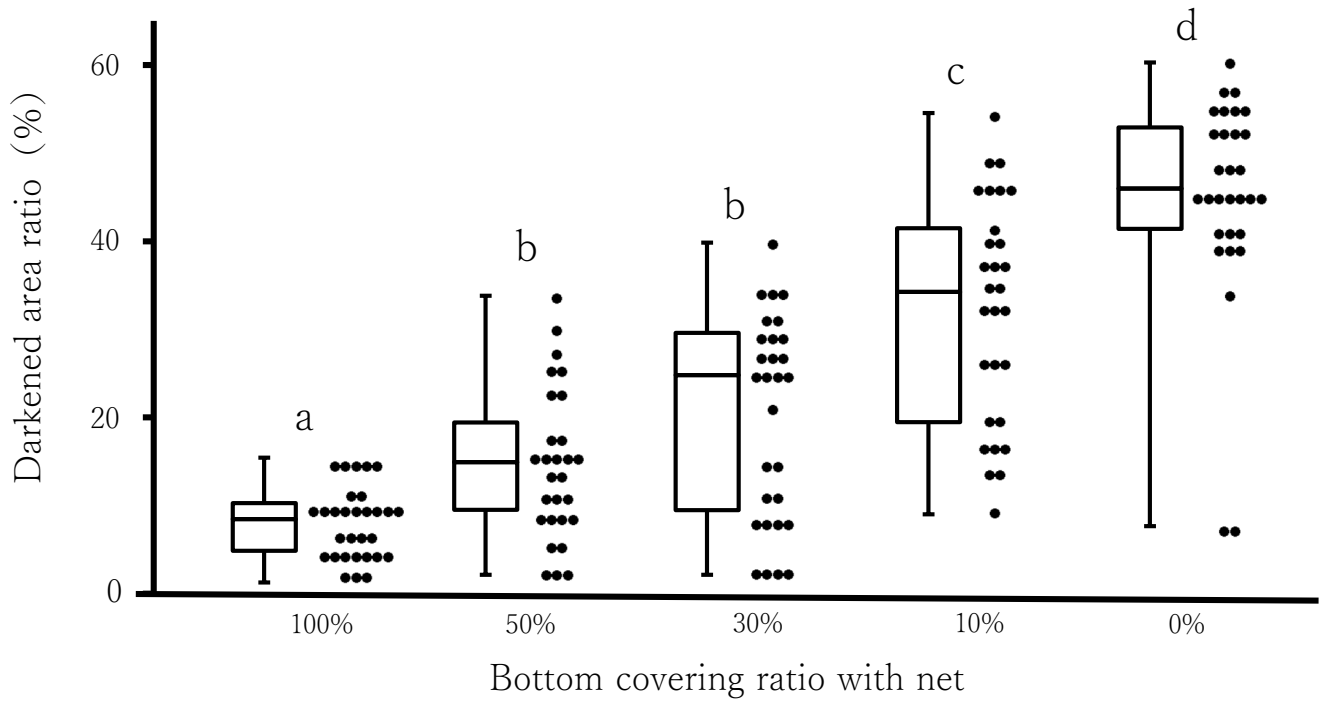


Fig. 6



100

50

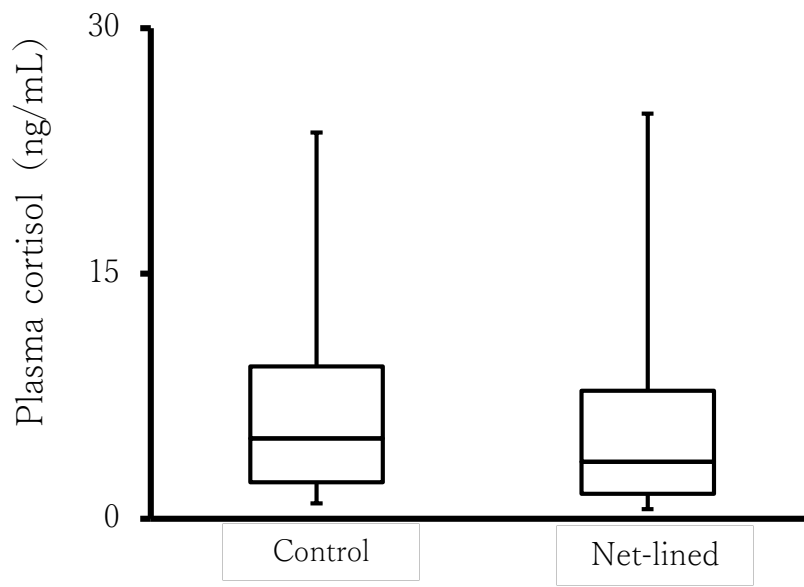
30

10

0

Bottom covering ratio with net (%)

Fig. 7



Article title:

An effective and practical method of net settings in rearing tank to suppress hypermelanosis in Japanese flounder

Journal name:

Fisheries Science

Author names:

Tsuyoshi Onoyama, Toshiyuki Yamada, and Masatomo Tagawa

Affiliation and e-mail address of the corresponding author

Corresponding author: Masatomo Tagawa

Affiliation: Division of Applied Biosciences, Graduate School of Agriculture, Kyoto University,

E-mail address: tagawa.masatomo.8m@kyoto-u.ac.jp

The fish covering ratio of the tank bottom was approximately 80% at the sixth month, as shown in Figure 1c. Therefore, some individuals cannot lie on the net in tanks having the net size of, and the size less than, 50%. The number of individuals that could not lie on the net (therefore on flat bottom, or possibly on other individuals) in each tank was calculated from the bottom coverage ratios of the net and the fish at the sixth month, and compared with the observed number of individuals with a greater darkened area ratio than the darkest individual in the 100% loose net tank, in which all individuals theoretically can fit on the net (Table 1 in this Online Resource 1). In this study, the two numbers matched fairly well. Although we cannot exclude the possibility of coincidence, this observation may suggest the possible prediction of the effectiveness of hypermelanosis prevention from the relationship between the total fish coverage and the net size at a certain time. In addition, it may be possible that individuals with a lower darkened area ratio “prefer” to be located on the bottom net, and spend more time on it. This point should be examined in behavioral studies, with individual identification of experimental juveniles as the next step in this line of research.

Table 1. Comparison of the calculated number of individuals unable to lie on the net material due to insufficient area and the observed number of individuals having a higher darkened area ratio than the maximum value in the 100% loose net tank

group	calculated number	observed number
50% loose net	9.25	13
30% loose net	17.75	17
10% loose net	26.25	26
control	30	28