

1 **Viral nature of the aquatic ecosystems**

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18 **Abstract**

19 Viruses infecting microorganisms are ubiquitous and highly abundant in
20 aquatic environments. They considerably affect the dynamics, diversity, and evolution
21 of their host microorganisms. In this review, we discuss the ecological implications of
22 viruses from the perspectives of the biogeochemical cycles, microbial diversity, and
23 virus–host coevolutionary dynamics in aquatic environments. Generally, viruses redirect
24 host metabolism toward reproduction through molecular host–virus interactions
25 characterized by the compositional and stoichiometric changes in intracellular
26 metabolites, which are eventually released into the environment when the infected host
27 cells are lysed, thus also changing the chemical composition of the water. Therefore, the
28 modulation of metabolite biosynthesis and promotion of their recycling are major viral
29 functions. Viruses also maintain microbial community diversity via increased infection
30 and lysis rates of the dominant taxa and genotypes in a frequency-dependent manner,
31 thereby allowing the co-existence of members with various competitive abilities. Finally,
32 viruses can expand their own genotypic diversity and that of the host through complex
33 defense and counter-defense interactions, including loss of host fitness due to the cost of
34 resistance and the possible need for antiviral defense-specific (e.g., intra- vs.
35 extracellular) changes in the hosts genome diversification. Continuous interactions drive

36 the coevolution of hosts and viruses, thereby increasing both the host and viral
37 micro-diversity. Hence, these fundamental functions are viral “raison d’etre” and are
38 essential for the functioning of aquatic ecosystems and its components.

39

40 **Keywords**

41 Virus-host interactions, Biogeochemical cycle, Microbial diversity, Coevolution, Lytic
42 infection, Lysogenic infection

43

44 **1.1 Introduction**

45 Viruses infecting microorganisms are ubiquitous and abundant in aquatic
46 ecosystems (Suttle 2005, 2007). They typically are small particles (generally 20–200
47 nm in length) comprised of nucleic acids (single- or double-stranded DNA or RNA) and
48 structural proteins and have no intrinsic metabolism. Thus, their reproduction depends
49 entirely on host cellular metabolism and replication machinery. Viral reproduction can
50 be classified as lytic or lysogenic (Guttman et al. 2004). During lytic infection, viruses
51 inject their genomes into host microorganisms, redirect host metabolism for efficient
52 viral genomic nucleic acid replication and protein synthesis, and are finally released
53 through host cell lysis (Guttman et al. 2004). In contrast, in lysogenic infection, the viral
54 genome is integrated into the host genome as a provirus (also called prophage if the

55 virus integrates into the bacterial chromosome) and is propagated vertically within the
56 host lineage until the induction of the lytic cycle under specific conditions (e.g.,
57 depending on host cell density or environmental conditions) (Howard-Varona et al.
58 2017a).

59 Both types of viral infections, lytic and lysogenic, have great potential to affect
60 microbial communities in aquatic ecosystems. For example, viral-mediated cell lysis
61 releases nutrients and organic matter from cells to the environment, thus stimulating
62 biogeochemical cycling (Fuhrman 1999; Suttle 2005, 2007). In addition, viruses affect
63 host microbial diversity in at least three different ways (Marston et al. 2012). First,
64 viruses contribute to the maintenance of host microbial diversity by
65 frequency-dependent infection, often seeming to have a greater effect upon those
66 microbial taxa and genotypes that either are highly abundant or most metabolically
67 active in the environment (Thingstad 2000). Second, viruses increase host genetic
68 diversity via the reciprocal co-evolution of host resistance and viral infectivity
69 (Buckling and Rainey 2002a). Lastly, viruses affect the genomic evolution and the
70 fitness of microbial hosts through horizontal gene transfer (HGT) including the presence
71 and movement of auxiliary metabolic genes (Breitbart et al. 2007; Hurwitz and U'Ren
72 2016), generalized transduction during lytic infection (Thierauf et al. 2009; Touchon et

73 al. 2017), and specialized transduction during lysogenic infection (Gottesman and
74 Yarmolinsky 1968; Fernandes et al. 1989; Campos et al. 2003; Touchon et al. 2017).

75 The ever-growing number of culturable viral isolates, together with recent
76 advances in sequencing technologies and bioinformatics, provides us with a deeper
77 understanding of the viral nature in aquatic ecosystems. This chapter summarizes
78 current understanding of viral effects on biogeochemical cycles, microbial diversity, and
79 virus–host evolutionary dynamics in aquatic ecosystems. The viral role in the promotion
80 of HGT that affects host genomic evolution and fitness has been reviewed extensively
81 elsewhere and is therefore not discussed here (e.g. Balcázar 2018; Yoshida et al. 2019;
82 Chapter E and therein).

83

84 **1.2 Viral influence on biogeochemical cycle**

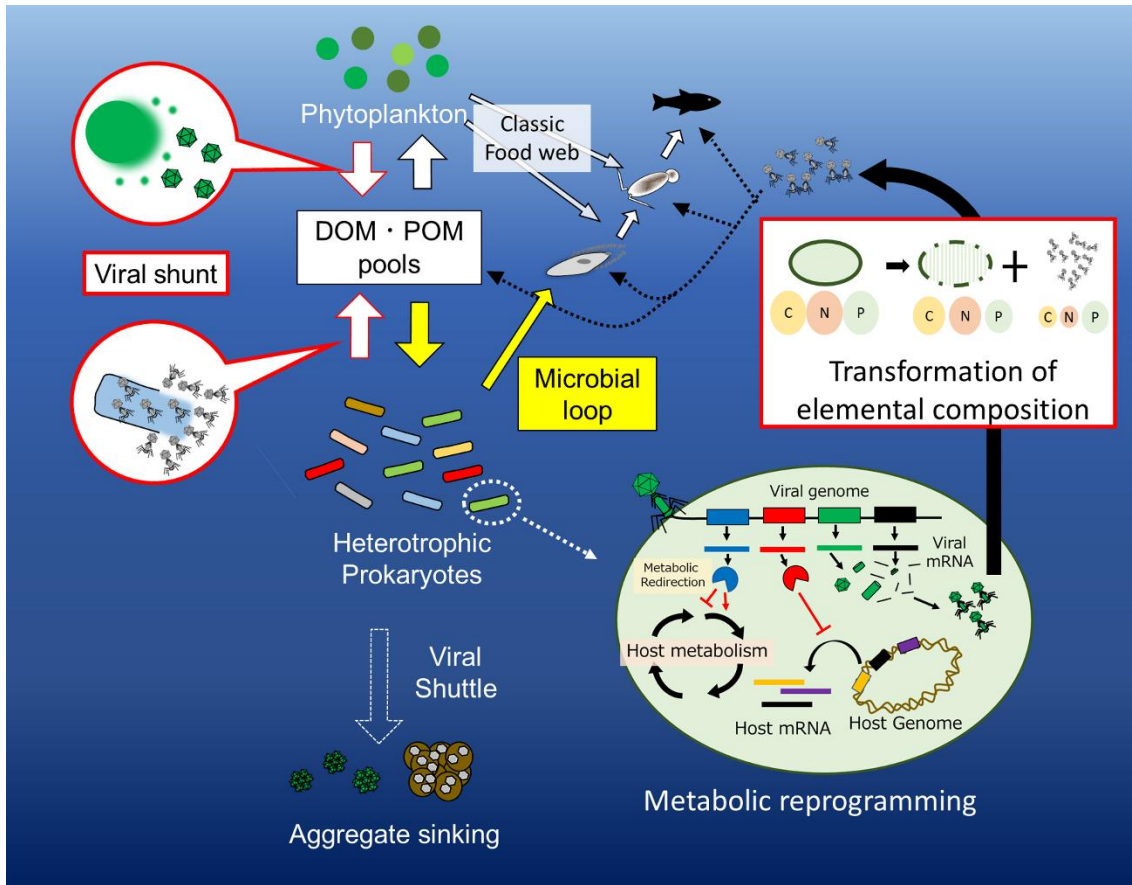
85 ***1.2.1 Viral modulation on patterns of geochemical cycling in the ocean***

86 In the ocean, approximately 10^{29} cells of different microorganisms form the
87 basis of the marine food web (Whitman et al. 1998). Photosynthetic eukaryotes and
88 prokaryotes contribute to up to 50% of the total net primary production on Earth (Field
89 et al. 1998). Approximately half of the resultingly fixed carbon is released into the
90 environment, re-mineralized by heterotrophic prokaryotes, and then incorporated into
91 higher trophic levels of the aquatic food web (Azam et al. 1983). This process of

92 recycling of photosynthetic products is called “microbial loop” and is an essential
93 pathway of biogeochemical cycling in the ocean (Azam et al. 1983).

94 Viruses outnumber prokaryotes, and up to 20% of marine microorganisms are
95 thought to be infected and lysed by viruses daily (according to a certain view in Suttle
96 2007). Lysis of infected cells leads to the release of organic matter and nutrients, which
97 would otherwise be incorporated into higher trophic levels by grazing (Fuhrman 1999;
98 Wilhelm and Suttle 1999). This pathway of carbon flux regulated by viruses is called
99 “viral shunt” (Wilhelm and Suttle 1999) (Fig. 1.2.1). Calculations based on the
100 presumed microbial biomass, its turnover rates, and the predicted daily lysis suggest
101 that viral shunts are responsible for the release of approximately 25% of primary
102 production in the surface ocean, which amounts to up to 3 gigatons of carbon into the
103 oceans per year (Wilhelm and Suttle 1999; Suttle 2005). However, the quantification of
104 virus-mediated carbon flux in natural environments remains challenging owing to
105 methodological limitations and ecosystem complexity. Recently, more advanced
106 nutrient–phytoplankton–zooplankton (NPZ) models, including heterotrophic bacteria
107 and viruses, have proposed that viral shunts accelerate organic matter recycling and
108 increase net primary productivity while reducing transfer to higher trophic levels (Weitz
109 et al. 2015). Moreover, the effect of virus-mediated carbon flux may depend on the

110 trophic status of the system and the limiting nutrients (Pourtois et al. 2020).



111

112 **Fig 1.2.1. Overview of virus-mediated biogeochemical cycling in aquatic microbial**
113 **ecosystem.** When viruses lyse their host, intracellular organic matter is released from
114 host cells to the particulate organic matter (POM) and dissolved organic matter (DOM)
115 pools, a process that has been termed a Viral shunt. This process is accompanied by
116 compositional and stoichiometric changes in chemical properties of intracellular
117 metabolites via viral metabolic redirection, i.e., hijacking host transcription-translation
118 systems and expression of viral auxiliary metabolic genes (AMG). Viral particles are
119 also a source of phosphorus-rich DOM (compared with host debris) and consumed by
120 direct grazing. Viral infection facilitates carbon export to the deeper layer (biological
121 pump) through particle aggregation driven by the release of lysis products and

122 virus-induced alterations in host physiology (Viral shuttle).

123

124 On the other hand, viruses can also contribute to carbon removal from the
125 surface ocean (Weinbauer 2004; Sullivan et al. 2017; Laber et al. 2018). Cellular debris,
126 including cytoplasmic material and components of the cell wall, released via viral lysis
127 can easily aggregate and sink to the deeper layers leading to carbon sequestration
128 (Weinbauer 2004; Laber et al. 2018). This alternative viral influence on the geochemical
129 flux that promotes the biological carbon pump is referred to as the “viral shuttle”
130 (Sullivan et al. 2017) (Fig. 1.2.1). For example, virus-induced carbon transportation has
131 been extensively studied in *Emiliania huxleyi* (haptophyte) and its known virus (Laber
132 et al. 2018). Several laboratory studies have reported that viral infection stimulates the
133 production of transparent exopolymer particles (TEP), which increases the stickiness of
134 cells, thus promoting aggregation (Rosenwasser et al. 2014; Nissimov et al. 2018). Field
135 studies monitoring *E. huxleyi* blooms in the North Atlantic have revealed that TEP
136 concentrations increase during the early stages of infection and that infected cells are
137 preferentially transported to the deep ocean (Sheyn et al. 2018). Considering the huge
138 abundance of *E. huxleyi* (reaching 10^7 cells/mL during their bloom period; Silkin et al.
139 2020), the virus-mediated sinking of its cells would thus have a significant impact on
140 the available carbon in the surface ocean. Moreover, laboratory experiments have

141 reported that the virus-infected culture of *Chaetoceros tenuissimus* (Diatomea) is up to
142 59-fold enriched in particulate organic matter compared with uninfected controls
143 (Yamada et al. 2018). At the global scale, a metagenomic study based on data from the
144 Tara Ocean has suggested that the infection and lysis of the widespread and abundant
145 cyanobacteria *Synechococcus* significantly contribute to carbon export compared with
146 other microorganisms (Guidi et al. 2016). Further studies using quantitative methods are
147 required for a better understanding of the effect of viral shuttles in the global ocean.

148 In addition, viral particles themselves contribute to the biogeochemical cycling
149 of carbon and other nutrients. For example, approximately 0.03 Gt C per year (Bar-On
150 and Milo 2019), and most of the viral biomass is attributed to the dissolved organic
151 matter (DOM) fraction ($< 0.45 \mu\text{m}$) due to the size of the virion (e.g., bacterial viruses
152 generally range between 20 and 200 nm) (Zsolnay 2003; Leenheer and Croué 2003;
153 Findlay and Parr 2017). Recently, a relatively large number of marine viruses have been
154 identified to attach to non-host organisms and particles (Yamada et al. 2020). Thus, if or
155 when these non-host organisms or particles are predated, their attached viruses
156 indirectly contribute to classical marine food webs even when they are not infecting any
157 organisms. Altogether, viral particles could contribute to both DOM and the particulate
158 organic matter (POM) pools in the ocean.

159 A previous study investigating the elemental composition of both virus
160 particles and viral lysates eluted from their host debris revealed that viral lysates tend to
161 be more depleted in phosphorus than are uninfected cells because the amount of
162 genomic nucleic acids contained in progeny viral particles results in those viruses being
163 relatively phosphorus-rich as compared with their amounts of carbon and nitrogen
164 (Jover et al. 2014). By extrapolating this model to the ecosystem scale, marine viruses
165 are predicted to constitute a comparatively high proportion (> 5%) of the total DOP pool
166 in the surface ocean (Jover et al. 2014). Thus, viruses themselves can be regarded as an
167 abundant nutrient source.

168 Predation of viral particles by predators has been demonstrated in several
169 studies using culture experiments (Suttle and Chen 1992; Bettarel et al. 2005; Lawrence
170 et al. 2018; Welsh et al. 2020). For example, a co-cultivation study exposing
171 *Phaeocystis globosa* and its virus to various predators has demonstrated that viruses can
172 be effectively removed (up to 98% within 24 h) from the water column by non-host
173 organisms, including sea anemones, polychaete larvae, sea squirts, crabs, cockles,
174 oysters, and sponges (Welsh et al. 2020). Therefore, although the rate of viral particle
175 removal by aquatic protists may vary depending on both the virus and predator strains
176 (Suttle and Chen 1992; Gonzalez and Suttle 1993; Lawrence et al. 2018; Welsh et al.

177 2020), the ingestion of viruses can serve as a possible source of nutrients, especially
178 phosphorus.

179 These viral influences on the biogeochemical cycles fluctuate across short- and
180 long-time scales. On a long-term scale, for instance, the microbial community exhibits
181 seasonal compositional changes (Cram et al. 2015; Parada and Fuhrman 2017;
182 Needham et al. 2018; Choi et al. 2020), which are followed by the seasonal dynamics of
183 their viruses (Needham et al. 2017; Ignacio-Espinoza et al. 2020). Furthermore, diverse
184 taxa of photosynthetic microorganisms and even some heterotrophic ones show diel
185 activity in culture and environmental studies, which is partly explained by the viral
186 infection cycle (Morimoto et al. 2020 and references therein). Both the seasonality and
187 the diel cycle activity of microorganisms and their viruses suggest that host–virus
188 interactions could generate temporal fluctuations in geochemical cycles in aquatic
189 environments.

190

191 ***1.2.2 Virus–host interactions-mediated modification of host cell metabolism***

192 Viruses switch their host metabolism from cellular replication to progeny
193 production. Compared with non-infected hosts, metabolically reprogrammed cells can
194 be generally distinguished based on changes in the host transcription program,

195 eventually leading to distinctiveness in the proportion of end-point products between
196 infected and uninfected cells (Ankrah et al. 2014; Jover et al. 2014; Rosenwasser et al.
197 2014; Ma et al. 2018) (Fig. 1.2.1). For instance, disproportioning of phosphorus
198 between infected and uninfected hosts (as discussed in the previous section) could be
199 attributed to a viral reprogramming mechanism in which viruses degrade host DNA and
200 utilize the resultant nucleic acids for the synthesis of viral DNA (Wikner et al. 1993;
201 Kutter et al. 2018).

202 Currently, cell metabolic reprogramming by viruses has been studied using
203 both transcriptomic and metabolomic analyses and is found to be a highly regulated
204 process. For instance, the infection strategy of T4-like viruses follows the three
205 temporal expression classes of early, middle, and late genes, corresponding to host
206 takeover, replication, and virion morphogenesis, respectively, and occurs in accordance
207 with the downregulation of genes related to host replication (Roucourt and Lavigne
208 2009). Such transcriptional regulation by viruses has also been investigated in several
209 lineages of marine and freshwater prokaryotic as well as eukaryotic phytoplankton
210 (Lindell et al. 2007; Rosenwasser et al. 2014; Bachy et al. 2018; Moniruzzaman et al.
211 2018; Morimoto et al. 2018; Ku et al. 2020) and heterotrophic bacteria (Ankrah et al.
212 2014; Howard-Varona et al. 2017b, 2020). The metabolic regulation of host cells may

213 also depend on host taxonomy and thus differ between various species (e.g.,
214 cyanoviruses with broad and narrow host range seem to have different infection
215 strategies; *E. huxleyi* virus EhV possesses five gene expression phases during infection)
216 (Lindell et al. 2005; Clokie et al. 2006; Doron et al. 2016; Morimoto et al. 2018; Ku et
217 al. 2020). In addition, it may be influenced by host physiological states, which in turn
218 depends on nutrient availability (e.g. phosphate) (Kelly et al. 2013; Lin et al. 2016;
219 Bachy et al. 2018). Thus, viruses could affect the proportion of end-point products in the
220 infected cells while those hosts either directly or indirectly are responding to
221 environmental conditions.

222 Viruses can also possess host-derived genes (also called as auxiliary metabolic
223 genes, AMGs) that are expressed during infection, thus altering host metabolism, and
224 increasing the efficiency of viral reproduction (Breitbart et al. 2007; Hurwitz and U'Ren
225 2016). These AMGs can largely be classified into two classes (Class I and II) based on
226 their function according to the Kyoto Encyclopedia of Genes and Genomes database
227 (Hurwitz and U'Ren 2016). Viral-encoded AMGs not only maintain cellular functions
228 necessary for viral DNA replication and virion production (e.g., ATP production and
229 nucleotide synthesis) during infection (Lindell et al. 2004, 2005), but also both down
230 and up-regulate a range of targeted metabolic pathways that can substantially alter cell

231 stoichiometry and nutrient metabolism (De Smet et al. 2016, 2017). Most AMGs that
232 have been identified to date are directly involved in either the utilization and uptake of
233 limiting nutrients or energy production (Enav et al. 2014; Hurwitz and U'Ren 2016),
234 which in turn may have (at least temporarily) a positive feedback effect on the host cell
235 by improving its fitness during infection (Zeng and Chisholm 2012): the acquisition and
236 metabolism of carbon (e.g., *psbA* and *psbD*; Lindell et al. 2004, 2005; Thompson et al.
237 2011), nitrogen (e.g., *amt*; Monier et al. 2017), and phosphorus (e.g., *pstS* and *phoA*;
238 Zeng and Chisholm 2012). New putative AMGs are continuously being discovered in
239 bacterial (Breitbart 2011; Crummett et al. 2016; Breitbart et al. 2018; Warwick-Dugdale
240 et al. 2019), eukaryotic (Schvarcz and Steward 2018; Needham et al. 2019), and archaea
241 viruses (Ahlgren et al. 2019) or the more broadly defined environmental viromes
242 (Williamson et al. 2008; Anantharaman et al. 2014; Hurwitz et al. 2015; Moniruzzaman
243 et al. 2020; Schulz et al. 2020; Kieft et al. 2020). Thus, considering that viral-encoded
244 AMGs are abundant and widespread in aquatic environments (Williamson et al. 2008),
245 AMG-mediated metabolic reprogramming can substantially contribute to major
246 biogeochemical cycles and ecosystem functioning at the global scale (Sieradzki et al.
247 2019).

248 Indeed, changes in metabolites mediated by virus–host interactions have been

249 observed in both eukaryotes and prokaryotes using a metabolomic approach
250 (Rosenwasser et al. 2014; Ma et al. 2018). For example, *E. huxleyi* virus EhV
251 downregulates host *de novo* sphingolipid genes and simultaneously promotes the
252 induction of a viral-encoded homologous pathway, resulting in the metabolic shift
253 toward viral sphingolipid production (Rosenwasser et al. 2014). A recent culture-based
254 study on *Synechococcus* and its viruses revealed that the composition of chemical
255 compounds (proteins, carbohydrates, and lipids) of organic matter differs between
256 infected and uninfected cells (Ma et al. 2018). Such compositional changes induced by
257 viruses have been detected during viral infection of *Sulfitobacter* (C:N ratio of host cell
258 shifted to nitrogen-rich state compared with uninfected cells) (Ankrah et al. 2014).

259 Thus, from an ecological perspective, the virus-induced metabolic
260 reprogramming depends on both the virus-host pair (and therefore on the diversity and
261 composition of microbial assemblages) and the host physiological state during infection
262 and can modulate the generation as well as the diversity of end-point products, thus
263 leading to altered biogeochemical cycling. It is also speculated that halting the viral
264 reproduction at various infection stages using antiviral responses, including signal
265 transduction, cell cycle regulation (Moniruzzaman et al. 2018), and metabolic pathway
266 (Rosenwasser et al. 2014), might establish metabolite diversity in the infected cells via

267 the generation of intermediate products of viral progeny (Zborowsky and Lindell 2019)
268 or unusual proteins in the uninfected cells.

269

270 **1.3 Viral infection shaping microbial community diversity**

271 *1.3.1 Contribution of lytic infection to microbial diversity maintenance*

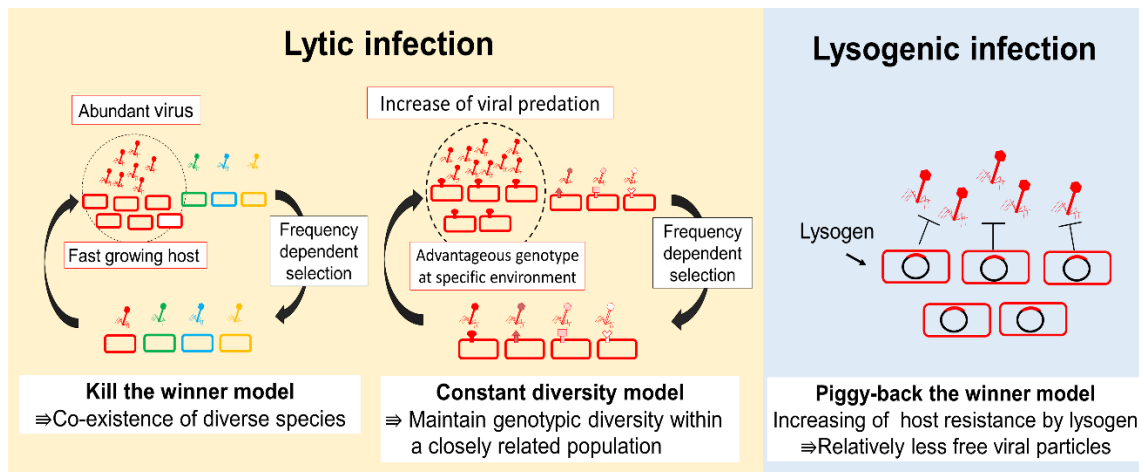
272 In aquatic ecosystems, diverse microorganisms compete for nutrient resources
273 but can co-exist and account for a large proportion of total aquatic biodiversity, a
274 concept that has been known as the classical question “paradox of the plankton”
275 (Hutchinson 1961). Viruses are currently thought to contribute to the co-existence of
276 microbial species and genotypes.

277 Basically, viruses are believed to infect their specific microbial hosts in a
278 frequency-dependent manner (Fuhrman and Suttle 1993). Therefore, viral infection
279 checkmates microbial species that become dominant through the competition among
280 co-existing microorganisms that possess different substrate affinity, and thereby enables
281 the co-existence of multiple competing microbial species (“Kill the Winner” hypothesis)
282 (Thingstad 2000) (Fig. 1.3.1). Indeed, several culture and environmental studies have
283 demonstrated that viral top-down control modulates microbial abundance, which is
284 consistent with the results expected from a Kill the Winner hypothesis (Tarutani et al.
285 2000; Schwalbach et al. 2004; Bouvier and Del Giorgio 2007; Yoshida et al. 2008a;

286 Rodriguez-Brito et al. 2010; Kuno et al. 2012; Parsons et al. 2012; Kimura et al. 2013;
287 Needham et al. 2013; Cram et al. 2016).

288 The viral-mediated co-existence mechanism, by which viruses are expected to
289 affect host diversity in a frequency dependent manner according to the Kill the Winner
290 hypothesis, could provide a mechanism that explains the continuing co-existence of
291 diverse genotypes within a single microbial species (or closely related lineage) rather
292 than the co-existence of diverse microbial species. Conventionally, the philosophy has
293 been that phenotypic and genotypic diversity within a microbial population was
294 expected to become homogenized to a greater level of fitness in the environment. The
295 microbial population (species or closely related lineage) that are genetically cohesive
296 and ecologically distinct are called an “ecotype” (ecotype hypothesis) (Maharjan et al.
297 2006; Cohan and Koeppel 2008). The ecotype had been believed to be periodically
298 replaced as fitter ecotypes emerged after profitable mutation or preferable
299 environmental changes (Maharjan et al. 2006; Cohan and Koeppel 2008). However,
300 metagenome sequence alignment analyses have demonstrated that several genomic
301 regions (metagenomic islands; MGIs) are underrepresented even within a single
302 microbial population in similar environments (Coleman and Chisholm 2007;
303 Cuadros-Orellana et al. 2007; Kettler et al. 2007; Wilhelm et al. 2007; Frias-Lopez et al.

304 2008; Rodriguez-Valera et al. 2009; Rodriguez-Valera and Ussery 2012). This suggests
305 that diverse genotypes coexist within a single microbial population. Furthermore, these
306 MGIs include diverse accessory genes, such as extracellular structure-related genes that
307 can be viral recognition sites (Reva and Tümmler 2008; Sharma et al. 2008;
308 Rodriguez-Valera et al. 2009; Rodriguez-Valera and Ussery 2012) and antiviral
309 response-related genes such as CRISPR in addition to genes that affect
310 restriction-modification (Sorek et al. 2008; Wilmes et al. 2009). Therefore, MGIs are
311 thought to play an important role in the ability of hosts to escape or survive from viral
312 infection, in which host genotypic diversity is driven by viral predation pressure,
313 thereby leading to the co-existence of multiple competing microbial genotypes
314 (“Constant Diversity dynamics” model) (Rodriguez-Valera et al. 2009) (Fig. 1.3.1).
315 Indeed, multiple genotypes of *Microcystis aeruginosa* possessing different CRISPR
316 arrays and its virus Ma-LMM01 coexist and oscillate during the massive bloom of this
317 nuisance and toxic species (Kuno et al. 2012, 2014; Kimura et al. 2013). Hence, lytic
318 viruses play important roles in not only the co-existence of microbial species but also
319 the maintenance of high diversity within a single microbial population.
320



321

322 **Fig 1.3.1. Schematic diagram of mechanisms in virus-mediated maintenance of**
 323 **microbial community diversity.** Preferential viral infection of abundant species
 324 enables the co-existence of diverse competing microbial species by preventing the
 325 dominance of only few species (“Kill the Winner” hypothesis). Similar viral top-down
 326 control is proposed as a mechanism to maintain high genotypic diversity within a single
 327 microbial population (“Constant Diversity dynamics” model). Lastly, the prevalence of
 328 lysogeny in dominant microbial population may be another potential mechanism that
 329 allows abundant host species to be dominant by taking advantages benefit from
 330 lysogenic conversion such as superinfection exclusion (“Piggyback-the-Winner”
 331 model).

332

333

334 ***1.3.2 Potential contribution of lysogenic infection to microbial diversity maintenance***

335 So far, we have focused on lytic viruses and described their contribution to the
336 maintenance of microbial diversity in aquatic environments. Another key question
337 related to viral impact on microbial diversity is whether temperate viruses also
338 contribute to shaping microbial community diversity.

339 Provirus integration can occur either through repeated random transposition
340 events or at specific integration sites (e.g., host tRNA genes; also called site-specific
341 recombination) and is associated with the immediate transcriptional suppression (e.g.
342 via specific virus repressors) of lytic promoters and genes associated with virion
343 production (Casjens and Hendrix 2015). This mode of viral infection that generates
344 lysogenic cells (Hobbs and Abedon 2016) is considered as an adaptive strategy of
345 temperate viruses to ensure their persistence in the environment, in which novel
346 phenotypic or metabolic advantages are sometimes conferred to host microorganisms
347 via concomitant effects by mechanisms such as HGT (Hendrix et al. 2000;
348 Howard-Varona et al. 2017a), the capability of up and down-regulation of host genes
349 (Argov et al. 2017), and integration-driven gene disruption (Feiner et al. 2015).

350 Although one study estimated that approximately half of 100 marine bacterial
351 isolates harbored temperate viruses in their genomes (Paul 2008), proviruses are rarely

352 found in the marine-dominant bacterial lineages (e.g., only one provirus was recently
353 reported in SAR11 clades; see Morris et al. 2020) possibly due to genome streamlining,
354 in which a bacterial genome is minimized to a highly constrained gene set that confers
355 maximum fitness (Touchon et al. 2016). Therefore, the ecological significance of
356 lysogenic infection on microbial diversity maintenance in marine ecosystems remains
357 under debate. Traditionally, it was believed that bacterial viruses control their host
358 abundances in a frequency-dependent manner as described above, and thus viral
359 abundance is typically 10-folds higher than that of prokaryotes (Wommack and Colwell
360 2000; Weinbauer 2004). Therefore, lysogenic infection is presumed to be the preferred
361 viral strategy under conditions of reduced host cell number and activity (Stewart and
362 Levin 1984; Sime-Ngando 2014; Brum et al. 2016). However, viral metagenomic and
363 metadata approaches have revealed that viral particles are relatively less abundant at
364 high microbial densities (Knowles et al. 2016; Wigington et al. 2016). Likewise, it was
365 demonstrated that the virus/host genome abundance ratio was negatively correlated with
366 the host abundance at the genus or phylum levels (Coutinho et al. 2017). Additionally,
367 the relative abundance of hallmark genes encoded by temperate viruses increased with
368 microbial density in a coral reef (Knowles et al. 2016). These findings suggest that
369 lysogenic infection may become dominant at high-cell densities because proviruses can

370 replicate quickly in a way that will keep pace with their fast-growing host and
371 provirus-mediated superinfection resistance might become increasingly important at
372 high cell densities (called “Piggyback-the-Winner” model) (Knowles et al. 2016, 2017;
373 Coutinho et al. 2017) (Fig. 1.3.1).

374

375 **Footnotes**

376 Paradox of the plankton: The concept arguing paradoxical situation of coexistence of
377 various plankton species competing for identical resources in homogeneous and
378 resource limited environment (Hutchinson 1961).

379 Kill the Winner hypothesis: A model proposing the dynamics of virus-host interactions
380 in which an increase of host population (winner) is accompanied with increasing of its
381 infectious viruses, and thereby viruses prevent their hosts from becoming dominant
382 through increased mortality of the winner (Thingstad 2000).

383 Constant Diversity dynamics model: A hypothetical model proposing that the diversity
384 of prokaryotic populations is maintained by viral predation, because the best-adapted
385 populations are selected by viral predation. The hypothesis assumes that each microbial
386 population has distinct viral receptor (Rodriguez-Valera et al. 2009).

387 Piggyback-the-Winner model: A hypothetical model proposing that lysogeny would be

388 favored when a bacterial host is dominated in the environment because a provirus can
389 replicate rapidly together with host DNA replication (Knowles et al. 2016).

390

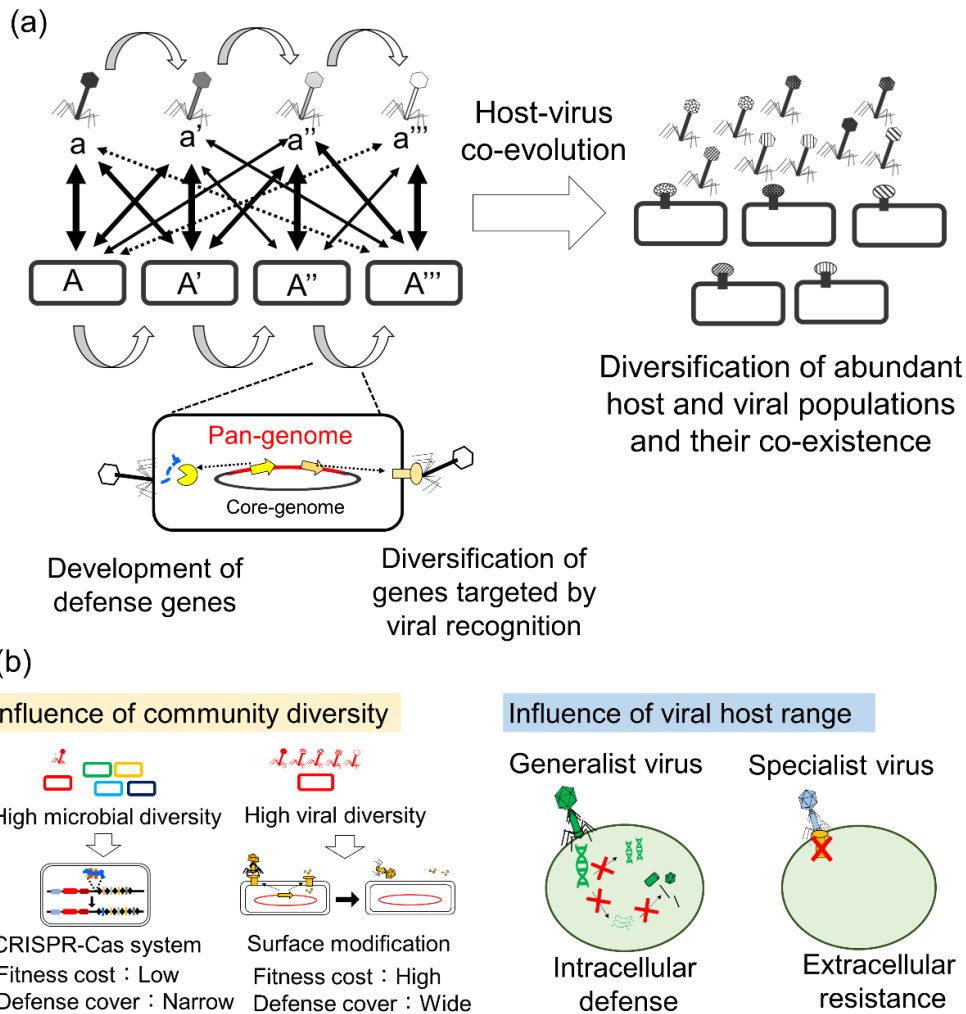
391 **1.4 Evolutionary roles of viruses that generate genotype-level microbial** 392 **diversification**

393 In short-term laboratory experiments, host–virus coevolution appeared to be
394 suppressed by the emergence of a viral-resistant genotype, which the virus could not
395 evolve to overcome (Dennehy 2012). In particular, *de novo* mutations that cause
396 changes in those bacterial cell-surface structures which serve as viral receptor sites are
397 one of the major factors that prevent viral attachment, and thereby confer the potential
398 host with resistance against viral infection (Lenski and Levin 1985). Therefore,
399 host–virus coevolution has been considered to be constrained by the asymmetry of
400 evolutionary potential between hosts and viruses (Cannon et al. 1971; Cowlshaw and
401 Mrsa 1975; Barnet et al. 1981; Lenski and Levin 1985; Waterbury and Valois 1993;
402 Middelboe et al. 2001; Wei et al. 2010, 2011).

403 On the other hand, long-term evolutionary laboratory studies have indicated
404 that the host and its virus undergo persistent coevolution over a prolonged period, as
405 evidenced with the soil bacterium *Pseudomonas fluorescens* (Buckling and Rainey
406 2002a, b; Brockhurst et al. 2007; Hall et al. 2011a, b) and *E. coli* (Mizoguchi et al.

407 2003). Similar co-evolutionary dynamics have also been observed in the marine bacteria
408 *Prochlorococcus* (Avrani et al. 2011), *Synechococcus* (Marston et al. 2012), and
409 *Cellulophaga baltica* (Middelboe et al. 2009).

410 The above-mentioned co-evolution scenario, based on the
411 one-virus-to-one-bacteria relationship, predicts that the genetic contents of both bacteria
412 and virus would converge over repeated interactions. As described above, however,
413 genotypic diversity is observed in MGIs even within a single microbial population
414 (Coleman and Chisholm 2007; Cuadros-Orellana et al. 2007; Kettler et al. 2007;
415 Wilhelm et al. 2007; Frias-Lopez et al. 2008; Rodriguez-Valera et al. 2009;
416 Rodriguez-Valera and Ussery 2012). Additionally, under-represented genomic regions
417 have been found in marine viruses (metaviromic islands) with a large fraction of their
418 identified genes (e.g., 59 out of 138) associated with host recognition of viruses
419 (Mizuno et al. 2014). Thus, the recent understanding of host–virus coevolution is based
420 on multiple genotype (strain)-level interactions within both the microbial host and viral
421 species which seem to represent defense and counter-defense strategies (e.g., CRISPR
422 and mutation in protospacer) or resistance and overcome of resistance (e.g., mutation in
423 cell surface and viral tail gene) (Fig. 1.4.1).



424

425 **Fig. 1.4.1 Proposed mechanisms in host pan-genome expansion via virus–host**

426 **interactions.** (a) Under the viral top-down control toward dominant species and

427 genotypes, “Red Queen” like host–virus co-diversification can be established in

428 abundant host and abundant viruses. Continuous arms race via intracellular defense (e.g.,

429 CRISPR-Cas system) and extracellular resistance (e.g., viral recognition sites) plays a

430 part in genomic diversification of both host and virus. (b) A complicated balance

431 between trade-off and specificities of each defense mechanism can affect the arms race.

432 For example, extracellular resistance (e.g., surface modification) is preferable to

433 intercellular defense (e.g., CRISPR-Cas) under conditions of high viral genetic diversity

434 because the former promotes resistance toward against broader viral genotypes.
435 However, because extracellular resistance may result in a higher cost, such as the
436 impairment of nutrient uptake ability, intercellular defense is possibly preferable under
437 competitive situations with host competitors.

438

439 In freshwater ecosystems, host-virus coevolution have been intensively studied
440 in the bloom-forming cyanobacterium *M. aeruginosa* and its viruses (Yoshida et al.
441 2005, 2008a, b; Kimura et al. 2012, 2013, 2018; Kuno et al. 2012, 2014;
442 Yoshida-Takashima et al. 2012; Morimoto et al. 2019). Interestingly, the most abundant
443 *Microcystis* CRISPR genotype is known to coexist with that derived by novel spacer
444 acquisitions from cyanoviruses in the environment. This finding suggested that both
445 abundant host and viral genotypes have diversified in the bloom without a complete
446 selective sweep (Kimura et al. 2018). Thus, the Red Queen like dynamics could be
447 established, to some extent, between the abundant host genotype and its cyanoviruses
448 under high viral contact rate; with reciprocal adaptation via defense and counter-defense
449 continuously occurring in multiple-to-multiple relationships (Koskella and Brockhurst
450 2014), thus subsequently increasing the diversity of host organisms and viruses (Fig.
451 1.4.1). Also, a recent metagenomic survey revealed the co-existence of highly
452 host-specific (narrow host range) and broad host range *Microcystis* viruses and the high

453 co-expression of antiviral defense and viral genes in the environment (Morimoto et al.
454 2019). Considering that they often induce antiviral responses, broad host range viruses
455 might be important for host genotype diversification (Morimoto et al. 2019).

456 In contrast with freshwater cyanobacteria typically having the greatest overall
457 numbers of defense genes (Makarova et al. 2011), marine prokaryotes rarely possess
458 distinctive defense genes such as CRISPR-Cas due to their genome streamlining
459 (Touchon et al. 2016). However, instances of co-existence between dominant marine
460 prokaryotes and their viruses were observed, which are presumably sustained by Red
461 Queen-like co-evolution dynamics. For example, the dominance of the SAR11 clade
462 bacteria and their viruses was predicted to be maintained by host rapid adaptation to
463 viruses. The rapid adaptation was achieved by high recombination rates among SAR11
464 in their variable genomic region that encoded genes involved in synthesis of cell surface
465 proteins, and this hypothesis is possibly supported by their high host cell density (King
466 of the Mountain hypothesis) (Zhao et al. 2013). Furthermore, constant turnover of
467 single-nucleotide polymorphism variants in relatively abundant marine viruses also
468 suggests that Red Queen-like virus–host coexistence could be established by perpetually
469 changing minor variants.

470 As described in this section, one of the major forces driving host and viral

471 genotypic diversification is the reciprocal defense and counter-defense that occur
472 through extracellular (e.g., *de novo* mutation of cell-surface structure) and intracellular
473 resistance (e.g., acquisition of novel CRISPR spacers). Comparative antiviral resistance
474 analyses in *Synechococcus* and *Prochlorococcus* provided new insights into
475 host-favored antiviral defenses in narrow and broad host range viruses (Doron et al.
476 2016; Zborowsky and Lindell 2019). Host cyanobacteria resist against narrow host
477 range viruses irrespective of the viral family by preventing viral entry into the cell,
478 whereas intracellular resistance arrests the infection cycle of broad host range viruses at
479 various infection stages (Doron et al. 2016; Zborowsky and Lindell 2019) (Fig. 1.4.1).
480 These differences in antiviral responses that seemingly occur according to viral host
481 range are speculated to be associated with fitness trade-offs in extracellular and
482 intracellular antiviral responses. In extracellular antiviral responses, mutations in
483 cellular surface structures can impair nutrient uptake and utilization but can protect
484 against diverse viral attacks, leading to an increase in host fitness trade-off favoring the
485 host (Winter et al. 2010; Avrani et al. 2012) (Fig. 1.4.1). On the other hand, choosing to
486 modify intracellular antiviral responses techniques may be energetically favorable,
487 especially the CRISPR-Cas system, because the cost of a new spacer acquisition is
488 speculated to be low, although it is possible that additional types of resistance costs may

489 exist (Thingstad et al. 2014) (Fig. 1.4.1). Indeed, recent studies focusing on the
490 differences in biotic complexity between *in vitro* and environments have revealed that
491 coexistence among human pathogens amplified the fitness trade-offs associated with
492 viral receptor mutations in *Pseudomonas aeruginosa* and therefore enhanced the
493 evolution of CRISPR-based resistance (Alseth et al. 2019). Higher viral genetic
494 diversity can also influence CRISPR-based evolution, with an example being that the
495 majority of a *Pseudomonas* population more favorably evolved based on the mutation
496 of viral receptors to resist a broader range of viral genotypes than CRISPR-based
497 specific resistance (Broniewski et al. 2020). Thus, fitness trade-offs in bacterial host
498 species and diversity of both bacterial host and viruses in the environment could be
499 another important factor that affects host–virus coevolution. From the perspective of
500 fitness trade-off, most recently, the “pan-immune system” concept has been proposed.
501 This states that a single strain can access immune defense mechanisms in closely related
502 strain via HGT, although it cannot possess all possible defense systems (Bernheim and
503 Sorek 2020).

504

505 **Footnotes**

506 Red Queen like dynamics: A hypothesis proposing co-evolutionary process between

507 competing species (Valen 1973); in the case of virus-host interactions, this hypothesis
508 explains continuous dynamics of resistance acquisition in microbial hosts and viral
509 avoidance to the host resistance (Brockhurst et al. 2014).

510 King of the Mountain hypothesis: A hypothesis proposing that high recombination rate
511 enables dominance of a competitive prokaryote in the ecosystem via horizontal transfer
512 of genes involved in resistance to viral infection (Zhao et al. 2013).

513

514 **1.5 Conclusion**

515 Viruses, which are highly abundant biological entities lacking their own
516 metabolism, can reprogram host cells toward the production of virus progeny, after
517 which the host cell is lysed, releasing new virions into the surrounding environment.

518 This reprogramming viral strategy can change the content and composition of host
519 metabolites and releases a large amount of organic matter, thus considerably affecting

520 biogeochemical cycles. In addition, viral infection checkmates not only microbial
521 species that become dominant but also abundant genotypes within a single microbial

522 species, and thereby enables the coexistence of diverse microbial species and genotypes
523 in the aquatic ecosystem. Thus, viral infection is one of the key factors shaping

524 microbial community diversity and maintaining high diversity within a single microbial
525 population. Likewise, because temperate viruses may be prevalent under specific

526 combinations of environment and conditions, they could affect microbial diversity via
527 superinfection exclusion during either the lysogenic cycle or induction of lytic cycle.
528 Meanwhile, microorganisms have evolved extracellular and intracellular antiviral
529 mechanisms with different fitness costs and specificities. Therefore, viruses with diverse
530 host range and genetic diversity interact with abundant hosts by a complex balancing
531 between fitness trade-offs and the specificity of extracellular- and intracellular antiviral
532 resistance in hosts. Together, this results in continuous virus–host coevolution, leading
533 to diversification in aquatic ecosystems. Collectively, viruses are important biological
534 entities that sustain and generate microbial diversity and control the biogeochemical
535 cycle in aquatic environments.

536

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