



The optimal strategy of incompatible insect technique (IIT) using *Wolbachia* and the application to malaria control

Taiga Matsufuji^a, Sungrim Seirin-Lee^{b,c,d,*}

^a Graduate School of Integrated Sciences for Life, Hiroshima University, Higashi-hiroshima 739-8530, Japan

^b Institute for the Advanced Study of Human Biology (ASHBi), Kyoto University Institute for Advanced Study, Kyoto University, Kyoto 606-8315, Japan

^c Department of Mathematical Medicine, Graduate School of Medicine, Kyoto University, Kyoto 606-8315, Japan

^d JST CREST, 4-1-8 Honcho, Kawaguchi, Saitama 332-0012, Japan

ARTICLE INFO

Keywords:

Wolbachia

Incompatible insect method

Malaria

Epidemic modeling

ABSTRACT

For decades, techniques to control vector population with low environmental impact have been widely explored in both field and theoretical studies. The incompatible insect technique (IIT) using *Wolbachia*, based on cytoplasmic incompatibility, is a technique that *Wolbachia*-infected male mosquitoes are incapable of producing viable offspring after mating with wild-type female mosquitoes. While the IIT method experimentally ensured its effectiveness in several field works, the failure of female mosquito population control by replacement owing to the accidental contamination of *Wolbachia*-infected female mosquitoes has been a concern and an obstacle in implementing the IIT method in nature. In this study, we develop a population-based IIT mathematical model using cytoplasmic incompatibility and evaluate the effectiveness of the IIT method in scenarios where contamination is present or absent. In addition, by extending the model to assess the disease infection status of the human population with malaria, we evaluate the optimal release strategy and cost for successful disease control. Our study proves that IIT could be a promising method to control mosquito-borne diseases without perfect suppression of vector mosquito population regardless of contamination.

1. Introduction

Vector-borne infectious diseases account for more than 17% of all infectious diseases and more than 700,000 people die each year from vector-borne diseases (WHO, 2020). The distribution of vector-borne infections is determined by a complex interaction of environmental and biological factors; therefore, the control of the vector population is critical for controlling the disease pandemic. For example, Malaria is a vector-borne disease transmitted by approximately 40 species of the *Anopheles* mosquito genus and is particularly devastating in the tropical and subtropical regions of the world (Phillips et al., 2017; WHO, 2019).

For decades, techniques for controlling the mosquito population with low environmental impact have been widely explored in both field and theoretical studies (Knippling, 1955; Thomas et al., 2000; Phuc et al., 2007; Vreysen et al., 2007; Alphey et al., 2010; Lacroix et al., 2012; Seirin-Lee et al., 2013a,b; Carvalho et al., 2015; Natiello and Solari, 2020). In particular, the strategy of releasing large numbers of male mosquitoes, which can reduce the mosquito population has been extensively applied. One classical control method is the sterile

insect technique (SIT), in which males that have been sterilized using radiation do not make wild females produce eggs even after mating (Knippling, 1955). The release of large numbers of sterile males results in a reduction of their reproductive output and potentially mosquito population abundance. As an improved method of SIT, transgenic technology, such as the release of insects carrying a dominant lethal gene (RIDL) (Thomas et al., 2000), has also been well explored for reducing mosquito populations (Lacroix et al., 2012; Carvalho et al., 2015). In RIDL, the released transgenic males are homozygous for a dominant lethal gene expressed in both male and female progeny that results from mating with wild-type insects. Genetically modified male mosquitoes are released and eggs laid by mating with wild females die in the larval stage, which consequently leads to the reduction of mosquito population. The control effectiveness for both classical SIT and RIDL has been verified by field experiments (Evans et al., 2019; Bouyer et al., 2020) and has been well studied in mathematical models so that possible control strategies have been examined and suggested under various situations (Phuc et al., 2007; Seirin-Lee et al., 2013a,b; Natiello and Solari, 2020).

* Corresponding author at: Institute for the Advanced Study of Human Biology (ASHBi), Kyoto University Institute for Advanced Study, Kyoto University, Kyoto 606-8315, Japan.

E-mail address: lee.seirin.2c@kyoto-u.ac.jp (S. Seirin-Lee).

<https://doi.org/10.1016/j.jtbi.2023.111519>

Received 26 September 2022; Received in revised form 12 March 2023; Accepted 27 April 2023

Available online 30 April 2023

0022-5193/© 2023 The Authors. Published by Elsevier Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

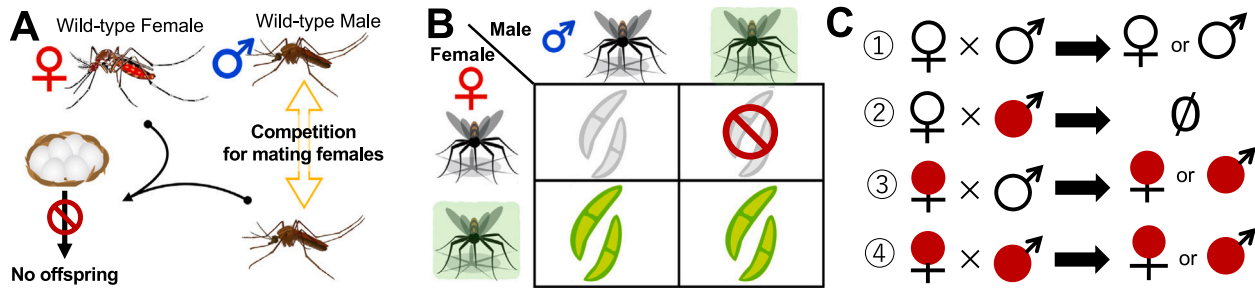


Fig. 1. IIT images and combinations. (A) Schematic representation of IIT based on cytoplasmic incompatibility. The offspring do not hatch when a male mosquito infected with *Wolbachia* mates with a female mosquito of the wild type. The wild-type and *Wolbachia*-infected male mosquitoes compete with each other in mating with female mosquitoes. (B) Results of mating between *Wolbachia*-infected (green shading) and uninfected (no shading) individuals. Infected females (bottom) always produced infected viable offspring, whereas uninfected females (top) produced uninfected viable offspring only when mating with uninfected males. In addition, mating between an uninfected female and an infected male result in non-viable offspring. (C) Four mating patterns are shown in (B). *Wolbachia*-infected/uninfected is marked by red/white color.

In recent years, the incompatible insect technique (IIT) has attracted considerable attention as an alternative to the classical SIT method (Hughes et al., 2014; Ritchie et al., 2018). This approach relies on *Wolbachia*-infected male mosquitoes that are incapable of producing viable offspring after mating with wild-type females (Fig. 1A) (Laven, 1967). *Wolbachia* is an intracellular symbiotic bacterium that infects a wide range of arthropods, including mosquitoes, and is thought to infect approximately 40% of the arthropod species in nature. Because it multiplies within the host cell, it cannot penetrate the sperm and is often localized in the ovaries of females; therefore, it is essentially transmitted from females to offspring via the egg (Werren et al., 2008). It causes a reproductive process in host insects termed cytoplasmic incompatibility by which breeding with an infected male interferes with the reproduction of an uninfected female by causing abnormal cell division in the early embryonic stage of the egg, resulting in the failure to hatch (Werren et al., 2008).

While cytoplasmic incompatibility between infected males and uninfected females causes the failure to hatch when infected females breed with uninfected males or infected females breed with infected males, the eggs hatch normally (Fig. 1B, C). Thus, *Wolbachia*-infected females are sure to leave offspring for the next generation (Yen and Barr, 1971). That is, if the strain of *Wolbachia* that already exists in the colony is used for the IIT method, when bred with wild females carrying the same strain, the eggs laid will hatch. In addition, if infected males with a new strain of *Wolbachia* are released with the contamination of infected females, the colony will spread a new strain of *Wolbachia* and replacement of the female mosquito population may occur (Yen and Failloux, 2020; Soh et al., 2022). Thus, for successful control using the IIT method, using the same strain of *Wolbachia* or the contamination of *Wolbachia*-infected females should be avoided.

Recently, the IIT method based on cytoplasmic incompatibility was experimentally investigated in Guangzhou, China (Zheng et al., 2019), California, USA (Crawford et al., 2020), and North Queensland, Australia (Beebe et al., 2021). In the former two field studies, a hybrid method of IIT and SIT was tested to compensate for the contamination problem of female mosquitoes. In these studies, both experiments showed a high suppression rate beyond 80% in the population of wild-type female mosquitoes. The latter fieldwork examined the IIT using bidirectional incompatibility and confirmed its high efficacy (above 80%) for mosquito suppression. Although these field studies proved the effectiveness of the IIT based on the hybrid method and the effectiveness of the IIT for short periods and in localized areas, the results suggest that the IIT could be a practical method for controlling mosquito populations. Thus, exploring the global effectiveness of the IIT method over a long time should be an important next step to consider, as sustained release strategies of large numbers over a

wide region are costly and careful consideration of alternative control strategies is required before implementation.

Conversely, strategies for controlling vector-borne diseases typically focus on the extinction of vector insects. However, the IIT method that uses *Wolbachia*, may not realistically address that objective because the contaminated *Wolbachia*-infected females produce offspring that continue spreading the disease. Differently expressed, we must reconsider the effectiveness of the IIT method for suppressing the mosquito population and consider controlling the disease by other means, such as reducing the number of infected people. Thus, the key question is whether disease transmission can be prevented even with only partial suppression using the IIT method, and if so, to what extent female mosquitoes must be suppressed to successfully prevent disease transmission.

To compensate for the field experiments' limitations and answer the key question concerning the effectiveness of the IIT method in disease control scenarios, we developed a mathematical model of a mosquito population subject to the *Wolbachia* IIT method to evaluate the effectiveness quantitatively by applying mathematically defined measures. Using *in silico* experiments with the mathematical model, we considered two scenarios, one in which the female mosquitoes were contaminated with *Wolbachia* and one in which they were not. In each scenario, we propose an optimal release strategy for the effective suppression of female mosquito populations by introducing a release cost. Furthermore, we extend the model to assess the disease infection status of human populations rather than vector populations, especially for malaria (Shaw et al., 2016). With the extended model, we primarily investigate the optimal release rate of the IIT strategy to reduce malaria infection rather than target the extinction of female mosquito populations. Our study suggests that IIT could be a promising method to control mosquito-borne diseases without perfect suppression of mosquito populations even in the contamination scenario.

2. Materials and methods

2.1. *Wolbachia* incompatible insect technique model (*Wolbachia* IIT model)

The life cycle of a mosquito is approximately 40 days. It takes approximately 15 days for the mosquito to reach adulthood via egg and larval stages (Carvajal-Lago et al., 2021). Thus, in this study, we assume that mosquito population growth proceeds via a stage-structured process and that density-dependent mortality acts on a pre-adult developmental stage. We choose the simple stage-structure model suggested by Dye (1984), in which the dynamics of wild-type

mosquitoes are given to

$$\begin{aligned} \frac{dF(t)}{dt} &= qrF(t-\tau)M(t-\tau)\phi(t) - \mu F(t), \\ \frac{dM(t)}{dt} &= (1-q)rF(t-\tau)M(t-\tau)\phi(t) - \mu M(t), \end{aligned} \tag{1}$$

where t is the time (day) and $F(t)$ and $M(t)$ are the population densities of wild-type female and male mosquitoes, respectively, r is the survival rate from egg to adult, q is the sex ratio of female mosquitoes, τ is the growth time from egg to adult mosquitoes, and μ is the death rate. Function ϕ defines the density-dependent effect in the larval stage and is given as follows:

$$\phi(t) = \exp[-\alpha \{E_0 F(t-\tau)M(t-\tau)\}^\beta],$$

where α is a density-dependent coefficient, E_0 is the egg production rate of adult mosquitoes without correction for density-independent survival between egg stage and adulthood, and β is a parameter derived from fitting empirical data, as detailed in Dye (1984).

We extend the model (1) with *Wolbachia*-infected mosquitoes and develop an IIT model based on cytoplasmic incompatibility. The offspring of a wild female mosquito and a *Wolbachia*-infected male mosquito cannot hatch normally (Fig. 1A). However, females infected by *Wolbachia* can lead to offspring regardless of the infectiveness of a male mosquito by *Wolbachia*, so that *Wolbachia* is inherited by the offspring (Fig. 1B and C). We denote the population densities of *Wolbachia*-infected female and male mosquitoes by $F_w(t)$ and $M_w(t)$, respectively. Subsequently, the IIT model, based on Fig. 1 is constructed as follows.

$$\begin{aligned} \frac{dF_w(t)}{dt} &= q \left(r_1 \frac{F_w(t-\tau)M(t-\tau)}{1+cM_w(t-\tau)} + r_2 \frac{F_w(t-\tau)M_w(t-\tau)}{1+cM(t-\tau)} \right) \phi_1(t) \\ &\quad - \mu F_w(t) + p\kappa, \end{aligned} \tag{2}$$

$$\begin{aligned} \frac{dM_w(t)}{dt} &= (1-q) \left(r_1 \frac{F_w(t-\tau)M(t-\tau)}{1+cM_w(t-\tau)} + r_2 \frac{F_w(t-\tau)M_w(t-\tau)}{1+cM(t-\tau)} \right) \phi_1(t) \\ &\quad - \mu M_w(t) + (1-p)\kappa, \end{aligned} \tag{3}$$

$$\frac{dF(t)}{dt} = qr_3 \frac{F(t-\tau)M(t-\tau)}{1+cM_w(t-\tau)} \phi_2(t) - \mu F(t), \tag{4}$$

$$\frac{dM(t)}{dt} = (1-q)r_3 \frac{F(t-\tau)M(t-\tau)}{1+cM_w(t-\tau)} \phi_2(t) - \mu M(t). \tag{5}$$

The first terms of Eqs. (2) and (3) are growth terms for mating mosquitoes and were modeled from (3) and (4) in Fig. 1C, in which we assumed that mating by infected/wild-type male mosquitoes decreases in proportion to the relative abundance of wild-type/infected male mosquitoes due to mating competition (Fig. 1A) (Knipling, 1955; Phuc et al., 2007). c ($0 < c \leq 1$) represents the reduced mating competitive ability of *Wolbachia*-infected male or wild-type male mosquitoes. r_1, r_2 , and r_3 are the survival rates from egg to adult. As wild-type offsprings are reproduced only by mating between wild-type mosquitoes (1 in Fig. 1C), the models for wild-type female and male mosquitoes, Eqs. (4) and (5), are given in the basic model of Eq. (1) with adding the term mating competitive ability with *Wolbachia*-infected males. The density-dependent functions, $\phi_1(t)$ and $\phi_2(t)$, are given to

$$\phi_1(t) = \exp \left[-\alpha \left\{ E_1 \frac{F_w(t-\tau)M(t-\tau)}{1+cM_w(t-\tau)} + E_2 \frac{F_w(t-\tau)M_w(t-\tau)}{1+cM(t-\tau)} \right\}^\beta \right], \tag{6}$$

$$\phi_2(t) = \exp \left[-\alpha \left\{ E_3 \frac{F(t-\tau)M(t-\tau)}{1+cM_w(t-\tau)} \right\}^\beta \right], \tag{7}$$

where E_1, E_2 , and E_3 are egg production rates for each mating combination. α and β are positive constants that determine the density-dependent effects.

The second terms of Eqs. (2)–(5) represent the natural mortality rate of mosquitoes. The terminal terms in Eq. (2) and (3) are the release policies for the *Wolbachia*-infected mosquitoes and we assume

a constant daily release of a fixed amount of mosquitoes and define it by a relative equilibrium density of wild-type female mosquitoes,

$$\kappa \equiv \omega F^*,$$

where ω is the release rate and F^* is the initial equilibrium density of the wild-type female mosquito. p ($0 \leq p < 1$) is the contamination ratio of the *Wolbachia*-infected female mosquitoes in the release policy. Note that the effect of reducing the number of offsprings using the IIT method (2 in Fig. 1C) is not explicitly included in our IIT model (2)–(5) because there is no hatching of eggs.

2.2. Application of the wolbachia IIT model to malaria (Malaria-IIT model)

We will examine the IIT strategy for malaria and consider the effectiveness of the IIT method in reducing infected cases. For this purpose, we combine a malaria transmission model comprising infected humans and mosquitoes.

A model to understand the dynamics of malaria transmission was first suggested by Ross (1911) and Lotka (1923). Subsequently, biological realism was given and plausible parameter values were estimated by Macdonald (1957). Ruan et al. (2008) extended the Ross–Macdonald model with combining the effect of the incubation period of malaria and updated the feasibility of model. Thus, we chose the Ross–Macdonald malaria model including the incubation periods of malaria.

By defining the malaria-infected human population as $H(t)$ and the malaria-infected female mosquito population as $V(t)$, the delayed Ross–Macdonald model with incubation periods is given by

$$\frac{dH(t)}{dt} = b_1 \frac{a}{H_{tot}} \{H_{tot} - H(t-\tau_2)\} V(t-\tau_2) - \gamma H(t),$$

$$\frac{dV(t)}{dt} = b_2 \frac{a}{H_{tot}} \{V_{tot} - V(t-\tau_3)\} H(t-\tau_3) - \mu V(t),$$

where H_{tot} is the total population density of humans, V_{tot} is the total population density of vector mosquitoes, a/H_{tot} is the average mosquito biting rate per individual, b_1 is the transmission rate of malaria from infected female mosquitoes to uninfected human individuals through blood sucking, b_2 is the transmission rate of malaria from infected individuals to uninfected female mosquitoes, γ is the recovery/mortality rate of infected patients, and μ is the mortality rate of malaria-infected female mosquitoes. τ_2 and τ_3 are the incubation periods for malaria in humans and mosquitoes, respectively.

Now, we incorporate the IIT strategy into the delayed Ross–Macdonald Malaria model by replacing the total population of vector mosquito V_{tot} with

$$V_{tot}(t) \equiv F_w(t) + F(t).$$

Note that only female mosquitoes bite humans; therefore, the male population is not included. Finally, we obtain the Malaria-IIT model as the following.

$$\begin{aligned} \frac{dH(t)}{dt} &= b_1 \frac{a}{H_{tot}} \{H_{tot} - H(t-\tau_2)\} V(t-\tau_2) - \gamma H(t), \\ \frac{dV(t)}{dt} &= b_2 \frac{a}{H_{tot}} \{F_w(t-\tau_3) + F(t-\tau_3) - V(t-\tau_3)\} H(t-\tau_3) - \mu V(t). \end{aligned} \tag{8}$$

2.3. Parameters and initial conditions

We first assume that the sex ratio of mosquitoes is 1:1 (namely, $q = 0.5$) because cytoplasmic incompatibility does not critically change the sex ratio. The delay parameters, developmental period of mosquitoes (τ) and incubation periods for humans (τ_2) and mosquitoes (τ_3), were chosen as values fitted from empirical data (Dye, 1984; Ohm et al., 2018; Shaw et al., 2020). We also confirmed that the impact of delay scales on the effectiveness of IIT method is negligible (See Appendix B in more detail). For other parameter values, we chose those from previous studies that were estimated through experiments or mathematical

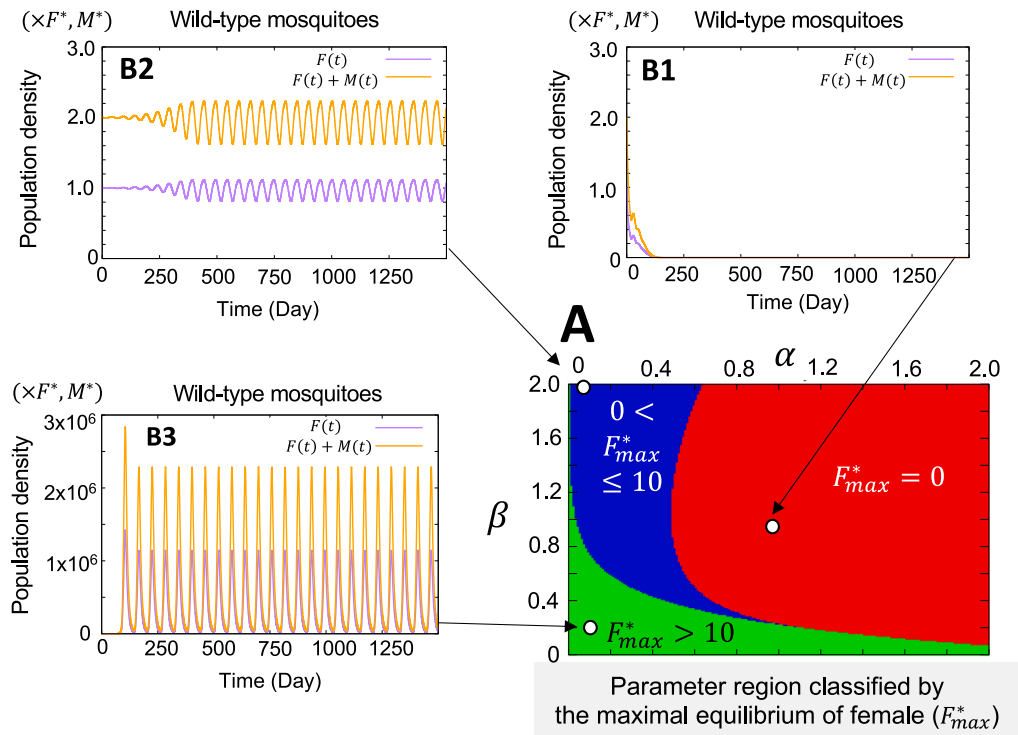


Fig. 2. Parameter region and dynamics of the wild-type mosquitoes depending on (α, β) . (A) The parameter region of (α, β) is classified according to the magnitude of the maximal equilibrium density F_{max}^* . The red region corresponds to the case in which $F^* = F_{max}^* = 0$ and $(F^*, M^*) = (0, 0)$ becomes a unique equilibrium. The blue and green regions correspond to the three equilibrium cases and satisfy a bistable state. Namely, $(F^*, M^*) = (0, 0)$, and (F_{max}^*, M_{max}^*) become stable equilibria. (B1–B3) The examples of the model (1) with $\tau = 18.84$ days. (B1) shows the case of $(\alpha, \beta) = (1.0, 1.0)$, (B2) shows the case of $(\alpha, \beta) = (0.14, 2.0)$, and (B3) is the case of $(\alpha, \beta) = (0.1, 0.2)$.

verification. The detailed parameter set and references are shown in Table 1.

However, the parameters determining the effect of density dependence (α and β) in Eqs. (6) and (7) are unknown, although they are likely to influence the model dynamics (Phuc et al., 2007). Thus, to determine the feasible values of α and β , we directly tested the wild-type mosquito model (1) over a wide range of α and β values. We found that model dynamics can be classified according to the number of wild-type mosquito equilibria. We first note that there exists either the case of a unique equilibrium $(0, 0)$ (red region of Fig. 2A), or the case of three equilibria, including $(0, 0)$ (blue and green regions in Fig. 2A). Furthermore, the equilibrium $(0, 0)$ is always locally stable, but the three equilibrium cases give rise to bi-stable states when we assume $\tau = 0$ in Eq. (1) (see Appendix A for the detailed analysis). In addition, we found that the maximal equilibrium density of female mosquitoes (F_{max}^*) varies greatly depending on α and β in case of bistability. As the delayed differential equation likely causes oscillating dynamics, we presume that unrealistic oscillating dynamics may occur as F_{max}^* increases. Indeed, we often found oscillating dynamics in the bi-stable equilibria region of α and β (blue and green regions in Fig. 2A).

Fig. 2 shows a representative result in which we investigated the parameter regions by $F_{max}^* = 0$, $0 < F_{max}^* \leq 10$, and $F_{max}^* > 10$. In the red region, the zero-equilibrium existed uniquely and mosquito populations were extinct when development period of mosquito ($\tau = 18.84$ days, (Dye, 1984)) was chosen in the model (1) (Fig. 2B1). Thus, we excluded this parameter region. On the other hand, for the green and blue regions corresponding to the bistable region of Eq. (1) with $\tau = 0$, the system showed oscillating dynamics when $\tau = 18.84$ (Fig. 2B2, B3). The green region showed oscillating dynamics with a large amplitude of a dozen days period (Fig. 2B3), which is not a plausible dynamic for reflecting wild-type mosquitoes. In contrast, we

found reliable parameter sets when F_{max}^* was not too large, where the mosquito populations oscillated with a low amplitude (Fig. 2B2). Thus, we chose α and β in a blue region of Fig. 2A.

Next, to define the initial conditions, we assumed that the *Wolbachia*-infected mosquitoes were non-existent in nature before release and the wild-type mosquitoes were at a stable non-zero equilibrium density, (F^*, M^*) . The details are given to

$$\begin{aligned} F_w(0) &= M_w(0) = 0.0 && \text{for } t \in [-\tau, 0], \\ F(0) &= F^*(1 + \epsilon s_1(t)) && \text{for } t \in [-\tau, 0], \\ M(0) &= M^*(1 + \epsilon s_2(t)) && \text{for } t \in [-\tau, 0], \end{aligned}$$

where $s_1(t)$ and $s_2(t)$ are chosen randomly in $[0, 1]$, and $\epsilon = 0.01$. Note that when the contamination ratio is zero ($p = 0$), the *Wolbachia*-infected female becomes $F_w(t) = 0$ for all $t \geq 0$ values.

Similarly, we assumed that the malaria-infected female mosquitoes and infected humans were in a state of co-existent equilibrium in the initial situation and then the IIT method was implemented. Thus, in the Malaria-IIT model, we adopted the following initial conditions:

$$\begin{aligned} H(0) &= H^*(1 + \epsilon s_3(t)) && \text{for } t \in [-\tau_2, 0], \\ V(0) &= V^*(1 + \epsilon s_4(t)) && \text{for } t \in [-\tau_3, 0], \end{aligned}$$

where $s_3(t)$ and $s_4(t)$ are chosen randomly in $[0, 1]$, and $\epsilon = 0.01$. H^* and V^* are asymptotic densities obtained from Eqs. (1) and (8) over a sufficiently long time.

We solved the model equations by using C to program code based on the Runge–Kutta method including time delay. The data were visualized using Gnuplot Version 5.4. The accuracy of the numerical code was confirmed by regenerating the same results with published data and checking the data consistency by changing the time grid size.

Table 1
Representative parameter set.

Parameters	Definition	Value
IIT model		
F^*	Equilibrium density of wild-type female mosquito	1.7179
M^*	Equilibrium density of wild-type male mosquito	1.7179
${}^a\tau$	Mosquito development period from egg to adult	18.84 [day]
${}^a\mu$	Mortality rate of adult mosquito	0.12 [day ⁻¹]
p	Contamination ratio of the <i>Wolbachia</i> -infected female mosquito in release	[0, 6](%)
bq	Sex ratio of mosquitoes	0.5
ω	Release rate of <i>Wolbachia</i> -infected male mosquitoes	[0, 2] [day ⁻¹]
${}^a r_1$	Survival rate from egg to adult by mating F_w and M	0.333 [day ⁻¹]
${}^a r_2$	Survival rate from egg to adult by mating F_w and M_w	0.326 [day ⁻¹]
${}^a r_3$	Survival rate from egg to adult by mating F and M	0.367 [day ⁻¹]
${}^c E_1$	Egg hatching rate by mating F_w and M	0.81
${}^c E_2$	Egg hatching rate by mating F_w and M_w	0.79
${}^c E_3$	Egg hatching rate by mating F and M	0.89
${}^c c$	Coefficient of mating competition	1.0
α	A coefficient determining density-dependent effect	[0, 2.0]
β	A coefficient determining density-dependent effect	[0, 2.0]
Malaria-IIT model		
H^*	Asymptotic density of malaria infected humans in endemic state before IIT strategy	0.2887
V^*	Asymptotic density of Malaria-infected mosquito in endemic state before IIT strategy	0.4556
${}^d\tau_2$	Incubation period of Malaria in human	13.0 [day]
${}^d\tau_3$	Incubation period of Malaria in mosquito	10.0 [day]
${}^e H_{tot}$	Total population density of human	0.5
${}^f a$	Average biting rate of female mosquito	0.15 [day ⁻¹]
${}^f b_1$	Transmission rate of Malaria from Malaria-infected mosquito to human	0.5
${}^f b_2$	Transmission rate of Malaria from Malaria-infected human to female mosquito	0.5
${}^f \gamma$	Mortality/Recovery rate of Malaria-infected human	0.05[day ⁻¹]

^aA parameter derived from fitting empirical data in Dye (1984).
^bA parameter chosen in a range empirically observed in Shaw et al. (2016).
^cA parameter chosen in a range empirically observed in Zheng et al. (2019).
^dA parameter chosen in a range empirically observed in Ohm et al. (2018) and Shaw et al. (2020).
^eA parameter mathematically assumed from the transmission rate of Malaria in Ruan et al. (2008).
^fA parameter chosen by Ruan et al. (2008) in a parameter range empirically and numerically estimated in Gu et al. (2003). The values of parameters whose notation is omitted are given by the numerical observations (See Section 2.3).

3. Results

3.1. The effect of the *Wolbachia* IIT in the scenario without the contamination of the *Wolbachia*-infected female mosquitoes

Here, we investigated whether the *Wolbachia* IIT method is effective in eradicating the population of female mosquitoes assuming that contamination of the *Wolbachia*-infected female is absent in the release of the *Wolbachia*-infected male. Note that $F_w(t) \equiv 0$, because $p = 0$ in this scenario (Fig. 3A; red lines).

We found that the IIT method was very effective when the release amount was sufficient (Fig. 3A and B). When the release rate (ω) was chosen to be less than 0.08, the wild-type female mosquitoes increased and the IIT method was ineffective (Fig. 3A, left panel). This was because the increase in male mosquitoes increased mating chances. Nonetheless, if the release rate was chosen to be a value larger

than or equal to 0.08, the population of wild-type female mosquitoes dramatically decreased and the IIT method was very effective (Fig. 3A, right panel). This result indicates that our model is plausible for evaluating the effectiveness of the IIT method as shown in a field experiment (Mains et al., 2016; Zheng et al., 2019). In addition, when the release rate was greater than 0.08, the time for extinction of the female mosquito after 750 days was not significantly dependent on the scale of the release rate (Fig. 3B). This suggests that if we choose a minimal release ratio to successfully suppress female mosquitoes, the release cost can be significantly reduced.

Next, we examined whether continuous release is necessary to make female mosquitoes extinct. Thus, for a given release rate in the range of successful suppression, we stopped releasing the *Wolbachia*-infected male mosquitoes when the female mosquito population approached a certain level. We define release stop ratio (θ), satisfying

$$\theta = \frac{F(t)}{F(0)}. \tag{9}$$

Specifically, θ refers to the population level of female mosquitoes compared to the initial population. We took $F(0) = F^*$ as the initial equilibrium density of the wild-type female mosquito. Unexpectedly, we found that we could obtain successful suppression even when we stopped the release at $\theta = 0.5$ and we need not continue the release of the *Wolbachia*-infected male mosquitoes (Fig. 3C). This result indicates that there exists an optimal stop point at which we can achieve successful suppression economically.

Thus, we investigated precisely how much the female mosquito population decreased depending on the release stop ratio θ . To explore it, we defined the strategy effectiveness by

$$[SE](\theta) = \frac{F(0) - F^\theta(\infty)}{F(0)} \times 100(\%), \tag{10}$$

where $F^\theta(\infty)$ is the asymptotic density of the female mosquito obtained over a sufficiently long time when the release of the *Wolbachia*-infected male mosquitoes is stopped at θ . In the simulation, $F^\theta(\infty)$ was calculated as

$$F^\theta(\infty) = \frac{1}{T} \int_{t_\infty}^{t_\infty+T} F(t) dt,$$

where t_∞ is a sufficiently long time, and we set $T = 280$ (day) in the simulations. For strategy effectiveness, there exists a threshold-like value (θ_c) of θ , and for $\theta < \theta_c$, the perfect suppression of female mosquitoes is possible (Fig. 3D, green shadow region). Furthermore, we found that we can have a certain amount of effectiveness even when $\theta > \theta_c$ (Fig. 3D, blue shadow region). However, when the release of the *Wolbachia*-infected male mosquitoes is stopped too early, the IIT method is ineffective and increases the density of female mosquitoes (Fig. 3D, yellow shaded region). Therefore, the IIT method can be a nature-friendly and cost-saving strategy to decrease vector mosquitoes effectively, at least, in the absence of contamination.

3.2. The effect of the *Wolbachia* IIT in a scenario with contamination of the *Wolbachia*-infected female mosquitoes

Here, we consider the *Wolbachia*-infected female mosquitoes are contaminated in the release of *Wolbachia*-infected male mosquitoes. We first tested the influence of contamination with the parameter set where the IIT method was successful in suppressing female mosquitoes when contamination of the *Wolbachia*-infected female was absent. As seen in Fig. 4A, the IIT method failed to eradicate female mosquitoes, even for a small contamination ratio (e.g., $p = 0.1\%$) (Fig. 4B). This is because the IIT method is based on cytoplasmic incompatibility. The offspring could survive by mating the *Wolbachia*-infected females (Fig. 1C), so that the total population of female mosquitoes completely replaced from wild-type to *Wolbachia*-infected female (Fig. 4A, second-line panels). However, the maximum density of female mosquitoes did not increase monotonically as the contamination ratio increased. For example, we

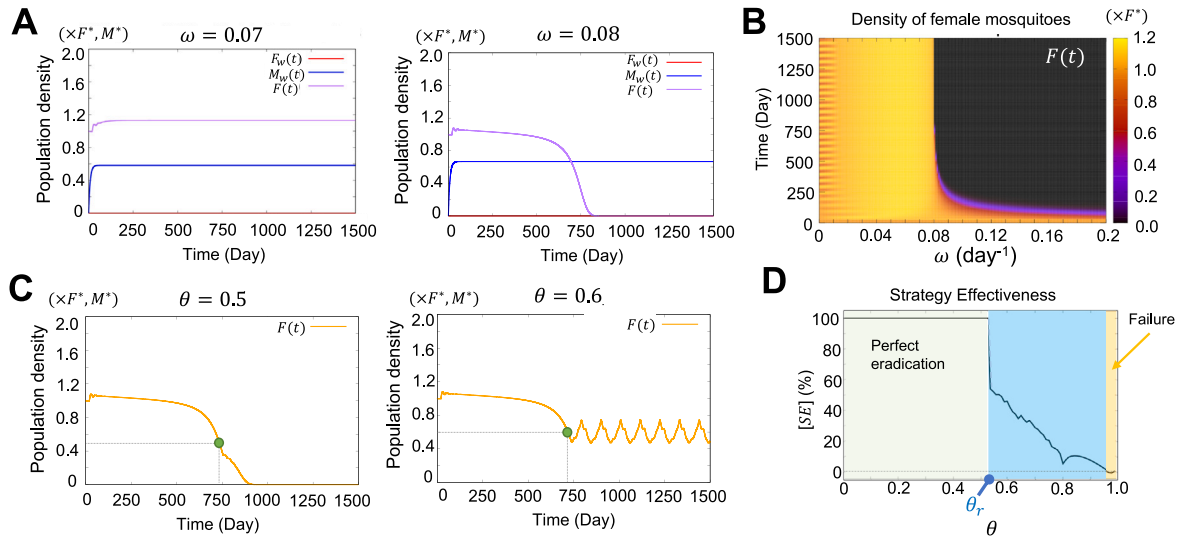


Fig. 3. The effectiveness of the IIT method in the contamination absent scenario ($p = 0$). (A–B) The results of population dynamics by the release of the *Wolbachia*-infected male mosquito with varying release rates ω . As $p = 0$, $F_w(t) = 0$ at all times. (C) Results obtained by choosing the release stop point (θ , Eq. (9)). The release was stopped at the 50% (left panel) and 60% point (right panel) of the initial density of the female mosquitoes. $\omega = 0.08$. Green circles indicate the release stop points. (D) Strategy effectiveness calculated using Eqs. (10). The threshold-like value at which $[SE] < 100\%$ was approximately $0.53 < \theta_r < 0.54$. $\omega = 0.08$.

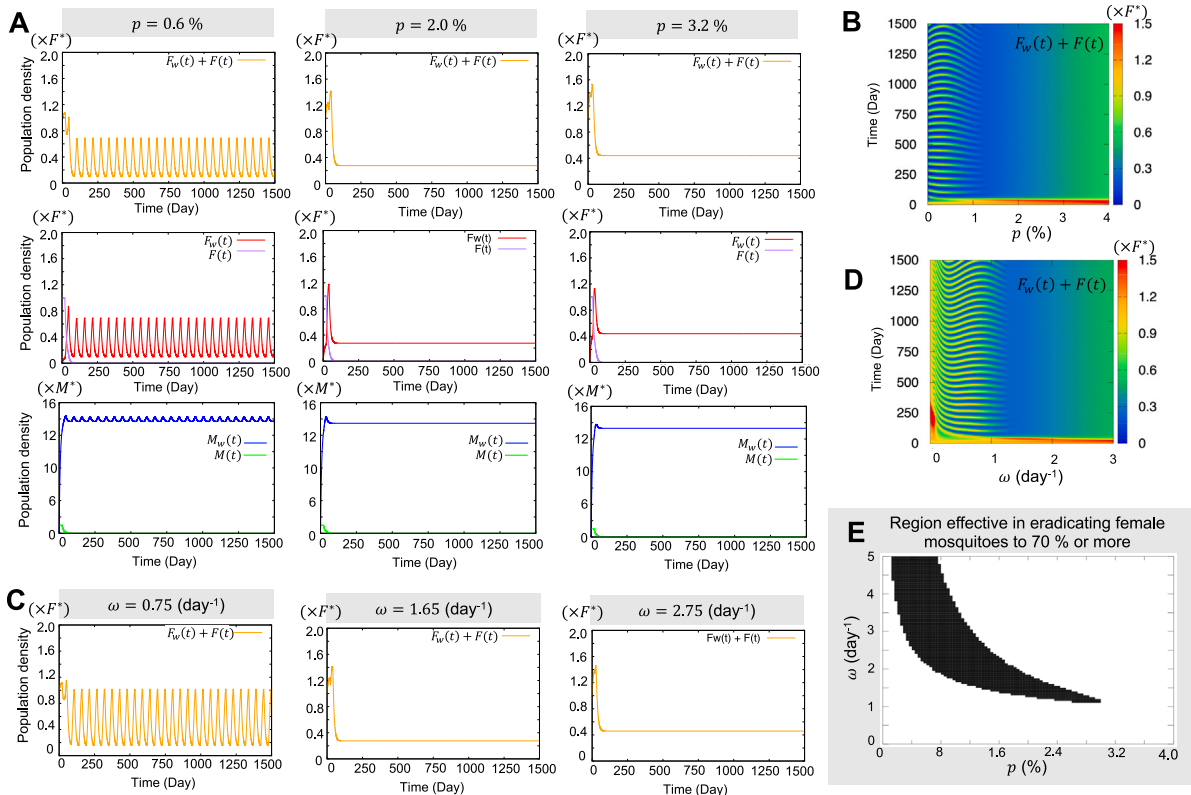


Fig. 4. The effectiveness of the IIT method in the contamination present scenario ($p > 0$). (A) Population dynamics of mosquitoes with increasing contamination ratios (p). $\omega = 1.65$. (B) Change in the maximum density of female mosquitoes as p increases. $\omega = 1.65$ (C) Population dynamics of female mosquitoes with increasing release rate (ω). $p = 2\%$. (D) Change in the maximal density of female mosquitoes with increasing release rate (ω). $p = 2\%$. (E) The black region satisfies condition $F_w(t^\infty) + F(t^\infty) < 0.3F^*$. t^∞ was calculated for 7000 days after the total density of mosquitoes was less than $0.3F^*$.

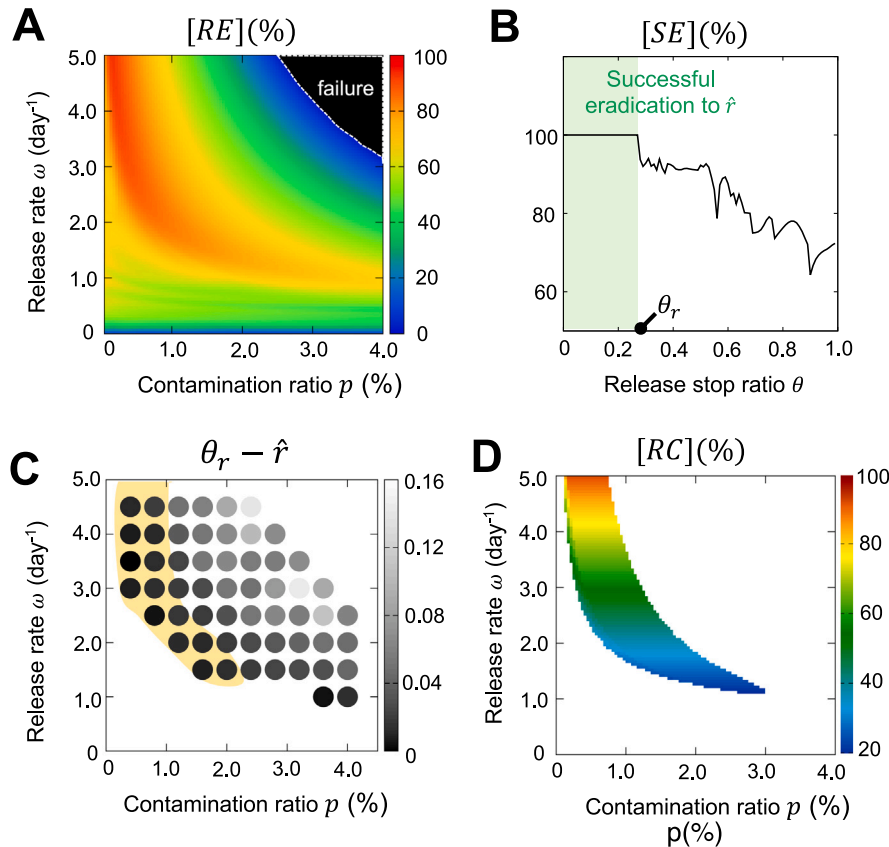


Fig. 5. Release effectiveness and cost to reduce the female mosquitoes. (A) Release effectiveness calculated using Eq. (11), depending on the contamination ratio (p) and release rate (ω). The failure area is the parameter region in which the number of female mosquitoes increase more than the initial density. (B) Strategy effectiveness calculated by Eq. (13) for the case of $p = 2\%$, $\omega = 1.67$. The asymptotic density of the female mosquito was $\hat{r} = 0.259$, and the threshold value of the release stop ratio was $\theta_r = 0.27$. (C) The effectiveness of the release stop strategy defined by the difference between the threshold value (θ_r) of the release stop ratio and asymptotic density (\hat{r}) of the female mosquito. The yellow shadow region corresponds to the region of high release effectiveness (red region in (A)). The dark color implies that the *Wolbachia*-infected male mosquitoes must be released continuously until the density of female mosquitoes reaches the asymptotic density. (D) Release costs calculated using Eqs. (14) when $[RE] \geq 70\%$ (i.e., $\hat{r} \leq 0.3$).

have a lower density of female mosquitoes in the case of $p = 2.0\%$ than in those with $p = 0.6\%$, 3.2% (Fig. 4A and B). Similarly, when we varied the release ratio (ω), we obtained a certain effectiveness for a range of release ratios (Fig. 4C and D). These results indicate that a perfect eradication is not possible when contamination occurs, while IIT is still effective in reducing the female mosquito density at a certain level by choosing an appropriate release rate with respect to a given contamination ratio.

Thus, we next investigated a parameter region of the contamination ratio and release rate in which the total density of female mosquitoes can be suppressed to a certain level, for example, 70% or more (Fig. 4E). We found that there was a successful reduction in the release rate in some regions when the contamination ratio was less than approximately 3%, while the IIT became ineffective easily when the contamination ratio was high. Therefore, a high release rate with respect to a high contamination ratio is insufficient for suppressing the density of female mosquitoes. This is because of the overabundance of *Wolbachia*-infected female mosquitoes caused by contamination in the large number of released mosquitoes. This proposes that the release rate should be carefully adjusted when the contamination exists.

3.3. Reduction effectiveness and release cost to reduce the female mosquitoes

In the previous section, we confirmed that contamination of *Wolbachia*-infected female mosquitoes is a fatal drawback of the IIT method

based on cytoplasmic incompatibility. Nonetheless, the IIT method was still effective in reducing the density of female mosquitoes to a certain level. Naturally, contamination cannot be controlled perfectly without a combination of other technologies (Zheng et al., 2019; Crawford et al., 2020). Thus, we propose a policy to reduce the total population of female mosquitoes effectively, although it will not be a perfect suppression.

We investigated the effectiveness of IIT in reducing the density of female mosquitoes and the release cost required to reduce the female mosquito population to a certain level. To evaluate the reduction effectiveness of IIT, we calculated the reduction effectiveness $[RE]$, defined by $[RE] \equiv (1 - \hat{r}) \times 100$ (%) where

$$\hat{r} \equiv \frac{F_w(\infty) + F(\infty)}{F_w(0) + F(0)}$$

is the asymptotic density of the female mosquito by IIT control, and we define \hat{r} in $0 \leq \hat{r} \leq 1$. That is, the reduction effectiveness implies how much we can decrease the density of the female mosquito to a level of $\hat{r}(F_w(0) + F(0))$ from the initial density, and is given by

$$[RE] = \frac{F_w(0) + F(0) - \{F_w(\infty) + F(\infty)\}}{F_w(0) + F(0)} \times 100 \text{ (%).} \tag{11}$$

We investigated $[RE]$ by varying the release rate (ω) and contamination ratio (p). In our model, we take $F_w(0) = 0$, $F(0) = F^*$. In Fig. 5A, we first noticed that release effectiveness does not simply increase as the release rate increases. There exists a proper range of the release rate

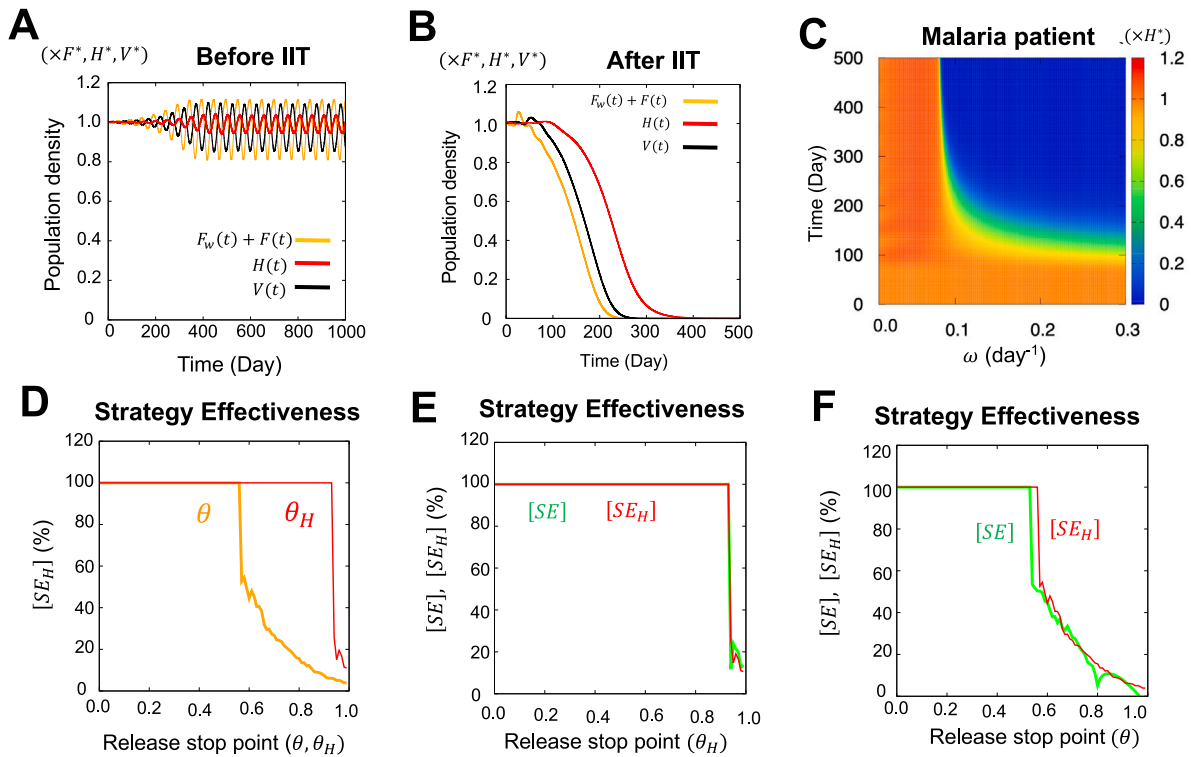


Fig. 6. Malaria-IIT model and strategy effectiveness in the contamination absent scenario ($\rho = 0$). (A) Population dynamics of female mosquitoes ($F_w(t) + F(t)$), Malaria-infected humans ($H(t)$), and Malaria-infected mosquitoes ($V(t)$) before conducting the IIT method. Model Eqs. (1) and (8) were simulated. (B) Population dynamics after IIT. The model Eqs. (2)–(5) and (8) were simulated with $\omega = 0.1$. (C) Effect of the IIT method on release ratio ω . (D) Strategy effectiveness ($[SE_H]$) with respect to release stop ratio θ (Eq. (12)) and θ_H (Eq. (15)). $\omega = 0.1$ (E–F) Strategy effectiveness ($[SE]$ and $[SE_H]$) with respect to release stop ratios θ_H and θ . $\omega = 0.1$.

in which we can obtain a high release effectiveness with respect to a given contamination ratio (red-colored area in Fig. 5A), indicating that there exists an optimal release rate at which the effectiveness reaches its maximum.

Next, we further explore the strategy effectiveness function (13) to confirm whether there exists an effective release stop point:

$$\theta = \frac{F_w(t) + F(t)}{F_w(0) + F(0)}, \quad (12)$$

such that the total population of female mosquitoes reduces to a targeted density without extra release, namely, to an asymptotic density level $F_w(\infty) + F(\infty) = \hat{r}\{F_M(0) + F(0)\}$, obtained after a sufficiently long time. The strategy effectiveness function is given as

$$[SE](\theta, \hat{r}) = \left[\frac{F_w(0) + F(0) - \{F_w^\theta(\infty) + F^\theta(\infty)\}}{F_w(0) + F(0)} + \hat{r} \right] \times 100 (\%) \quad (13)$$

where $F_w^\theta(\infty)$ and $F^\theta(\infty)$ are the asymptotic densities of the female mosquitoes after a sufficiently long time and when the release of the *Wolbachia*-infected male mosquitoes is stopped at θ . This measure implies that the reduction effectiveness reaches to 100% when $F_w^\theta(\infty) + F^\theta(\infty) = F_w(\infty) + F(\infty)$.

We first found that a release stop ratio also exists in the scenario with contamination (Fig. 5B). Thus, we next explored the effectiveness of the release stop strategy in the case of imperfect eradication. We evaluated the measure of $\theta_r - \hat{r}$, where θ_r is the threshold value of the release stop ratio. $\theta_r > \hat{r}$ implies that we can have the same $[RE]$ level without extra releases when the release is stopped at $F_w(t) + F(t) = \theta_r F^* > F_w(\infty) + F(\infty) = \hat{r} F^*$. On the contrary, $\theta_r \approx \hat{r}$ implies that the release stop strategy is not useful. We found that the release stop strategy is not useful especially in the parameter region where $[RE]$ is

high (the yellow shadow region in Fig. 5C). This result is in contrast with the scenario without contamination.

Because the release stop strategy was not effective, we finally evaluated the release cost defined by

$$[RC] \equiv \int_0^{t_f} \kappa dt = \int_0^{t_f} \omega F^* dt, \quad (14)$$

where t_f is a minimal time point satisfying $F_w(t_c) + F(t_c) = \hat{r}\{F_w(0) + F(0)\}$. The release cost provides a measure of the number of *Wolbachia*-infected male mosquitoes that should be released to reduce the density of female mosquitoes to a certain level, namely, \hat{r} . We calculated the release cost for the parameter region satisfying $[RE] \geq 70\%$ and found that the release cost increases as the release ratio increases. This was because the time t_f did not depend significantly on the release rates or the contamination ratios. Therefore, choosing a release rate where the $[RE]$ becomes maximal in a given contamination is likely to be the optimal strategy to reduce the number of female mosquitoes without wasting costs.

3.4. Dynamics of malaria infections after the *Wolbachia* IIT without contamination

The original goal of the insect strategy was to decrease the number of infected humans and control vector-borne diseases. Thus, we explored the effectiveness of the IIT method in decreasing the number of malaria cases. We first tested the dynamics of malaria patients and malaria-infected mosquitoes when *Wolbachia* mosquitoes were not introduced (Fig. 6A). The results showed that spread of malaria and its patients persisted as long as the population of female mosquitoes persisted. In contrast, the population of patients dramatically decreased

once IIT was performed (Fig. 6B). With IIT, the number of patients decreased followed by a decrease in malaria-infected mosquitoes while there was a time gap in complete extinction. We also investigated how the release rate (ω) affects the effectiveness of the case and found that the IIT becomes effective when the release rate is larger than the same threshold release rate at which female mosquitoes become perfectly suppressed (Fig. 6C, and 3B). This indicates that we can have successful suppression in both cases and in female mosquitoes.

Next, we explored strategy effectiveness in a manner similar to that in Fig. 3D with respect to the release stop point (θ) for the female mosquitoes. In addition, we explored the release stop point (θ_H) for an infected human such that

$$\theta_H = \frac{H(t)}{H(0)}. \tag{15}$$

We also defined the strategy effectiveness based on the population of infected humans as follows:

$$[SE_H](\theta_H) = \frac{H(0) - H^{\theta_H}(\infty)}{H(0)} \times 100(\%), \tag{16}$$

where $H^{\theta_H}(\infty)$ is the asymptotic density of the patients after a sufficiently long time and when the release of the *Wolbachia*-infected male mosquitoes is stopped at θ_H . In the simulation, $H^{\theta_H}(\infty)$ was calculated as

$$H^{\theta_H}(\infty) = \frac{1}{T} \int_{t_\infty}^{t_\infty+T} H(t)dt,$$

where t_∞ is a sufficiently long time and T is taken as 280 days.

We first investigated the strategy effectiveness $[SE_H]$ with respect to the two release stop points, θ and θ_H in Fig. 6D, respectively. We found that both θ and θ_H can provide perfect effectiveness when $\theta \leq 0.56$ or $\theta_H \leq 0.93$. This suggests that we can control the pandemic without releasing the *Wolbachia*-infected mosquitoes until the female mosquitoes are completely suppressed or the large number of infected humans decrease.

Finally, to compare which strategy effectiveness should be chosen as a measure that incurs less release costs, we investigate $[SE]$ and $[SE_H]$ with respect to both θ_H and θ (Fig. 6E and F). We found no notable difference between the effectiveness of the two strategies for both θ_H and θ , indicating that we may choose either release stop measures or strategy effectiveness measures at least under the scenario of no contamination.

3.5. The effectiveness of the *Wolbachia* IIT on malaria and optimal release cost under the scenario of contamination present

Section 3.2 reports that a certain level of strategic efficacy was achieved depending on the release rate (ω) and contamination ratio (p) of *Wolbachia*-infected female mosquitoes. Thus, we examined the effectiveness of IIT in reducing the population of malaria-infected cases in the presence of contamination. We first investigated the dynamics of malaria cases by varying ω and p (Fig. 7A and B). Unexpectedly, we found that we can have a perfect extinction of malaria-infected cases, even though female mosquitoes were not perfectly suppressed (Fig. 7A and B, the second panels, and Fig. 7E). However, once the female mosquitoes persist for more than a certain population level, the IIT method could not lead to a perfect eradication of malaria, although we still could have a certain level of effectiveness in reducing the population of malaria-infected cases (Fig. 7A and B, first and third panels, and Fig. 7C and D).

Finally, we calculated the release cost and explored the optimal release ratio values for a given contamination ratio. Because we were able to obtain a perfect eradication cases using the IIT method as shown in Fig. 7E, we first investigated the release cost, $[RC_H]$, defined by

$$[RC_H] \equiv \int_0^{t_e} \kappa dt = \int_0^{t_e} \omega F^* dt, \tag{17}$$

where t_e is the extinction time of cases, that is, the minimal time point at which the malaria-infected cases of humans are extinct. To avoid additional calculations owing to numerical errors, we set $t_e = \min\{t|H(t_e) \leq \epsilon H^* (\epsilon \ll 1)\}$.

The result of Fig. 8A shows that the release cost is not simply determined by the release rate monotonously and there exists an optimal release rate at which the release cost becomes minimal for a given contamination ratio. Thus, we further investigated where the minimal release cost was obtained in the range of the release rate shown in Fig. 8B. We found that the minimal release cost increased as the contamination ratio increased, whereas the optimal release rate satisfying the minimal release cost decreased. This indicates that when the contamination ratio is high, we must choose a lower release rate. On the other hand, a long time was required for cases to become extinct when we chose a low release rate, as shown in Fig. 8C. If the disease persists for a long time, the number of deaths will increase. Thus, when contamination is large, the policy of minimal release cost is not always the best.

Taken together, we concluded that the IIT method can be a very effective strategy to eradicate malaria but the optimal policy to reduce the cost should be carefully considered depending on the contamination ratio and extinction period of the cases.

4. Conclusion

Incompatible insect technique (IIT) using maternally inherited endosymbiotic bacteria *Wolbachia* is considered a promising method that can avoid an influence on male mating competitiveness, survivability, and nature (Werren et al., 2008). Thus, the IIT method was chosen as an alternative to the sterile insect technique (SIT); in particular, it has been considered an effective method for the suppression of mosquito populations (Curtis et al., 1982). However, the contamination problem of the *Wolbachia*-infected female mosquito in the IIT method has been a concern because the *Wolbachia*-infected female mosquito leads to the replacement of the wild-type field mosquito population with the *Wolbachia*-infected mosquitoes, which results in the failure of disease control.

In this study, we developed a mathematical model based on the population dynamics of *Wolbachia*-infected mosquitoes and explored the extent to which the IIT method and the release strategy of *Wolbachia*-infected male mosquitoes can effectively suppress female mosquitoes in the two scenarios of the contamination being absent or present. As shown in previous field studies (Curtis et al., 1982; Mains et al., 2016; Zheng et al., 2019), the IIT method was highly effective in eradicating wild-type female mosquitoes when contamination was absent. In contrast, the perfect eradication policy failed easily even if a small percentage of *Wolbachia*-infected female mosquitoes are contaminated by the release of *Wolbachia*-infected male mosquitoes, whereas a decrease in the total number of female mosquitoes at a certain level is possible. Our study also found that *Wolbachia*-infected male mosquitoes may need not be released until the female mosquitoes become extinct and there exists an optimal stopping point of the release at which we can obtain sufficiently successful control. This result suggests a new insight into reducing the economic burden when it comes to actual policy implementation. However, our study also suggests that we need a different policy under the scenario of contamination. In this scenario, the release stop strategy has little effect on reducing the release costs. Furthermore, the release cost was determined by the scale of the release rate. These theoretical observations could be validated by field experiments in a limited local area and may be extended to the real world by further development of mosquito sampling methods and analysis of sample data over a wider area.

Similarly with the previous field work (Zheng et al., 2019), our modeling study also suggested that IIT cannot be an effective method to eradicate female mosquitoes perfectly when contamination is present. However, our study using the Malaria-IIT model found that female

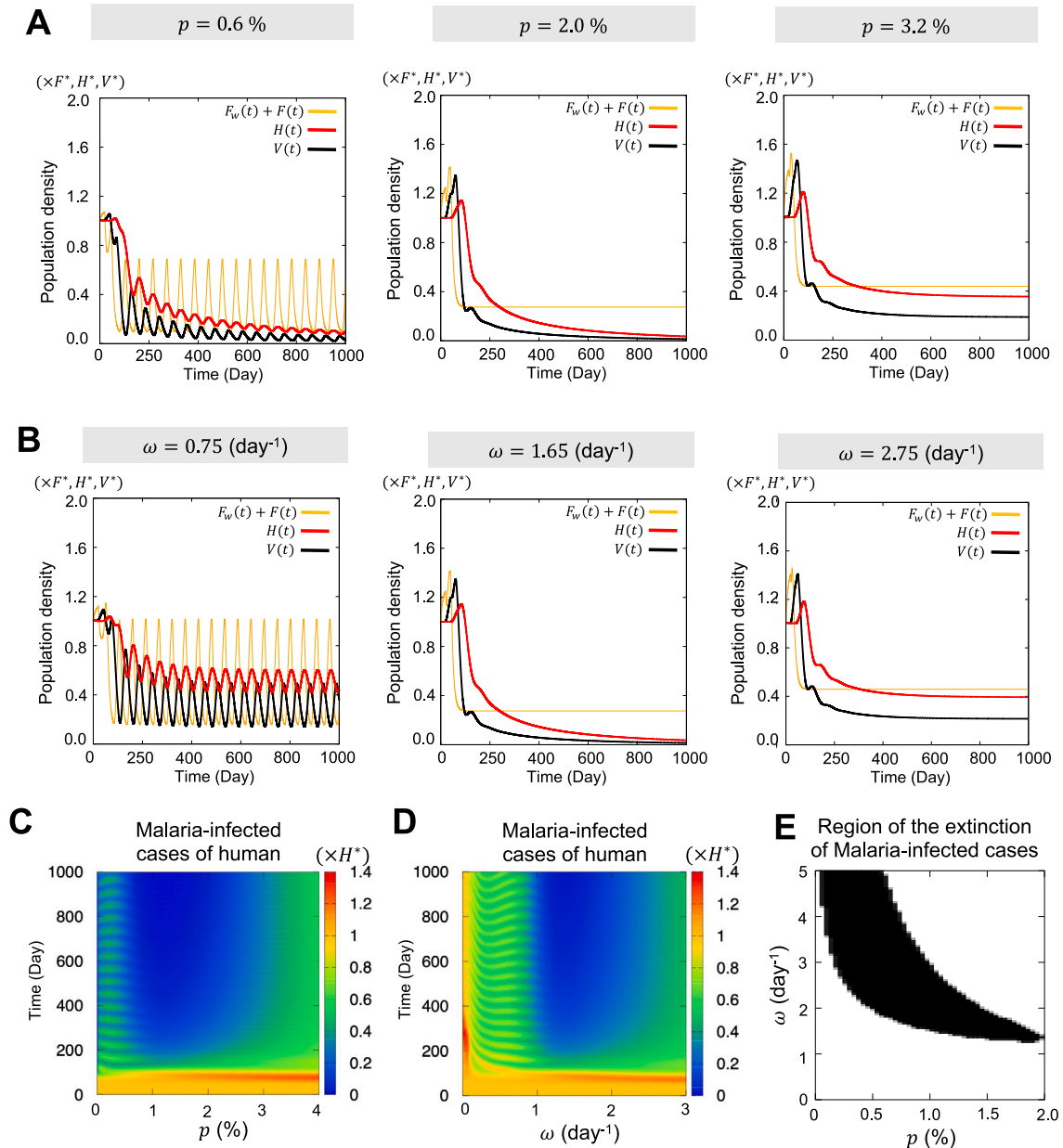


Fig. 7. The effectiveness of the *Wolbachia* IIT under the scenario of contamination present. (A) Population dynamics of Malaria-infected female mosquitoes with varying contamination ratios (p). $\omega = 1.65$ was selected. The parameters of the IIT model are the same as those in Fig. 4. (B) Population dynamics of malaria infected cases and female mosquitoes with varying release rates (ω). $p = 2.0\%$ was chosen. The parameters of the IIT model are the same as those in Fig. 4. (C–D) Population dynamics of Malaria-infected humans with varying contamination ratios and release rates. $\omega = 1.65$ was fixed in (C) and $p = 2.0\%$ was fixed in (D). (E) Parameter region where malaria-infected cases are extinct. The region was calculated using the condition satisfying $H(t)/H^* \leq 0.001$ for a sufficiently long time.

mosquitoes need not be eradicated perfectly to extinguish malaria and we can obtain a successful control result even with contamination occurring. We also found that there exists an optimal release rate at which the release cost becomes minimal, which should be smaller when the contamination ratio is higher. On one hand, the extinction time is prolonged when we choose a lower release rate. Therefore, a measure should be carefully chosen which is optimal in designing a control strategy for preventing mosquito-borne diseases.

In this study, we excluded the case of pathogen-blocking phenotype *Wolbachia*. Some *Wolbachia* strains are known to block pathogen transmission (Caragata et al., 2013; O’Neill et al., 2019). Thus, the

contamination of the *Wolbachia*-infected female mosquito may not be a problem for controlling certain diseases, and population replacement may be an effective control strategy. However, *Wolbachia* can be affected by the environment, and its pathogen-blocking phenotype may weaken over time, allowing it to become a new source of disease transmission. Therefore, the ideal strategy for suppressing the female mosquito population remains an important issue.

Every year, many people are infected and lose their lives because of vector-borne diseases. However, it is difficult to implement a control policy because field experiments are spatially and temporally restricted. In particular, the IIT method has been shunned owing to contamination.

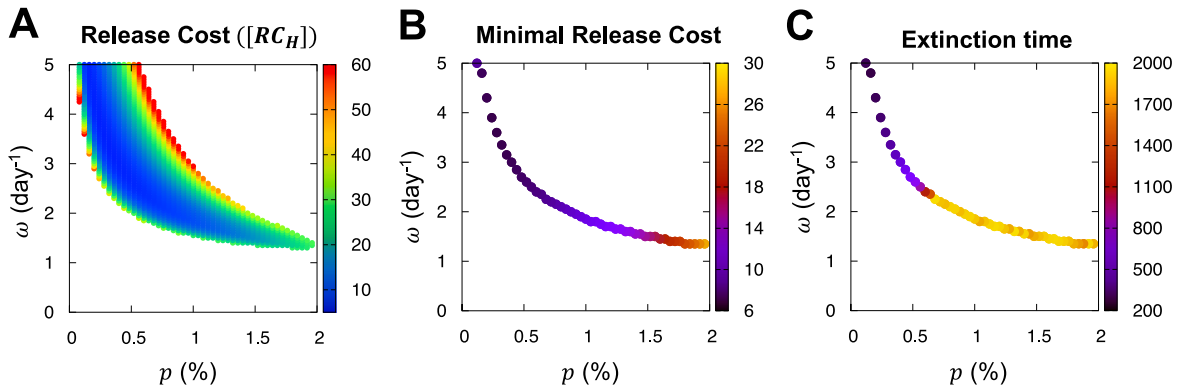


Fig. 8. Release cost and optimal release ratio. (A) The release cost ($[RC_H]$) given by Eq. (17) was calculated by varying the release rate(ω) and contamination ratio (p). (B) The minimal release cost plotted from (A) for a given contamination ratio. (C) Extinction time t_e plotted from (A) for a given contamination ratio.

Our study suggests that the IIT method is very promising, even though perfect suppression is not possible in real and it might be valuable to be tested in the real field of endemic. In this study, we did not consider spatial effects, such as geographical conditions and spatial spreading, which we consider as a future work.

CRedit authorship contribution statement

Taiga Matsufuji: Conceptualization, Software, Validation, Formal analysis, Investigation, Resources, Data curation, Writing – original draft, Visualization. **Sungrim Seirin-Lee:** Conceptualization, Methodology, Validation, Formal analysis, Investigation, Resources, Writing – original draft, Writing review & editing, Visualization, Supervision, Project administration, Funding acquisition.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Data availability

All relevant data are within the manuscript.

Acknowledgments

This work was partially supported by a Grant-in-Aid for Scientific Research KAKENHI B (Japan Society for the Promotion of Science(JSPS), JP19H01805), and Japan Science and Technology Agency(JST) CREST (JPMJCR2111) to S.S.L.

Appendix A. Equilibria and linear stability of wild-type mosquitoes

The equilibria (F^*, M^*) of the wild-type mosquitoes satisfy the equations;

$$0 = qr_3FM \cdot \exp(-\alpha E_3^\beta F^\beta M^\beta) - \mu F,$$

$$0 = (1 - q)r_3FM \cdot \exp(-\alpha E_3^\beta F^\beta M^\beta) - \mu M.$$

With taking $q = 0.5$, we have $F^* = M^*$ and

$$\frac{1}{2}r_3F^{*2} \exp(-\alpha E_3^\beta \cdot F^{*2\beta}) - \mu F^* = 0.$$

Next, we analyze the stability of the equilibrium. With setting

$$f(F, M) = \frac{1}{2}r_3FM \cdot \exp(-\alpha E_3^\beta F^\beta M^\beta) - \mu F,$$

$$g(F, M) = \frac{1}{2}r_3FM \cdot \exp(-\alpha E_3^\beta F^\beta M^\beta) - \mu M,$$

we obtain the Jacobian matrix

$$J(F, M) = \begin{pmatrix} \frac{\partial}{\partial F} f(F, M) & \frac{\partial}{\partial M} f(F, M) \\ \frac{\partial}{\partial F} g(F, M) & \frac{\partial}{\partial M} g(F, M) \end{pmatrix}.$$

and the eigenvalue equation of $\lambda^2 - (\lambda_1 + \lambda_2)\lambda + \lambda_1 \cdot \lambda_2 = 0$, where

$$\lambda_1 + \lambda_2 = \frac{\partial}{\partial F} f(F^*, M^*) + \frac{\partial}{\partial M} g(F^*, M^*)$$

$$= \frac{1}{2}r_3 \cdot \exp(-\alpha E_3^\beta F^{*\beta} M^{*\beta})(F^* + M^*)(1 - \alpha\beta E_3^\beta F^{*\beta} M^{*\beta}) - 2\mu, \quad (18)$$

$$\lambda_1 \cdot \lambda_2 = \frac{\partial}{\partial F} f(F^*, M^*) \cdot \frac{\partial}{\partial M} g(F^*, M^*) - \frac{\partial}{\partial M} f(F^*, M^*) \cdot \frac{\partial}{\partial F} g(F^*, M^*)$$

$$= \frac{1}{2}r_3\mu \cdot \exp(-\alpha E_3^\beta F^{*\beta} M^{*\beta})(F^* + M^*)(\alpha\beta E_3^\beta F^{*\beta} M^{*\beta} - 1) + \mu^2. \quad (19)$$

We numerically evaluated the stability by judging the sign of the Eqs. (18) and (19) with given parameter values.

Appendix B. Additional simulations on the effect of delay scales

To see the effect of the delay scale in the IIT model, we investigated the pre-/post-implementation state of the IIT method numerically for four cases (Fig. 9); (A) Change of the mosquito development period (τ) in the IIT model, (B) Change of incubation period of Malaria in humans (τ_2) with a fixed incubation period of Malaria in mosquitoes (τ_3) in the Malaria-IIT model, (C) Change of incubation period of Malaria in mosquitoes (τ_3) with a fixed incubation period of Malaria in humans (τ_2) in the Malaria-IIT model, and (D) Change of incubation period of Malaria in both humans and mosquitoes (τ_2 and τ_3) in the Malaria-IIT model.

The results show that the effect of the IIT method is less sensitive to the mosquito development period (τ), although the elimination time becomes longer as the development period becomes longer, as shown in Fig. 9A. However, we could not see a significant prolongation of elimination durations in the larger scale of mosquito development period which should be considered to the extent biologically feasible. Thus, we concluded that the delay scales in the IIT model could be negligible.

Next, we also explored the impact of incubation period scales on the Malaria-IIT model (Fig. 9B–D) with respect to the empirically estimated mosquito development period, $\tau = 18.84$ days (Dye, 1984). The simulation results show that the IIT method is effective and there is no marked difference in its effectiveness for several variations of the incubation period scales (Fig. 9B–D, lower panels). On the other hand, interesting results were obtained for very short incubation periods (the first panels in Fig. 9B–D), which resulted in longer pandemic durations. This observation indicates that temporal effects should be considered when applying the IIT method to other vector-borne diseases with short incubation periods.

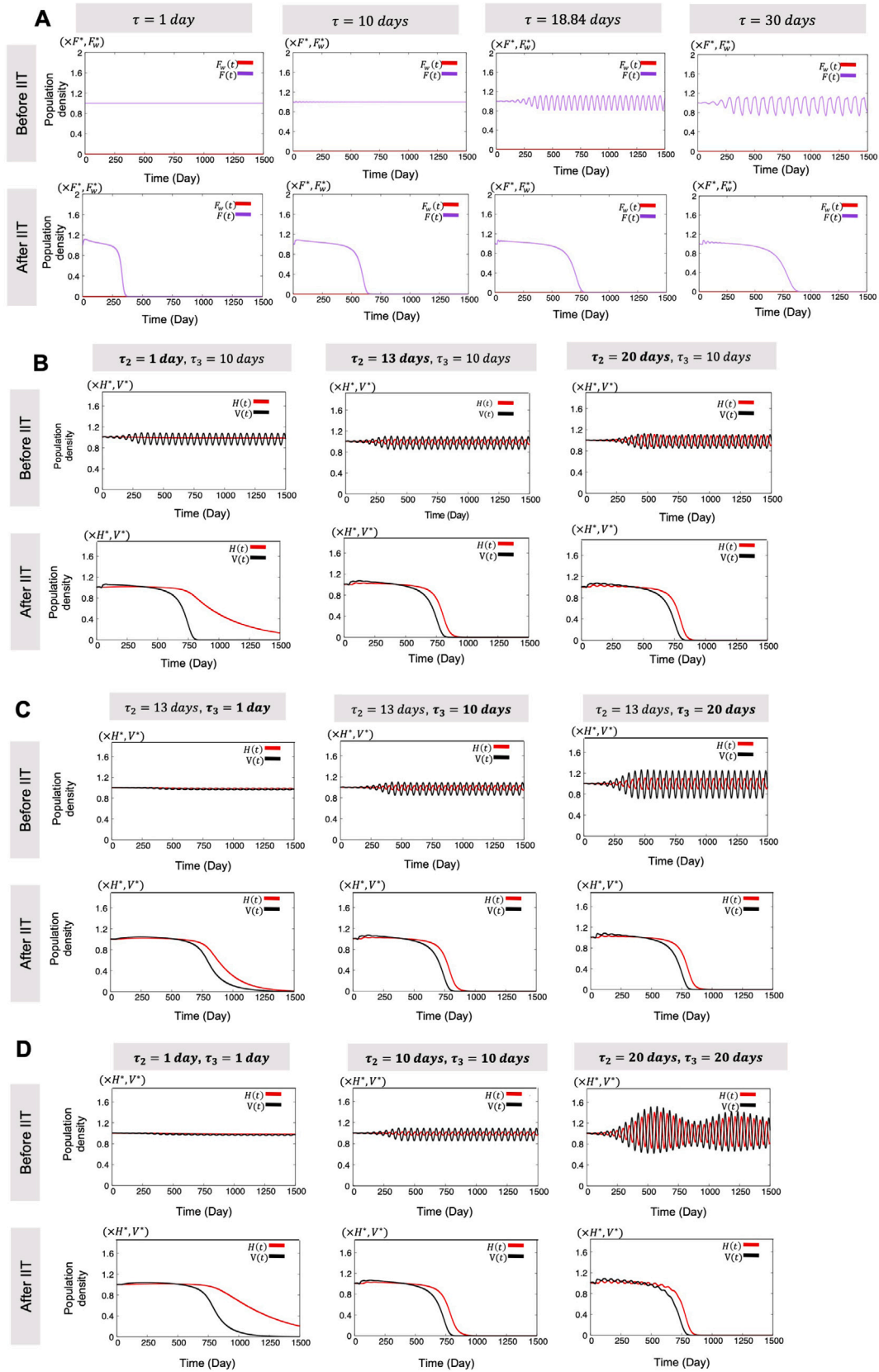


Fig. 9. The effect of delay size before/after IIT method. (A) IIT model with various mosquito development period(τ). (B–D) Malaria-IIT model with various incubation periods in either/both human(τ_2) or/and mosquito(τ_3) for $\tau = 18.84$ days. Before IIT and After IIT correspond to $\omega = 0.0$ and 0.08 , respectively. The contamination is absent ($p = 0$).

References

- Alphey, L., Benedict, M.Q., Bellini, R., Clark, G.G., Dame, D., Service, M., Dobson, S., 2010. Sterile-insect methods for control of mosquito-borne diseases: An analysis. *Vector Borne Zoonotic Dis.* 10, 295–311.
- Beebe, N.W., Pagendamb, D., Trewin, B.J., Boomer, A., et al., 2021. Releasing incompatible males drives strong suppression across populations of wild and *Wolbachia*-carrying *Aedes aegypti* in Australia. *Proc. Natl. Acad. Sci. USA* 118 (41), 1–12.
- Bouyer, J., Culbert, N.J., Dicko, A.H., Pacheco, M.G., et al., 2020. Field performance of sterile male mosquitoes released from an uncrewed aerial vehicle. *Science Robotics* 5 (43), <http://dx.doi.org/10.1126/scirobotics.aba6251>.
- Caragata, E.P., Rance, E., Hedges, L.M., Gofton, A.W., Johnson, K.N., O'Neill, S.L., McGraw, E.A., 2013. Dietary cholesterol modulates pathogen blocking by *Wolbachia*. *PLOS Pathog.* 9 (6), e1003459.
- Carvajal-Lago, L., Ruiz-Lopez, M.J., Figuerola, J., de la Puente, J.M., 2021. Implications of diet on mosquito life history traits and pathogen transmission. *Environ. Res.* 195 (110893).
- Carvalho, D.O., et al., 2015. Suppression of a field population of *Aedes aegypti* in Brazil by sustained release of transgenic male mosquitoes. *PLOS Negl. Trop. Dis.* <http://dx.doi.org/10.1371/journal.pntd.0003864>.
- Crawford, J.E., Clarke, D.W., Criswell, V., Desnoyer, M., et al., 2020. Scaled deployment of *Wolbachia* to protect the community from dengue and other *Aedes* transmitted arboviruses. *Nature Biotechnol.* 38, 482–492.
- Curtis, C.F., Brooks, G.D., Ansari, M.A., et al., K.K.G., 1982. A field trial on control of *Culex quinquefasciatus* by release of males of a strain integrating cytoplasmic incompatibility and a translocation. *Entomol. Exp. Appl.* 31, 181–190.
- Dye, C., 1984. Models for population dynamics of the yellow fever mosquito, *Aedes aegypti*. *J. Anim. Ecol.* 53, 247–268.
- Evans, B.R., Kotsakiozi, P., da Silva, A.L.C., Ioshino, R.S., Garziera, L., Pedrosa, M.C., Malavasi, A., Virginio, J.F., Capurro, M.L., Powell, J.R., 2019. Transgenic *Aedes aegypti* mosquitoes transfer genes into a natural population. *Sci. Rep.* 9 (13047).
- Gu, W., Killeen, G.F., Mbogo, C.M., Regens, J.L., Githure, J.I., Beier, J.C., 2003. Rethinking the extrinsic incubation period of malaria parasites. *Trans. R. Soc. Trop. Med. Hyg.* 97, 43–50.
- Hughes, G.L., Rivero, A., Rasgon, J.L., 2014. *Wolbachia* can enhance *Plasmodium* infection in mosquitoes: Implications for malaria control? *PLOS Pathog.* 10 (9), e1004182.
- Knipling, E.F., 1955. Possibilities of insect control or eradication through the use of sexually sterile males. *J. Econ. Entomol.* 48, 459–462.
- Lacroix, R., et al., 2012. Open field release of genetically engineered sterile male *Aedes aegypti* in Malaysia. *PLoS One* 7 (8), e42771.
- Laven, H., 1967. Eradication of *Culex pipiens fatigans* through cytoplasmic incompatibility. *Nature* 216 (5113), 383–384.
- Lotka, A.J., 1923. Contribution of the analysis of malaria epidemiology. *Am. J. Hyg.* 3 (Supp), 1–37.
- Macdonald, I.G., 1957. *The Epidemiology and Control of Malaria*. Oxford University Press.
- Mains, J.W., Brelsfoard, C.L., Rose, R.I., Dobson, S.L., 2016. Female adult *Aedes albopictus* suppression by *Wolbachia*-infected male mosquitoes. *Sci. Rep.* 6, 33846.
- Natiello, M.A., Solari, H.G., 2020. Modelling population dynamics based on experimental trials with genetically modified (RIDL) mosquitoes. *Ecol. Model.* 424, 108986.
- Ohm, J.R., Baldini, F., Barreaux, P., Lefevre, T., Lynch, P.A., Suh, E., Whitehead, S.A., Thomas, M.B., 2018. Rethinking the extrinsic incubation period of malaria parasites. *Parasites and Vectors* 11 (178), <http://dx.doi.org/10.1186/s13071-018-2761-4>.
- O'Neill, S.L., Ryan, P.A., Turley, A.P., Wilson, G., et al., 2019. Scaled deployment of *Wolbachia* to protect the community from dengue and other *Aedes* transmitted arboviruses. *Gates Open Res.* 2 (36), 1–28.
- Phillips, M.A., Burrows, J.N., Manyando, C., van Huijsduijnen, R.H., Van Voorhis, W.C., Wells, T.N.C., 2017. *Malaria*. *PRIMER* 3 (17050), 1–24.
- Phuc, H., Andreasen, M., Burton, R., Vass, C., Epton, M., Pape, G., Fu, G., Condon, K., Scaife, S., Donnelly, C., Coleman, P., White-Cooper, H., Alphey, L., 2007. Late-acting dominant lethal genetic systems and mosquito control. *BMC Biol.* 5 (11), 1–11.
- Ritchie, S.A., et al., 2018. Mission accomplished? We need a guide to the 'post release' world of *wolbachia* for *aedes*-borne disease control. *Trends Parasitol.* 34 (3), 217–226.
- Ross, R., 1911. *The Prevention of Malaria*. Jhon Murray.
- Ruan, S., Xiao, D., Beier, J.C., 2008. On the delayed ross-macdonald model for malaria transmission. *Bull. Math. Biol.* 70, 1098–1114.
- Seirin-Lee, S., Baker, R.E., Gaffney, E.A., White, S.M., 2013a. Modelling *Aedes aegypti* mosquito control via transgenic and sterile insect techniques: endemics and emerging outbreaks. *J. Theoret. Biol.* 331, 78–90.
- Seirin-Lee, S., Baker, R.E., Gaffney, E.A., White, S.M., 2013b. Optimal barrier zones for stopping the invasion of *Aedes aegypti* mosquitoes via transgenic or sterile insect techniques. *Theor. Ecol.* 6, 427–442.
- Shaw, W.R., Holmdahl, I.E., Itoe, M.A., Werling, K., Marquette, M., Paton, D.G., Singh, N., Buckee, C.O., Childs, L.M., Catteruccia, F., 2020. Multiple blood feeding in mosquitoes shortens the *Plasmodium falciparum* incubation period and increases malaria transmission potential. *PLOS Pathog.* 16 (12).
- Shaw, W.R., Marcenac, P., Childs, L.M., Buckee, C.O., Baldini, F., Sawadogo, S.P., Dabire, R.K., Diabate, A., Catteruccia, F., 2016. *Wolbachia* infections in natural *Anopheles* populations affect egg laying and negatively correlate with *Plasmodium* development. *Nature Commun.* 7 (11772).
- Soh, S., Ho, S.H., Ong, J., Seah, A., Dickens, B.S., et al., 2022. Strategies to mitigate establishment under the *Wolbachia* incompatible insect technique. *Viruses* 14, 1132.
- Thomas, D.D., Donnelly, C.A., Wood, R.J., Alphey, L.S., 2000. Insect population control using a dominant, repressible, lethal genetic system. *Science* 287, 2474–2476.
- Vreysen, M.J.B., Robinson, A.S., Hendrich, J. (Eds.), 2007. *Area-Wide Control of Insect Pests*. IAEA, Springer, Netherlands.
- Werren, J.H., Baldo, L., Clark, M.E., 2008. *Wolbachia*: master manipulators of invertebrate biology. *Nat. Rev.* 6, 741–751.
- WHO, 2019. *World Malaria Report 2019*. World Health Organization.
- WHO, 2020. *Vector-Born Diseases*. World Health Organization, <https://www.who.int/news-room/fact-sheets/detail/vector-borne-diseases>.
- Yen, J.H., Barr, A.R., 1971. New hypothesis of the cause of cytoplasmic incompatibility in *Culex pipiens* L. *Nature* 232 (5313), 657–658.
- Yen, P.-S., Failloux, A.-B., 2020. A review: *Wolbachia*-based population replacement for mosquito control shares common points with genetically modified control approaches. *Pathogens* 9 (404), 1–14.
- Zheng, X., Zhang, D., Li, Y., Yang, C., Wu, Y., et al., 2019. Incompatible and sterile insect techniques combined eliminate mosquitoes. *Nature* 572, 56–61.