Review

The Chemistry on Diterpenoids in 1978. Part-II¹⁾

Eiichi Fujita*, Kaoru Fuji, Yoshimitsu Nagao, and Manabu Node

Received September 5, 1979

I. INTRODUCTION

This is one of a series of our annual reviews on diterpenoids chemistry. The classification of the compounds is the same as that adopted in our reviews since 1969. This review covers the literatures published between July and December 1978 and also omissions in Part-I.

II. PODOCARPANE DERIVATIVES

Podocarpane

Copper-catalyzed 1,6-addition of benzyl Grignard reagents 1a and 1b to a dienone 2 gave rise to 3a and 3b. Acid-catalyzed cyclization of the products gave 1:1 mixture of A/B trans- and cis-2-oxopodocarpa-8,11,13-trienes 4a and 4b respectively, while cyclization of the allylic alcohol 5 gave the A/B cis-isomer 6 in good yield.²⁾

Meo
$$(1a) para-OMe$$

$$(1b) meta-OMe$$

$$(3a) R = P-OMe$$

$$(34) R = m-OMe$$

^{*} 藤田栄一,富士 薫, 長尾善光, 野出 学:Laboratory of Physiological Activity, Institute for Chemical Research, Kyoto University, Uji, Kyoto-Fu 611.

$$(44) R = 12-0Me$$

$$(44) R = 13-0Me$$

$$(6)$$

A stereocontrolled total synthesis of (\pm) -19 α ,20 α -(acetylimino)-12-hydroxy-5 β , 10 α -podocarpa-8,11,13-triene (7), a degradation product of atisine, was reported.³⁾ Dieckmann cyclization of 8 in refluxing 1% aqueous methanolic KOH gave 9 and 10 in 85% yield and 11 in 13% yield.⁴⁾ The conformation of 12 in crystalline state was determined by X-ray analysis. Ring A in 12 is in a distorted chair conformation, whereas ring B is in a distorted boat conformation.⁵⁾

OH

$$A_{c}$$
 A_{c}
 A_{c}

III. LABDANE DERIVATIVES

Labdane

Labdanoid diterpenes 13–18 from Helichrysum confertum, 19–21 from H. albirosulatum, and 13, 16 and 17 from Denekia capensis were isolated.⁶⁾ Root of Critonia daleoides was found to contain a furanoditerpene 22.⁷⁾ Labdane derivatives 23 and 24 were isolated from Nidorella auriculata ssp. polycephala.⁸⁾

(+)-Polyalthic acid (25) was isolated from Sequoia semperivirens.⁹⁾ A new diterpene, 13-hydroxyballonigrinolide (27) was isolated from Ballota lanata, together with a known diterpene, ballonigrin (26).¹⁰⁾ Investigation by gas liquid chromatography of a small subfraction of Oriental tobacco condensate (Nicotiana tabacum L.) led to the identification of norlabdane derivatives 28–34.¹¹⁾ 13,17-Epithio-8,13-epoxy-14,15-dinorlabdane (35) was synthesized.¹²⁾

(13)
$$R^1 = R^2 = Me$$
 (16) (17)

(14) $R^1 = co_2 Me$; $R^2 = H$ (15) $R^1 = H$; $R^4 = co_4 Me$

(18) (19) $R^1 = R^2 = H$ (117)

(19) $R^1 = R^2 = H$ (117)

(118) (19) $R^1 = R^2 = H$ (117)

(19) $R^1 = R^2 = H$ (117)

(119) (119) $R^1 = R^2 = H$ (111) $R^1 = H$ (1111) R^1

Hydroxymethylation of 36 led to the synthesis of a new series of intramolecular acetals related to known perfumery compounds.¹³⁾ A stable ozonide 37 was isolated from the ozonolysis of manool. Another major ozonolysis product was 8β ,13:8,14-diepoxy-15,17-dinorlabdan-14 β -ol (38).¹⁴⁾

IV. CLERODANE DERIVATIVES

Clerodane

Populifolic acid (39) and its derivatives 40–45 were isolated from Cistus populifolius as their methyl esters. ¹⁵⁾ cis-Kolavenic acid (46) and its dihydro-derivative were isolated from Fleischmannia sinclairii. ⁷⁾ Since stereochemistry at C-13 of populifolic acid (39) and dihydrokolavenic acid has not been determined and direct comparison of these compounds has not been done, identity of these diterpenes remains uncertain. The structure of a new diterpene isolated from Haplopappus ciliatus was elucidated to be 47 by X-ray diffraction techniques. ¹⁶⁾

$$(39) R^{1} = R^{2} = H$$

$$(40) R^{1} = H; R^{2} = 0H$$

$$(41) R^{1} = H; R^{2} = 0Ac$$

$$(42) R^{1} = 0H; R^{2} = H$$

$$(43) R^{1} = 0Me; R^{2} = H$$

$$(44) R^{1} = H; R^{2} = 0Me$$

$$(45) R^{1}, R^{2} = 0$$

$$(46)$$

$$(47)$$

$$(48)$$

$$(49)$$

$$(49)$$

$$(49)$$

$$(49)$$

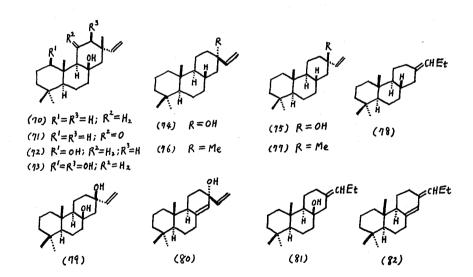
Three new diterpenes 48–50 from Nidorella agria and two new diterpenes 51 and 52 from N. resedifolia were isolated.⁸⁾ Isocrotocaudin (53) was isolated from Croton caudatus.¹⁷⁾ The hexane extract of the Colombian medicinal plant Baccharis tricuneate yielded four new ent-clerodanes, bacchotricuneatins A–D (54–57).¹⁸⁾ Teucrins Hl¹⁹⁾-H4 (58–61) were isolated from Teucrium hyrcanicum and their structures were determined²⁰⁾. Croton diasii was found to contain a new diterpene diasin (62).²¹⁾ Montanin C (63)²²⁾ and D (64)²³⁾ were isolated from Teucrium montanum.

The absolute stereochemistries of clerodin, caryoptin, and 3-epicaryoptin were corrected to the enantiomeric forms of the previously reported formulas 65, 66, and 67, respectively, from the results of some chiroptical data.²⁴⁾ Diels-Alder approach to the synthesis of the antifeedant, ajugarin I (68) was reported.²⁵⁾ A potential intermediate 69 in the synthesis of friedolabdanes was prepared and its reactions with peracid, ozone, and diborane were studied.²⁶⁾

V. PIMARANE AND ISOPIMARANE DERIVATIVES

Pimarane and Isopimarane

The sandaracopimara-15-enes 70–73 were isolated from *Premna latifolia*.²⁷⁾ In the presence of bis(triphenylphosphine)nickel dichloride both vinylcarbinols 74 and 75 reacted with methylmagnesium bromide affording a ca. 74:4:24 mixture of hydrocarbons 76, 77, and 78 (two isomers), respectively. The first olefin 76 was transformed into hibaene in five steps. In contrast to 74 and 75, on reaction with methylmagnesium bromide vinylcarbinols 79 and 80 exclusively afforded the terminally methylated olefins 81 and 82, respectively.²⁸⁾



Pimaradiene 83 was converted into D-norsteroids 84 and 85.²⁹) Intramolecular C-alkylation of a diazoketone 86 derived from pimaradiene 83 gave a mixture (60:40) of the two isomeric ketones 87 and 88. On the other hand, diazoketone 89 gave 90 as the sole product.³⁰) Solvolysis of virescenol B 19-tosylate (91) in DMSO gave 92 and 93.³¹)

(390)

$$A_{c}O + H O + H$$

Boron trifluoride etherate catalyzed rearrangement of methyl $8,14\beta$ -epoxysandara-copimarate (94) afforded a mixture of 95 (33%), 96 (9%), and 97 (40%).³²⁾ The cleavage of methyl pimarate $8,14\alpha$ -epoxide (98) with a variety of acidic reagents was studied and a suggestion that the pimaranes are not the biogenetic precursors of the tetracyclic diterpenes was provided.³³⁾ Compound 99 was comparatively efficiently incorporated into rosenonolactone (100).³⁴⁾

VI. ABIETANE DERIVATIVES

Abietane

In organic matter in the sediments in Hiro Bay near outlet of pulp mill waste

water, retene (101), dehydroabietane (102), and dehydroabietic acid (103) were identified by GC-MS.³⁵⁾ Dehydroabietic acid (103) and its homolog 104 were found in the lignite extract.³⁶⁾

Two new compounds, pisiferic acid (105)³⁷⁾ and premnolal (106),³⁸⁾ were isolated from *Chamaecyparis pisifera* and *Premna latifolia*, respectively. The isolation of a new diterpenoid ibozol (107) together with the known 7α -hydroxyroyleanone (108) from *Iboza riparia* was reported.³⁹⁾

Three new diterpenoids, 109 (nellionol), 110, and 111, were isolated from *Premna latifolia*.⁴⁰⁾ Methylenetanshinquinone (112) was isolated from *Salvia miltiorrhiza* along with the known tanshinones. Its sulfonate derivative 113, which was useful in treatment of angina pectoris, was prepared.⁴¹⁾ From *Plectranthus edulis*, edulon A (114) which may be regarded as a 4,5-seco-abietane derivative was isolated.⁴²⁾

(109) (110)
$$R = H$$
 (112) $R^1 = H$; $R^2 = CH_2$ (114) (114) $R = OH$ (113) $R^1 = SO_3 Na; R^2 = Me_2$

The carbon-13 NMR spectra of abietic acid and its methyl ester were published.⁴³⁾ Kinetics⁴⁴⁾ of nucleophilic substitution of chlorine in 3-chloro-2-hydroxypropyl dehydroabietate (115) by secondary amines and photolysis⁴⁵⁾ of N-acylimidazoles (116 and 117) were investigated. The irradiation of 116 yielded compounds 118 and 119.

Syntheses of taxodione (121), royleanone (122), and their analogs (123 and 124) from the 12-hydroxy ester 120 were accomplished.⁴⁶⁾ Epoxidation studies with compound 125 and synthesis of 126 with C-ring functionality and stereochemistry corresponding to triptonide (127) were reported.⁴⁷⁾

(118)
$$R = \frac{1}{N}$$
(118) $R = \frac{1}{N}$
(117)
(116) $R = -N = \frac{1}{N}$
(117)
(116) $R = -N = \frac{1}{N}$
(120)
(121) $R = Me$
(122) $R = Me$
(123) $R = Co_2 Me$
(124) $R = Co_2 Me$
(125)
(126)
(127)

Epoxides 129 related to triptolide (130) was prepared from levopimaric acid (128) according to the sequence shown in Chart 1.48)

The N-(alkylaminomethyl)imides of maleopimaric acid, 131–134, isolated as their amine salts, were prepared in 79~90% yields by treatment of 135 with the corresponding amine.⁴⁹⁾ Ozonolysis of phenolic dehydroabietic acid derivatives 136–140 was investigated.⁵⁰⁾

E. Fujita, K. Fuji, Y. Nagao, and M. Node

(131)
$$R = -N \bigcirc 0$$

(132) $R = N \in t_2$
(133) $R = -N \bigcirc$
(134) $R = N (cH_2cH_2OH)_2$
(135) $R = OH$
(137) $R' = R^2 = H$; $R^3 = OH$
(138) $R' = R^2 = H$; $R^3 = OH$
(139) $R = H$
(140) $R = OH$

Total syntheses of (\pm) -ferruginol (141) and (\pm) -hinokinone (142) were achieved, in which the tricyclic ring system was assembled in the order of C \rightarrow BC \rightarrow ABC. The outline is shown in Chart 2.⁵¹⁾

In a review on chemical constituents of the Celastraceae family isolated up to 1 March 1978, some abietane type diterpenoids were discussed.⁵²⁾

VII. TOTARANE DERIVATIVES

The Chemistry on Diterpenoids in 1978. Part-II

The total syntheses of natural tricyclic diterpenes, (—)-dispermone (143), (+)-dispermol (144), and (+)-maytenoquinone (145), were achieved starting from (R)-(—)- α -cyclocitral (146) and the proposed structure 147 for dispermol was revised to 12-hydroxy-13-methoxytotara-8,11,13-triene (144).⁵³⁾

In the foregoing review,⁵²⁾ maytenoquinone, dispermol, and dispermone were described.

VIII. CASSANE DERIVATIVES

Cassane

No papers have been published on the title topics in this period.

IX. KAURANE DERIVATIVES

Kaurane

Two new diterpene acetates were isolated from *Jungermannia infusca*. Their respective structures were determined to be *ent*-15-oxokauran-11a-yl acetate (148) and *ent*-15-oxokaur-16-en-11a-yl acetate (149) based on the chemical and spectral evidence.⁵⁴⁾ Isolations of new diterpenes, 150 from *Bedfordia salicina*, ⁵⁵⁾ and 151 and 152 from *Lagases rigida*⁵⁶⁾ were published.

Three new diterpenes have been isolated from South African *Helichrysum* species. 69 Thus, compounds 153 and 154 from *H. cooperi*, compounds 152, 153, and 154 from *H. aureum var. monocephalum* have been isolated together with known diterpenoids kaurenoic acid and compound 155.

From *Montanoa pteropoda*, three new diterpenes (156–158) have been isolated together with a number of known kaurene type diterpenoids.⁵⁷⁾

$$R^{2}$$
 $Co_{2}R^{3}$
 $Co_{2}H$
 $Co_{2}H$
 $Co_{2}H$
 $Co_{2}H$
 $Co_{2}H$
 $Co_{2}H$
 $Co_{2}H$
 $Co_{2}H$
 $Co_{3}H$
 $Co_{4}H$
 $Co_{5}H$
 Co

Isolations of the known kaurene type diterpenes (159 from *Sciadocephala schultze-rhonhofiae* and kaurenoic acid and alcohol 160 from *Critonia daleoides*) were reported.⁷⁾ From *Stevia ovata*, there were isolated five known kaurene type ester-glucosides, paniculosides-I(161), -II(162), -III(163), -IV(164), and V(165).⁵⁸⁾

HO

H

OH

H

CO2Gluc

(159)

(160)

(161)

$$R' = H; R^2 = OH$$

(162)

 $R' = R^2 = OH$

(163)

(164)

(165)

 $R' = H; R^2 = OGluc$

(163)

(164)

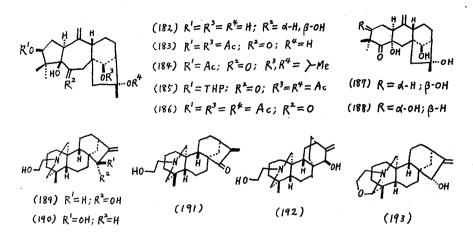
From *Pteris plumbaea*, five new diterpenes (166–170) were isolated together with the known *ent*-kaurane-derivatives 171 and 172.⁵⁹⁾ A new atractyligenin glucoside was isolated from green coffee-beans. Its structure was determined by spectral data and chemical reactions to be 173.⁶⁰⁾ A new diterpene glucoside, grayanoside A (174) was isolated from *Leucothoe grayana*.⁶¹⁾

The Chemistry on Diterpenoids in 1978. Part-II

Carbon-13 NMR spectroscopy of atisine and veatchine-type C_{20} -diterpenoid alkaloids from Aconitum and Garrya species was reported. In this paper, it was indicated that the C-20 epimers of atisin did not exist in an equilibrium mixture in solution and were not interconvertible via a zwitterion as reported earlier.⁶²⁾ 15 α -Fluorokaurenoic acid (175) was prepared and fed to fermentations of Gibberella fujikuroi; the products have been shown to include 15 α -fluoro-7 β -hydroxykaurenolide (176), 15 α -fluorofujenal (177), and gibberellin derivatives (178–180).⁶³⁾ In a review on "transportcatalyst in biomembrane illustrated by example of ADP, ATP-carriers of mitochondria", carboxy-atractylate 181 was cited.⁶⁴⁾

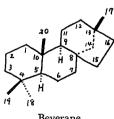
Acyloin rearrangement of 6-dehydro derivatives of grayanotoxin-II (182) was investigated and compounds (183–186) were converted to a 6/6 membered ring system (187 and 188) in alkaline media.⁶⁵⁾

A simple method using active MnO₂ for converting the -N-CH₂-CH₂-OH group-containing alkaloid derivatives (189–192) into their iso-oxazolidine ring-containing alkaloids (e.g. garryine 193) was reported.⁶⁶⁾



A simple and efficient method for converting the oxazolidine-ring containing alkaloids, e.g., ovatine (194), garryfoline (195), and veatchine (196), into their corresponding imine derivatives was published.⁶⁷⁾ (See Chart 3.)

X. BEYERANE DERIVATIVES



Beyerane

From Montanoa pteropoda, the known beyerane-type diterpene 197 was isolated together with kaurene-type diterpenes.⁵⁷⁾ Isolation of two new secobeyerene diterpenes (198 and 199) from Beyeria calycina was reported. This paper describes also a feeding experiment indicating quite distinct pathway to the seco acid 200 and the 6hydroxydihydro analog 198.

The photolysis of 3-oxo-beyerene derivative 201 occurred with retention of configuration of C-4 to give the 4S-3,4-seco acid 202. In this phototransformation, the C-2 axial hydrogen is transferred preferentially to C-4.69)

$$0 \xrightarrow{H} H O_1 \xrightarrow{C} H \xrightarrow{H} H O_2 \xrightarrow{C} H_2 O H$$

$$(201) \qquad (202)$$

XI. GIBBERELLANE DERIVATIVES

Two new gibberellins A_{50} (203) and A_{52} (204) were isolated from seeds of *Lagenaria leucantha var. clavata*. A novel gibberellin glucoside was isolated from immature seed of *Dolichos lablab* and its structure was determined to be 3-O- β -D-glucopyranosyl gibberellin A_1 (205).⁷¹⁾

The inhibitory effects of 1-alkylimidazoles on the gibberellin biosynthesis of Gibberella fujikuroi were reported. Microbiological production of fluorogibberellins (178–180) was reported. 7-Homogibberellin A₃ (207) was partially synthesized from gibberellin A₃ (206). Thus, the latter was treated with (COCl)₂ and CH₂N₂ to give a diazoketone, which was photochemically rearranged and then hydrolyzed with NaOMe. 73)

Stereospecific total synthesis of gibberellic acid (206) was achieved by Harvard team.^{74,75)} The synthetic pathway is shown in Chart 4.

Chart 4

XII. ATISANE DERIVATIVES

From the roots of Margotia gummifera, a new diterpenoid, gummiferolic acid (208) was isolated in very high yield (2% of the dry plant), together with the known ent-kaur-16-en-19-oic acid. (76) A satisfactory method for quantitative determination of a series of diterpene alkaloids in Japanese aconite roots was developed, using the combination of dual wave length thin-layer chromatography scan and gas chromatography. (77) Carbon-13 NMR spectra of some ent-atisene derivatives (209–214) were reported.

The Chemistry on Diterpenoids in 1978. Part-II

$$(209) R' = H_2; R^2 = Me$$

$$(210) R' = H_2; R^2 = CH_2OH$$

$$(211) R' = H_2; R^2 = CO_2Me$$

$$(212) R' = O; R^2 = CO_2Me$$

$$(208)$$

$$(213) R' = d + H_2 + OA_c; R^2 = CO_2Me$$

$$(214)$$

Carbon-13 NMR spectroscopy of atisine-type alkaloids from *Aconitum* species⁶²⁾ and X-ray crystallography of four C₂₀-diterpenoid alkaloids (192, 196, 215, and 216)⁷⁹⁾ were investigated. In these papers, epimerization and isomerization in their alkaloids were discussed. A simple and efficient method for the degradation of the oxazolidine ring of C₂₀-diterpene alkaloids [216, 217—218] was published,⁶⁷⁾ as mentioned above.

Dihydroatisine (192) was converted to isoatisine (216) using active MnO₂.⁶⁶⁾ An aconite alkaloid, kobusine (219) was converted to a C₁₄-C₂₀ bond cleaved derivative 221 by a novel fragmentation reaction *via* a chloramine 220. Furthermore, the C₁₄-C₂₀ bond regeneration in compound 221 was also accomplished by intramolecular Grignard type reaction.⁸⁰⁾ (See Cahrt 5.)

XIII. ACONANE DERIVATIVES

A method for the quantitative analysis of a series of diterpene alkaloids in Japanese aconite roots was developed.⁷⁷ The structure of gigactonine (222) isolated as a new base from *Aconitum giga* was determined.⁸¹ The diterpene alkaloids of *Delphinium brownii* were re-examined. As the result, it was found that, in addition to 223 and 224,

browniine 14-O-acetate (225) occurs in the bases extracted from aerial portions of the plant.⁸²⁾ Ambiguine (226) and dihydroajaconine, two new minor diterpene alkaloids obtained from *Consolida ambigua* were characterized by ¹³C NMR.⁸³⁾ The structure of ranaconitine, a new alkaloid of *Aconitum ranunculaefolium*, was reported as 227.⁸⁴⁾

HO A H H OH OH OH OH OH OH OH (224)
$$R^1 = R^2 = H$$
; $R^2 = Me$ OH OH OH OH (224) $R^1 = R^2 = H$; $R^2 = Me$ OH OH (225) $R^1 = R^2 = Me$ OH OH OH OH (225) $R^2 = R^3 = Me$ OH OH (227)

XIV. TAXANE DERIVATIVES

Taxane

No reports have been reported on the title topics in this period.

XV. THE OTHERS

The structures of the previously reported irienol A (228) and iriediol (229) were further refined as shown. The structures of irieol B (230), C (231), D (232), E (233), F (234), G (235), and irieol (236) were reported.⁸⁵⁾ They are new dibromo-diterpenoids of a unique skeletal class from the red seaweed *Laurencia irieii*.

Br
$$\beta r$$
 βr β

Two new cembranoid lactones, flexibilide (237) and dihydroflexibilide (238), were isolated from an Australian collection of the soft coral *Sinularia flexibilis*. The structure and relative configuration of flexibilide (237) was determined by single-crystal X-ray diffraction method.⁸⁶⁾ The structure of a novel bicyclic diterpene alcohol obtained from an unknown species of soft coral was reported and shown to be 239 by the chemical and crystallographic study.⁸⁷⁾ The structure of a novel nor-cembranoid

diterpene isolated from the soft coral Sinularia leptoclados was determined as 240 by X-ray analysis.⁸⁸⁾

$$(237) \quad R = CH_2$$

$$(238) \quad R = Me, H$$

$$(239) \quad (240)$$

The isolations of epoxyisoneocembrene-A (241) from the soft coral *Sinularia grayi* and of isoneocembrene-A (242) from the soft coral *Sarcophyton ehrenbergi* were reported. Their structures were demonstrated by chemical and spectroscopic means.⁸⁹⁾

The structure and absolute configuration of verticillol (243), a macrocyclic diterpene alcohol from the wood of *Sciadopitys verticillata*, were assigned on the basis of physicochemical studies. ⁹⁰⁾ Four new diterpenes which were named cyafrin A_4 (244), cyafrin B_4 (245), allocyafrin B_4 (246), and cyafrin A_5 (247) were isolated together with the known compounds from the liquid culture of the bird's nest fungus *Cyathus africanus* Brodie. ⁹¹⁾ The Carbon-13 NMR spectra of cyathin A_3 (248) and several related compounds were measured and the chemical shifts were assigned. ⁹²⁾

The isolation and identification of aphidicolin (249) contained in the culture filtrate of the fungus *Harziella entomophilla* as well as the biological activity of this compound as a root growth inhibitor were reported.⁹³⁾ This compound prevented mitotic cell division by interfering with the activity of DNA polymerase-α.⁹⁴⁾ Phorbol derivative 250 was described in the research news about tumor promoters.⁹⁵⁾ In a review on cocarcinogen, several highly irritant diterpenes, phorbol (251), 16-hydroxyphorbol (252), 4-deoxy-16-hydroxyphorbol (253), and their esters were discussed in detail.⁹⁶⁾

Two minor diterpenes, 254 and 255, were isolated from Euphorbia latex.⁹⁷⁾ Three new diterpenes, coleonol-B, -C, and deoxycoleonol were isolated from Coleus forskohlii.⁹⁸⁾ A new diterpene analog, β -springene (256), was isolated from the dorsal gland of the springbok Antidorcas marsupialis.⁹⁹⁾ Aplidiasphingosine (257), an antimicrobial and antitumor terpenoid, was isolated from an Aplidium species.¹⁰⁰⁾ Fuscol (258), a new elemene-type diterpene alcohol, was isolated from the gorgonian Eunicea fusca.¹⁰¹⁾

The isolation of four new diterpenes, epoxydilophone (259), dilopholone (260), epiacetoxydilopholone (261), and acetoxydilopholone (262), from the brown alga *Dilophus prolificans* was reported. Two new diterpenes, xeniolide-A (263) and -B (264), were isolated from the soft coral *Xenia macrospiculata*. 103)

$$(260) \quad R' = R^2 = H$$

$$(261) \quad R' = 0Ac; \quad R^2 = H$$

$$(262) \quad R' = H; \quad R^2 = 0Ac$$

The structures 265 and 266 were assigned to the compounds isolated from the frontal glands of *Nasutitermes costalis* soldiers. The structure and absolute configuration of a new dibromo-diterpene, angasiol, isolated from the South Pacific Ocean sea hare *Aplysia angasi* was determined as 267 by X-ray analysis. Three new mono-

acetates of diterpene diols were isolated from *Mylia verrucosa* and the structures were determined as 268, 269, and 270 on the basis of chemical and spectral evidence. ¹⁰⁶⁾

$$R$$
 (265) $R = α-OH$, $β-OH$ (267) (268) (269) $R' = Ac$, $R^2 = H$ (270) $R' = H$, $R^2 = Ac$

Striatins A, B, and C, novel diterpenoid antibiotics, were isolated from *Cyanthus striatus*. Striatin A (271) was shown by X-ray analysis to contain a cyathin skeleton triple linked to a pentose unit. This result allowed the assignment of structures to the closely related striatins B (272) and C (273).¹⁰⁷⁾ Mechanism of geranylgeranyl pyrophosphate cyclization in fusicoccin (274) biosynthesis was discussed by use of deuterium as a tracer with ¹³C NMR spectroscopy.¹⁰⁸⁾

(±)-Ligantrol (276), isolated from *Liatris elegans*, was synthesized from the hypothetical precursor, 18-hydroxygeranylnerol (275) which was an isomer of 18-hydroxygeranylgeraniol isolated from *Croton sublyratus* as an antigastric ulcer principle. 109) The synthetic route is shown in Chart 6.

A synthesis of (\pm) -norisoambreinolide (277) and (\pm) -isoambrox (278) was reported. Syntheses of (\pm) -2,6-dimethyl-10-(p-tolyl)undeca-2,6-(E)-diene (279), the diterpene of *Salvia dorisiana* were reported. The acid-catalyzed cyclization

of diazomethyl 5,8-dimethoxy-1,2,3,4-tetrahydro-2-naphthyl ketone (280) into compound 281 or 282 was reported.¹¹³⁾

A rearrangement of bicyclo [2.2.2] octene precursors 283 and 284 into bridgehead-substituted bicyclo [3.2.1] octene derivatives 285 and 286 was investigated, which may be available for construction of the C-D ring system of gibberellic acids, beyerenes, and grayanotoxins.¹¹⁴⁾ The mechanistic route is shown in Chart 7.

(283)
$$\stackrel{\text{R}}{R}$$
 $\stackrel{\text{Nu}}{\longrightarrow}$ $\stackrel{\text{Nu}}{R}$ $\stackrel{\text{Nu}}{\longrightarrow}$ $\stackrel{\text{Nu}}{$

The naturally occurring cembrenoids, 287, 288, and 289 were successfully synthesized, demonstrating unequivocally the assigned structure.¹¹⁵⁾

ADDENDA

Refluxing the compound 290 having the skeleton of dolabradiene with formic acid for 30 hours followed by chromatography gave 291–293 in 60, 10, and 11.4% yields, respectively.¹¹⁶⁾

$$(290) \qquad (291) \qquad (292) \qquad (293) \qquad (293) \qquad (294)$$

$$(294) \qquad (294) \qquad (294) \qquad (294)$$

$$(297) \qquad R^{1} = H; \quad R^{2} = 0H \qquad (300)$$

$$(297) \qquad R^{1} = H; \quad R^{2} = 0H \qquad (300)$$

$$(297) \qquad R^{1}, \quad R^{2} = 0$$

$$(301) \qquad R^{1} = H; \quad (300) \qquad (297) \qquad R^{1}, \quad R^{2} = 0$$

The structure of teucrin H₂ (294), isolated from *Teucrium hyrcanicum*, was confirmed by NMR, IR, UV, and mass spectroscopy and by reduction, hydrogenation, and oxidation products.¹¹⁷⁾ Two new diterpenoid dilactones related to nagilactone A, isolated from *Podocarpus nagi*, were shown to have structures 295 and 296 according to their NMR spectra and chemical correlations.¹¹⁸⁾ The mass spectra of gibberellins 297–302 were reported. All the compounds showed the same skeletal fragmentation.¹¹⁹⁾ Diterpene synthon 303 was prepared by cyclocondensation of cyclohexanone pyrrolidine enamine with CH₂=CHCOCH₂CO₂Et, hydrogenation of the octahydronaphthalenone 304, and methylation of the naphthalenol 305 with methyl iodide. Compound 305 was reduced to the corresponding alcohol, the structure of which was determined by X-ray crystal structure analysis.¹²⁰⁾ A new diterpene, coleosol, was isolated from *Coleus forskohlii*.¹²¹⁾

Cyafrin A_4 (244), cyafrin B_4 (245), allocyafrin B_4 (246), and cyafrin A_5 (247) isolated from *Cyathus africanus*, a bird's nest fungus, were described again, and metabolites of other bird's nest fungi were reviewed. Mass spectral fragmentation patterns were determined for teucrin H_1 - H_4 (58–61) isolated from *Teucrium hyrcanicum*. 123)

REFERENCES

- (1) For 1978 Part-I, see E. Fujita, K. Fuji, Y. Nagao, and M. Node, Bull. Inst. Chem. Res., Kyoto Univ., 57, 260 (1979).
- (2) B. R. Davis and S. J. Johnson, J. C. S. Chem. Comm., 614 (1978).
- (3) U. R. Ghatak, S. K. Alam, and J. K. Ray, J. Org. Chem., 43, 4598 (1978).
- (4) S. Ghosh, S. Chakrabarty, and U. R. Ghatak, Indian J. Chem., Sect. B, 16B, 723 (1978). (Chem. Abstr., 90, 55128 m [1979].)
- (5) M. Mondal and S. Guha, J. C. S. Perkin Trans. II, 968 (1978).

E. Fujita, K. Fuji, Y. Nagao, and M. Node

- (6) F. Bohlmann, C. Zdero, E. Hoffmann, P. K. Mahanta, and W. Dorner, Phytochemistry, 17, 1917 (1978).
- (7) F. Bohlmann, P. Zitzkowski, A. Suwita, and L. Fiedler, ibid., 17, 2101 (1978).
- (8) F. Bohlmann and U. Fritz, ibid., 17, 1769 (1978).
- (9) K. Ohta and T. Nawamaki, Agr. Biol. Chem., 42, 1957 (1978).
- (10) G. Savona, F. Piozzi, and J. R. Hanson, Phytochemistry, 17, 2132 (1978).
- (11) E. Denole and P. Enggist, Helv. Chim. Acta, 61, 2318 (1978).
- (12) P. K. Grant and H. T. L. Liau, Austral. J. Chem., 31, 1777 (1978).
- (13) P. K. Grant and C. K. Lai, ibid., 31, 1785 (1978).
- (14) P. K. Grant and H. T. L. Liau, ibid., 31, 1791 (1978).
- (15) J. De Pascual Teresa, J. G. Urones, and J. Agustin Herrero, An Quim., 74, 476 (1978). (Chem. Abstr., 90, 104154 c [1979].)
- (16) M. L. Bittner, V. Zabel, W. B. Smith, and W. H. Watson, Phytochemistry, 17, 1797 (1978).
- (17) A. Chatterjee, A. Banerjee, and F. Bohlmann, ibid., 17, 1777 (1978).
- (18) W. Wagner, R. Seitz, H. Lotter, and W. Herz, J. Org. Chem., 43, 3339 (1978).
- (19) G. B. Oganesyan and V. A. Mnatsakanyan, Arm. Khim. Zh., 31, 768 (1978). (Chem. Abstr., 90, 168775 p [1979].)
- (20) E. Gács-Baitz, L. Radics, G.B. Oganessian, and V.A. Mnatsakanyan, Phytochemistry, 17, 1967 (1978).
- (21) M. A. de Alvarenga, H. E. Gottlieb, O. R. Gottlieb, M. T. Magalhães, and V. O. de Silva, ibid., 17, 1773 (1978).
- (22) P. Y. Malakov, G. Y. Papanov, N. M. Mollov, and S. L. Spassov, Z. Naturforsch., 33b, 789 (1978).
- (23) P. Y. Malakov, G. Y. Papanov, N. M. Mollov, and S. L. Spassov, ibid., 33b, 1142 (1978).
- (24) N. Harada and H. Uda, J. Am. Chem. Soc., 100, 8022 (1978).
- (25) D. J. Goldsmith, G. Srouji, and C. Kwong, J. Org. Chem., 43, 3182 (1978).
- (26) A. Ardon-Jimenez and T. G. Halsall, J. C. S. Perkin Trans. I, 1461 (1978).
- (27) C. B. Rao and T. N. Rao, Curr. Sci., 47, 577 (1978). (Chem. Abstr., 89, 215599a [1978].)
- (28) B. L. Buckwalter, I. R. Burfitt, H. Felkin, M. Joly-Goudket, K. Naemura, M. F. Salomon, E. Wenkert, and P. M. Wovkulich, J. Am. Chem. Soc., 100, 6445 (1978).
- (29) P. Ceccherelli, M. Tingoli, M. Curini, and R. Pellicciari, Tetrahedron Lett., 3869 (1978).
- (30) P. Ceccherelli, M. Tingoli, M. Curini, and R. Pellicciari, ibid., 4959 (1978).
- (31) P. Ceccherelli, M. Curini, R. Pellicciari, G. V. Buddeley, M. S. Raju, and E. Wenkert, J. Org. Chem., 43, 4244 (1978).
- (32) B. Delmond, M. Taran, and J. Valade, Tetrahedron Lett., 4791 (1978).
- (33) J. W. ApSimon and S. F. Hall, Can. J. Chem., 56, 2156 (1978).
- (34) B. Dockerill and J. R. Hanson, Phytochemistry, 17, 1119 (1978).
- (35) Y. Yamaoka, Nippon Kagaku Kaishi, 1706 (1978).
- (36) R. Hayatsu, R. E. Winans, R. G. Scott, L. P. Moore, and M. H. Studier, Nature, 275, 116 (1978).
- (37) H. Fukui, K. Koshimizu, and H. Egawa, Agr. Biol. Chem., 42, 1419 (1978).
- (38) C. B. Rao and T. N. Rao, Curr. Sci., 47, 498 (1978). (Chem. Abstr., 89, 180185h [1978].)
- (39) R. Zelnik, E. Rabenhorst, A. K. Matida, H. E. Gottlieb, D. Lavie, and S. Panizza, Phytochemistry, 17, 1795 (1978).
- (40) C. B. Rao, T. N. Rao, and E. K. S. Vijayakumar, Curr. Sci., 47, 455 (1978). (Chem. Abstr., 89, 163778m [1978].)
- (41) M.-K. Chien, P.-T. Young, W.-H. Ku, Z.-X. Chen, H. T. Chen, and H.-C. Yeh, Hua Hsueh Hsueh Pao, 36, 199 (1978). (Chem. Abstr., 90, 138047k [1979].)
- (42) G. Buchbauer, P. Rüedi, and C. H. Eugster, Helv. Chim. Acta, 61, 1969 (1978).
- (43) W. B. Smith, Org. Magn. Reson., 11, 427 (1978). (Chem. Abstr., 90, 138046j [1979].)
- (44) A. Tardenaka and J. Zandersons, Latv. PSR Zinat. Akad. Vestis, Kim. Ser., 606 (1978). (Chem. Abstr., 90, 55131g [1979].)
- (45) S. Iwasaki, Helv. Chim. Acta, 61, 2843 (1978).
- (46) Y. Ohtsuka and (the late) A. Tahara, Chem. Pharm. Bull. 26, 2007 (1978).
- (47) D. M. Frieze, G. A. Berchtold, and J. F. Blount, Tetrahedron Lett., 4607 (1978).
- (48) H. Koike and T. Tokoroyama, ibid., 4531 (1978).
- (49) D. Svikle and A. Prikule, Latv. PSR Zinat. Akad. Vestis, Kim. Ser., 593 (1978). (Chem. Abstr., 90, 72363b [1979].)
- (50) H. Akita and T. Oishi, Tetrahedron Lett., 3733 (1978).

The Chemistry on Diterpenoids in 1978. Part-II

- (51) D. L. Snitman, R. J. Himmelsbach, and D. S. Watt, J. Org. Chem., 43, 4758 (1978).
- (52) R. Brüning and H. Wagner, Phytochemistry, 17, 1821 (1978).
- (53) T. Matsumoto and S. Usui, Chemistry Lett., 897 (1978).
- (54) A. Matsuo, S. Uto, J. Kodama, M. Nakayama, and S. Hayashi, Nippon Kagaku Kaishi, 1680 (1978).
- (55) F. Bohlmann and N. Le Van, Phytochemistry, 17, 1173 (1978).
- (56) F. Bohlmann and J. Jakuponic, ibid., 17, 1677 (1978).
- (57) F. Bohlmann and N. Le Van, ibid., 17, 1957 (1978).
- (58) N. Kaneda, H. Kohda, K. Yamasaki, O. Tanaka, and K. Nishi, Chem. Pharm. Bull., 26, 2266 (1978).
- (59) N. Tanaka, K. Nakatani, T. Murakami, Y. Saiki, and C.-M. Chen, ibid., 26, 3260 (1978).
- (60) H. Richter and G. Spiteller, Chem. Ber., 111, 3506 (1978).
- (61) J. Sakakibara, N. Shirai, T. Kaiya, and H. Nakata, Phytochemistry, 17, 1672 (1978).
- (62) N. V. Mody and S. W. Pelletier, Tetrahedron, 34, 2421 (1978).
- (63) B. E. Cross and A. Erasmuson, J. C. S. Chem. Comm., 1013 (1978).
- (64) M. Klingenberg, Naturwissenschaften, 65, 456 (1978).
- (65) R. Iriye, Agr. Biol. Chem., 42, 1495 (1978).
- (66) S. W. Pelletier, N. V. Mody, and J. Bhattacharyya, Tetrahedron Lett., 5187 (1978).
- (67) N. V. Mody and S. W. Pelletier, ibid., 3313 (1978).
- (68) E. L. Ghisalberti, P. R. Jefferies, and M. A. Sefton, Phytochemistry, 17, 1961 (1978).
- (69) E. L. Ghisalberti, P. R. Jefferies, and M. A. Sefton, Tetrahedron, 34, 3337 (1978).
- (70) H. Fukui, K. Koshimizu, and R. Nemori, Agr. Biol. Chem., 42, 1571 (1978).
- (71) T. Yokota, S. Kobayashi, H. Yamane, and N. Takahashi, ibid., 42, 1811 (1978).
- (72) K. Wada, ibid., 42, 2411 (1978).
- (73) E. P. Serebryakov, M. Lischewskii, and G. Adam, Izv. Akad. Nauk SSSR, Ser. Kim., 2181 (1978). (Chem. Abstr., 90, 23309e [1979].)
- (74) E. J. Corey, R. L. Danheiser, S. Chandrasekaran, P. Siret, G. E. Keck, and J.-L. Gras, J. Am. Chem. Soc., 100, 8031 (1978).
- (75) E. J. Corey, R. L. Danheiser, S. Chandrasekaran, G. E. Keck, B. Gopalan, S. D. Larsen, P. Siret, and J.-L. Gras, ibid., 100, 8034 (1978).
- (76) M. Pinar, B. Rodriguez, and A. Alemany, Phytochemistry, 17, 1637 (1978).
- (77) F. Kurosaki, T. Yatsunami, T. Okamoto, and Y. Ichinohe, Yakugaku Zasshi, 98, 1267 (1978).
- (78) B. Rodriguez, A. Alemany, and M. Pinar, Tetrahedron Lett., 3069 (1978).
- (79) S. W. Pelletier, W. H. De Camp, and N. V. Mody, J. Am. Chem. Soc., 100, 7976 (1978).
- (80) T. Yatsunami, S. Furuya, and T. Okamoto, Chem. Pharm. Bull., 26, 3199 (1978).
- (81) S. Sakai, N. Shinma, S. Hasegawa, and T. Okamoto, Yakugaku Zasshi, 98, 1376 (1978).
- (82) V. N. Aiyar, M. Benn, Y. Y. Huang, J. M. Jacyno, and A. J. Jones, Phytochemistry, 17, 1453 (1978).
- (83) S. W. Pelletier, R. S. Sawhney, and N. V. Mody, Heterocycles, 9, 1241 (1978). (Chem. Abstr., 89, 215615c [1978].)
- (84) S. W. Pelletier, N. V. Mody, A. P. Venkov, and N. M. Mollov, Tetrahedron Lett., 5045 (1978).
- (85) B. M. Howard and W. Fenical, J. Org. Chem., 34, 4401 (1978).
- (86) R. Kazlauskas, P. T. Murphy, R. J. Wells, P. Schönholzer, and J. C. Coll, Austral. J. Chem., 31, 1817 (1978).
- (87) B. F. Bowden, J. C. Coll, S. J. Mitchell, G. J. Stokie, and J. F. Blount, ibid., 31, 2039 (1978).
- (88) B. F. Bowden, J. C. Coll, S. J. Mitchell, J. Mulder, and G. J. Stokie, ibid., 31, 2049 (1978).
- (89) B. F. Bowden, J. C. Coll, W. Hicks, R. Kazlauskas, and S. J. Mitchell, ibid., 31, 2707 (1978).
- (90) B. Karlsson, A.-M. Pilotti, A.-C. Söderholm, T. Norin, S. Sundin, and M. Sumimoto, *Tetrahedron*, 34, 2349 (1978).
- (91) W. A. Ayer, T. Yoshida, and D. M. J. van Schie, Can. J. Chem., 56, 2113 (1978).
- (92) W. A. Ayer, T. T. Nakashima, and D. E. Ward, ibid., 56, 2197 (1978).
- (93) K. Kawada, Y. Kimura, K. Katagiri, A. Suzuki, and S. Tamura, Agr. Biol. Chem., 42, 1611 (1978).
- (94) S. Ikegami, T. Taguchi, and M. Ohashi, Nature, 275, 458 (1978).
- (95) J. L. Marx, Science, 201, 515 (1978).
- (96) E. Hecker, Naturwissenschaften, 65, 640 (1978).
- (97) R. J. Schmidt and F. J. Evans, Phytochemistry, 17, 1436 (1978).
- (98) J. S. Tandon, P. K. Jauhari, R. S. Singh, and M. M. Dhar, Indian J. Chem., Sect. B, 16, 341 (1978).
- (99) V. B. Burger, M. le Roux, H. S. C. Spies, V. Truter, and R. C. Bigalke, Tetrahedron Lett., 5221 (1978).
- (100) G. T. Carter and K. L. Rinehart, Jr., J. Am. Chem. Soc., 100, 7441 (1978).

E. FUJITA, K. FUJI, Y. NAGAO, and M. NODE

- (101) Y. Gopichand and E. J. Schmitz, Tetrahedron Lett., 3641 (1978).
- (102) R. Kazlauskas, P. T. Murphy, R. J. Wells, and J. F. Blount, ibid., 4155 (1978).
- (103) Y. Kashman and A. Groweiss, ibid., 4833 (1978).
- (104) J. Vrkoč, M. Buděšínský, and P. Sedmera, Collect. Czech. Chem. Comm., 43, 2478 (1978).
- (105) G. R. Pettit, C. L. Herald, J. J. Finck, L. D. Vanell, P. Brown, and D. Gust, J. Org. Chem., 43, 4685 (1978).
- (106) S. Hayashi, A. Matsu, H. Nozaki, M. Nakayama, D. Takaoka, and M. Hiroi, *Chemistry Lett.*, 953 (1978).
- (107) H.-J. Hecht, G. Höfle, W. Steglich, T. Anke, and F. Oberwinkler, J. C. S. Chem. Comm., 665 (1978).
- (108) A. Banerji, R. Hunter, G. Mellows, K. Sim, and D. H. R. Barton, ibid., 843 (1978).
- (109) S. Takahashi, E. Kitazawa, and A. Ogiso, Chem. Pharm. Bull., 26, 3416 (1978).
- (110) S. Torii, K. Uneyama, and H. Ichimura, J. Org. Chem., 43, 4680 (1978).
- (111) T. K. John and G. S. K. Rao, Proc. Indian Acad. Sci., Sect. A, 87A, 235 (1978). (Chem. Abstr., 90, 23308d [1979].)
- (112) O. P. Vig, I. R. Trehan, and R. Kumar, Indian J. Chem., Sect. B, 16B, 773 (1978). (Chem. Abstr., 90, 168773m [1979].)
- (113) D. W. Johnson and L. N. Mander, Austral. J. Chem., 31, 1561 (1978).
- (114) S. A. Monti, S.-C. Chen, Y.-L. Yang, S.-S. Yuan, and O. P. Bourgeois, J. Org. Chem., 43, 4062 (1978).
- (115) M. Suzuki, A. Shimada, and T. Kato, Chemistry Lett., 759 (1978).
- (116) A. K. Banerjee, E. Boliver, and M. Narvaez, Gazz. Chim. Ital., 108, 505 (1978). (Chem. Abstr., 90, 187160a [1979].)
- (117) G. B. Oganesyan and V. A. Mnatsakanyan, Arm. Khim. Zh., 31, 776 (1978). (Chem. Abstr., 91, 20772k [1979].)
- (118) Y. Hayashi, T. Matsumoto, and T. Sakan, Heterocycles, 10, 123 (1978). (Chem. Abstr., 91, 20788v [1979].)
- (119) D. Voigt, G. Adam, J. Schmidt, and P. Franke, Org. Mass Spectrom., 13, 599 (1978). (Chem. Abstr., 90, 204286r [1979].)
- (120) F. Orsini, F. Pelizzoni, and R. Destro, Gazz. Chim. Ital., 108, 639 (1978). (Chem. Abstr., 91, 20793t [1979].)
- (121) P. K. Jauhari, S. B. Katti, J. S. Tandon, and M. M. Dhar, Indian J. Chem., Sec. B, 16, 1055 (1978).
- (122) W. A. Ayer, L. M. Browne, S. Fermandez, D. E. Ward, and T. Yoshida, Rev. Latinoam. Quim., 9, 177 (1978). (Chem. Abstr., 91, 39673a [1979].)
- (123) V. A. Mnatsakanyan and G. B. Oganesyan, Khim. Prir. Soedin., 727 (1978). (Chem. Abstr., 91, 39674b [1979].)