Terpenoids. LV The Structure and Absolute Configuration of Macrocalyxoformin E

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On the basis of chemical evidence and spectroscopic investigations, the structure and absolute configuration of macrocalyxoformin E, which had been isolated from Rabdosia macrocalyx (Dunn.) Hara form (Labiatae) were established as ent-6,20: 7,20α-diepoxo-6α, 15α-dihydroxy-7-oxo-6,7-seco-16-kaurene (5).

KEY WORDS: Robdosia macrocalyx (Dunn.) Hara form/ Diterpenoid/ Macrocalyxoformin E/ 6,7-Secokaurene/ A Ring Conformation/ Absolute configuration/ Hydride Shift/

From the leaves of Rabdosia macrocalyx (Dunn.) Hara form (Labiatae), five new diterpenoids, macrocalyxoformin A-E (1-5), have been isolated and their structures have been previously reported2). Among these, the structure of macrocalyxoformin E (5) has been speculated without sufficient chemical or spectral evidence, leaving ambiguity in the configuration of several functional groups. We reexamined the structure of macrocalyxoformin E and clarified the whole structure including the A ring conformation and the absolute configuration. Here, we describe the structure elucidation of macrocalyxoformin E.

In the previous report3), it was speculated that macrocalyxoformin E (5) has an ent-6,7-seco-16-kaurene skeleton and, has an internal acetal group at C-20 form-

\[ R = 0 \]
\[ R = α-OH, β-H \]
ing a lactone ring, a hemiacetal group at C-6, and an allyl alcohol on the D ring. However, there is no decisive evidence for the stereochemistry of the functional groups at C-6, C-15, and C-20 and for the absolute configuration of 5. At first, we examined several chemical reactions to get some chemical evidence for the structures of macrocalyxoformin E. The oxidation of 5 with manganese oxide afforded α, β-unsaturated ketone 6. This reaction proved the existence of an allylic alcohol system on the D ring. If the hydroxyl group of the allylic alcohol system is located on the endo (β)-side of the C/D ring system, the allylic alcohol system will be converted to the methyl ketone 7 by the Garryfoline-Cuauchichicine rearrangement under acidic conditions. The reaction of 5 with p-toluenesulfonic acid in methanol caused the above rearrangement, involving the stereoselective hydride shift from the exo-H at C-15 to the exo-H at C-16 through the nonclassical carbonium ion 9, to give methyl ketone 8 with concomitant acetalization at C-6. Because of this rearrangement, the stereochemistry of the hydroxy group at C-15 was proved to be endo (β-configuration). Methyl acetalization under acidic conditions should proceed via the oxonium ion 10 which is attacked by the nucleophile (MeOH) from the less hindered (β)-side to give the acetal 8. Since the coupling constants of C-6-H in compounds 5 and 8 showed the same value (ca. 6 Hz) in 1H-NMR spectra, the above speculation leads to a β-configuration for C-6-OH of 5. Attempted catalytic hydrogenations of 5 afforded the ketone 7 unexpectedly through the same type of rearrangement.

Next, we conducted the nuclear Overhauser enhancement (NOE) experiment on 5 to determine the configuration of C-20. Two kinds of ring A conformation (A and B) have been shown to exist in the Rabdosia diterpenoids of the spirolactone-type. Each conformer is distinguishable from the other by the NOE between 18-H3 and 9-H or between 19-H3 and 20-H. The chemical shifts of the protons required for the NOE inspection is easily determined from the 1H-1H correlation spectroscopy (COSY) NMR spectrum (Fig. 1). The results of NOE experiment on 5 are
shown in Table 1. The NOEs (12% and 2%) between 18-H$_3$ and 9-H (run 1 and 4) indicate that the A ring of 5 exists in the A conformation. The NOEs between C-5-H and C-9-H, and between C-20-H and C-14-H are expected in the structure C, while the NOEs between C-5-H, C-9-H and C-20-H are also expected in the

![Diagram A](image)

![Diagram B](image)

![Diagram C](image)

![Diagram D](image)

Fig. 1. $^1$H--$^1$H COSY Spectrum of Macrocalyxoformin E(5) in CDCl$_3$–C$_6$D$_5$N(2: 1).
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Table 1. NOE data for macrocalyxoformin E (5)*

<table>
<thead>
<tr>
<th>run</th>
<th>Monitor proton (δ, ppm)</th>
<th>Observed protonb (Intensity increase)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>18-H3</td>
<td>5-H, 9-H</td>
</tr>
<tr>
<td></td>
<td>(0.97)</td>
<td>(8.6%) (11.7%)</td>
</tr>
<tr>
<td>2</td>
<td>19-H3</td>
<td>5-H, 6-H, 18-H3</td>
</tr>
<tr>
<td></td>
<td>(1.03)</td>
<td>(7.8%) (13.7%)</td>
</tr>
<tr>
<td>3</td>
<td>5-H</td>
<td>9-H, 18-Hs</td>
</tr>
<tr>
<td></td>
<td>(1.86)</td>
<td>(12.5%) (1.2%) (1.4%)</td>
</tr>
<tr>
<td>4</td>
<td>9-H</td>
<td>5-H, 18-Hs</td>
</tr>
<tr>
<td></td>
<td>(2.75)</td>
<td>(14.3%) (1.8%)</td>
</tr>
<tr>
<td>5</td>
<td>20-H</td>
<td>14α-H</td>
</tr>
<tr>
<td></td>
<td>(5.50)</td>
<td>(6.7%)</td>
</tr>
<tr>
<td>6</td>
<td>14α-H</td>
<td>20-H, 13-H</td>
</tr>
<tr>
<td></td>
<td>(1.71)</td>
<td>(9.8%)</td>
</tr>
</tbody>
</table>

a, Spectra at 400 MHz NMR; solution in CDCl₃-D₅N (2:1).
b, The existence of the observed protons without any % of intensity increase was indicated by the subtractive spectra from the NOE experiment.

structure D. The NOE experiment in Table 1 (run 3–6) coincides with the former structure C. Thus, the relative configuration of all functional groups in the structure 5 was established.

Finally, in order to determine the absolute configuration of macrocalyxoformin E, the circular dichroism (CD) spectrum of its rearranged product 9 was measured. On the basis of a negative Cotton effect at 306 nm on the CD curve, the absolute configuration of macrocalyxoformin E was established as that shown as 5. Consequently, the structure of macrocalyxoformin E should be represented as ent-6, 20:7,20α-diepoxo-6α,15α-dihydroxy-7-oxo-6,7-seco-16-kaurene (5).

EXPERIMENTAL SECTION

The melting points were determined on a Yanagimoto melting points apparatus and are uncorrected. Infrared (IR) spectra were recorded with a JASCO IR-810 spectrophotometer, and ultraviolet (UV) spectra were taken with a JASCO UV-DEC-610C spectrophotometer. Optical rotations were measured with a JASCO DIP-181 digital polarimeter, and CD spectra were measured with a JASCO J-600 spectropolarimeter. Mass spectra (MS) and the fast atom bombardment mass spectra (FAB-MS) were determined on a JEOL JMS-DX300 mass spectrometer. ¹H-NMR spectra and ¹³C-NMR spectra were measured with a JEOL GX-400, Varian VXR-200, and JEOL FX-100, and recorded in δ values with tetramethylsilane as an internal reference. Kieselgel 60 (0.05–0.2 mm, Merk) was used for column chromatography and precoated silica gel plates F₂₅₄ (0.25 and 0.5 mm in thickness) were used for thin layer chromatography (TLC).

Isolation—Dried leaves (5Kg) of R. macrocalyx (Dunn.) Hara form collected in
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The plant material was further extracted 2 times in the same manner. The combined ethanol extract was concentrated under reduced pressure to 20 l. The extract was treated with activated charcoal (350 g), and evaporated under reduced pressure to give a sirupy residue which was dissolved in boiling ethylacetate (8 l). The ethylacetate solution was shaken with 5% aqueous Na₂CO₃ (2 l) to remove any acidic substances. After being washed with water and dried with anhydrous Na₂SO₄, the ethylacetate extract was concentrated under reduced pressure to sirupy residue (200 g), which was chromatographed on a silica gel (1.2 Kg) column with chloroform-acetone. The eluates (1.5 g) from chloroform-acetone (9:1) were rechromatographed on silica gel (100 g) with dichloromethane-acetone as eluants. The eluates (500 mg) from dichloromethane-acetone (9:1) was recrystallized from methanol to give macroclyxoformin E (5) (100 mg).

Macrocalyxoformin E (5): colorless needle, mp 242-244°C, [α]D -164.5° (c=0.23, MeOH). IR νmax cm⁻¹: 3490, 3400, 1740, 1665, 1055, 1000. ¹H-NMR (GX-400, CDCl₃: C₅D₅N=1:1) δ: 0.95, 1.04 (each 3H, s, 18-H₃ and 19-H₃), 2.76 (1H, br.s, 13-H), 2.81 (1H, d, J=6.8, 9-H), 5.08 (1H, d, J=2.4, 17-Hₐ), 5.35 (1H, br.s, 15-H), 5.45 (1H, br.s, 17-Hₜ), 5.57 (1H, s, 20-H), 5.63 (1H, d, J=5.9, 6-H).

¹³C-NMR (FX-100, CD₃N) δ: 19.3 (t), 20.9 (t), 26.8 (t), 27.3 (q, C-19), 30.1 (s), 30.9 (q, C-18), 33.0 (t), 33.8 (t), 34.0 (t), 35.7 (d), 39.9 (d), 48.2 (s), 51.9 (s), 60.7 (d), 75.6 (d, C-15), 101.7 (d, C-20), 105.9 (t, C-17), 107.2 (d, C-6), 156.0 (s, C-16), 173.6 (s, C-7). FAB-MS m/z: 371 (M⁺+Na). Anal. calcd for C₂₀H₂₈O₅: C, 68.96; H, 8.04. Found: C, 68.72; H, 8.18.

Oxidation of macrocalyxoformin E (5)——To a solution of macrocalyxoformin E (5) (10 mg) in acetone (3 ml) was added active MnO₂ (50 mg). The mixture was stirred for 4h at room temperature, then filtered and washed with acetone. After evaporation of the filtrate under reduced pressure, the residue was chromatographed on a preparative TLC plate with CHCl₃-Me₂CO (8:2) to afford 6 (5 mg). Recrystallization with methanol gave 6 as colorless prisms, mp 215-217°C. IR νmax cm⁻¹: 3570, 1765, 1720, 1645, 1300. UV λmax μm: 235 (ε = 7,200).

¹H-NMR (VXR-200, CD₃N) δ: 0.85, 1.08 (each 3H, s, 18-H₃ and 19-H₃), 2.55 (1H, d, J=6, 5-H), 3.06 (1H, br.s, 13-H), 5.30 (1H, s, 17-Hₜ), 5.62 (1H, s, 20-H), 5.68 (1H, d, J=6, 6-H), 6.05 (1H, s, 17-Hₜ). High MS Calcd for C₂₀H₂₈O₅(M⁺): 346.1779. Found 346.1763.

Acidic Treatment of 5——To a solution of macrocalyxoformin E (5) (10 mg) in methanol (3 ml) was added p-toluenesulfonic acid (8 mg). The mixture was refluxed for 15h under nitrogen atmosphere and the mixture was poured into ice-water, and extracted with CH₂Cl₂. After the usual treatment, the extract was concentrated in vacuo to give a residue which was chromatographed on a preparative TLC plate with CHCl₃-Me₂CO (9:1) to give 8 (8 mg). The product was recrystallized with MeOH, mp 196-198°C. CD (c=0.2, MeOH) [θ]²⁵ (nm): -1, 450 (306) (negative maximum). IR νmax cm⁻¹: 1770, 1726, 1250, 1100. ¹H-NMR (FX-100, CDCl₃) δ: 0.94 (6H, s, 18-H₃ and 19-H₃), 1.14 (3H, d, J=7, 17-Hₜ), 3.43 (3H, s, -OMe), 4.90 (1H, d, J=6, 6-H), 5.40 (1H, s, 20-H). High MS Calcd for C₂₁H₂₉O₅ (97)
Rearrangement Reaction of 5——To a solution of macrocalyxoformin E (5) (10 mg) in methanol (3 ml) was added a catalytic amount of 10% Pd-carbon and the mixture was stirred under the atmospheric pressure of hydrogen at room temperature overnight. The catalyst was filtered off, and the filtrate was evaporated in vacuo to give a crystalline product 7 (9 mg), which was recrystallized from MeOH to give colorless prisms, mp 238-240°C, IR ν max cm⁻¹: 3575, 1770, 1725, 1245, 1127, 982. 1H-NMR (VXR-200, C₅D₅N) δ: 0.82, 1.10 (each 3H, s, 18-H₃ and 19-H₃), 1.08 (3H, d, J=7, 17-H₃), 5.67 (1H, s, 20-H), 5.72 (1H, d, J=6, 6-H). High MS Calcd for C₂₀H₂₃O₅ (M⁺): 348.1925. Found 348.1919.

REFERENCES AND NOTES

(6) It is well known that the absolute configuration of 15-oxokaurane derivatives bearing endo methyl at C-16 is determinable by the sign of the Cotton effect in the CD or ORD spectra; J. MacMillan and E.R.H. Walker, J.C.S. Perkin I, 1972, 986.